This is the author's final, peer-reviewed manuscript as accepted for publication. The publisher-formatted version may be available through the publisher's web site or your institution's library.

# Plant-mediated interactions: considerations for agent selection in weed biological control programs

Lindsey R. Milbrath and James R. Nechols

# How to cite this manuscript

If you make reference to this version of the manuscript, use the following information:

Milbrath, L. R., & Nechols, J. R. (2014). Plant-mediated interactions: Considerations for agent selection in weed biological control programs. Retrieved from http://krex.ksu.edu

# **Published Version Information**

**Citation**: Milbrath, L. R., & Nechols, J. R. (2014). Plant-mediated interactions: Considerations for agent selection in weed biological control programs. Biological Control, 72, 80-90.

**Copyright**: © 2014 Published by Elsevier Inc.

Digital Object Identifier (DOI): doi:10.1016/j.biocontrol.2014.02.011

#### Publisher's Link:

http://www.sciencedirect.com/science/article/pii/S104996441400036X

This item was retrieved from the K-State Research Exchange (K-REx), the institutional repository of Kansas State University. K-REx is available at <a href="http://krex.ksu.edu">http://krex.ksu.edu</a>

# Plant-mediated interactions: considerations for agent selection in weed biological control programs

Lindsey R. Milbrath<sup>a,\*</sup> and James R. Nechols<sup>b</sup>

<sup>a</sup>United States Department of Agriculture, Agricultural Research Service, Robert W. Holley
 Center for Agriculture and Health, 538 Tower Road, Ithaca, NY 14853-2901, USA
 <sup>b</sup>Department of Entomology, Kansas State University, 123 W. Waters Hall, Manhattan, KS
 66506-4004, USA

\*Corresponding author e-mail: Lindsey.Milbrath@ars.usda.gov, fax: 1-607-255-1132, phone: 1-607-254-7268

#### Abstract

1

Plant-mediated indirect interactions among herbivores (arthropods and pathogens) are common 2 and extensively reported in the ecological literature. However, they are not well-documented 3 with respect to weed biological control. Such interactions between biological control agents can 4 5 have net positive or negative impacts on total weed suppression depending on the strength of the interaction(s), the relative importance of the agent indirectly impacted, and the combined weed 6 suppression that results. A better understanding of plant-mediated interactions may improve 7 decision-making about which agents to introduce in classical biological control programs for 8 9 greatest impact on invasive weeds. This paper reviews the subject, including examples from the 10 biological control literature; outlines the need for research on indirect effects of herbivores on other herbivores; discusses how such knowledge may strengthen classical biological control 11 12 programs for invasive weeds; and provides recommendations for the kind of studies that should be done and how information about plant-mediated interactions could be integrated into agent 13 evaluation protocols, to assist in decision-making about agents for importation and release. 14 15 Keywords: Indirect interactions; Weed biological control; Pre-release evaluation; Agent 16 selection 17

#### 1. Introduction

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

33

34

35

36

37

38

39

40

During the past two decades there has been considerable discussion and debate about how to evaluate candidate agents for classical biological control of weeds so that decisions to release agents will pose a minimal risk to non-target organisms (e.g., Briese and Walker, 2008; Follet and Duan, 2000; Heard, 2000; Louda et al., 2003, 2005; Pemberton, 2000; Sheppard et al., 2005; Wajnberg et al., 2001; Withers et al., 2000). An outcome of this discussion has been a more detailed review of candidate agents which, likely, has reduced the rate of agents released. Adopting a more deliberate and parsimonious approach for releasing biological control agents also increases the need for making better predictions about which agents are most likely to become established, and to have the greatest impact on invasive weeds. In that regard, recent reviews have called for more extensive research on the biology and impact of candidate agents, weed life histories and population dynamics, and environmental conditions affecting weedherbivore interactions (Mills and Kean, 2010; Morin et al. 2009; Müller-Schärer and Schaffner 2008). A potentially important, but neglected, category of weed-herbivore interactions in research and in pre- and post-release evaluations of prospective biological control agents concerns indirect effects of herbivorous agents on other agents resulting from induced changes in host plant quality, or in the quantity of resources available. Because the effects of these plantmediated interactions (hereafter, P-MIs) on agents may be positive or negative (or not occur at all), they have the potential for increasing or decreasing the level of biological control and, thus, the impact on weed populations. To date, experimental evidence for P-MIs in the weed biological control literature is limited (see references in Table 1). However, extensive documentation of a wide range of plant-mediated effects involving herbivores and pathogens in

the ecological literature (see reviews by Agrawal et al., 1999; Damman, 1993; Denno et al., 1995; Hatcher, 1995; Hatcher and Paul, 2001; Karban and Baldwin, 1997; Masters and Brown, 1997; Ohgushi et al., 2007; Wootton, 1994) suggests that these indirect interactions are common among organisms used to control invasive weeds. Further, the intensity of indirect interactions can increase with potentially higher densities of introduced herbivores and pathogens due to decreased natural enemy loads (Denno et al., 1995). How frequently P-MIs occur, what impact they have on agents, and ultimately how this effects weed populations, is unknown. Research is needed to provide answers to these questions and, in so doing, will help to determine whether studies of plant-mediated effects should be incorporated into evaluation protocols for biological control programs. In addition to providing practical information, investigations of P-MIs would also benefit theory. For example, the Multiple Stress Hypothesis, which has been used widely in biological control programs, assumes that most weeds require more than one stressor for effective suppression, such as multiple species of biological control agents targeting different locations and/or plant functions (Harris, 1981, 1991). A second, related assumption is that agents that feed in a complementary manner, i.e., on different organs or in ways that partition the weed resource (e.g., James et al., 1992), will not compete because they do not interact directly. Based on what is now known about P-MIs, the second assumption needs to be examined more rigorously. Therefore, even if research suggests that multiple agents may improve weed suppression, a prerelease assessment of agents for possible negative P-MIs should lead to a better introduction strategy and greater overall success (see 4. 'Research needs and recommendations'). We also expect that research on P-MIs targeting weeds will add to our fundamental knowledge by revealing new mechanisms underlying indirect interactions, and by providing a more complete

41

42

43

44

45

46

47

48

49

50

51

52

53

54

55

56

57

58

59

60

61

62

63

understanding of the complexity of existing interactions. Furthermore, as the list of empirical examples of P-MIs accumulates, new patterns, predictions and hypotheses will result, thus contributing to ecological theory.

