STUDIES ON THE SPECIES OF VINEGAR GNATS (DIPTERA: D OSOPHILIDAE) IN KANSAS

by

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B. A., Iowa Wesleyan College, 1950

A THESIS

submitted in partial fulfillment of the

requirements for the degree

MASTER OF SCIENCE

Department of Entomology

KANSAS STATE COLLFGE OF AGRICULTURE AND APPLIED SCIENCE

# TABLE OF CONTENTS

Documents LD 2668 T4 1952 Y6 C.2

07-22-52

•	•	•	•	•	•	1
			•	•		2
E OCCURI	ING I	N KAN	SAS			7
f Drosop	hilid	ae in	Kans	R.S		7
During t	his S	tudy				8
			ith F	our S	tates	
tate	•	•	•	۰		9
				•	•	18
Kansas				•		21
FION						23
ledia					•	23
Fruits			•			23
Temper	eture	Most	Fevo	able		
•	•	•			•	40
						43
						47
ol Test			•			47
oplicat	ions	with 3	Three			
•		•	•	•	•	47
	•	•	•	•	•	51
•		•	•			54
•			•			57
						58
	f Drosop During t as Droso tate Kansas FION Media Fruits i Temper	f Drosophilid During this S as Drosophili tate . Kansas . FION . Media . Fruits . i Temperature	f Drosophilidae in During this Study as Drosophilidae w tate Kansas . FION Media Fruits i Temperature Most  	During this Study . as Drosophilidae with Formation of the second secon	f Drosophilidae in Kansas During this Study as Drosophilidae with Four State Kansas Konsas HOM Kadia Fruits d Temperature Most Favorable  	f Drosophilidae in Kansas . During this Study as Drosophilidae with Four States tate Kansas Kansas HOM Hedia Arequire the favorable .  

## INTRODUCTION

During late summer through the fall season, there are many species of vineger gnats which belong to the Dipterous family Drosophilidae in and around the homes. The flies are not of economic importance as far as direct damage is concerned, but rather because of the enormous numbers which at times become an annoyance. They are especially troublesome around homes in late summer during the canning season. They also enter the kitchen and sometimes accidentally fall into foods. They deposit eggs on the lid of a fruit jar that may not have been sealed tightly. These flies seek their food where it is most available to them and require only 11 days to produce a generation under favorable conditions.

The first problem in studying these flies was to collect species of Drosophilidae under the various ecological conditions existing in Kansas; the second, to determine how many species were actually known to exist in this state. Physiology and behavior patterns must be considered in selecting the best methods for collecting these flies.

Some of the important aspects of trapping flies are considered here, such as the type of baits used, the time of flies collected, and the temperature and the humidity requirement for maximum number of flies present in the baits.

Mass rearings on infested fruits especially those showing possibilities of Drosophilid infestation are a significant phase

## of the study.

Most of the work was done in the Manhattan area during the past 16 weeks of school in the fall of 1950 and 1951.

Every home is equipped with either a garbage can or a garbage pail in the house and outdoors next to the kitchen. A few garbage can lids may be somewhat damaged and may not fit tightly. Under these conditions, the flies readily enter the garbage can to feed and deposit eggs for the coming generation. This is one of the reasons for the increase of species of vinegar gnats around the homes.

As a means of control, experiments were conducted under field conditions to test such means to prevent the flies from increasing within the garbage can.

# REVIEW OF LITERATURE

Vinegar gnats are objectionable, not so much because of the damage they really do, but rather because of the enormous number which at times fairly take possession of an establishment.

Pavan, Dobzhansky and Burla (1950) reported a diurnal behavior pattern of <u>Drosophila pseudoobscura</u> Prolowa with a close correlation to weather conditions, such as clear, warm, and dry days. The flies are active in the morning and before sunset and mostly quiescent during the middle of the day. No activities of flies during nights have been observed. In a humid rain forest with equable temperatures, or on rainy days, flies are active. Methods of collecting species of <u>prosophila</u> have been studied by Patterson (1943). He showed that the different species could be found in the following habitats: (1) parasitic on Gercopids; (2) fungi; (3) coralise of large flowers; (4) decaying fruits; (5) general scavengers--rotten potatoes, excrement, and stale formalized meat. Bonds (1939) reported two species, <u>D. funebris</u> and <u>D. busckii</u>, to be associated with decaying potatoes in Maine. Ditman et al. (1937) studied <u>D. repleta</u> on fermenting tomatoes with relation to canning of tomatoes. They found that the female of <u>D. repleta</u> laid 430 eggs during a week, averaging 14.84 eggs per day at 77° to 88° F.

Dobzhansky and Epling (1944) established a standard technique of collecting by means of baited traps. The success of a trap depends on the nature and the quantity of the bait, on the condition of the flies, on the weather, and on the presence or absence of other sources of attraction in the environment. Attractive radius of a banana trap on <u>D</u>. <u>pseudobscura</u> was 40 or 60 meters at two stations in the pine and pine-oak forests of the Southern California mountains. The best catches were made during a period of 30 to 50 per cent humidity and  $47^{\circ}$  to  $90^{\circ}$  P. range in temperatures.

Sister Geisler (1942) studied the effects of humidity on <u>D. melanogaster</u> pupae and found that lower humidity increased the mortality to a greater extent than did 100 per cent humidity.

Kalmus (1945) carried out a 14 months study on the popula-

tion of equal numbers of wild type and the abony type of  $\underline{P}$ . <u>melanogaster</u>. High temperature was found to favor the wild type and low temperature the abony mutant type. Also, heterozyotes are superior to both homozygotes.

In 1943, Northrop found that an insufficient quantity of food retarded the growth of <u>Drosophila</u> flies. Sterile fruits, or fruits containing no yeast, caused a vitamin deficiency in the flies.

Spencer (1937) proposed a new technique for studying <u>Droso-</u> <u>phila</u> eggs and larvae. He constructed a tiny metal ring cage where a single femals fly can oviposit her eggs.

Parker (1835), at the University of Texas laboratory, reported the use of commercial mold preventive in <u>Drosophils</u> culture media. Moldex-A (a commercial trade name) is composed of the sodium salt of para-hydroxybenzoic acid. A small amount of this substance added to the food will stop the mold growth. Also, mites in the culture could be eliminated by heat sterilization.

Spieth (1951) reported that <u>D. lacicola</u> Patterson in Minnesota is known to breed in a narrow fringe about a pond where the rotting phloem of Aspen, <u>Populus tremuloides</u> Michx., is to be found. The larvae live on the yeasts which are formed in the rich aspen bark.

In 1937, Galtsoff reported on the principal methods of making culture media for <u>Drosophila</u>. Bridges (1921) formulated the first simple banana culture media consisting of 100 grams of banana, 100 cc of water, and 2 grams of agar-agar. Bridges and Darby (1933) found a cheaper method of making a standard medium for laboratory use. The following is their formula: 75 cc of water, 10 grams of cornmeal, 13.5 cc of molasses, and 1.5 grams of agar.

Lewis (1942) during the war substituted tomato-paste medium for banana. He used 1,000 cc of water, 100 grams of tomatopaste (canned), 100 grams of white corn syrup, 20 grams of granulated agar-agar, and one gram of Moldex. The culture lasted for four weeks without drying out.

Blaufus (1936) discussed rearing method for <u>D</u>. <u>melanogaster</u> by the use of quart milk bottles.

Kalmus (1943) experimented on the mineral requirement of <u>D. melanogaster</u>. He reported that KgHP04 and MgS04 were necessary for their dist. He designed the following media for <u>Drosophila</u> culture: 891 grams of H<sub>2</sub>O, 20 grams of agar, 5 grams of tartaric acid, 80 grams of sucrose, 1.5 grams of K<sub>2</sub>HP04, 2 grams of (NH4)<sub>2</sub>S04, and 0.5 gram of MgS04.7 H<sub>2</sub>O.

Spencer (1937) reported that <u>D</u>. <u>funebris</u> was attracted to human and animal excrement. Also Dove (1937) reported on a case of human intestinal myiasis with <u>D</u>. <u>funebris</u> larvae as the infesting insect. Smart (1943) reported that <u>Drosophila</u> maggots could be swallowed in food and thus be found in cases of intestinal myiasis.

DeCoursey (1925) points out that <u>D</u>. <u>melanogaster</u> becomes a pest in groceries and homes. He suggested a practical control in the form of a trap baited with fermenting banana around homes. In 1935, McDaniel reported on the life cycle and control of species of <u>Drosophila</u>. She suggested three ways of decreasing the population of <u>Drosophila</u> in the community: (a) proper disposal of fruit waste; (b) don't leave fruits lying on the ice-box or near window--keep the fruits covered in a receptacle or put them in ice-box; and (c) fruits canned while in process should be sealed with rubber rings or paraffin. For insecticide control, she recommended a kerosene and pyrethrum mixture consisting of eight ounces of Pyrethrum to one gallon of kerosene mixed and settled for 24 hours. Then, pour off the clear liquid at the top and use for spraying.

According to McLeod (1944) the uso of nicotine sulphate resulted in a higher mortality rate for the female than the male <u>Drosophila</u>.

Sloan (1945) recommended a dust consisting of 10 parts of lead arsenate, 6 parts of sulfur, and 4 parts of cupric carbonic acid for field use.

Metcalf (1948) used the fluorine analogue of DDT, commonly known as DPDT--fluoro DDT or fluogogessarol. The tests were made on adult <u>D. melanogaster</u> Meigen by impregnating 9-cm filter paper with 1 ml of standard acetone solution of DFDT. In 24 hours, the mortality rate was 63 per cent "knockdown" with 0.16 microgram concentration of toxicant applied per square centimeter of filter paper.

SPECIES OF DROSOPHILIDAE OCCURRING IN KANSAS

Previous Records of Drosophilidae in Kansas

Snow (1903) listed <u>Prosophila ampelophila</u> Loew (synonym of <u>Drosophila melanogaster</u> Meigen) as the only known species in Kansas in 1903. Crevecceur (1905) added <u>Phortica vittata</u> Coq. to the list of Drosophilidae in Kansas. The latter species is now placed in the genus <u>Scaptomyza</u>. Tucker (1906) has added four additional species to the list of Drosophilidae. The following were the ones in Tucker's list:

> Drosophila (Scaptomyza) adusta Loew. Drosophila (Chymomyza) amoena Loew. Drosophila funebria Fab. Drosophila (Scaptomyza) graminum Fal.

He listed <u>Scaptomyza</u> as a subgenus of <u>Drosophila</u>. In the recent list of family Drosophilidae by Sturtevant (1921), <u>Scaptomyza</u> has been considered to be a separate genera. The <u>Drosophila amoena</u> Loew. recorded by Tucker has been placed in the genus <u>Chymomyza</u>. All species have been confirmed in the present study and added to the list of Drosophilidae in Kansas. Sturtevant (1921) listed several genera and species in Kansas under "geographical distribution". The following were the genera and species in his Kansas list:

> Aulacigaster leucopeza (Meigen) Chymomyza amoena (Loew.)

<u>Chymomyza prochemia</u> Williston <u>Leucophenga maculosa</u> (Dufour) <u>Leucophenga varia</u> (Walkor) <u>Scaptomyza adusta</u> (Loev.) <u>Scaptomyza graminum</u> (Fallon) <u>Drosophila affinis</u> Sturtevant <u>Drosophila busckii</u> Coquillett <u>Prosophila funebris</u> (Fabricius) Drosophila melanogaster Meigen

# Species Collected During this Study

Species of the following four genera were collected during the course of this investigation: <u>Mycodrosophila</u>, <u>Chymomyza</u>, <u>Scaptomyza</u>, and <u>Brosophila</u>. In all except genus <u>Brosophila</u>, only one or two species were collected. But under the genus <u>Brosophila</u>, there were 16 species. Among these 19 species which have been collected during the investigation, one genus and 13 species have not been recorded previously in the list of Diptera in Kansas. The following were the previously unrecorded species now added to those known to occur in Kansas (see Table 1 for host records and locality):

Mycodrosophila dimidiata Loew.

