

# Determining potential for on-farm fecal collection for DNA extraction

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## Introduction

- Grazing cattle diet composition determination methods include: field observations, ruminal or esophageal contents, fecal microhistological analysis, and “Markers” (saturated hydrocarbons, alkanes) but can be labor intensive, costly, and under/over predicts some plants in the diet (summarized by Ho et al., 2010)
- Wildlife researchers have used fecal samples and DNA to determine mammalian intake of feeds, but quantification of exact diet composition was not determined (Jarman et al., 2002; Deagle et al., 2005; Deagle and Tollit, 2007)
- Ruminant plant fecal identification has been completed with moderate success, but diet quantification was not attempted (Pegard et al., 2009; Joo et al., 2014; Mohammadzadeh et al., 2014)
- (DeMay et al., 2013), but Fecal DNA quantity can be reduced by temperature and UV radiation in rabbit fecal DNA analysis has not been completed

## Objective

The intention of this project is to determine DNA concentration differences based on fecal physical composition, time of collection, and animal classification.

## Experimental Procedures

- Holsteins (n=4), heifer calves (n=3), and dry, pregnant beef cows (n=4) were fed a diet at 1.68% of BW for 21-d
  - 20% alfalfa, 50% fescue, 20% corn, and 10% soybean meal (DM basis)
- On d 22-24 at 7 AM and 3 PM individual fecal samples were collected and on d 25 at 7 AM
  - FRESH: sample collected directly from cow or from ground when observing defecation
  - DRY: dry sample collected from pen. Samples were ~90% dry in the fecal pile
- Samples frozen immediately at -20°C
- DNA was extracted following kit protocols (Quiagen Quick Spin Fecal DNA kit, illustrated in Figure 1)
- Purity and quantity were assessed through a spectrophotometer.
- Samples with a low 280/260 ratio underwent a DNA contamination clean-up (Zymo DNA PCR Clean-up & Concentration Kit)

## Statistics

- The study is a completely randomized block design with repeated measures
- Quality (280/260 ratio) and quantity (ug/mL) were analyzed using appropriate t-tests and generating 90 and 95% confidence intervals for:
  - Freshness – FRESH vs DRY
  - Time of day for collection – AM vs PM
  - Animal classification – Cow, Holstein, or Heifer
  - Interactions

References:  
 Deagle, B. E., D. J. Tollit, S. N. Jarman, M. A. Hindell, A. W. Triles, and N. J. Gates. 2006. Molecular scatology as a tool to study diet: analysis of prey DNA in scats from captive Stellar sea lions. *Mol. Ecol.* 14:1831-1842.  
 Deagle, B. E. and D. J. Tollit. 2007. Quantitative analysis of prey DNA in pringled faeces: potential to estimate diet composition? *Cons. Gen.* 8:743-747.  
 DeMay, S. M., P. A. Becker, C. A. Edson, J. L. Rachlow, T. R. Johnson, and L. P. Waits. 2013. Evaluating DNA degradation rates in faecal pellets of endangered pygmy rabbit. *Mol. Ecol. Resour.* 13:654-662.  
 Ho, K. W., G. L. Krebs, P. McCafferty, S. P. van Wyngaarden, and J. Addison. 2010. Using faecal DNA to determine consumption by kangaroos of plants considered palatable to sheep. *Animal.* 4:282-288.  
 Jarman, S. N., N. J. Gates, M. Terrey, P. C. Gill, and N. G. Elber. 2002. A DNA-based method for identification of kill species and its application to analyzing the diet of marine vertebrate predators. *Mol. Ecol.* 11:2879-2890.  
 Joo, S., D. Han, E. J. Lee, and S. Park. 2014. Use of length heterogeneity polymerase chain reaction (LH-PCR) as non-invasive approach for dietary analysis of Svalbard reindeer, Rangifer tarandus platyrhynchus. *PLoS One.* 11:e01562.  
 Mohammadzadeh, H., D. R. Yanez-Ruiz, G. Martinez-Fernandez, and L. Abadia. 2014. Molecular comparative assessment of the microbial ecosystem in rumen and faeces of goats fed alfalfa hay alone or combined with oats. *Anaerobe.* 29:52-58.  
 Pegard, A., C. Miquel, A. Valentini, E. Coissac, F. Bouvier, D. Francois, P. Taberner, E. Engel, and F. Pompanon. 2009. Universal DNA-based methods for assessing the diet of grazing livestock and wildlife from feces. *J. Agric. Food Chem.* 57:5700-5706.