64

65

66

67

68

69

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

The scope of this paper encompasses P-MIs between herbivorous agents, which we define broadly to include both arthropods and plant pathogens. Discussion and examples of effects of indirect interactions involving weeds and different species of herbivores are given. The focus is on single season interactions; but we include cases where P-MIs carry over to the next growing season (e.g., winter annual, biennial, and perennial weeds). We also consider situations where life cycles of agents span more than one season (e.g., species that have an intervening period of dormancy). Most of the examples are for agents that have one generation per year (univoltinism); however, P-MIs can occur when one or both species has two or more generations. Because plantmediated effects can be unidirectional or bidirectional, we discuss cases where one or both agents is responsible for inducing plant-mediated effects. And as P-MIs may occur in agents that are spatially or temporally isolated from one another, we cover both and include a related discussion about persistence (duration) of effects and pervasiveness (local vs. systemic) within plants. We exclude all direct interactions among agents (e.g., direct forms of competition, intraguild predation, etc.), indirect exploitative competition involving agents that are present at the same time and place (e.g., Berube, 1980), and cases involving systemic pathogens (e.g., Kruess, 2002; see section 3.1). With respect to introduction strategies in classical biological control programs, discussion and recommendations about P-MIs apply to all situations, including: (1) multiple agents being considered for importation (Gerber et al., 2007); (2) additional agents considered to supplement previously imported species (Swope and Parker, 2010); and (3) importation and release of agents in environments where resident herbivores

(either indigenous or fortuitously introduced) are present (e.g., Caesar, 2003; Hatcher, 1995; Moran, 2005).

The primary purpose of our paper is to highlight the role that P-MIs may play in weed biological control programs. In the following sections we review types of P-MIs, summarize known examples of P-MIs in weed biological control programs, discuss research needs to better utilize knowledge about P-MIs for enhancing biological control, and provide recommendations for evaluating outcomes of different categories of P-MIs in relation to agent selection.

# 2. Overview of plant-mediated interactions

Plant-mediated interactions among herbivores pertain to a particular category of indirect interactions in which feeding or other activities by an arthropod or a plant pathogen induces changes in a plant which then affect other species feeding on the same plant (see reviews by Ohgushi, 2005; Ohgushi et al., 2007; Wootton, 1994). Interspecific interactions among plant-feeding organisms can occur within and between feeding guilds, and with taxonomically-related or unrelated species. Effects on an herbivore may be positive, negative, or neutral depending on how the herbivore responds to the altered plant. Denno and Kaplan (2007) describe herbivore-induced effects that result in positive or negative outcomes for other herbivores as induced susceptibility (facilitation) or induced resistance, respectively.

Plant responses to herbivory that result in indirect plant-mediated effects are diverse and include changes in structure (e.g., texture, tissue hardness, growth form, etc.), physiology (e.g., growth rate and productivity), and/or chemistry. The latter may involve primary products such as nutrients or secondary chemicals, especially those used for plant defense against herbivores (Karban and Baldwin, 1997). In turn, induced effects on plants influence herbivores by a range of mechanisms which include ecological (e.g., food resources, altered risk of natural enemy

attack, shelter [Damman, 1993; Marquis and Lill, 2007]), behavioral (Milbrath and Nechols, 2004a), phenological (synchrony between herbivores and plant resources [Milbrath and Nechols, 2004a]), and physiological (e.g., herbivore growth and development, survival, reproduction [Hunt-Joshi and Blossey, 2005; Simelane, 2006]). Indirect P-MIs occur in plant-feeding species that are separated spatially (Blossey and Hunt-Joshi, 2003; Buccellato et al., 2012; Campanella et al., 2009; Hunt-Joshi and Blossey, 2005; Masters and Brown, 1997; Raghu et al., 2012; Simelane, 2006), temporally (Damman, 1989; Wold and Marquis, 1997), or both (Gerber et al., 2007; Milbrath and Nechols, 2004a; Swope and Parker, 2010; Swope and Stein, 2012; Van Hezewijk and Bourchier, 2012). Thus, species that interact indirectly fall into one of three categories: spatial overlap but temporal separation; spatial separation but temporal overlap; spatial and temporal separation (Denno and Kaplan, 2007; Denno et al., 1995; Ohgushi, 2005). The extent to which plant-mediated effects impact an herbivore population may depend on several factors, including (a) how much temporal overlap exists between its population and the one that induces the plant effects, (b) the amount of time separation of each species on the host plant, (c) the amount of time the life stage(s) of the inducing herbivore is present, (d) the persistence of the herbivore-induced plant effect(s) in relation to the plant and/or herbivores' life histories, (e) the strength/magnitude of the effect(s), and (f) the type and distance of spatial separation (see below; Damman, 1993). Another consideration as to what constitutes spatial isolation depends on whether herbivoreinduced effects are local or systemic (i.e., influencing the entire plant, affecting source-sink dynamics, etc.) and also on whether herbivores share a feeding guild. For example, although whiteflies and leafminers both feed on leaves and may occur in close physical proximity to one another, the fact that one is a phloem-feeder whereas the other chews through the mesophyll may

111

112

113

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

allow spatial separation, at least in terms of P-MIs. The fact that negative indirect interactions commonly occur refutes the outdated notion that partitioning plant resources spatially or temporally is sufficient for herbivores to avoid competition. Thus, indirect P-MIs have a bearing on how niche separation is defined.

Herbivore-induced indirect interactions appear to be quite common in nature. For example, Denno et al. (1995) noted that 53% of the interspecific interactions surveyed in their study involved plant-mediation. They also found that a majority of the indirect interactions was asymmetrical, suggesting that P-MIs are largely unidirectional or, if bi-directional, one herbivore has greater influence than the other.

Plant-mediated effects on herbivores are not fixed but, rather, are condition-dependent. Some of the factors known to influence the elicitation, magnitude, and/or persistence of plant-mediated responses are 1) the manner in which herbivores feed, and the feeding sites; 2) the sequence of attack (i.e., 'priority effects' in Erb et al. 2011); 3) other plant stressors (e.g., abiotic); 4) herbivore densities (e.g., threshold or graded responses) (Crowe and Bourchier, 2006 vs. Smith and Mayer, 2005); and 5) the species of herbivores involved in the interaction (Agrawal et al., 1999 [various chapters]; Sabelis et al., 2007). Although all of these factors are of potential importance, we view herbivore densities (or pathogen titers) as a key consideration in assessing and predicting P-MIs in weed biological control (see Section 3).

# 3. Plant-mediated interactions in weed biological control

We examined the primary literature and review articles for examples of P-MIs in weed biological control programs in which agents were separated in space and/or time. A general overview of these papers is shown in section 3.1, and Table 1 summarizes the key features of

each study in relation to the others. Section 3.2 and Table 2 provide an in-depth analysis of a case study we did on weevils introduced to control musk thistle, which relates P-MIs to weed impact. Section 3.3 discusses problems in predicting P-MIs, and how this relates to weed impact. The relevance and applicability of investigating P-MIs in weed biological control are covered in sections 3.4 and 3.5.