<u>Drosophila algonguin</u> Sturtevant and Dobzhansky <u>Drosophila athabasea</u> Sturtevant and Dobzhansky <u>Drosophila americana</u> Spencer <u>Drosophila immigrans</u> Sturtevant <u>Drosophila guttifera Walker</u> <u>Drosophila hydei</u> Sturtevant <u>Drosophila melanica</u> Sturtevant <u>Drosophila putrida</u> Sturtevant <u>Drosophila repleta</u> Wollaston <u>Drosophila robusta</u> Sturtevant <u>Drosophila tripunetata</u> Loew. <u>Drosophila simulana</u> Sturtevant

The external morphology of the species <u>D</u>. <u>melanogaster</u> and <u>D</u>. <u>simulans</u> is very similar, except the length of the compound eye vertically is slightly longer in <u>D</u>. <u>simulans</u>. Lots of specimens either trapped or reared and included under <u>D</u>. <u>melanogaster</u> probably included both <u>D</u>. <u>melanogaster</u> and D. simulans.

Members of the <u>repleta</u> group are composed of about 30 known species characterized by grayish spotted mesonotum, with each hair and bristle arising from a dark brown or black spot. There are several species under the subgroup of <u>repleta</u> which are difficult to separate.

#### Comparison of Kansas Drosophilidae with Four States Bordering the State

The geographical distribution of the various species of Drosophilidae may be discussed here in comparison with those known from the four states bordering Kansas.

Missouri. In 1941 (Patterson, 1943), there were 18 species

of <u>Drosophila</u> recorded from Missouri. Among the total number of species, 10 were the same as those known from Kansas. There were 14 species from Kansas not recorded from Missouri. These are as follows: <u>Mycodrosophila dimidiata</u>, <u>Aulacigaster lecucopeza</u>, <u>Leucophenga maculosa</u>, <u>Leucophenga varia</u>, <u>Chymomyza amoena</u>, <u>Chymomyza procnemis</u>, <u>Scaptomyza graminum</u>, <u>Scaptomyza adusta</u>, <u>D. algonquin</u>, <u>D. athabasca</u>, <u>D. americana</u>, <u>D. immigrans</u>, <u>D.</u> <u>guttifera</u>, and <u>D. simulans</u>.

The 10 species similar to those found in Kansas were: D. busckii, D. melanogaster, D. affinis, D. putrida, D. repleta, D. tripunctata, D. funebris, D. hydei, D. robusta, and D. melanica.

Only one species from Missouri unknown to Kansas was <u>D</u>. macrospina.

Oklahoma. Collections recorded in 1941 (Patterson, 1943) included 22 species represented from Oklahoma. Among the total number of flies represented, there were 12 species of Drosophilidae identical to the species from Kansas. The following were the species recorded: <u>Chymomyza amoena</u>, <u>D. busckii</u>, <u>D.</u> <u>melanogaster</u>, <u>D. simulans</u>, <u>D. affinis</u>, <u>D. putrida</u>, <u>D. tripunctata</u>, <u>D. funebris</u>, <u>D. replets</u>, <u>E. hydei</u>, <u>D. robusta</u> and <u>D. melanica</u>.

There were 12 species from Kansas not recorded from Oklahoma. These were as follows: <u>Mycodrosophila</u> <u>dimidiata</u>, <u>Aulacigaster lecucopeza</u>, <u>Leucophenga maculosa</u>, <u>Leucophenga varia</u>, <u>Chymomyza procnemis</u>, <u>Scaptomyza graminum</u>, <u>Scaptomyza adusta</u>, <u>D</u>. <u>algonquin</u>, <u>D</u>. <u>athabasea</u>, <u>D</u>. <u>americana</u>, <u>D</u>. <u>immigrans</u>, and <u>D</u>. guttifera.

Several Oklahoma species unknown to Kansas were as follows: <u>Gitona americana</u>, <u>D. duncani</u>, <u>D. transversa</u>, <u>D. texana</u>, <u>D. mulleri</u>, <u>D. macrospina</u>, <u>D. aldrichi</u>, <u>D. nigromelanica</u>, and <u>D</u>. oarbonaria.

<u>Colorado</u>. For Colorado, collection records (Patterson, 1943) included 14 species of Drosophilidae. There were 9 out of 14 species similar to those from Kansas. The 9 species are: <u>Scaptomyza adusta</u>, <u>D. busckii</u>, <u>D. melanopaster</u>, <u>D. simulans</u>, <u>D.</u> <u>athabasca</u>, <u>D. funebria</u>, <u>D. rapleta</u>, <u>D. hydei</u>, and <u>D. immigrans</u>.

There were 15 species from Kansas not recorded from Colorado. These were as follows: <u>Mycodrosophila dimidiata</u>, <u>Aulacigaster lecucopeza</u>, <u>Leucophenga maculosa</u>, <u>Leucophenga varia</u>, <u>Chymomyza amoena</u>, <u>Chymomyza procnemis</u>, <u>Scaptomyza graminum</u>, <u>D</u>. <u>robusta</u>, <u>D</u>. <u>americana</u>, <u>D</u>. <u>algonquin</u>, <u>D</u>. <u>tripunctata</u>, <u>D</u>. <u>gutti-</u> <u>fera</u>, <u>D</u>. <u>putrida</u>, <u>D</u>. <u>melanica</u> and <u>D</u>. affinis.

Several Colorado species unknown to Kansas were as follows: <u>D. pseudoobscura</u>, <u>D. suboccidentalis</u>, <u>D. subquinaria</u>, <u>D. montana</u>, and <u>D. macroptera</u>.

<u>Nebraska</u>. Williams (1948) collected and listed 22 species of Drosophilidae from Nebraska. From his listed species, 16 species were identical to Kansas species. These were as follows: <u>Chymomyza amoena</u>, <u>Aulacigaster</u> sp. <u>Scaptomyza graminum</u>, <u>Chymomyza</u> <u>proceeding</u>, <u>D. melanogaster</u>, <u>D. affinis</u>, <u>D. algonquin</u>, <u>D. funebris</u>, <u>D. melanica</u>, <u>D. hydei</u>, <u>D. busckii</u>, <u>D. guttifera</u>, <u>D. robusta</u>, <u>D.</u> putrida, and <u>D. simulans</u>.

There were 7 species from Kansas not recorded from Nebraska.

These were as follows: <u>Mycodrosophila dimidiata</u>, <u>Leucophenga</u> <u>maculosa</u>, <u>Leucophenga varia</u>, <u>D. americana</u>, <u>D. athabasea</u>, <u>D.</u> <u>repleta</u>, and <u>D. tripunctata</u>. The majority of the species caught in Nebraska were similar to those occurring in Kansas.

Several Nebraska species unknown to Kansas were as follows: <u>D. macrospina</u>, <u>D. pseudoobscura</u>, <u>D. quinaria</u>, <u>D. transversa</u>, <u>D. pseudomelanica</u>, <u>D. victoria</u>, <u>D. macrospina</u>, and <u>D. cinera</u>.

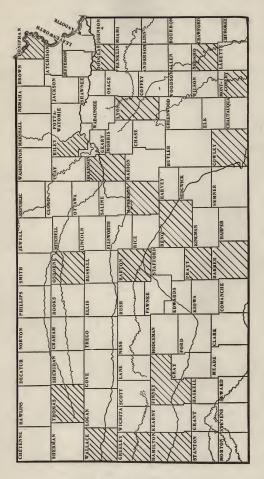
There were four species of Drosophilidae collected in Kansas but not recorded in any of the four states bordering the state of Kansas. These species are: <u>Mycodrosophila dimidiata</u>, Leucophenga maculosa, Leucophenga varia, and <u>Drosophila</u> americana.

The species <u>Scaptomyza adusta</u>, <u>D</u>. <u>trinunctata</u>, <u>D</u>. <u>athabasca</u>, and <u>D</u>. <u>repleta</u> represent the farthest north that these species have been found.

The most commonly known species from Kenses and also from Missouri, Oklahoma, Colorado, and Nebraska are <u>D</u>. <u>funebris</u>, <u>D</u>. <u>melanogaster</u>, <u>D</u>. <u>simulans</u>, <u>D</u>. <u>putrida</u>, <u>D</u>. <u>busckii</u>, and D. hydei.

The Fatterson (1943) and Sturtevant (1921) taxonomic papers have been very useful as references in the systematic classification of Drosophilidae.

At present there are approximately 118 species listed in the classification of the genus <u>Drosophila</u>. Specimens of nearly all of the species listed in the collections have been sent to Dr. Marshall R. Wheeler of Texas University for determination or verification.



Kansas counties from which Drosophilidae have been recorded. Fig. l.

ountles, the date of collec-	tion, and number of substrates from which they were either reared or trapped in Kansas.
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Species	tion, Kansa
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Table	

Genus and species	: Dete : collected	: sollected:	County in : Kansas :	Substrate
Mycodrosophila dimidiata Loew.	Sept. 26/50	50 4	Riley	Fungus
		51 3	Riley	Fungus
Sceptomyza adusta Loew.		50 I	RILey	Cucumber
Chymomyza amoena Loew.		34 1	McPherson	Apple
		51 3	Mc Pherson	Apple
		51 2	Lyons	Pear
Drosophila affinis Sturt.		51 3	Hiley	Banana
		51 2	Barton	Crab apple
	Sept. 28/	51 2	Filey	Crab apple
		51 I	Riley	Banane
		51 1	Lyons	Persimon
		[2] T	Riley	Garbage
		51 1		Pers imnon
		51 2		Persimon
Prosophile algonquin Sturt.		51 2		Banana
and Dobzk.		51 1		Crab apple
Drosophila americana Spencer		51 1		Banana
		1 1		Garbage
		1 1		Banana
	Sept. 24/51	51 I		Persimon
		10 T		Apple
Drosopulla atnabasca Sturt.		T TC		Banana
and Dobzk.		24/51 1		Crab apple
		1 1		Banana
Drosophila busckii Coq.		19/51 1		Banana
		e Te		Garbage
		50 5	Doniphan	Pear
Drosopulla lunebris Fab.		1 20	Riley	Persimon
		TOC	RILey	Grape
	0ct. 6/	6/50 1	Riley	Pear

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Table 1.