## Experimental Results and Methods

Figure 1: Method for rapidly obtaining inhibitor-free DNA isolate from bovine fecal samples

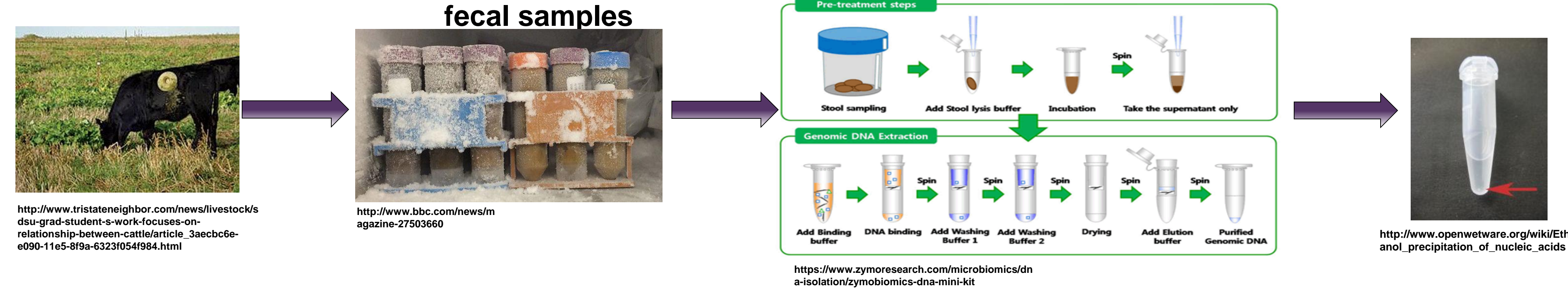


Table 1: Quality (280/260) comparisons of fecal type, time of day for fecal collection, and animal classification.

Treatment	Mean	SE <sup>1</sup>	Lower <sup>2</sup>	Upper <sup>3</sup>	P - value <sup>4</sup>
<b>Comparison of FRESH fecal samples to DRY pen-grabbed fecal samples</b>					
DRY	1.57	0.23	1.51	1.63	NS
FRESH	1.64	0.17	1.60	1.69	NS
<b>Comparison of time of day for fecal collections</b>					
AM	1.53	0.22	1.52	1.63	NS
PM	1.66	0.17	1.60	1.71	NS
<b>Comparison of fecal samples collected based on animal classification</b>					
Cow	1.66	0.20	1.60	1.73	NS
Heifer	1.59	0.18	1.52	1.65	NS
Holstein	1.57	0.22	0.149	1.64	NS