# 3.1. Examples of P-MIs in weed biological control (Table 1)

Until recently relatively few published papers have dealt explicitly with P-MIs between biological control agents. Furthermore, as noted by Hatcher and Paul (2001), evaluations of weed biological control programs have generally considered either the impacts of herbivores/pathogens on weeds (but not necessarily all possible combinations of agents), or interactions among herbivores/pathogens, but not both. Our search found fourteen papers representing ten weeds that examined P-MIs. Six weed examples (seven papers) involved agents separated spatially but present at the same time, and four weed examples (seven papers) involved agents separated in space and time. These are listed in Table 1, which includes the target weed name, the interacting biological control agents, the type of interaction observed, a summary of the combined agent impact to the plant relative to individual agent impact (only six of the ten weed examples included a plant impact assessment), and the experimental venue. Three of the examples involved an insect-pathogen interaction. We could not find appropriate examples involving agents that feed on the same plant parts and are only separated temporally from each other.

The studies were a mixture of greenhouse and field (including common garden) experiments, and most were retrospective (i.e., they involved agents that had already been released in the

field), with the exception of Gerber et al. (2007) and Raghu et al. (2012) which were done prospectively (i.e., prior to agent releases). Only three of the studies were conducted in the native range (see Table 1). Interactions between spatially-separated agents, in which a two-way interaction is possible, were variable although two papers did not examine both possible interactions. The paper by Raghu et al. (2012) considered P-MIs among three herbivore species, making for two sets of a two-way interaction. As a result, out of seven pairs of herbivores (14) possible interactions) in this category, P-MIs were fully characterized for only five pairs of herbivores. One of the agents usually had no apparent effect on the other species (four of the five herbivore pairs that were fully examined, Table 1). In contrast, the effect of the second species on the first agent could vary depending on the parameter measured or the type of experiment (e.g., Hunt-Joshi and Blossey, 2005). The P-MIs in this case were primarily negative (five of seven herbivore pairs) and ranged from strong to weak, i.e., only one of the measured parameters was affected (Table 1). Only Campanella et al. (2009) and Raghu et al. (2012) noted potential positive (facilitative) interactions among spatially-separated herbivores (see Table 1), although the specific mechanisms were unknown. Interactions between agents separated in space and time, in which a one-way interaction occurred, were neutral (one weed example), negative (two), or mixed (studies of yellow starthistle, Centaurea solstitialis L., Table 1). Low densities of some agent species may not have allowed for the full range of P-MIs to be characterized. For example, Hunt-Joshi et al. (2004) noted that the experimental densities achieved for the weevil Hylobius transversovittatus Goeze were much lower than field infestations in Europe, which could partly explain the relative lack of impact of the weevil on the leaf beetle Galerucella calmariensis L. or their shared host plant.

180

181

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

The combined agent impact on the plant that was reported tended to be equal to that of a single agent, but could vary according to the plant parameter measured or environmental condition such as soil type (Table 1). For example, the combination of the rust fungus *Puccinia* jaceae Otth var. solstitialis and seed-head insects caused a greater reduction in seed of yellow starthistle than either type of herbivore alone for plants growing on serpentine soils but not on non-serpentine soils (Swope and Stein, 2012). Plants growing on non-serpentine soils were much larger and fecund than those on serpentine soils. An unanticipated effect reported by Swope and Parker (2010) and Swope and Stein (2012) was that the presence of the rust fungus reduced seed feeding by various insect larvae via an unknown mechanism without generally altering survival rates of the larvae. This was hypothesized to be due to either a decrease in seed quality from an induced defensive response (negative effect) or an increase in seed nutrient quality (positive effect). As a result, the negative effect of the fungus on seed production of yellow starthistle growing on non-serpentine soils was cancelled out by the reduced seed predation. Interference (of the fungus with the insect agents) can occur despite the lack of a strong signal of competition. In one case it was difficult to distinguish whether a response was indirect and plant-mediated or a direct response to the agent itself. In the study by Simelane (2006) there were clear-cut P-MI effects of leaf-feeding lace bugs that negatively influenced root quality for larvae of the flea beetle Longitarsus bethae Savini & Escalona. However, Simelane (2006) also showed that adult flea beetles avoided lantana plants that were infested with high densities of lace bug nymphs. This may represent a plant-mediated behavioral response, but alternatively may reflect a direct response to the presence of the other herbivore. Similarly, a study by Kruess (2002), which we do not cite as a P-MI example, found that when thistles were systemically infected by the fungal pathogen *Phoma destructiva* (Plowr.), this resulted in negative effects on both adult feeding and

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

oviposition behavior, as well as preimaginal development, survival and body weight of the leaf beetle *Cassida rubiginosa* Müller. However, because the fungal pathogen is distributed throughout the plant, it is unclear whether the leaf beetle responded indirectly to changes in plant quality, directly to the fungal pathogen, or a combination of both (see similar study by Kluth et al. [2001] involving a different systemic fungal pathogen of *Cirsium arvense* (L.) Scop. whose effects on herbivores cannot be distinguished). While we have excluded direct interactions from consideration in this paper, it is clearly important to know if/how the influence of one herbivore on another, whether from direct or indirect interactions, influences agent populations and performance and, thus, the impact on target weeds.

# 3.2. Musk thistle case study (Table 2)

The interaction between musk, or nodding, thistle, *Carduus nutans* L., and its two primary biological control agents - the weevils *Rhinocyllus conicus* Froelich (head weevil) and *Trichosirocalus horridus* (Panzer) (rosette weevil) - represents perhaps the best known example of plant-mediated indirect effects for weed biological control agents (see Milbrath and Nechols, 2004a,b for full details). Musk thistle is a Eurasian/North African plant introduced into North America in the mid-1800s, where it became a noxious weed in pasture and rangeland (Dunn, 1976; Stuckey and Forsyth, 1971). It grows primarily as a biennial or winter annual and is propagated only through seed (McCarty and Scifres, 1969). *Trichosirocalus horridus* adults oviposit into vegetative rosettes from late fall through spring. Larvae feed in the root crown and destroy the apical meristem, which may alter subsequent plant architecture and flowering and therefore indirectly seed set (Cartwright and Kok, 1985; Woodburn, 1997). *Rhinocyllus conicus* adults oviposit onto flower heads (capitula) produced in the spring and early summer, and the

larvae feed on the receptacle tissue, reducing seed production (Kok, 2001; McCarty and Lamp, 1982; Shorthouse and Lalonde, 1984; Surles and Kok, 1976). Thus, the two species of herbivores attack different plant parts at different times of the year, leading to a one-way (asymmetric) interaction of *T. horridus* on *R. conicus*.