Genus and species	: Date : collected	ie cteč	:No. flies:	County in :	Substrate
				0 01 01 01 01 01 01 01 01 01 01 01 01 01	
Prosophila funebris Fab.	Sept.	26/50	1	Rilev	Witheria
		13/51	-	10110	and and a
		19/51	1 14	Dilow	TOTIN TO
		10/01	0 0	Louis Contract	Danana
		10/00	21	Darron	Banana
		TQ/82	0	Filey	
		25/51	03	Riley	
Drosophila guttifera Walker	Sept.	26/50	-	Rilav	Miss of him on the
Drosophila hydel Sturtevant		19/51	-	Rilev	THON THE END OF
		21/51	1	Bilev	Carbana
		29/51	-	Riley	Combooo
	Sent.	26/50	1 65	Dontahon	Decre
		00/00		D OLLA PLANEL	JRAI
		ne /at	. 52	HILOY	Persimmon
	Sept.	26/20	-1	Riley	Fungus
		17/50	1	Riley	Persimon
		18/50	1	Filey	Fersiamon
		26/50	1	Doniphan	Pear
Prosophila melanica Sturt.		19/51	I	Riley	Ranana
		24/51	4	HILEY	(Trab anole
Drosophila melanogaster Meigin		18/50	4	Kiley	Ranana
		18/50	5 C	Riley	Cucumbar
		18/50	4	Bilav	Miri a hreadon
		18/50	4	Rilev	Pometo
		19/50	10	Thomas	Devere
		10/00		2 440/21/00 C	Devitering
		ne'lar	22	Greeley	Banana
		6/50	CV3	Riley	Persiamon
		6/50	C3	Riley	Pokewaed
		18/50	1	Wallace	Prickly near
		17/50	02	Riley	Persimon
	Oct.	17/50	01	Riley	Grape

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Genus and species	: col	Late collected	:No. Tles: :collected:	County in : Ransas :	Substrate
Drosophila melanogaster Meigen	Sept.	26/50	г	Riloy	Apple
	Sept.	26	03	Riley	Persimon
	Oct.	18	in	Finney	Banana
	Sopt.	26	-	Riley	Apple
	May		63	Riley	Garbage
	May	27/51	2	Riley	Garbage
	May	28/51	63	RILey	Garbage
	May	29/51	S	Riley	Garbage
	May	30/51	01	Riley	Garbage
	May	51/51	4	Riley	Garbage
	Sept.	19	01	Riley	Garbage
	Sept.	0	12	Montgomery	Banana
	Sept.	0	9	Cowley	Banana
	Sept.	0	4	McPherson	Banana
	Sept.	0	5	Barton	Eanana
	Oct.	5	9	Doniphan	Crape
	Oct.	5	8	Doniphan	Apple
	Sept.	24	-	Riley	Crab apple
	Sept.	0	63	Lyons	Apple
	Sept.	0	4	Lyons	Grape
	Sept.	o	11	Neosho	Banana
	Sept.	19	5	Riley	Banana
	Sept.	25/51	4	Riley	Crab apple
Drosophila melanogaster and D.	Oct.	19/50	8	Thomas	Banana
	Sept.	26/50	4	Riloy	
	Sept.	26/50	CV3	Riley	Pokeweed
	Oct.	6/50	63	Riley	Apple
	Oct.	6/50	63	Riley	Grape
	Oct.	26/50	-1	Doniphan	Pear
	Oct.	7/51	-	Doniphan	Grape
	Oct.	18/50	4	Wallace	Prickly pear

Table 1. (concl.)

Troosophilamelanogesterand\$\frac{1}{2.1milans}\$\$\frac{1}{36}\frac{5}{5}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{1}{3}\frac{5}{5}\frac{1}{3}\frac{1}{3}\frac{5}{5}\frac{1}{3}	Genus and species	: Date : collected	:No. flies: d :collected:	County in : Kansas :	Substrate
Sept. 19/50         A Miloy           Sept. 19/50         B Miloy           Sept. 19/50         B Miloy           Sept. 19/50         B Miloy           Sept. 26/50         B Miloy           Sept. 26/50         B Miloy           Sept. 26/50         B Miloy           Sept. 26/51         B Miloy	nelanogaster	18,	50 22	Riley	Cucumber
Supt. 19/50         5 Miley           Cutride Sturt.         Sept. 19/50         3 Miley           Sept. 10/52         1 Douglas           Sept. 10/52         1 Douglas           Sept. 10/52         1 Douglas           Sept. 26/50         1 Douglas           Sept. 10/52         1 Douglas           Sept. 10/52         1 Douglas           Sept. 26/50         1 Miley           Sept. 26/50         1 Miley           Sept. 26/51         1 Miley           Sept. 26	simulans	18	50	Riley	Mushroom
Dutride         Surt.         Sept. 19/50 out.         7         811ey surt.           Dutride         Surt.         500.         1         811ey surt.           Cort.         5/20         1         111ey surt.         111ey surt.           Cort.         5/20         1         111ey surt.         111ey surt.           Sept. 26/50         2         111ey surt.         111ey surt.         111ey surt.           group         Sept. 26/51         1         111ey surt.         111ey surt.         111ey surt.           group         Sept. 26/51         1         111ey surt.         111ey surt.         111ey surt.           group         Sept. 25/51         1         11ey surt.         111ey surt.         111ey surt.           group         Sept. 25/51         1         111ey surt.         111ey surt.         111ey surt.           surt.         20/51         1         111ey surt.		18	20	Filey	Tomato
Dutrida         Sturt.         Sept. 24/51         I Miler           Sept. 2/50         I Miler           Sept. 28/51         Miler <td></td> <td>18</td> <td>-</td> <td>Riley</td> <td>Banana</td>		18	-	Riley	Banana
Oct.         10/52         1         Douglas           Sept.         5/50         1         Nouglas           Sept.         5/50         2         1         Nuglas           Sept.         18/50         1         Nuglas         1           group         Sept.         18/51         1         Nuglas           Sept.         26/51         1         Nuglas         1           Sturt.         25/51         1         Nuglas         1           Sturt.	Drosophila putrida Sturt.	24		Riley	Banana
Sept.         8/50         1         N119           Troplets         Wollaston         Sept.         8/50         1         N119           Troplets         Wollaston         Sept.         8/50         1         N119           Froup         Sept.         8/50         1         N119         N119           Group         Sept.         19/51         1         N119         N119           Group         Sept.         19/51         1         N119         N119           Sept.         19/51         2         1         N119         N119           Sept.         19/51         2         1         N119         N119           Sturt.         30/50         1         N119         N119         N119           Sturt.         30/50         1         N119         N119         N119           Sturt.         30/51         1         N119         N119         N119           Sturt.         Sept.         10/51         1         N119         N119           Sturt.         Sept.         25/51         1         N119         N119           Sept.         25/51         1         1         N119		10		Douglas	
Other         5/38         1         Douglas           repleta         Wollaston         Sapt. 8/50         1         Nouglas           Sapt. 18/50         18/50         1         1189           Sapt. 18/50         18/50         1         1189           Sapt. 18/50         18/50         1         1189           Sapt. 28/51         1         1199         1199           Sapt. 18/51         1         1199         1199           Sapt. 28/51         1         1199         1199           Sapt. 28/51         1         1199         1199           Sapt. 28/51         1         1199         1199           Sturt.         3050         1         1199           Sturt.         25/51         1         1199           Sapt. 28/51         1         1199         1109           Sapt. 28/51         1         1199         1109           Sapt. 28/51         1         1199         1109           Sapt. 28/51         1         1199         1199           Sapt. 28/51         1         1199         11199           Sapt. 28/51         1         1199         11199				Riley	Cucumber
Sept. 26/50         E #11ey           Tepleta Wollaston         Sept. 26/50         E #11ey           Sept. 36/50         E #11ey         Hiley           Sept. 36/51         Hiley         Hiley           Sturt.         36/51         Hiley           Sturt.         36/51         Hiley           Sturt.         36/51         Hiley           Sturt.         36/51         Hiley           Sturt.         3601.19/51         Hiley           Sturt.         3601.19/51         Hiley           Sturt.         3601.19/51         Hiley           Sturt.         Sept. 36/51         Hiley           Sturt.         Sept. 36/51         Hiley           Sturt.         Sept. 36/51         Hiley           Sturt.         Sept. 36/51         Hiley           Sept. 36/51         Hiley         Hiley <td></td> <td></td> <td></td> <td>Douglas</td> <td></td>				Douglas	
Tepleta Wollaston         Sept. 18/50         1         Miley           Tepleta Wollaston         Sept. 7/51         1         Pulley           Sept. 7/51         1         Filey         Pulley           Sept. 19/51         1         Filey         Pulley           Sept. 20/50         1         Filey         Pulley           Oct. 20/50         1         Filey         Pulley           Sturt.         20/50         1         Filey           Sturt.         20/51         1         Filey           Sturt.         20/51         1         Filey           Sept. 25/51         1         Filey         Filey </td <td></td> <td></td> <td></td> <td>Piley</td> <td>Mushroom</td>				Piley	Mushroom
Teplete         Wollaston         Sppt. 8/49         1         Hiloy           group         Sept. 8/51         1         Hiloy           group         Sept. 8/51         1         Hiloy           group         Sept. 8/51         1         Hiloy           sept. 8/51         1         Hiloy         Hiloy           ott         8/56         1         Hiloy         Hiloy           sept. 85/51         1         Hiloy         Hiloy         Hiloy           sept. 85/51         1         Hiloy         Hiloy         Hiloy           sept. 85/51         1         Hiloy         Hiloy         Hiloy           sept. 19/51         1         Hiloy         Hiloy         Hiloy           sept. 85/51				Riley	Cucumber
Septe 27/51         4         Domiphan           group         Septe 27/51         1         1110           group         Septe 27/51         1         1110           septe 27/51         1         110         110           septe 27/51         1         110         110           septe 27/51         1         110         110           septe 27/50         1         110         110           septe 27/50         1         110         110           septe 27/51         1         110         110           septe 28/51         1         110         11				Riley	
Sept. 29/51         1         8110           group         58pt. 19/51         1         8110           sspt. 29/51         1         810         9           oct. 30/50         1         8110         9           oct. 30/50         1         810         9           oct. 25/51         1         810         9           oct. 25/51         1         810         9           sept. 25/51         1         811.9         8           oct. 25/51         1         8         8           oct. 25/51				Doniphan	Grape
group 5ept. 19/51 2 Hiley 6roup 06t. 19/51 2 Hiley 06t. 20/50 1 Hiley 06t. 30/50 1 Hiley 06t. 30/50 1 Hiley 06t. 30/50 1 Hiley 06t. 25/51 2 Hiley 06t. 25/51 1 Hiley 9ept. 25/51 1 Hiley 06t. 25/51 2 Hiley 06t. 25/51 2 Hiley 06t. 25/51 1				Riley	Crab apple
group (19/50 1 Riley				Riley	Banana
attract         attract         attract         attract           cote:         30/56         1         attract           cote:         30/56         1         attract           cote:         30/56         1         attract           cote:         25/56         1         attract           cote:         25/56         1         attract           cote:         25/51         1         attract           cote:         26/51         2         attract           cote:         26/51         1         attract           cote:         26/51         2         attract           cote:         26/51         1         attract           cote:         26/51         1         attract           cote:         26/51         1 <td>D. repleta group</td> <td>17</td> <td>50 1</td> <td>Riloy</td> <td>Persimmon</td>	D. repleta group	17	50 1	Riloy	Persimmon
Sturt. 20,500 1 Riley 000: 30,500 1 Riley 000: 30,500 1 Riley 000: 22,51 28,51 1 Riley 000: 25,51 Riley 000: 25,51 Riley			23	Riley	
Sturt. 25/50 1 Miley 000: 30/50 1 Miley 001: 30/50 1 Miley 001: 25/51 2 Miley 001: 25/51 2 Miley 001: 25/51 1 Miley 001: 25/51 1 Miley 001: 25/51 2 Miley 001: 25/51 2 Miley 001: 25/51 1 Miley 001: 25/51 2 Miley 001: 25/51 1 Miley 001: 25/51 Miley 001: 25/51 Miley 001: 25/51 Miley 001: 25/51			20 1	Riley	Banana
Sturt. 20/50 1 61197 Sturt. 20/51 1 6000 004: 23/51 2 61197 397: 23/51 1 81197 397: 23/51 1 81197 397: 23/51 1 81197 397: 19/51 1 81197 397: 23/50 2 81197 597: 23/50 1 81197 597: 19/51 1 81197 597: 19/51 1 81197 597: 19/50 2 81197		30/	50 1	Riley	Grape
Sturt. 22 1 Reno Sturt. 22 30t. 25/51 2 Hiley 30t. 25/51 1 Riley 30t. 25/51 1 Riley 30t. 25/51 1 Riley 30t. 12/51 1 Riley 30t. 12/51 1 Riley 30t. 25/50 2 Riley 50t. 25/50 1 Riley 30t. 15/51 1 Riley 50t. 15/51 1 Riley 30t. 15/50 2 Riley 50t. 15/50 2 Riley 30t. 15/50 2 Riley		30	1 20	Riley	Tomato
Sturt. 000. 23/51 2 Hiley Spt. 25/51 2 Hiley Sept. 25/51 1 Hiley Sept. 25/51 1 Hiley 0.00. 17/51 2 Hiley 3.00. 17/51 2 Hiley Sept. 25/50 1 Hiley Sept. 25/50 1 Hiley Way 20/51 1 Hiley Sept. 15/50 2 Hiley Sept. 15/50 2 Hiley Sept. 15/50 2 Hiley		22		Reno	
Sept. 25/51 1 641ey 3 2012 23/51 1 641ey 3 2012 19/51 1 741ey 3 2012 19/51 2 741ey 3 2012 25/50 2 741ey 5 2012 25/50 1 741ey 7 29/51 1 741ey 8 2012 19/50 2 741ey 8 2012 19/50 2 741ey	robusta	23		Riley	Porsimaon
Sept. 25/51 1 Riley 0.04. 13/51 1 Riley 0.04. 13/51 2 Riley 5.04. 24/51 2 Riley 5.04. 25/50 2 Riley 5.04. 25/51 1 Riley 5.04. 13/51 1 Riley 5.04. 13/51 2 Riley 5.04. 13/50 2 Riley		25	51 1	Riley	Banana
0et.         17/51         1         141ey           Sept.         19/51         2         141ey           Sept.         28/751         1         111ey           Sept.         28/551         1         111ey           Sept.         28/551         1         111ey           Villey         29/51         1         111ey           Villey         29/51         1         111ey           Villey         29/51         1         111ey           Sept.         19/50         2         111sy		28		Riley	Porsimnon
Sopt. 24/51 2 Miley simulans Sturt. Sopt. 24/51 1 Miley Sopt. 25/50 2 Miley Eripunctata Loew. Sopt. 25/51 1 Miley Sopt. 15/51 2 Miley Sopt. 15/50 2 Miley		17		Riley	
simulans Sturt. Sept. 24/51 1 111e7 Sept. 25/50 1 1 11e7 Tripunotate Loew. Sept. 19/51 1 11e7 NW 28/51 1 11e7 Sept. 15/50 2 711e7 Sept. 15/70 2 711e7		19		Riley	Banana
simulans Sturt. Sopt. 25/50 2 Riley tripunctata Loew. Sopt. 19/51 1 Riley Nay 29/51 1 Riley Sept. 19/50 2 Riley		24		Riley	
Uripurotata Loew. Sept. 19/51 1 Riley 3 29/51 1 Riley 3 Sept. 15/50 2 Riley		23		Riley	Mushroom
29/51 1 Riley t. 19/50 2 Riley		•		Riley	Banana
18/50 2 Riley				Riley	Garbage
				Riley	Tomato