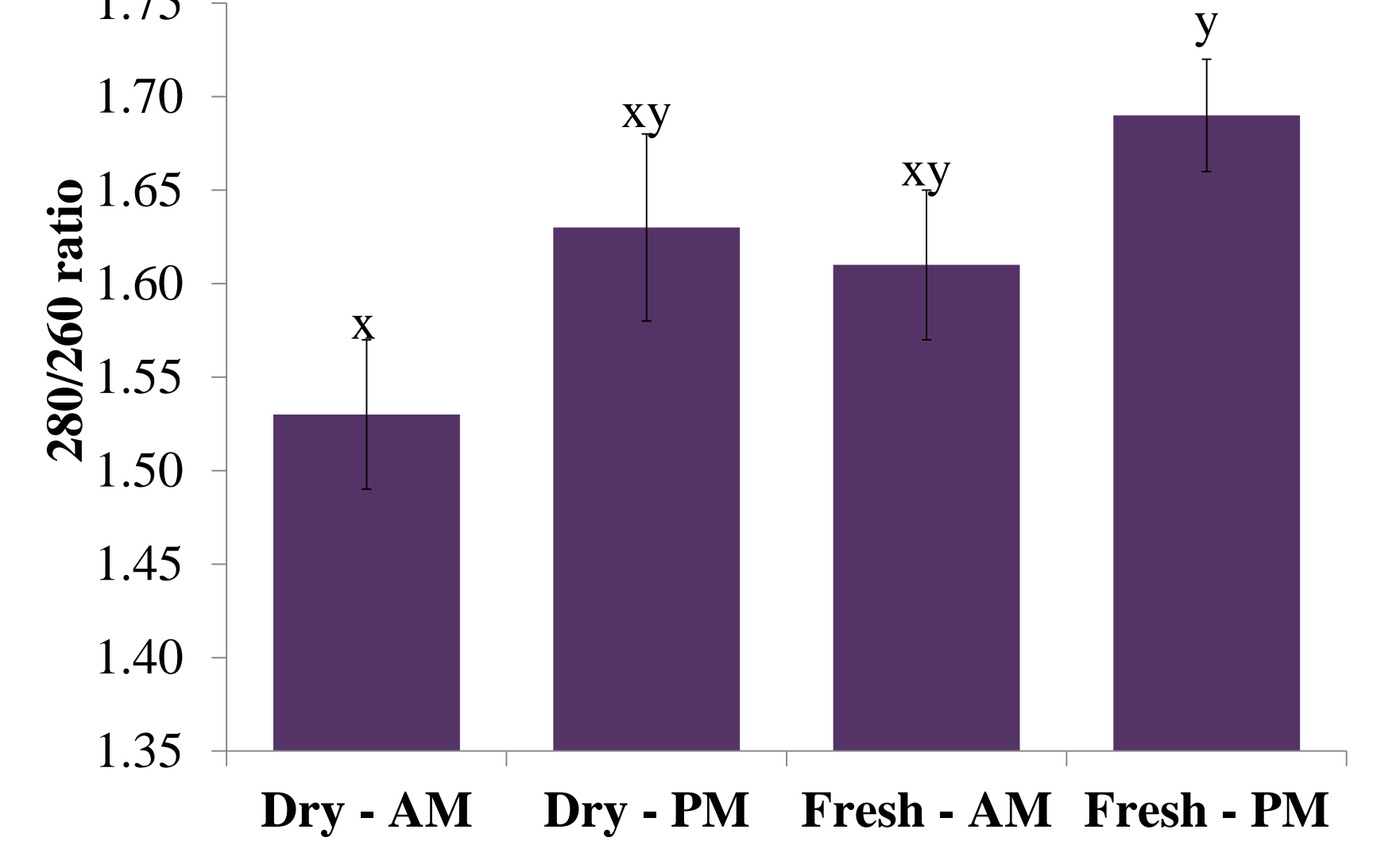
<sup>1</sup> SE: standard error  
<sup>2</sup> Lower limit of confidence interval  
<sup>3</sup> Upper limit of confidence interval  
<sup>4</sup> P-value of NS means non-significant with  $P > 0.10$   
 \* If confidence intervals are overlapping then treatments are the same with  $P > 0.10$

Table 2: Quantity (µg/mL) comparisons of fecal type, time of day for fecal collection, and animal classification.

Treatment	Mean	SE <sup>1</sup>	Lower <sup>2</sup>	Upper <sup>3</sup>	P - value <sup>4</sup>
<b>Comparison of FRESH fecal samples to DRY pen-grabbed fecal samples</b>					
DRY	36.21	18.81	30.96	41.46	NS
FRESH	45.51	22.42	39.31	51.72	NS
<b>Comparison of time of day for fecal collections</b>					
AM	36.59	19.28	31.73	41.45	NS
PM	47.39	22.35	40.41	54.37	NS
<b>Comparison of fecal samples collected based on animal classification</b>					
Cow	40.10	21.92	32.87	47.33	NS
Heifer	47.47	22.53	39.03	55.90	NS
Holstein	36.43	18.20	30.45	42.50	NS

<sup>1</sup> SE: standard error  
<sup>2</sup> Lower limit of confidence interval  
<sup>3</sup> Upper limit of confidence interval  
<sup>4</sup> P-value of NS means non-significant with  $P > 0.10$   
 \* If confidence intervals are overlapping then treatments are the same with  $P > 0.10$