A series of field and greenhouse experiments were conducted to retrospectively address two questions in parallel: does early vegetative feeding by *T. horridus* alter musk thistle as a resource for the later-arriving *R. conicus*, and how might this influence the individual and combined effects of *R. conicus* and *T. horridus* on musk thistle seed production? The P-MI was hypothesized to be negative and occur via four mechanisms (Table 2) in which altered plants would affect: the seasonal synchrony between ovipositing *R. conicus* and musk thistle flower head availability (phenological effect); the acceptability of musk thistle flower heads for oviposition by *R. conicus* (behavioral effect); the quantity of plant tissue available to developing *R. conicus* larvae (resource limitation effect); and the quality of musk thistle heads for developing *R. conicus* larvae (physiological effect).

A negative P-MI was confirmed although it was influenced by the density of *T. horridus* larvae (see Table 2 for summary). Low densities (<20 larvae per plant) generally had no effect on musk thistle phenology or flower and seed production (although it did create multiple-stemmed plants) and no effect on adult *R. conicus* oviposition and subsequent production of new adults compared to thistles not previously infested by *T. horridus* (Milbrath and Nechols 2004a,b). In contrast, high larval densities of *T. horridus* (66 per plant) resulted in thistles producing fewer flower heads which were also delayed in their first appearance by one week compared to non-infested thistles. A cascade of plant-mediated effects ensued - there was a reduction in 1) *R. conicus*-musk thistle flower head synchrony, 2) the acceptability of *T*.

horridus-infested musk thistle to ovipositing *R. conicus*, 3) the quantity and 4) quality of resource available to *R. conicus* larvae. As a result, the production of new *R. conicus* adults was reduced 63% (Milbrath and Nechols, 2004a). This study revealed for the first time multiple and diverse plant-mediated effects between spatially- and temporally-isolated herbivores in a weed biological control system.

Despite the large, negative effect of *T. horridus* on the recruitment of new *R. conicus*, the reduction in musk thistle viable seed was still greater when both weevils were present (59%) compared to *R. conicus* alone (45%) (Table 2; Milbrath and Nechols, 2004b). Hence, the outcome for biological control programs may not necessarily be adverse even with plant-mediated competition because of compensatory trade-offs concerning the relative impacts of the two herbivores on the weed. Regional environmental conditions also may alter the relative abundance of the two weevils, thus affecting their interaction and combined impact on musk thistle.

# 3.3 Problems in making predictions of P-MIs and weed impact

Because of the low number of published studies concerning P-MIs for weeds, general predictions about whether a plant-mediated response will occur, and, if so, what kind, are not possible at the present time. Besides including a limited taxonomic range and number of feeding guilds among known examples, most if not all studies are incomplete in one or more respects, including not having evaluated reciprocal effects of interacting herbivores, conducting experiments that were either too short or could not be done under realistic field conditions, and not assessing organisms over a range of densities (threshold effects). Despite these limitations, a comparison of studies in which similar taxonomic groups and feeding guilds of herbivores

elicited and were affected by P-MIs, suggest that the types of plant-mediated effects produced also may be similar. For example, in both studies where leaf feeders indirectly affected root feeders (spatial separation: Hunt-Joshi and Blossey, 2005; Simelane, 2006), the authors reported reduced survival of root-feeding larvae. As noted by Blossey and Hunt-Joshi (2003), this is only a partial fulfillment of the predictions of the model of Masters et al. (1993) regarding interactions between above- and below-ground herbivores, and contradictory examples are known. Milbrath and Nechols (2004a) and Van Hezewijk and Bourchier (2012) demonstrated that when crown- and root-feeding weevils altered plant structure and floral productivity, the behavior and resources available to spatially- and temporally-separated seed feeding weevils were negatively impacted. Finally, an evaluation of the impact of a leaf rust pathogen on several later-arriving weevil and tephritid fly species, which attack flower heads of thistles, resulted in reduced feeding in all species (Swope and Parker, 2010; Swope and Stein, 2012).

Besides known examples of P-MIs from the weed literature, a review of plant-mediated herbivore responses by Denno and Kaplan (2007) also suggested that certain P-MI responses of herbivores that feed and live in particular ways may be predictable. For example, phloem feeders commonly alter the flow of nutrients by creating localized sinks at feeding sites, and other phloem feeders located distally to the inducing species or at other plant locations may suffer from reduced nutrients (Inbar et al., 1995; Larson and Whitham, 1991). We suggest that knowing how early-colonizing herbivores impact plant structure, chemistry, and growth as well as where and how they feed may lead to predictions about the type of plant-mediated responses that might occur, which can then be tested experimentally.

#### 3.4. General relevance to weed biological control

The study of P-MIs could improve decision-making regarding single versus multiple agent introductions, or which agents to consider, especially in cases where a competitive or even neutral interaction may occur without the benefit of additional impact on the target weed (Hatcher and Paul, 2001; Swope and Parker, 2010; Swope and Stein, 2012). Issues of agent complementarity, which may involve plant-mediated indirect effects among agents, are acknowledged in programs in New Zealand (see Barratt et al., 2010). Alternatively, as pointed out by Swope and Parker (2010), it could be beneficial to identify facilitative or synergistic interactions in order to prioritize such combinations of agents for release. However, to do so will require experimental evaluation of single and multiple agents for indirect interactions and weed impact. As already noted, there are few studies of P-MIs for weeds that have experimentally evaluated the impact of each agent, and the two agents together, for the target weed. Likewise, the issue of what threshold densities are required to trigger a plant-mediated response has been tested only rarely (e.g., Hunt-Joshi et al., 2004; Milbrath and Nechols, 2004a). Both prospective and retrospective studies are needed to assess general patterns of P-MIs in weed biological control. Interactions of exotic or indigenous species with previously-released agents or resident species are also amenable to this approach, and therefore can be used in classical, augmentative (including bioherbicides), or conservation biological control programs, including hybrid versions of these approaches.

335

336

337

338

339

334

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

# 3.5. Relationship to efficacy filter-first approaches

Weed biological control workers have advocated for the assessment of a candidate agent's efficacy before proceeding to host range tests in order to minimize the release of ineffective agents (e.g., Balciunas and Coombs, 2004; McClay and Balciunas, 2005; Raghu et al., 2006).

One common method to investigate efficacy involves impact studies on individual plants using live insects or simulated herbivory. Our suggested protocol (outlined below) is an expansion of an impact study approach. A second method involves demographic modeling of target weed populations in order to identify guilds of potentially effective agents. In particular, matrix population models are used to identify plant life stage transitions that have the greatest impact on population growth rates of the target weed and should be targeted for disruption (Caswell, 2001). Researchers typically incorporate known impacts of candidate agents in the models to assess their potential effects on plant population growth (e.g., Davis et al., 2006; Raghu et al., 2006). If multiple agents are being modeled simultaneously, their impacts may not be additive, i.e., independent, in the case of plant-mediated competition or facilitation. Our expanded impact approach can provide more precise information for the perturbation analyses (although impact studies on all relevant plant life stages will be needed).