### Habitat of Species

Several species of Drosophilidae are known to have distinct kinds of food on which they feed and breed. The foods around the Midwest region are most plentiful during middle and late summer. The natural production of food supply from cultivated farms, fields, and forests in this area can furnish these flies with a variety of foods. Also, garbage pails in homes and garbage cans outdoors can be good feeding places as well as breeding places for the flies. Most of the species of Drosophilidae can be best collected by attention to their natural selection of foods. Therefore, the collection of Drosophilidae may be difficult unless it is a common species such as <u>D. melanogaster</u> that feeds on most of the available fruits in Kansas.

Flies are more apt to feed on semi-liquid to liquid foods. All of the food represented in Table 1 were in these two categories. Some of the more common species of <u>Drosophila</u> caught were recorded in Table 2. The species <u>Drosophila melanogaster</u> Meigen for example showed its great abundance by the number of times caught during September to October of 1950 and 1951. Usually there were at least 50 per cent <u>D</u>. <u>melanogaster</u> in every trap.

The species collected in Kansas may be placed into three general divisions. There were the general scavengers, the fruitfeeders, and the fungus-feeders.

C = Species collected R = Species reared

	Total		日本1000000000000000000000000000000000000	110
		00 10	03 44	60
	CLED SPPLE	0	NH HN H	5
		•	d	
	Wild grape	0	-	-
	Virginia creeper	C F:	02	03
	ь	C F.	Q	03
	Frickly pear	0 20	44°	03
	ь	C R:		01
1	** ** ** **			
1	Banana	0	SHOULH HOL HOLD	24
	** ** ** **	**	4 H	5
	etqqA	U	ri 🔉	N)
	** ** ** **	÷.	ннн	10
1.5	Creamper	0		03
	** ** ** **	÷.		2
tes	Mushroom	0	6 F R	44
tra (	40 00 00 00	8.	03	03
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Jo	endung	0		
80	** ** ** **	R.	H 20 20 H	9
AP	nomutereq	0	H H H H H H	11
0	olanoT	12	CO CO CO	9
-	·· ·· ·· ··	0	H H H	5
In	Grape	84	-	-
us	* * * * *	0	H 10100 H	6
reared using 15 types of substrates.	80 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0		affinia algonutin algonutin algonutin buski buski funbris futtifer futtifer funtifer molance puttie puttie foluste folosit fol	Total
1		1		L

Table 2. Survey of species populations of Drosophilidae in Kansas with number of times trapped or

The general scavengers fed on decaying vegetables (such as potatoes, tomatoes, cucumbers) and refuse in garbage cans (such as bacon, meat, gravy, etc.). The species found on decaying vegetables were <u>D</u>. <u>funebris</u>, <u>D</u>. <u>tripunctata</u>, <u>D</u>. <u>putrida</u>, <u>D</u>. <u>melanogaster</u>, and <u>Scaptomyza adusta</u>. This group of species has been collected either through rearing from the infested host fruits or trapping near the decaying food. Those species feeding on the refuse in the garbage can were <u>D</u>. <u>funebris</u>, <u>D</u>. <u>americana</u>, <u>D</u>. <u>busckii</u>, <u>D</u>. <u>affinis</u>, <u>D</u>. <u>hydei</u>, and <u>D</u>. <u>melanogaster</u>. All of these species was first collected in 1951 during the month of May. The fruit-feeders, such as <u>D</u>. <u>melanogaster</u>, <u>D</u>. <u>affinis</u>, <u>D</u>. <u>hydei</u> and <u>D</u>. <u>americana</u> were attracted to garbage cans probably due to unavailable native foods during early summer.

The fruit-feeders fed largely on the available fruits either native or cultivated; but the majority of this group of species concentrated on the food easily found in Kansas. The common fruits were pear, peach, persimmon, apple, wild grape, cultivated grape, crab apple, prickly pear, pokeweed, banana, and Virginia creeper. <u>Drosophila melanogaster</u> was the only species collected from all the available foods in Kansas. This is indicated in Table 1 under the subheading "substrate", in which there are 15 different food items. The fruit-feeders were the largest among flies collected in Kansas.

The fungus-feeders were found on bracket fungus and mushrooms in Manhattan. Apparently most of the fungus species fed and bred on warm summer months because few were found and collected during the late summer and early fall. This group included the following species: <u>D. putrida</u>, <u>D. hydei</u>, <u>D</u>. affinis and Mycodrosophila dimidiata.

#### List of Species in Kansas

- Mycodrosophila dimidiata Loew. 1862 Berl. Ent. Zeit. 6; 231. This species was described as <u>Drosophila dimidiata</u> Loew.; later Sturtevant transferred as <u>Mycodrosophila dimidiata</u> Loew. (1918. Jour. N. Y. Ent. Soc., 26; 38).
- Aulacigaster lecucopeza Meigen. 1830 Syst. Beschr. 6; 100. The European species were referred to the ephydrine genus Notiphila by Fallen (1910. Sp. Ent. nov. Dipt. 22; ), and to the Geomyzine genus <u>Diastata</u> by Meigen (1830. Syst. Beschr. Zweifl. Insekt. 6; **34**.
- Leucophenga maculosa Coquillett. 1895 Proc. Acad. Nat. Sci. Phila. 47; 317. This species is the European <u>Drosophila</u> maculata Dufour. Kahl transferred the species to <u>Leucophenga</u> (1917. Ann. Carnegle Mus., 11; 364).
- Leucophenga varia Walker. 1849 List. Dipt. Ins. 4; <sup>4</sup>his species was also described as <u>prosophila</u> <u>quadri-</u> maculata Walker. (1856 Dipt. Saund., 4; ).
- Chymomyza amoena Loew. 1862 Berl. Ent. Zeit. 6; 230. This species was described as <u>prosphila amoena</u> Loew. (1862 Ent. Zeit. 6; 230); Sturtevant transferred it to Chymomyza (1916. Ann Ent. Soc. Amer., 9; 325).
- Chymomyza procnemis Williston. 1896 Trans. Ent. Soc. London, 412.
- Scaptomyza adusta Loew. 1862 Berlin ent. Zeit. 6; 231. This species was described as <u>prosophila adusta Loew</u>. Coquillett transferred to <u>Scaptomyza</u> (1910. Froc. U. S. Nat. Mus. 37; 603).
- Scaptomyza graminum Fallen 1823. Dipt. Suec. Geomyz 2; 8. This species was described as <u>presophila graminum</u> Fallen; later Coquillett transferred it to <u>scaptomyza</u> (1910. Proc. U. S. Nat. Mus., 37, 603).

## Drosophila

- D. affinis Sturtevant. 1916 Ann. Ent. Soc. Amer. 9; 334.
- <u>D</u>. <u>algonquin</u> Sturtevant and Dobzhansky 1936. Amer. Nat. 70: 575.
- <u>p. (virila) americana</u> Spencer 1938. Geneticis 23; 169. Spencer described this form as a subspecies of Drosophila virilis Sturtevant.
- D. athabasca Sturtevant and Dobzhansky 1936. Amer. Nat. 70; 576.
- <u>D. buschij</u> Coguillett. 1901. Ent. News. 12; 16. This species was described as <u>D. rubrostriata</u> Becker and <u>D. plurillmeata Villeneuve; now Knab referred</u> these two species to <u>buschii</u> (1918. Bul. Amer. Mus. Nat. Hist. 33; 445.).
- D. funebris Fabriscius 1787. Mant. Ins. 2; 345.
- <u>D. guttifera</u> Walker 1849. List. Dipt. Ins., 4; 1110. This species was also described as D. multipuncta Loew. (1866 Berlin ent. Zeit., 10; 50).
- D. hydei Sturtevant 1921. Pub. Carneg. Inst. Wash., Pub. 301; 101.
- <u>D. immigrans</u> Sturtevant 1921. Pub. Garneg. Inst. Wash., Pub. 301; 83. This species was described as D. tripunctata (1918, Bul. Amer. Mus. Nat. Hist., 38; 445). by Sturtevant.
- D. melanica Sturtevant 1916. Ann. Ent. Soc. Amer. 9; 332.
- D. melanogaster Medgen. 1830 Syst. Beschr. 6; 85. This species has also been known as D. algoriventris Zetterstedt (1847. Dipt. Scand. 6; 2557), D. uvarum Rondani (1875. Bul. Com. Agr. Parm.), and D. ampelophila Loew. (1862. Berlin ent. Zeit. 6; 231.).
- D. putrida Sturtevant. 1916. Ann. Ent. Soc. Amer. 9; 339.
- D. repleta Wollaston 1858. Ann. Mag. Nat. Hist. 41; 117.
- P. robusta Sturtevant. 1916. Ann. Ent. Soc. Amer. 9; 331.
- D. simulans Sturtevant. 1919. Psyche 26; 153.
- <u>tripunctata</u> Loev. 1862. Berlin ent. Zeit., 6; 231. This species was also described as D. modesta Sturtevant 1916 (Ann. Ent. Soc. Amer., 9; 338).