Figure 3: Quality of DNA based on fecal freshness and time of day for collection



\* Different letters are different at  $0.05 < P < 0.10$   
 \* Classifications are sample freshness (DRY or FRESH) then time of day for collection (AM or PM)

Figure 2: Photos illustrating examples of fecal physical characteristics

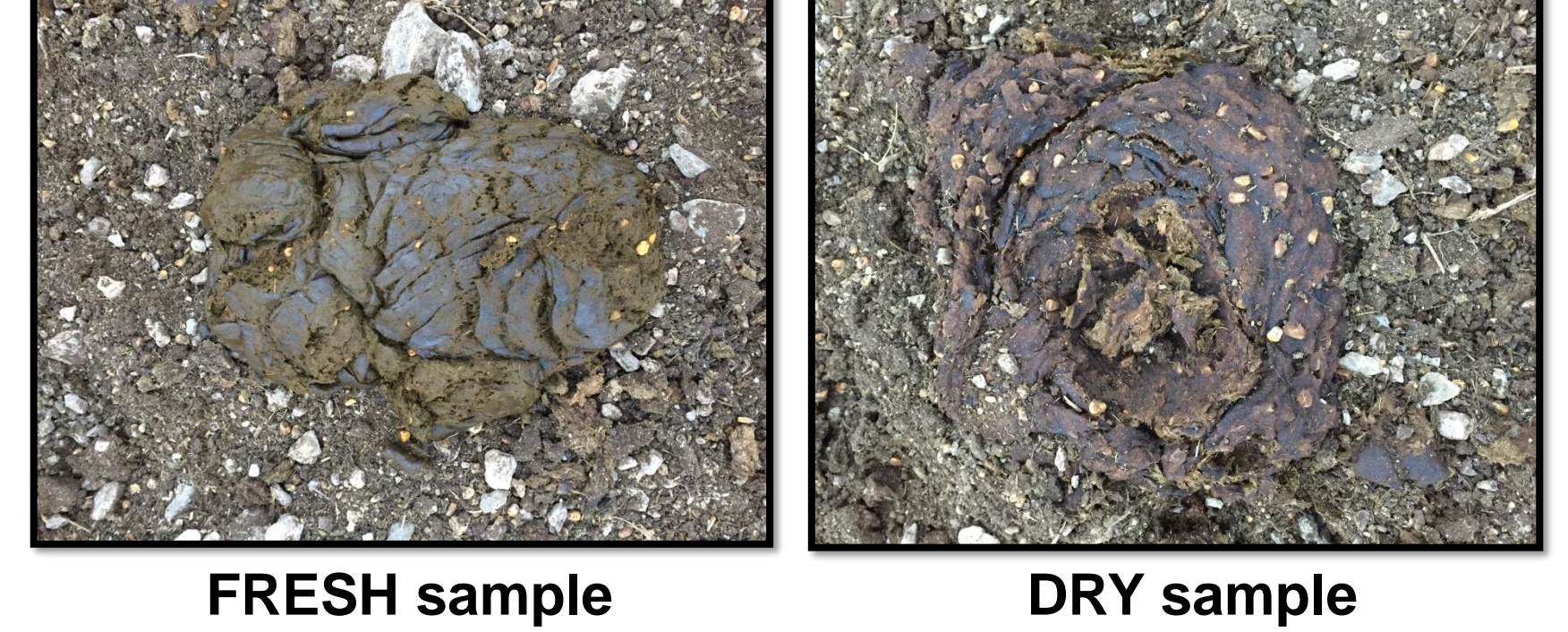
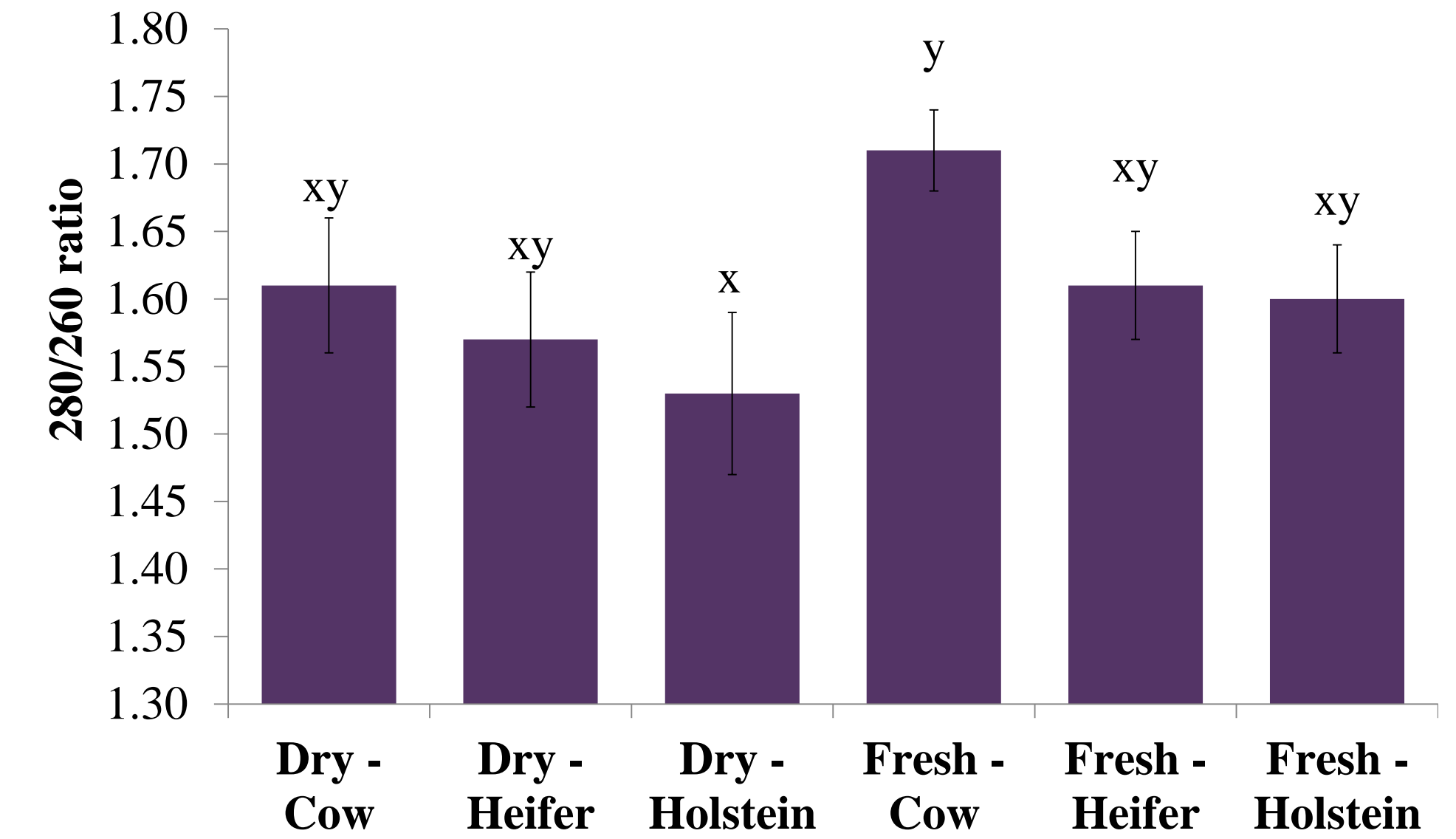
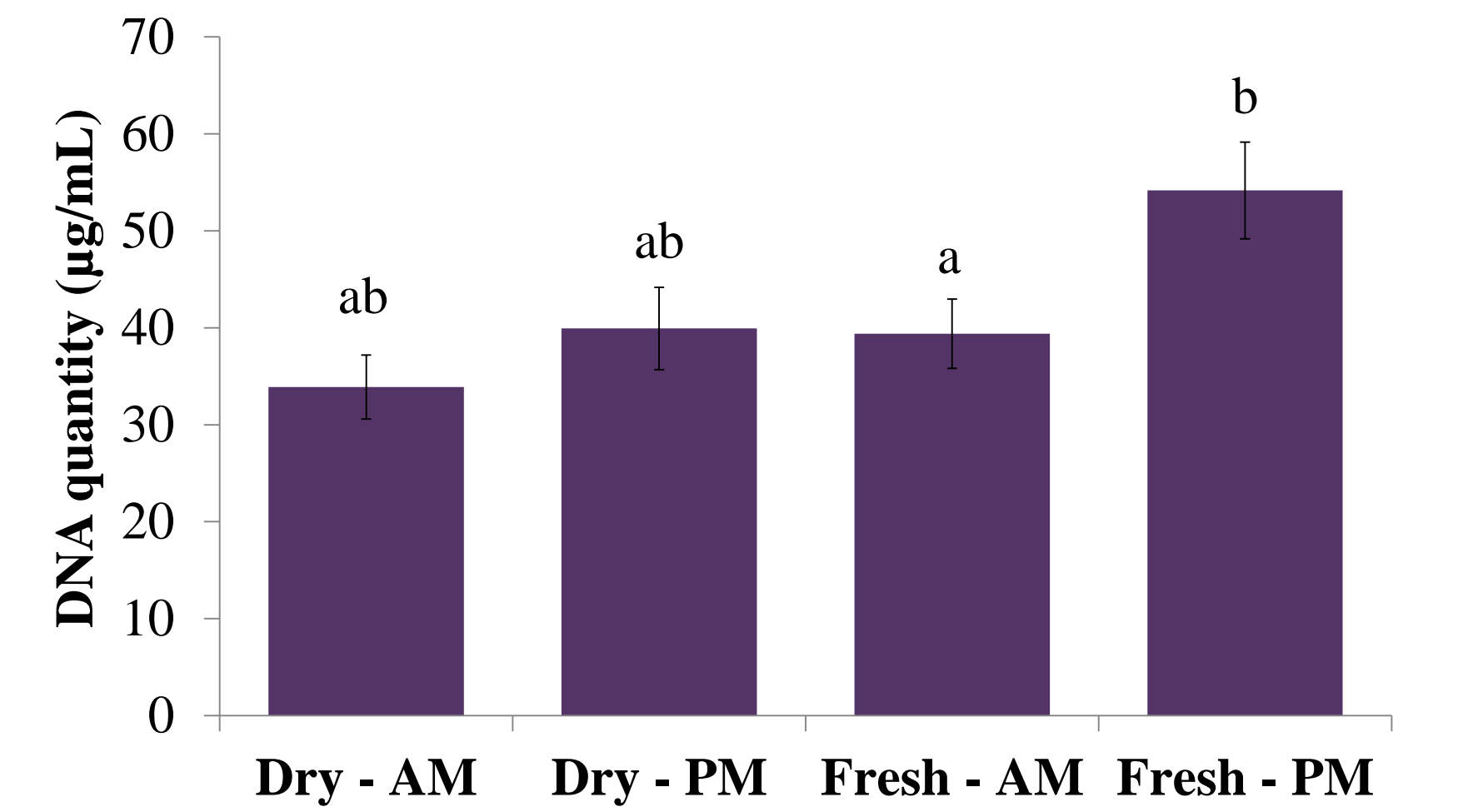