#### 4. Research needs and recommendations

Several questions need to be explicitly addressed in experimental evaluations of P-MIs to generate fundamental information on the frequency and type of P-MIs between biological control agents and the outcome for weed control. Among them are: 1) Do P-MIs occur among specific agents, and if so, are they positive or negative? 2) How strong is the effect? 3) What is the impact, and hence potential contribution to control, of each agent individually on the target weed, and does the level of impact change with a combination of agents? 4) If only one agent elicits a plant-mediated effect and it is negative, does it compensate for any reduced impact by the other agent? 5) If more than one agent is affected simultaneously, are there off-setting positive and negative effects? 6) Are plant responses triggered at critical agent densities?

In addition, it will be important to know how climatic and other environmental conditions affect herbivore populations (and therefore the relative abundances of interacting species), resulting plant-mediated responses, and ultimately the level of biological control achieved. One area of research that has received little attention involves plant-mediated effects across growing seasons (Denno and Kaplan, 2007). Given the number of perennial weed targets, this is a crucial gap in our knowledge. It is also of value for short-term studies, especially if conducted in a quarantine-laboratory or greenhouse, to be validated post-release given concerns of how accurately such studies translate to long-term field populations of the agents and the weeds (Hunt-Joshi and Blossey, 2005; Rayamajhi et al., 2010).

We conclude that investigating P-MIs will lead to better decisions about which agents to release, at least for some weed biological control programs. In that regard, the recommendations for evaluation and release which follow should provide a useful framework for investigators who wish to incorporate plant-mediated effects into candidate evaluation protocols. In putting forth these protocols, we acknowledge that they do not cover the full scope of possible interactions. For example, the scenarios we developed are for only two candidate agents because including more would make the experimental evaluation much more complex (e.g., see study by Kluth et al. (2001) involving multiple weed-feeding herbivores that exhibited numerous effects ranging from positive to negative to neutral, at least some of which may have been plant-mediated). In addition, our examples cover plant-mediated effects that occur within plant growing seasons, but not all cases of indirect effects that might occur between growing seasons. Specifically, we include one-way interactions whereby an agent feeding in one growing season induces either delayed or persistent plant changes that impact a second agent in a subsequent growing season. Two-way interactions between temporally-separated agents utilizing a perennial host that occur

between growing seasons (e.g., Strauss, 1991) are not considered in this paper because of the complexity of developing evaluation protocols. However, carefully-designed long-term studies accompanied by single- vs. dual-agent experimental evaluations could help document whether such interactions impact weed suppression positively or negatively.

We do not minimize the challenges of experimental evaluations in the field which, although more realistic, can be subject to uncontrolled variables versus logistical issues regarding the ecological realism of quarantine-based studies and hence the post-release predictably of results (Lonsdale et al., 2001). Ultimately, practitioners should document the outcomes of programs utilizing release decisions that were derived from our recommended protocols, in order to validate the usefulness (predictability) of this approach.

# 4.1. P-MI experimental protocols

The following protocols expand upon the design of plant impact studies widely used in biological control of weeds programs and apply to cases where two agents are separated in space and/or time. Numbered items represent a logical sequence.

- 1. Evaluate impact of candidate species on target weed, and on each other, experimentally. Design should evaluate candidate agents singly and in combination along with controls (no agents). Studies could be done concurrently with host specificity testing if desired, but prior to agent release. Field tests in the native range are preferred (e.g., common garden approach of Gerber et al., 2007), but logistical problems may require that evaluations be done under quarantine conditions.
- 2. Incorporate a range of densities for both agents in the design, including zero (agents not present), as it is difficult to predict field densities that will result following a release (Gassmann,

1996; Zalucki and van Klinken, 2006) or those that are needed to elicit a plant-mediated indirect response (Milbrath and Nechols, 2004a). This might require the use of cages, such as in open field experiments in the native range, to ensure high densities are maintained. We expect that, as agent densities increase or decrease, the relative impact of an individual agent, as well as the indirect interaction between agents (and hence the combined impact of the two agents), will change. Threshold effects are likely.

- 3. Use introduced plant genotypes in the experiment, due to known or potential differences between introduced and native populations in genetics (e.g., Treier et al., 2009), defensive chemistry (e.g., Wheeler et al., 2003) and/or tolerance (e.g., Wang et al., 2011). If multiple genotypes exist in the invaded range, consider including this as an experimental variable (e.g., Campanella et al., 2009).
- 4. Other factors that can alter P-MIs, such as plant size, plant competition, nutrient status, mycorrhizal fungi (Ohgushi et al., 2007), and soil type (Swope and Stein, 2012), should be considered but are at the researchers' discretion and will not be discussed further.
- 5. When conducting experiments, release agents at phenologically appropriate stages of plant development (e.g., Smith and Mayer, 2005 compared to Crowe and Bourchier, 2006).
- 6. If possible, allow agents to make choices, especially the adult stage. Not only does this reflect a realistic scenario in many field situations where some but not all plants have been exposed to an agent, it may also reveal plant-mediated effects on herbivore behavior. For example, in our field experiment with musk thistle, *R. conicus* females could choose between *T. horridus*-infested and non-infested thistles. The experimental outcome of reduced recruitment of new *R. conicus* adults was heavily influenced by the initial choice of oviposition sites of female *R. conicus*. Choice experiments with different levels of herbivory, including none, could be done

in a pair-wise design or a multiple-choice design (e.g., Simelane, 2006). In addition, no-choice tests could be conducted to determine if plant-mediated effects result in outright rejection (dispersal). However, experiments would need to be carefully designed to avoid restrictions on dispersal resulting from cages. For example, it is possible that the results of Gerber et al. (2007), which involved individual caged plants, would have differed if *Ceutorhynchus alliariae* Brisout had been allowed a choice to leave plants previously infested by *C. scrobicollis* Nerensheimer & Wagner.

7. For temporally isolated agents, assess if recruitment of Agent 2 is altered (positively, negatively, or not) based on prior presence of Agent 1. If the agents are separated in space only, then assess if recruitment of each agent is altered by the simultaneous presence of the other species. If multivoltine agents are being tested, recruitment can be assessed at each generation. Recruitment can refer to colonization, feeding and oviposition by adults, as well as larval development and survival, disease incidence, etc. All life stages should be assessed because indirect effects may target different stages of an herbivore's life cycle (Swope and Parker, 2010), including affecting a non-damaging adult stage that in turn could alter the density of a damaging larval stage (Briese, 1991; Milbrath and Nechols, 2004a).