## METHODS USED FOR COLLECTION

Baits and Culture Media

As attractants or baits, peach, banana, apple, grape and tomato were used; among these, fermenting bananas were found to be the best. They are considered best because of the natural odor, outlasting other fruits and containing less water in the fruit.

From time to time, cluture media were used as a substitute for the natural bait. Eridges' laboratory culture was extensively used. The formula is as follows: 100 grams of banana (peaches, pears or similar fruits were used), 100 cc of water, 2 grams of agar-agar. The medium was made by first heating the water, then adding the agar-agar and stirring until it dissolved. Then the crushed banana pulp was added to the heated agar. At the same time 0.5 gram of yeast was added. Then this mixture was allowed to stand for 24 hours before using. This gave time for the medium to cool and the accumulated gas in the jar to escape.

#### Traps and Infested Fruits

The first problem was to collect these flies. There were several ways in which they were collected. On September 15, 1950 infested fruits of pears, peppers, and tomatoes were collected at a garden in Manhattan. They were taken to the insectary in mayonnaise jars or similar vessels for rearing the flies (Table 3). Samples of flies reared were taken and pinned on minuten nadeln for further investigation.

Another method used was the banana trap. Ripe bananas were placed in the mayonnaise jar with two folded paper towels on the bottom of the jar. These were to absorb the excess moisture. The jars were exposed in various places for varying lengths of time from six hours to three days.

A dozen fly traps were placed in a vegetable garden next to the insectary. A stick was driven into the ground, and a jar tied to it about a foot from the ground so that other insects of the crawling type could not get into it.

Flies were also collected by sweeping over the infested fruits with an insect net. Flies were collected on infested or decaying cucumber, tomatoes, and squash by this method. Samples of flies were taken and pinned for determination.

At the garden near the insectary, ink-cap mushrooms of the black-spored gill fungi (<u>Coprinus</u> species) were found to have a few flies infesting the "ink". There were also houseflies (<u>Muscidae</u>) on the mushrooms. A cyanide bottle was used to collect a few specimens and the mushrooms were brought to the laboratory to rear the infesting insects (Table 3).

Other infested fruits brought in by friends increased the study area in Manhattan. Addresses sometimes were not clearly known, but the approximate locality has been marked on the city map (Fig. 2).

Table	3.	Mass rearings from infested fruits and uninfested
		fruits show the possibilities of Drosophilidae
		infestation from five counties during September
		to October, 1950.

Kind of fruit Loc	ality#	Species** and abundance of each
Green pepper	R	None
Tomato	R	(d) 14, (e) 5, (h) 3
Pear	R	(d) 55
Cucumbor	R	(d) 90, (e) 20, (g) 10
Mushroom	R	(d) 20, (g) 10, (1) 5
Bracket fungus	R	(a) 7, (b) 12, (c) 6
Peach	R	(d) 200
Dogweed	P	None
Pokeweed	P	(d) 50, (e) 12
Virginia creeper (Psedera, N. sp.)	P	(d) 15
Wild rose (Rosa L. sp.)	P	None
Wild grape (Vitis L. sp.)	P	(d) 10
Euonymus americanus L.	R	None
Euonymus atropurputeus Jaq.	R	None
Elacagnus umbellata L.	R	None
Vulgare L.)	T	None
Persimmon	R	(d) 29, (b) 7, (c) 11. (f) 4
Prickly pear	H	None
(Copuntia sp.)		
Prickly pear	W	(d) 23, (e) 7
(Copuntia sp.)		
Apple (Staymen)	R	(d) 16, (e) 5
Crab apple (Malus	R	(d) 14
ioensis)		
Hope crab	R	(d) 10
(Malus baccata)		

\*\*

H = Hamilton P = Pottawatomie R = Riley T = Thomas W = Wallace

(a)	Mycodrosopl	nila dimidiata
(b)	Drosophila	funebris
(c)	Drosophila	
(d)	Drosophila	melanoraster
(e)	Drosophila	melano aster and
	- simula	
(1)	Drosophila	repleta group
(g)	Trosophila	putrida
(h)	Prosophila	tripunctata
(1)	Drosophila	simulans

A colony of bracket fungus growing on an old tree stump at the southeast end of the campus was collected and insects reared from it (Tables 3 and 4). A few specimens of <u>Drosophils</u> from the fungus were etherized and placed in the collection box. Species of the family Muscidae were also represented in rearings from the fungus.

Next to the apiary, there was an area planted with corn belonging to the Department of Entomology. Four fly traps were placed under the trees and along the wire fence next to a wooded area nearby (Table 5). These traps were placed in the shade because the day time temperatures were high. The baits would have desiccated in the sum and there would have been little chance of catching the flies. A few specimens of <u>Drosophila</u> were taken from each jar for the collection or identification.

On October 6, some specimens were collected on the Horticultural Farm. Several jars with crushed grapes and apples were placed in the orchards. The following day, the jars were collected and brought back to the laboratory. The only fruits available at that time in the orchard were Staymen apples and grapes, since cherry and peach were out of season. Samples were collected earlier from Concord and Urbana grapes.

Fruits and berries in the neighborhood of the entomology greenhouse were caged for possible infestation. One of several traps was made by picking up ripe persimmons from the ground and putting them in collecting jars (Table 5). Three of these jars were strung from the branches of near-by shrubs to

Table 4. Mass rearings from infested fruits and with uninfested fruits show the possibilities of Drosophilidae infestation from four counties during September to October, 1951.

Kind of fruit Lo	cali	ty*:	Spec	ies	an	d ab	unda	nce	10	each
Green pepper	R		Non	8						
Tomato	R		(c)	20,	(d)	10,	(1)	5		
Pear	R		(c)	38						
Peach	R		(c)	29						
Bracket fungus	R		(a)	3						
Virginia creeper	R		(c)	8						
(Psedera, N. sp.)										
Crab apple	R		(b)	8,	(c)	6				
(Malus icensis)										
Hope crab	R		(b)	6,	(c)	14				
(Malus baccata)										
pple (Delicious)	L		(c)	34						
Frape (Concord)	L		(c)	14						
Luonymus americanus L.	R		Non							
uonymus atropurputeus	R		Non	3						
Jag.										
Elaeagnus umbellata L.	R		None	8						
Persimmon	R		(c)	25,	(0)	7				
(pple (Delicious)	D			23	(-)					
Apple	C			13						
C = Cloud	49-49-	(a)	Myes	odros	oph	ila d	11m1	tist		
D = Dickinson		(b)				affin				
L = Lyons		(0)				nelar		star		
R = Riley		(a)				nelar				bd
		( ~ )			nula		TOP EL	1001	, au	1.04
		(0)	Dros			hyde				
		(1)		soph						

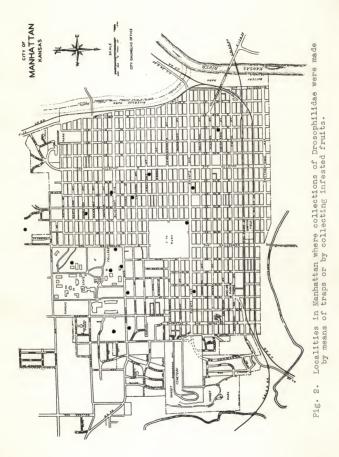


Table	5. Tre	Trapping data on in the fall of records of the		a sp. using midity and ate College	ills sp. using various fruits from di Humidity and temperature were taken State Colloge Department of Physics.	lts from di vere taken of Physics.	fferen from ti	Prosophila sp. using various fruits from different localities in Kanaas 1950. Humidity and temperature were taken from the hydrothermograph Kanaas State College Department of Physics.
Type & trap b	& number of: baits used ;	of: Kansas ed: locality	: Time :/ :exposed: : hrs. :	: Time : Approx. no.: :exposed: specimens : : hrs. : caught :	: Time of : collection	: fumidity:ature	:Temper-	.: :Species represented.
Banana		Insectary garden	36	58	9:30 a.m.	87	60.5	D. melanogaster
	02		37	55	10:30 a.m.	88 • 5	19	D. melanogaster D. busck11
	0		28	34	12:15 noon	87	70	D. melenogaster D. melenogaster & simulans
	41		43	60	4:30 p.m.	85	17	D. melanogaster Scaptomyza adusta
	н		36	40	5:30 p.m.	35	70.5	D. melanogaster & <u>D. melanogaster</u> &
	CV		33	45	5:30 p.m.	35	70.5	D. melanogaster D. melanogaster & Simulans
	63		362	25	6;00 p.m.	55	60.5	D. melanogaster
Pear	1	Manhattan	24	88	11:30 a.m.	82.5	80	D. melanogaster & D. melanogaster & simulans
Poar	64	Manha ttan (College Cafeteria)	24	105	10:30 a.m.	83	78.5	D. melanogaster

	1.1				tion	lon	tion
	-: :Species represented. : Hemarks	D. putrida D. melenogaster	D. <u>hydel</u> D. <u>melanogaster</u>	D. affinis	D. melenogester & <u>simulans</u> D. <u>melenogester</u> pay before collection was very windy.	D. melenogester & simulans D. melenogester D. <u>function</u> Dey before collection was very windy.	D. melanocaster Day before collection was very windy.
	: Humidity:ature :	74	72	82	62	62	62
	: Huni.	64	59	53	52	22	52
Q	of tion	• E • d	a.m.	noon	8 • 111 •	R •III •	8.•m.•
	Time of : collection :	4:00 p.m.	10:00 a.m.	12:00 noon	10:00 a.m.	10;30 a.m.	10:30 a.m.
- Aller	Time : Approx. no.: xposed: specimens : hrs. : caught :	Q	57	33	19 19	44 8	54
	: Time : Approx. no. :exposed: specimens : hrs. : caught	24	24	24	8	58	80
		Manhattan (school campus)	Manhattan (school campus)	Manhattan (school campus)	Manhattan (Hort. Farm)	Manhattan (Hort. Farm)	Manhattan (Hort. Farm)
	ar of :	r.	-	03			
1 • 411001 • • • • • • • • • • •	Type & number of: Kansas trap baits used : locality	Persimon			Staymen apple	Concord Erape	Urbana grape

Table 5. (cont.)

Table 5. (cont.)

: Ilme :Approx. r Type & number of: Kansaa :exposed: specime trap baits used : locality : hrs. : caught	ber of	Kansas locality	: Time :/ :exposed: : hrs. :		Time of collection	: Humidity:ature : . % : °F. :	sr-: a :Species represented. : Remarks
Banana	н	Manhattan (school. campus east)	36	50	12:00 noon	45 82	D. melanogaster
Benana	01	Manhattan (school campus west)	33	50	1:00 p.m.	44	D. melanogastor
Pear		Don1phan	*	80			D. 1.mulgrens D. busekii D. hydei D. melanogaster
Banana peels		Colby	over- night	22	early morning	Av. 48	48 D. melanogaster
		Syracuse	over- night	30	early morning	Av.	50 D. melanogaster
		Garden City	1	32	12:00 noon	AV.	70 D. melanogaster
		Goodland	over- night	none	early morning	Av. 42	42 L. melanogaster
Banana media		Osborne	*	10	*	AV. 35	25 D. melenogaster Culture media was too dry. Needed moisture.