Figure 4: Quality of DNA based on fecal freshness and animal classification



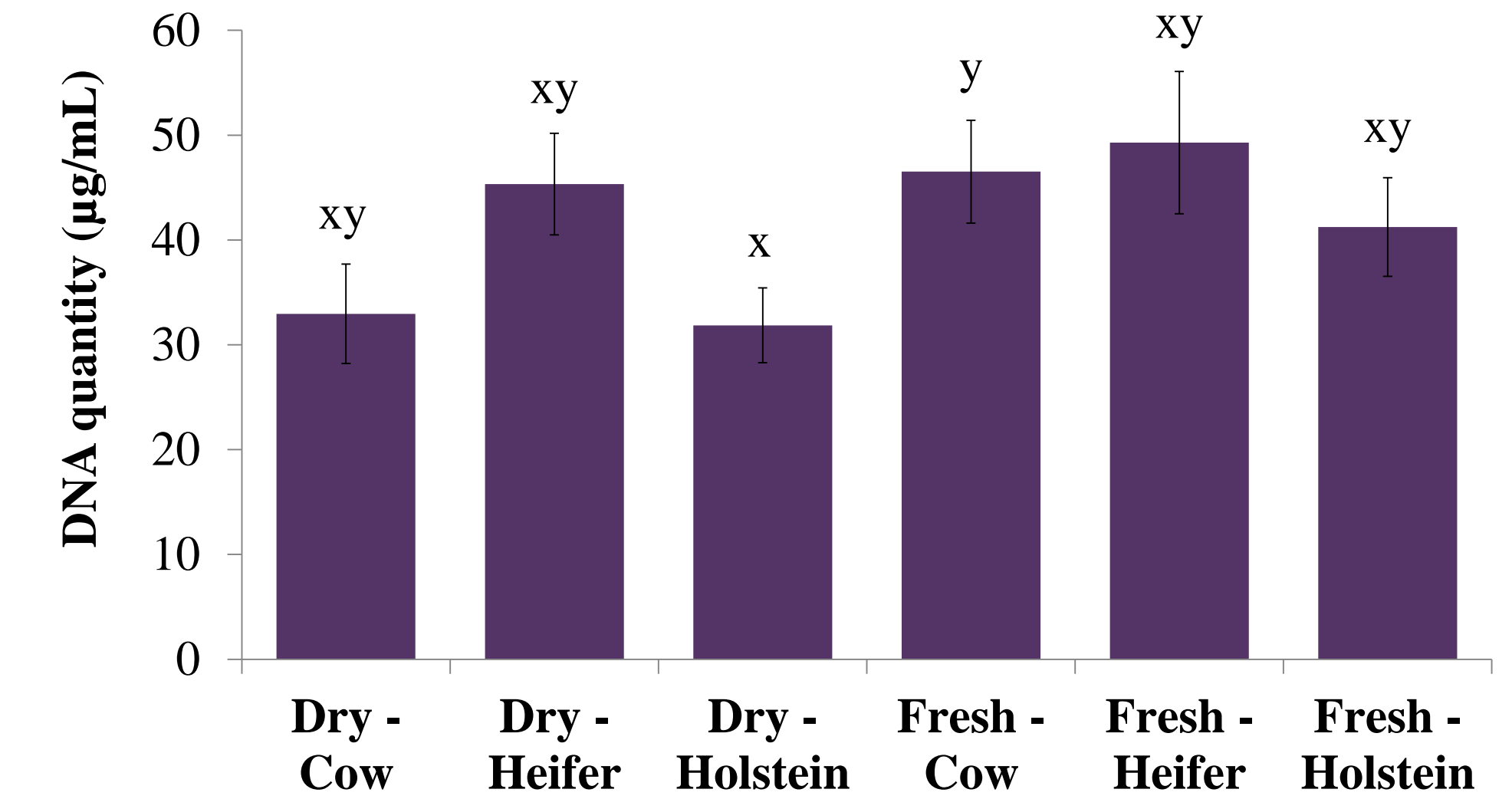
\* Different letters are different at  $0.05 < P < 0.10$   
 \* Classifications are sample freshness (DRY or FRESH) then animal classification (Cow, Heifer, or Holstein)

Figure 5: Quantity of DNA based on fecal freshness and time of day for collection



\* Different letters are different at  $P < 0.05$   
 \* Classifications are sample freshness (DRY or FRESH) then time of day for collection (AM or PM)

Figure 6: Quantity of DNA based on fecal freshness and animal classification



\* Different letters are different at  $0.05 < P < 0.10$   
 \* Classifications are sample freshness (DRY or FRESH) then animal classification (Cow, Heifer, or Holstein)

## Conclusions and Future Work

- Of the 105 samples analyzed, 89 had to complete a secondary clean-up to increase the 280/260 spectral reading to be  $>1.4$ .
- There was no difference in the quantity of DNA extracted from samples that were collected FRESH or DRY; in the AM or PM; nor differences based on cattle classification.
- There is evidence that a fresh sample collected in the afternoon would net the highest quality and yields of DNA to be used for downstream use in qRT-PCR.
- Possible explanation is that afternoon samples occurred 7 hr after daily feeding which might correspond in this diet to the time of maximal ruminal digestion and undigested turnover
- Overall, as with other species the fresher the sample collected the higher quality and quantity of DNA. However, for on-farm projects samples that have some moisture will still generate potentially usable DNA.