8. Quantify plant impact and rate as 'greater', 'same', or 'less' when comparing one vs. two agents (see Tables 3 and 4). Several plant parameters may be measured and assessed separately (e.g., Buccellato et al., 2012). The researcher will need to rank the importance of different parameters for control. With respect to the categories of Hatcher (1995) as modified by Turner et al. (2010), greater = synergistic and additive outcomes, same = equivalent outcome of the highest impacting agent acting alone, and less = all other outcomes including equivalent to the lowest impacting agent when acting alone or inhibitory.

4.2. Release decisions (Tables 3 and 4)

Once experimental data on plant impact and indirect agent interactions are obtained for the various treatment combinations, a sequential action model such as what we propose in Tables 3 and 4 could be used to guide release decisions for the two agents. Table 3 was developed for temporally-separated agents (which may or may not be separated in space) and Table 4 is for agents separated in space only. Literature examples from section 3.1 (Table 1) are also noted in Tables 3 and 4 to help illustrate where such studies may fit in our scheme.

We illustrate how release decisions could be made using the musk thistle study. As discussed under section 3.2, because the musk thistle system involves plant-mediated effects that are one-way (*T. horridus* negatively impacts *R. conicus* but not the reverse) and because the two agents are both temporally- and spatially-separated, we refer to Table 3.

At low densities, *T. horridus* (Agent 1) had no measurable impact on musk thistle, and no P-MI between *T. horridus* and *R. conicus* (Agent 2) was discernible. Thus, the only impact on musk thistle was from *R. conicus*. In cases like this, if no plant impact was observed for any tested density of Agent 1 (Table 3, last scenario), we would either discontinue working with Agent 1 or consider running tests at higher densities. The value of testing agents over a range of densities is seen in the very different experimental results we obtained at higher *T. horridus* densities. Here, the relative impact of *T. horridus* alone was less than that of *R. conicus* alone, and a negative P-MI was evident; but despite that, the combined impact of the two agents was greater than that of *R. conicus* alone (Table 3, second to last scenario). Based on these results, either both species or Agent 2 alone could be recommended for release, although additional study might be recommended in light of the competitive interaction observed. This is the only

case in Table 3 where additional assessment may be warranted by including a greater range of Agent 1 densities in a follow-up study. Specifically, in this case Agent 1 (*T. horridus*) would be characterized as a competitively superior species but an inferior agent (similar reasoning applies for two of the recommendations in Table 4). Therefore, higher densities of *T. horridus* (Agent 1) could further interfere with *R. conicus*' (Agent 2) ability to utilize musk thistle, although it is presently unknown if this would lead to an overall increase or decrease in plant impact, depending on gains and losses in impact from *T. horridus* and *R. conicus*, respectively.

To our knowledge no studies have been published comparing the individual and combined impacts of the two weed biological control agents, as well as documenting plant-mediated competition, under these conditions. However, Woodburn (1997) reported that under very high *T. horridus* densities in Australia, twice the levels we observed in Kansas, flowerhead production was delayed by two weeks instead of the one week we observed in Kansas. This result, if transferrable to our situation in Kansas, potentially could exacerbate the negative indirect competitive effect of *T. horridus* on *R. conicus*. Interestingly though, in Woodburn's study the relative impact of *T. horridus* alone was greater than the previously reported impact of *R. conicus* alone (72% versus 40% seed reduction, respectively). Thus, experimental evaluations must include a wide range of densities in order to reveal potential interference between agents, and net impacts on weed biological control.

#### 5. Conclusion

Plant-mediated indirect interactions undoubtedly occur commonly among organisms that attack weeds. However, very little is known about how these interactions change, and what the consequences are for weed biological control, when new assemblages of agents are created via

classical biological control efforts. Part of the problem is that, apart from host range tests and basic biological studies of candidate agents, relatively few additional pre-release experimental evaluations are done. Thus, the release of a new agent may or may not result in a net gain in the impact on a weed population. As we have discussed, plant-mediated effects that reduce the efficacy of a newly-introduced or existing agent can be compensated for by the agent that induces a negative indirect interaction. However, only by conducting experimental evaluations that take into consideration impacts of agents on the weed and on each other, will it be possible to understand the complex interactions involving target weeds and herbivorous candidate agents. If these more comprehensive evaluations are done, better predictions about agent selection for specific programs should result, and may also reveal general patterns that can be extrapolated to future programs.

# Acknowledgements

We thank anonymous reviewers for their comments which improved this manuscript. We also thank Tom Dudley, University of California Santa Barbara, and Peter McEvoy, Oregon State University, for helpful suggestions. Mention of trade names or commercial products in this article is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the U.S. Department of Agriculture. USDA is an equal opportunity provider and employer.

#### References

- Agrawal, A.A., Tuzan, S., Bent, E. (Eds.), 1999. Induced Plant Defenses against Pathogens and Herbivores. American Phytopathological Society Press, St. Paul, MN, USA.
- Balciunas, J.K., Coombs, E., 2004. International code of best practices for biological control of weeds, in: Coombs, E., Clark, J.K., Piper, G.L., Cofrancesco, A.F. (Eds.), Biological Control of Invasive Plants in the United States. Oregon State University Press, Corvallis, OR, pp. 130–136.
- Barratt, B.I.P., Howarth, F.G., Withers, T.M., Kean, J.M., Ridley, G.S., 2010. Progress in risk assessment for classical biological control. Biological Control 52, 245-254.
- Berube, D.E., 1980. Interspecific competition between *Urophora affinis* and *U. quadrifasciata* (Diptera: Tephritidae) for ovipositional sites on diffuse knapweed (*Centaurea diffusa*: Compositae). Zeitschrift für angewandte Entomologie 90, 299-306.
- Blossey, B., Hunt-Joshi, T.R., 2003. Belowground herbivory by insects: influence on plants and aboveground herbivores. Annual Review of Entomology 48, 521–547.
- Briese, D.T., 1991. Current status of *Agrilus hyperici* (Coleoptera: Buprestidae) released in Australia in 1940 for the control of St John's Wort: Lessons for insect introductions. Biocontrol Science and Technology 1, 207-215.
- Briese, D.T., Walker, A., 2008. Choosing the right plants to test: the host-specificity of *Longitarsus* sp. (Coleoptera: Chrysomelidae) a potential biological control agent of *Heliotropium amplexicaule*. Biological Control 44, 271–285.
- Buccellato, L., Byrne, M.J., Witkowski, E.T.F., 2012. Interactions between a stem gall fly and a leaf-spot pathogen in the biological control of *Ageratina adenophora*. Biological Control 61, 222-229.