-
-
100
12
1.0
0
12
0
-
10
24.5
- 42
-
100
100
123

Table 5. (concl.)	(•					
Type & number of: Kanaas :exposed: specimens trap baits used : locality : hrs. : caught	: Kensas : locality	: Time : .exposed: . hrs. :		: Time of :Humic : collection : %	: flumidity:ature : % . or.	: :Temper-: Time of :Humidity:ature :Species represented. collection : % : Op. : Remarks
Benana modia	Concordia	*	ω	*		D. melanogastor Medla needed mois- turre. It was too cold.
Banana močia	Lyons	*	none		49	Needed moisture content in culture media. Too cold and hardly any files around.

Trapped by county agents. The average temperature has been taken from the Climatological Data of Kansas.

救

try to obtain Drosophilidae eggs for rearing.

A professor brought in some fruits, and flies trapped on banana peels during a trip to western Kansas. He also collected ripened prickly pears fro Hamilton and Wallace counties and brought them back to be caged for possible Drosophilidae infestations. The localities from which specimens were collected were Garden City, Colby, Syracuse, Sharon Springs, and Goodland, Kansas. Banana peels, and garden refuse were the chief items used as trap baits (Table 5).

Several times while the traps were being collected. there were butterflies (<u>Styridae</u>, <u>Pieridae</u>), houseflies (<u>Musoidae</u>), bluebottle flies (<u>Calliphoridae</u>), and honeybees (<u>Apidae</u>) present together with the Drosophilidae.

With the aid of Mr. Dell Gates, Extension Entomologist, contact was made with county agents for collecting flies in their respective counties. Instructions regarding an ideal spot where flies could be found were given. This material was then given to six county agents.

In middle of May, 1950, the first species of Drosophilidae have been seen around Manhattan. There were large populations of these flies concentrated in and around the garbage can outside the homes between Laramie and Moro Street in the 900 block. Traps were set near the garbage cans and the first collection of these flies was made on May 26, 1950 (Table 6).

A collection trip to eastern Kansas from September 8 through 11 was made through the aid of a friend. Some infested

emper-: ature ;Species represented. 07. ; Remarks	D. melanoraater D. <u>affinis</u> Cloudy & cool day.	<u>P. melanogaster</u> Faining very hard	D. melanogaster	None	D. melanogaster It was dark & raining very hard	D. melanogaster	D. melanoraster	D. melanogaster It was a chilly morning.	D. melanogaster	D. melanogaster
:Temper-: : ature :S : oF. :	Av. 68	65	65	65	102	10 OL	10 10	04	70	10
Time of collection	4:20 p.m.	9:00 a.m.	9:00 8.00.	9:00 a.m.	4:00 p.m.	4:00 p.m.	4:00 p.m.	9:00 в.т.	9:00 a.m.	9:00 a.m.
specimens : caught :	Ω	თ	11	ŝ	4	9	C2	ß	.00	63
: Time : Approx. no :exposed: specimens : hrs. : caught	25	over- night	over- night	over- night	¢1	03	C3	over- night	over- night	-19AO
: Kansas : locality	Olpe	Chanute	Chanute	Chanute	Caney	Caney	Caney	Arkansas City	Arkansas City	Arkansas
& number of: baits used :	Ч	Ч	C1	63	Ţ	03	ю	н	C1	63
Type & trap ba	Banana									

	Species ropresente E. <u>melanogaster</u> It was a hot & humid It was a hot & humid It was a bright & suu afternoon. It was a bright & suu afternoon. It was a bright & suu afternoon. <u>E. unebris</u> <u>E. melanogaster</u> <u>D. affinis</u> <u>effinis</u> <u>effinis</u>	1 Temport - 1 2 Contract - 1	Time of collection 9:00 a.m. 1:30 p.m. 1:30 p.m. 3:30 p.m. 5:30 p.m. 5:00 p.m. 5:00 p.m.	a	<pre>: "fine :Approt. no : :xyosed: specimens : :xyosed: specimens over- 7 nicht 7 là none là none l none l none l none 2 4 2 4 2 5 2 5 2 6</pre>	: Kanses : Locality Arkanses City Medicine Lodge Fratt Bend	A mumber of batta used batta used a 1 1 3 3 3 3 4 1 1 3 3 4 1 1 3 3 4 1 1 3 4 1 1 1 3 4 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	
Wolhaven over 2 0.00 n n 60 h		65.5			01		10	
2 0 5:00 p.m. 65.5 D.		65.5	5:00 p.m.		01		CN	
2 5 5:00 peille 65.5 D 2 5:00 peille 65.5 D 2 5:00 peille 65.5 D 2 0 5:00 peille 65.5 D		65.5			01	Great Bend	H	
Great         2         4         5:00 p.m.         65.5         D           Bend         2         5         5:00 p.m.         65.5         D           2         5         5:00 p.m.         65.5         D         D           2         5         0         5:00 p.m.         65.5         D           2         0         5:00 p.m.         65.5         D           2         0         5:00 p.m.         65.5         D	oid	67	3:30 p.m.		F		63	
1         none         3:30 peme         67         It was a bright & affermoon.           Great         2         4         5:00 peme         65.5         0         unbubris           Band         2         5         5:00 peme         65.5         0 <u>prinburis</u> 2         5         5:00 peme         65.5         0 <u>prinburis</u> 2         0         5:00 peme         65.5         0 <u>prinburis</u> 2         0         5:00 peme         65.5 <u>printing</u> 2         0         5:00 peme         65.5 <u>printing</u>	a bright & noon.	67	3;30 p.m.		н		C3	
1     none     3:30 p.m.     67     It was a bright & afternoon.       1     none     3:30 p.m.     67     It was a bright & afternoon.       1     none     3:30 p.m.     67     It was a bright & afternoon.       Creat     2     4     5:00 p.m.     65.5 <u>P. unbbris</u> Bend     2     5     5:00 p.m.     65.5 <u>P. unbbris</u> 2     0     5:00 p.m.     65.5 <u>D. aftinis</u> 2     0     5:00 p.m.     65.5 <u>D. aftinis</u> 2     0     5:00 p.m.     65.5 <u>D. aftinis</u>	was a bright & fternoon.	67			н	Pratt	Ч	
Fratt         1         none         3:30 p.m.         67         It was a bright & affernoon.           1         none         3:30 p.m.         67         It was a bright & affernoon.           1         none         3:30 p.m.         67         It was a bright & affernoon.           1         none         3:30 p.m.         67         It was a bright & affernoon.           1         none         3:30 p.m.         67         It was a bright & affernoon.           1         none         3:30 p.m.         65.5 <u>p. fundoria</u> 2         5         5:00 p.m.         65.5 <u>p. fundoria</u> 2         5:00 p.m.         65.5 <u>p. affinia</u> 2         0         5:00 p.m.         65.5 <u>p. affinia</u>	was a hot & humid	69		ouou	12		63	
13     none     1:50 p.m.     59     It was a hot Sint       Fratt     1     none     5:50 p.m.     67     It was a bright       1     none     3:30 p.m.     67     It was a bright       1     none     3:30 p.m.     67     It was a bright       1     none     3:30 p.m.     67     It was a bright       1     none     3:30 p.m.     67     It was a bright       1     none     3:30 p.m.     67     It was a bright       1     none     3:30 p.m.     67     It was a bright       1     none     3:30 p.m.     67     It was a bright       1     none     3:30 p.m.     67     1       1     none     5:00 p.m.     65.5 <u>p. fundbrids</u> 2     5:00 p.m.     65.5 <u>p. affinis</u> 2     0     5:00 p.m.     65.5 <u>p. affinis</u>	was a hot &	69			122		C3	
13     none     1:30 p.m.     69     It was a hot & humid       13     none     1:30 p.m.     69     It was a hot & humid       14     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     67     It was a bright & su       1     none     3:30 p.m.     65.5     P. <u>unibridestor</u> 1     bend     2:00 p.m.     65.5     P. <u>milinogestor</u>	WAS R	69	l:30 p.m.		1.2	Medicine Lodge	ы	Banana
1       Wedicine       14       none       1:30 p.m.       69       It was a hot & humid         2       14       none       1:30 p.m.       69       It was a hot & humid         3       14       none       1:30 p.m.       69       It was a hot & humid         1       Fratt       1       none       1:30 p.m.       67       It was a bright & au         2       1       none       5:30 p.m.       67       It was a bright & au         2       1       none       5:30 p.m.       67       It was a bright & au         2       1       none       5:30 p.m.       67       It was a bright & au         3       1       none       5:30 p.m.       67       It was a bright & au         4       5:00 p.m.       67       It was a bright & au         8       4       5:00 p.m.       65.5 <u>ftinobris</u> 9       5:00 p.m.       65.5 <u>ftinis</u> <u>ftinis</u> 8       5:00 p.m.       65.5 <u>ftinis</u> <u>ftinis</u> 8       5:00 p.m.       65.5 <u>ftinis</u> <u>ftinis</u> 9       1       ftinis       ftinis       ftinis       ftinis		70			over- night	Arkanses City	H	Pear
1     Arkenses     over- night     7     9:00 a.m.     70     0.     0.     malanogester       a     1     Modicine     1%     none     1:50 p.m.     69     1t was a hot & humid       2     1%     none     1:50 p.m.     69     1t was a hot & humid       3     1%     none     1:50 p.m.     69     1t was a hot & humid       1     Prett     1     none     1:50 p.m.     69     1t was a bright & au       2     1     none     3:50 p.m.     69     1t was a bright & au       2     1     none     3:50 p.m.     67     1t was a bright & au       2     1     none     3:50 p.m.     67     1t was a bright & au       3     1     none     3:50 p.m.     67     1t was a bright & au       4     5:00 p.m.     67     1t was a bright & au       2     2     4     5:00 p.m.     67     1t was a bright & au       3     3     3     3     3     1t was a bright & au       4     5:00 p.m.     67     1t was a bright & au       5     5     0     5     1t was a bright & au       6     5     1t was a bright & au     1     1       7     <		:Temper- : ature : or.	Time of collection	pprox. no.: specimens : caught :	: Time :A :exposed: : hrs. :		mber of	Type & nu trap bait

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43
27
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200
0
CS.
54

Type & number of trap baits used	& number of: baits used :	Kansas Iocality	: Time : Approx. no. :exposed: specimens : hrs. : ceucht	Approx. no.: specimens : cencht :	Time of collection	ature : or.	: Species represented. : inemarks
Banana	60	McPherson	over- night	10	9:00 a.m.	68	D. melanogaster
Apple	ы	McPherson	over night	15	9:00 a.m.	68	D. melanogaster Chymomyza amoena
Banana	ч	Manhattan	over- night	10	9:30 a.m.	61	<pre>D. funebris <u>1. busckii</u> <u>nelenogaster</u> </pre>
Banana.	63	Manhattan	over- night	18	9:30 a.m.	61	D. americana D. melanogaster
Banana	CI	Manhattan	over- night	81	9°30 a.m.	61	D. robusta D. repleta D. melanogaster
Persimon	Ч	Manhattan	over- night	11	4:00 p.m.	56	D. melanogaster
Persimon	02	Manhattan	over- night	13	4:00 p.m.	56	D. melanogaster
Persimon	0	Kanhattan	over- night	O3	4:00 p.m.	56	D. melanogaster D. <u>affinia</u> <u>americana</u>
Banana	ы	Manhattan	over- night	15	9:30 a.m.	65	D. repleta D. robusta D. melanogaster