- Caesar, A.J., 2003. Synergistic interaction of soilborne plant pathogens and rootattacking insects in classical biological control of an exotic rangeland weed. Biological Control 28, 144–153.
- Campanella, D.M., McEvoy, P.B., Mundt, C.C., 2009. Interaction effects of two biological control organisms on resistant and susceptible weed biotypes of *Chondrilla juncea* in western North America. Biological Control 50, 50-59.
- Cartwright, B., Kok, L.T., 1985. Growth responses of musk and plumeless thistles (*Carduus nutans* and *C. acanthoides*) to damage by *Trichosirocalus horridus* (Coleoptera: Curculionidae). Weed Science 33, 57-62.
- Caswell, H. 2001. Matrix Population Models: Construction, Analysis and Interpretation. Sinauer, Sunderland, Massachusetts, USA.
- Crowe, M.L., Bourchier, R.S., 2006. Interspecific interactions between the gall-fly *Urophora affinis* Frfld. (Diptera: Tephritidae) and the weevil *Larinus minutus* Gyll. (Coleoptera: Curculionidae), two biological control agents released against spotted knapweed, *Centaurea stobe* L. ssp. *micranthos*. Biocontrol Science and Technology 16, 417-430.
- Damman, H., 1989. Facilitative interactions between two lepidopteran herbivores of *Asimina*. Oecologia 78, 214-219.
- Damman, H., 1993. Patterns of herbivore interaction among herbivore species, in: Stamp, N.E., Casey, T.M. (Eds.), Caterpillars: Ecological and Evolutionary Constraints on Foraging.

  Chapman and Hall, New York, USA, pp. 132-169.
- Davis, A.S., Landis, D.A., Nuzzo, V., Blossey, B., Gerber, E., Hinz, H.L., 2006. Demographic models inform selection of biocontrol agents for garlic mustard (*Alliaria petiolata*).Ecological Applications 16, 2399-2410.

- Denno, R.F., Kaplan, I., 2007. Plant-mediated interactions in herbivorous insects: mechanisms, symmetry, and challenging the paradigms of competition past, in: Ohgushi, T., Craig, T.P., Price, P.W. (Eds.), Ecological Communities: Plant Mediation in Indirect Interaction Webs. Cambridge University Press, Cambridge, UK, pp. 19-50.
- Denno, R.F., McClure, M.S., Ott, J.R., 1995. Interspecific interactions in phytophagous insects: competition revisited and resurrected. Annual Review of Entomology 40, 297-331.
- Dunn, P.H., 1976. Distribution of *Carduus nutans*, *C. acanthoides*, *C. pycnocephalus*, and *C. crispus*, in the United States. Weed Science 24, 518-524.
- Erb, M., Robert, C.A.M., Hibbard, B.E., Turlings, T.C.J., 2011. Sequence of arrival determines plant-mediated interactions between herbivores. Journal of Ecology 99, 7-15.
- Follet, P.A., Duan, J.J. (Eds.), 2000. Nontarget Effects of Biological Control. Kluwer Academic Publishers, Norwell, MA. 316 pp.
- Gassmann, A., 1996. Classical biological control of weeds with insects: a case for emphasizing agent demography, in: Moran, V.C., Hoffmann, J.H. (Eds.), Proceedings of the IX International Symposium on Biological Control of Weeds. University of Cape Town, Rondebosch, South Africa, pp. 171-175.
- Gerber, E., Hinz, H., Blossey, B., 2007. Interaction of specialist root and shoot herbivores of *Alliaria petiolata* and their impact on plant performance and reproduction. Ecological Entomology 32, 357–365.
- Harris, P., 1981. Chap. 22: Stress as a strategy in the biological control of weeds, in: Papavizas,G.C. (Ed.), Biological Control in Crop Protection. Beltsville Symposia in AgriculturalResearch 5. Allanheld, Osmun & Co., Totowa, NJ, pp. 333-340.

- Harris, P., 1991. Classical biocontrol of weeds: its definition, selection of effective agents, and administrative-political problems. Canadian Entomologist 123, 827-849.
- Hatcher, P.E., 1995. Three-way interactions between plant pathogenic fungi, herbivorous insects and their host plants. Biological Reviews 70, 639–694.
- Hatcher, P.E., Paul, N.D., 2001. Plant pathogen–herbivore interactions and their effects on weeds, in: Jeger, M.J., Spence, N.J. (Eds.), Biotic Interactions in Plant–Pathogen
   Associations. CABI Publishing, Wallingford, UK, pp. 193–225.
- Heard, T.A., 2000. Concepts in insect host-plant selection behavior and their application to host specificity testing, in: Van Driesche, R., Heard, T., McClay, A., Reardon, R. (Eds.),
  Proceedings of Session: Host-Specificity Testing of Exotic Arthropod Biological Control Agents The Biological Basis for Improvement in Safety. X International Symposium on Biological Control of Weeds, 8 July 1999, Bozeman, MT. USDA Forest Service, FHTET-99-1, Morgantown, WV, pp. 1-10.
- Hunt-Joshi, T.R., Blossey, B., 2005. Interactions of root and leaf herbivores on purple loosestrife (*Lythrum salicaria*). Oecologia 142, 554–563.
- Hunt-Joshi, T.R., Blossey, B., Root, R.B., 2004. Root and leaf herbivory on *Lythrum salicaria*: implications for plant performance and communities. Ecological Applications 14, 1574–1589.
- Inbar, M., Eshel, A., Wool, D., 1995. Interspecific competition among phloem-feeding insects mediated by induced host-plant sinks. Ecology 76, 1506-1515.
- James, R.R., McEvoy, P.B., Cox, C.S., 1992. Combining the cinnabar moth (*Tyria jacobaeae*) and the ragwort flea beetle (*Longitarsus jacobaeae*) for control of ragwort (*Senecio jacobaea*): an experimental analysis. Journal of Applied Ecology 29, 589-596.