2     Machattan     over- night     15     9:50 a.m.       3      over- night     19     0:00 a.m.       10     1      0ver- night     6     2:00 p.m.       2      over- night     6     2:00 p.m.       3      0ver- night     5     2:00 p.m.       4       5     2:00 p.m.       5       5     2:00 p.m.       6       5     0.00 p.m.       7       5     3:00 p.m.       6       5     0.m.       7       5     0.m.	Type & number of: trap baits used :	r of:	Kansas locality	: Time :A :exposed: : hrs. :	: Time : Approx. no.: exposed: specimens : hrs. : caught :	Time of : collection:	: temper-: s ature : : or.	Species represented. Ramarics
3     over light     19     9:30 a.m.     65     10.       10     1     night     6     2:00 p.m.     57     10.       2     night     8     2:00 p.m.     57     10.       3     over     5     2:00 p.m.     57     10.       4     1     over     5     2:00 p.m.     57     10.       5     over     5     3:00 p.m.     57     10.       6     1     over     5     3:00 p.m.     57     10.       1     over     5     3:00 p.m.     57     10.       1     over     5     3:00 p.m.     57     10.       1     budge city     0.     3:00 p.m.     57     10.	Banana		ianhattan	over- night	13	9:30 a.m.	65	
10       1       over- night       6       2:00 p.m.       57       10- 10- 10- 10- 10- 10- 10- 10- 10- 10-	Banana	10		over night	19	9:30 a.m.	65	
2         over- nicht         8         2:00 p.m.         57         0.0 0.0           5         over- nicht         5         2:00 p.m.         57         0.0           6         1         over- nicht         5         3:00 p.m.         57         0.0           2         over- nicht         5         3:00 p.m.         57         0.0           2         over- nicht         9         3:00 p.m.         57         0.0           2         over- nicht         9         3:00 p.m.         57         0.0	010	-1		over- night	Q	2:00 р.ш.	57	
3         over- night         3         2:00 p.m.         57         2: 5           b         1         over- night         5         3:00 p.m.         57         2: 5           2         over- night         9         3:00 p.m.         57         2: 5           b         0ver- night         9         3:00 p.m.         57         2: 5           b         0ver- night         9         3:00 p.m.         57         2: 5		63		over- n1ght	ω		57	
over- night     5     3:00 p.m.     57     D. D.       over- night     9     3:00 p.m.     57     D.       Dodge City     over- night     9     3:00 p.m.     57     D.		64		over- night	cu		57	
2 over 9 3:00 p.m. 57 D. D. Dodge City over none 52 Non	٩	н		over- night	ŝ	3:00 р.т.	57	
Dodge City over none 52 night		CN		over- night	03	3:00 р.ш.	57	
	pple	had	bodge city	over night	none		52	None

Species represented. Pemarks		repleta melanogaster funetris	funebris affinis	melano_aster			hydel fundtata fundtata fundtata amoriena amoriena affinis fundta fundtata fundta weig mand 2 colloc- tions man daw
Spec.	None	D. Fui	D. fur	D. mel	None	enon	
: Temper-: : ature : : oF.	58	unknown	unknown	unknown	umoun	unknown	60 62 75 75 75 75
Time of collection		3:00 p.m.	3:00 p.m.	3:00 p.m.			9;30 а.ш. апа 4;30 р.ш.
:Approx. no.: : specimens : : caught :	anone	35	32	23	anone	none	33
: Time :Approx. no. :exposed: specimens : hrs. : caught	over- night	72	72	72		over- night	MOOK
Kansas locality	McPherson	<b>Wathena</b>	Kathena	wathena	Norton	Edmond	Manhattan
& number of: baits used :		Ч	03				
Type & number of Kansas trap baits used : locality	Apple	Grape -	Grape	Apple	Banana peels	Banana peels	Banana

An average daily temperature ranging from 60° to 79° F. for six days.

fruits and trapped flies on bananas and apples were collected from Lyon, Neosho, Montgomery, Cowley, Barber, Stafford and McPherson counties. The traps were distributed in and out of the city limit of each town or city. Some traps were placed near fruit stands and others under fruit trees and garbage cans. The traps were either exposed overnight or two hours in each county (Table 6).

A trip to Wathena, Kansas was made on October 15, 1951 to collect Drosophilidae primarily on apples and grapes. In the apple orchards, countless numbers of decaying or rotting apples were on the ground and many of them were infested with these flies. Traps were exposed and collected by Mr. Elbert Eshbaugh around the apple orchards and grape vineyards at Wathena (Table 6).

During trips to northwest and southwest Kansas, Dr. R. H. Painter exposed traps with banana peels and apples in Norton, Washington, McPherson and Ford counties.

Various host fruits infested by species of Drosophilidae gathered during the fall of 1950 and 1951 are indicated in Tables 3 and 4. The common hosts that were infested in the state of Kansas were used in a comparative study. Some of the common fruits were pear, peach, persimmon, apple, wild grape, cultivated grape, crab apple and prickly pear. Other hosts used were vegetables, fungi, and berries.

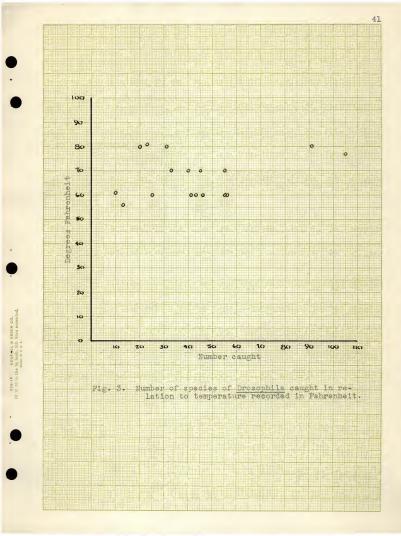
# Time, Humidity, and Temperature Most Favorable for Trapping

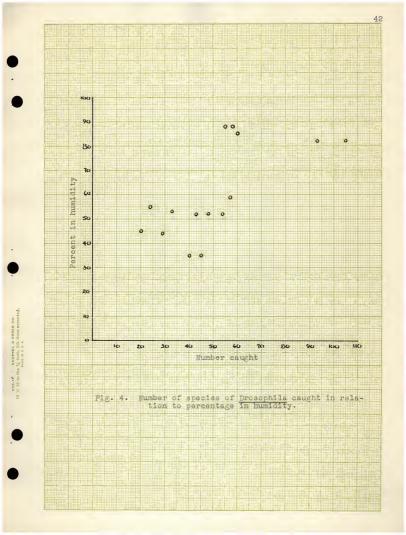
Trapping of Drosophilidae requires a knowledge of their ecology and physiology. The distribution of species can be determined partly through a study of collection data which is based on particular species and their habitat.

A slightly shaded area close to a decaying vegetable or fruit was found to be best for collecting Drosophilidae. The best time for collecting flies was found to be from 8:30 to 10:30 a.m. and from 4:00 to 5:00 p.m. Also, when the day was cloudy with high humidity at a temperature of  $80^{\circ}$  F., there was an increase in the numbers of the flies caught. One of the limiting factors in trapping the flies is rain. The rainfall, varying in amount, can ruin the catch and trap by filling the jar with water. It is difficult to find a completely sheltered area where a trap can be placed, unless beneath a roof of a house or similar structure.

All during this time of collecting, the temperature from September 15 to September 30 ranged from 60° to 85° Fahrenheit. The temperature best suited for collecting flies ranged between 75° to 85° Fahrenheit. Although time and humidity are mentioned separately, all three (time, humidity and temperature together) play an important role in the abundance of flies.

The temperature (Fig. 3) and the humidity (Fig. 4) were





plotted against the trapping of flies during the fall of 1950 from September to October in Manhattan area. A closer relationship is indicated between the number of species caught in relation to percentage of humidity than to that of temperature. The humidity factor, as well as lowering of the temperature, probably has much to do with the fact that flies come to traps in large numbers immediately before or after a rain. These flies were unable to venture to the traps during a high wind.

The temperature and humidity data are derived from hydrothermograph charts at Kansas State College Department of Physics.

### Rearing

In the laboratory each day, the jars were examined for a possible new generation from the infested fruits. After three to four days, varying numbers of tiny (0.5 mm) larvae were seen crawling on the fruit in the jars. They fed on the food for about four to six days at  $75^{\circ}$  to  $80^{\circ}$  F. room temperature until they were ready to pupate. The full grown larvae were white, legless, and 1 mm to 1.5 mm long. The full grown larvae attached themselves to the paper towel by the secretion of a silk-like substance. The cuticular wall of the larva changed into a puparium. Five to six days were approximately the length of the puparial stage. A single generation required from 14 to 16 days. The length of the life cycle depended on the quality of

the media and the temperature.

Ripened fruits or berries were collected for possible dormant eggs or other stages and were brought back to be reared under favorable conditions. The fruits were placed in the jars and a dash of water was sprinkled upon them. The jars were tightly covered with cheese cloth. Under room temperature, the eggs would be expected to hatch into larvae. Two weeks later, molds began to grow in one of the jars; and day by day, the molds increased in size. The wild grapes did not develop any mold infestation but instead, several <u>Drosophila</u> flies emerged. A tentative reason for the lack of development of flies on other berries and fruits was first, the berries were too dry and second, <u>Drosophila</u> require a fermenting fruit or a berry undergoing decay (Flate I).

A rearing oven was used in some cases to keep the culture at a constant temperature because the temperature in the room waried between 10 to 20 degrees Fahrenheit. The rearing oven may have been a failure because the carbon dioxide did not escape freely from the oven. All of the flies in the oven were dead. Whenever the oven was opened, the pungent odor spread over the entire room. This experiment was repeated several times by placing new cultures in the oven.

# EXPLANATION OF PLATE I

Methods used in the study of Drosophilidae

- Fig. 1. Fruit infested by Drosophilidae in a glass con-tainer enclosed by cheese cloth.
- Fig. 2. Culture media used as baits for trapping Drosophilidae.



# METHOD OF CONTROL

# Insecticidal Control Test

The insecticides used in this experiment were from three commercial brands. The first was the Cooper-Chlor #4.

The second insecticide used in the experiment was Lindex We-10 (containing lindane).

The third was Pestroy, 25 per cent DDT concentrate with a resin base.

# Small Scale Field Applications with Three Insecticides

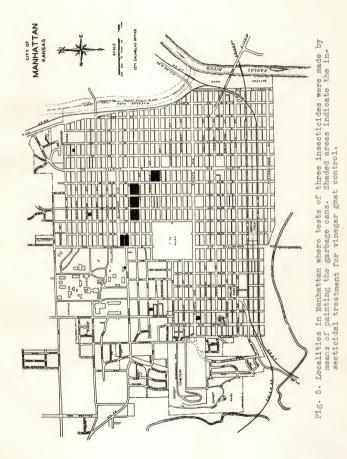
During the first test a 0.5 per cent water emulsion of DDT prepared from 25 per cent concentrate of DDT, 0.2 per cent water emulsion of Lindane, and 0.2 per cent water emulsion of Chlordane were used. In each case, alternate insecticides were used in each city block. These solutions were applied at the rate of approximately 100 milligrams per square foot by painting the inside of the lid of the garbage can and around the outside of the top of the garbage can. Most garbage cans used were of three sizes; 12 inches, 14 inches, and 20 inches diameter.

The insecticides were painted on the garbage cans with paint brushes. The treated street blocks with insecticides are designated on Fig. 5. They were distributed mainly throughout the central section of the city of Manhattan. The street blocks used were the 800, 900, and 1000 blocks between Moro and Laramie (east to west through the alley). Between Moro and Bluemont in 1200 block garbage cans were also painted. Other places were only designated home and neighbors of 1801 Poyntz Avenue, 1015 Thurston Street, 1729 Fairchild Street and 715 Fremont Street where three insecticides were used. Also the garbage cans were painted at the College Cafeteria, Mar's Cafe and Goodnow Courts. The applications of insecticides were made at two different times. The first application was made during the latter part of September, 1951 and the second in the earlier part of October, 1951.

The insecticides were applied during relatively warm days with little precipitation. The average precipitation during the week September 24 through September 30, 1951 was .075. The week following from September 30 to October 8, 1951 it was .061. And the average temperature for the week ending September 30 was 68° F. with the highest for the week 85° F. and the lowest,  $34^{\circ}$  F. The average for the following week, from September 30 through October 8, 1951 was 60° F. with the highest for the week 88° F. and the lowest  $34^{\circ}$  F.<sup>1</sup>

Methods used in measuring fly densities after insecticidal

<sup>1</sup> Temperature and rainfall were taken from Climatological Data of Kansas and Kansas State College Department of Physics.



treatment were compared with the density of the pre-control population. After 48 hours, observations were made at each garbage can to detect the reduction of the fly population. The 0.5 per cent water emulsion of DDT showed a marked reduction of the fly population whereas 0.2 per cent lindane and chlordane gave less reduction of the fly population. To indicate the relative percentage of control, in several garbage cans there were approximately 15 flies prior to the treatment but next day there were zero to 10 living flies in the garbage cans. This was only a partial indication of what might be the effectiveness of the toxicants but it provided some evidence. However, all flies that entered the garbage can may not have died immediately. The chlorinated hydrocarbon insecticides have been relatively slow in action against the flies by means of contact (Shepard, 1951). Considering these facts, it appeared that there may have been 50 to 75 per cent control.

In a second application of insecticides, 0.3 per cent of lindane and chlordane were used. There were marked differences in the reduction of the fly populations. It was impossible to obtain 100 per cent control in all cases.

For further study, three empty five gallon lard cans were used to simulate garbage cans. Inside the cans, a mixture of ripe bananas, pears, and peaches was placed in quart bottles approximately in the center of the lard can. The inside of the lid and around the outside of the top of the garbage cans were painted with a mixture of 0.5 per cent DDT, 0.3 per cent

lindene and 0.3 per cent chlordene. The lids of the cans were left open just enough for the flies to crawl in and out the cans.

The average counts after 48 hours using 0.5 per cent DDT have been 3.2 dead flies in the fruit jar and 2.0 dead flies inside the can. With the 0.3 per cent lindane, there was an average of 3.4 dead flies in the fruit jar and 4.4 dead flies inside the can. The third can using 0.3 per cent chlordane there was an average of 2.8 dead flies in the fruit jar and 4.2 dead flies in the can.

The garbage can and the lard can experiments constituted evidence that about equal control can be secured from all three insecticides. A more extensive painting or spraying of insecticides might have caused better results.

# CONCLUSION

The traps and naturally infested fruits were the chief sources for collecting species of Drosophilidae. Many types of baits were used, but fermented banana gave the best results. Some of the common fruits used were tomato, pear, peach, persimmon, apple, grape, crab apple and various berries. Other hosts were plant parts and fungi. In rearing, there were a few species, such as <u>D</u>. <u>melanogaster</u> which were reared from several substrates but others were closely related to a single substance. The substrates differed from one another by their longer lasting quality. Also, certain substrates, such as persimmon, peach, apple, etc. attracted more flies than others. There were close relationships between substrates gathered during the fall of 1950 and 1951 which have had similar adult emergence of Drosophilidae.

There were several materials on which rearings were attempted which gave no indication of fly infestation. One possible reason for this failure might have been the temperature fluctuation during the time of the rearings. The temperature was extremely high during the day and very low during the night in the rearing laboratory. Also condensation of toxic matter might have inhibited the growth of Drosophilidae. Many of the materials during the time of rearing accumulated a great abundance of mold, due primarily to excess moisture in the rearing jars and to less air circulation within the jars.

The total number of specimens reared from wild hosts during the fall of 1950 and 1951 totaled 1,050 of which 455 were <u>D. melanogaster</u>. Roughly 50 per cent of the total number reared were <u>D. melanogaster</u>. Among approximately 1,469 specimens trapped during the fall of 1950 and 1951, at least 75 per cent were D. melanogaster.

<u>Prosophila melanogaster</u> flies were either trapped or reared 72 different times while <u>Drosophila funebris</u> was the second most common species with 8 different collections.

There were more flies in the fly traps brought in during the morning than during the afternoon hours. An interesting species collected during the investigation was Mycodrosophila dimidiata which fed on fungus.

The study of the relationship of humidity and temperature with trapping was made at Manhattan during the fall of 1950. The highest percentage of flies was caught when the humidity ranged from 40 to 65 per cent. In other words, the humidity and the number of specimens caught during the period of study showed a positive relationship whereas any relationship between the temperature and the number of specimens caught was less evident within the range studied. Between 9:30 to 10:30 a.m. was the best time for collecting flies because a great number of Drosophilidae came to the baits at that time.

The field work with three insecticides on garbage cans was applied by painting the inside of the lid and outside of the top of the can. The evidence showed that some control can be secured by such means. To prevent Drosophilid infestation around homes, it is necessary to destroy the breeding places and adult females. The satisfactory and logical means of controlling the insects is the use of insecticides since satisfactory sanitation cannot be secured.

Since 1903, five genera and ll species have been recorded in the list of Drosophilidae in Kansas. Among the four genera and 19 species collected during 1950 and 1951, there was one genus and 13 species not previously recorded in the list of species from Kansas. The latter include: <u>Mycodrosophila</u> <u>dimidiata</u> Loew., <u>Drosophila algonquin</u> Sturtevant and Dobzhansky, <u>Drosophila athabasca</u> Sturtevant and Dobzhansky, <u>Drosophila</u> <u>americana</u> Spencer, <u>Drosophila guttifera</u> Walker, <u>Drosophila</u> <u>hydei</u> Sturtevant, <u>Drosophila immigrans</u> Sturtevant, <u>Drosophila</u> <u>melanica</u> Sturtevant, <u>Drosophila putrida</u> Sturtevant, <u>Drosophila</u> <u>robusta</u> Sturtevant, <u>Drosophila repleta</u> Wollaston, <u>Drosophila</u> tripunctata Loew. and Drosophila simulans Sturtevant.

There were four species of Drosophilidae collected in Kansas but not recorded in any of the four states bordering the state of Kansas. These species are: <u>Mycodrosophila dimidiata</u>, Leucophenga maculosa, Leucophenga varia, and Drosophila americana.

The most commonly known species from Kansas and also from Missouri, Oklahoma, Solorado, Nebraska were <u>D. funebris</u>, <u>D.</u> <u>hydei</u>, <u>D. melanogaster</u>, <u>D. simulans</u>, <u>D. putrida</u>, and <u>D. busckii</u>.

### SUMMARY

The species of vinegar gnats (Drosophilidae) and their control have been studied under the conditions prevailing in Kansas.

The Drosophilidae are not of economic importance as far as direct damage is concerned, but because of their great abundance in homes they become very annoying.

During the fall of 1950 and 1951, baits were used for trapping Drosophilidae. As attractants (baits), nine types of fruits were used. Because of the natural attraction of the insect to the odor and the longer lasting of the fruit, banana

was selected as the best bait. Also culture media were made to use in the traps procedure. The fruits were placed in the mayonnaise jars as traps in the Manhatten area and in several counties in Kansas.

Flies were also collected by sweeping the net over the infested or decaying vegetables and fruits.

Various hosts, such as fruits, fungi and plant parts, infested by Drosophilidae were collected in September and October 1950 and 1951. Some of the common fruits used were tomato, pear, peach, persimmon, apple, grape, crab apple and various berries. Most species of Drosophilidae, except <u>D. melenogaster</u> and D. simulans, were collected only on specific substrates.

An interesting species collected during the investigation was Mycodrosophila dimidiata which fed and bred on fungus.

Time, humidity, and temperature were the important elements concerned in successful trapping of Drosophilidae. Between 9:30 to 10:30 a.m. was the best time for collecting flies because of great numbers of Drosophilidae coming to the baits at that time.

The study of the relationship of humidity and temperature with trapping was made at Manhattan during the fall of 1950. The humidity and the number of specimens caught during the period of study showed a relationship whereas any relationship between the temperature and the number of specimens caught was less evident within the range studied.

Among the 72 collections from traps, there were only 8 without flies or approximately 10 per cent unsuccessful collections. . Among 22 different kinds of materials from which rearings were attempted, flies emerged from 10 substrates. This is approximately 55 per cent without infestation by Drosophilidae.

The field work with three insecticides on garbage cans was applied by painting the inside of the lid and the outside of the top of the can. The evidence showed that some control can be secured by such means. To prevent Drosophilidae infestation around homes, it is necessary to destroy the breeding places and adult females. The satisfactory and logical means of controlling the insect requires the use of insecticides when satisfactory sanitation cannot be secured.

During these studies, four genera and 19 species have been collected and reared. From the total number of species, there were 13 species not previously recorded from Kansas including one belonging to a previously unreported genus.

There were four species of Drosophilidae collected in Kansas but not recorded in any of the four states bordering the state of Kansas. These species were: <u>Mycodrosophila</u> <u>dimidiata</u>, <u>Leucophenga maculosa</u>, <u>Leucophenga varia</u>, and <u>Drosophila americana</u>.

The species <u>Scaptomyza</u> <u>adusta</u>, <u>D</u>. <u>tripunctata</u>, <u>D</u>. <u>atha-</u> <u>basca</u>, and <u>D</u>. <u>repleta</u> represent the farthest north that these species have been found.

The most commonly known species from Kansas and also from Missouri, Oklahoma, Colorado, and Nebraska were: <u>D</u>. <u>funebris</u>, <u>D</u>. <u>melanomaster</u>, <u>D</u>. <u>simulans</u>, <u>D</u>. <u>putrida</u>, <u>D</u>. <u>busckii</u>, and <u>D</u>. <u>hydei</u>.

# ACKNOT LEDGMENT

Assistance, suggestion, and cooperation have been received from numerous persons during the experiments and studies of this work.

Grateful appreciation is expressed to Dr. Reginald H. Painter, under whose supervision this work has been conducted. His valuable advice, interest and continual help in every phase of the work are appreciated.

Special credit is due Dr. Roger C. Smith for his undivided attention and critical suggestion which helped greatly in this work.

Gratitude to Dr. Marshall R. Wheeler for his valuable work in determining the species of Drosophilidae.

Many thanks to Dr. Paul A. Dahm and Dr. Howard E. Evans whose valuable contributions and suggestions have been administered toward the work.

Others whose cooperation or important contributions toward the making of this work are Dell E. Gates, Elbert L. Eshbaugh, Kenneth D. Havel, Professor Leon R. Quinlan, Roland L. Fischer, Joseph E. Pankaskie, Mrs. Ester Dominick, and various county agents.

Sincere thanks to James Y. Murashige, a Kansas State College student, who assisted with the field studies.

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STUDIES ON THI SPECIES OF VINEGAR GRATS (DIPTERA: DROSOPHILIDAE) IN KANSAS

by

CARL MASARU YOSHINOTO

B. A., Iowa Wesleyan College, 1950

AN ABSTRACT OF A THESIS

submitted in partial fulfillment of the

requirements for the degree

MASTER OF SCIENCE

Department of Entomology

KANSAS STATE COLLEGE OF AGRICULTURE AND APPLIED SCIENCE

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