- Karban, R., Baldwin, I.T., 1997. Induced Responses to Herbivory. University of Chicago Press, Chicago, IL, USA.
- Kluth, S., Kruess, A., Tscharntke, T., 2001. Interactions between the rust fungus *Puccinia punctiformis* and ectophagous and endophagous insects on creeping thistle. Journal of Applied Ecology 38, 548–556.
- Kok, L.T., 2001. Classical biological control of nodding and plumeless thistles. Biological Control 21, 206-213.
- Kruess, A., 2002. Indirect interaction between a fungal plant pathogen and a herbivorous beetle of the weed *Cirsium arvense*. Oecologia 130, 563–569.
- Larson, K.C., Whitham, T.G., 1991. Manipulation of food resources by a gall-inducing aphid: the physiology of sink-source interactions. Oecologia 88, 15-21.
- Lonsdale, W.M., Briese, D.T., Cullen, J.M., 2001. Risk analysis and weed biological control, in: Wajnberg, E., Scott, J.K., Quimby, P.C. (Eds.), Evaluating Indirect Ecological Effects of Biological Control. CABI Publishing, CAB International, Wallingford, UK, pp.185-210.
- Louda, S.M., Pemberton, R.W., Johnson, M.T., Follett, P.A., 2003. Nontarget effects the Achilles' heel of biological control? Retrospective analyses to reduce risk associated with biocontrol introductions. Annual Review of Entomology 48, 365-396.
- Louda, S.M., Rand, T.A., Russell, F.L., Arnett, A.E., 2005. Assessment of ecological risks in weed biocontrol: input from retrospective ecological analyses. Biological Control 35, 253-264.
- Marquis, R.J., Lill, J.T., 2007. Effects of arthropods as physical ecosystem engineers on plant-based trophic interaction webs, in: Ohgushi, T., Craig, T.P., Price, P.W. (Eds.), Ecological

- Communities: Plant Mediation in Indirect Interaction Webs. Cambridge University Press, Cambridge, UK, pp. 246-274.
- Masters, G.J., Brown, V.K., 1997. Host-plant mediated interactions between spatially separated herbivores: effects on community structure, in: Gange, A.C., Brown, V.K. (Eds.),
  Multitrophic Interactions in Terrestrial Systems. Blackwell Science, Oxford, UK, pp. 217-237.
- Masters, G.J., Brown, V.K., Gange, A.C., 1993. Plant mediated interactions between above- and below-ground insect herbivores. Oikos 66, 148-151.
- McCarty, M.K., Lamp, W.O., 1982. Effect of a weevil, *Rhinocyllus conicus*, on musk thistle (*Carduus thoermeri*) seed production. Weed Science 30, 136-140.
- McCarty, M.K., Scifres, C.J., 1969. Life cycle studies with musk thistle. Research Bulletin 230, University of Nebraska, Agricultural Experiment Station, Lincoln, NE.
- McClay, A.S., Balciunas, J.K., 2005. The role of pre-release efficacy assessment in selecting classical biological control agents for weeds—applying the Anna Karenina principle.

  Biological Control 35, 197–207.
- Milbrath, L.R., Nechols, J.R., 2004a. Indirect effect of early-season infestations of *Trichosirocalus horridus* on *Rhinocyllus conicus* (Coleoptera: Curculionidae). Biological Control 30, 95-109.
- Milbrath, L.R., Nechols, J.R., 2004b. Individual and combined effects of *Trichosirocalus* horridus and *Rhinocyllus conicus* (Coleoptera: Curculionidae) on musk thistle. Biological Control 30, 418-429.
- Mills, N.J., Kean, J.M., 2010. Behavioral studies, molecular approaches, and modeling: methodological contributions to biological control success. Biological Control 52, 255-262.

- Moran, P.J., 2005. Leaf scarring by the weevil *Neochetina eichhorniae* and *N. bruchi* enhances infection by the fungus *Cercospora piaropi* on waterhyacinth, *Eichhornia crassipes*.

  BioControl 50, 511–524.
- Morin, L., Reid, A.M., Sims-Chilton, N.M., Buckley, Y.M., Dhileepan, K., Hastwell, G.T., Nordblom, T.L., Raghu, S., 2009. Review of approaches to evaluate the effectiveness of weed biological control agents. Biological Control 51, 1–15.
- Müller-Schärer, H., Schaffner, U., 2008. Classical biological control: exploiting enemy escape to manage plant invasions. Biological Invasions 10, 859–874.
- O'Brien, J.M., Kyser, G.B., Woods, D.M., DiTomaso, J.M. 2010. The effect of *Puccinia jaceae* var. *solstitialis* on the yellow starthistle biological control insects *Eustenopus villosus* and *Chaetorellia succinea*. Biological Control 52, 182–187
- Ohgushi, T., 2005. Indirect interaction webs: herbivore-induced effects through trait change in plants. Annual Review of Ecology, Evolution, and Systematics 36, 81–105.
- Ohgushi, T., Craig, T.P., Price, P.W. (Eds.), 2007. Ecological Communities: Plant Mediation in Indirect Interaction Webs. Cambridge University Press, Cambridge, UK.
- Pemberton, R.W., 2000. Predictable risk to native plants in weed biological control. Oecologia 125, 489-494.
- Raghu, S., Wilson, J.R., Dhileepan, K., 2006. Refining the process of agent selection through understanding plant demography and plant response to herbivory. Australian Journal of Entomology 45: 308–316.
- Raghu, S., Brown, B., Purcell, M.F., 2012. Searching for the signal of competition in plant-mediated interactions among coexisting gall insects on broad-leaved paperbark. Arthropod-Plant Interactions 6, 83–91.

- Rayamajhi, M.B., Pratt, P.D., Center, T.D., Van, T.K., 2010. Insects and a pathogen suppress *Melaleuca quinquenervia* cut-stump regrowth in Florida. Biological Control 53, 1–8.
- Sabelis, M.W., Takabayashi, J., Janssen, A., Kant, M.R., Van Wijk, M., Sznajder, B., Aratchige,
  N.S., Lesna, I., Belliure, B., Schuurink, R.C., 2007. Ecology meets plant physiology:
  herbivore-induced plant responses and their indirect effects on arthropod communities, in:
  Ohgushi, T., Craig, T.P., Price, P.W. (Eds.), Ecological Communities: Plant Mediation in
  Indirect Interaction Webs. Cambridge University Press, Cambridge, UK, pp. 188-217.
- Saner, M.A., Jeanneret, P., Müller-Schärer, H., 1994. Interaction among two biological control agents and the developmental stage of their target weed, Dalmatian toadflax, *Linaria dalmatica* (L.) Mill. (Scrophulariaceae). Biocontrol Science and Technology 4, 215-222.
- Sheppard, A.W., van Klinken, R.D., Heard, T.A., 2005. Scientific advances in the analysis of direct risks of weed biological control agents to nontarget plants. Biological Control 35, 215-226.
- Shorthouse, J.D., Lalonde, R.G., 1984. Structural damage by *Rhinocyllus conicus* (Coleoptera: Curculionidae) within the flowerheads of nodding thistle. Canadian Entomologist 116, 1335-1343.
- Simelane, D.O., 2006. Effect of herbivory by *Teleonemia scrupulosa* on the performance of *Longitarsus bethae* on their shared host, *Lantana camara*. Biological Control 39, 385–391.
- Smith, L., Mayer, M., 2005. Field cage assessment of interference among insects attacking seed heads of spotted and diffuse knapweed. Biocontrol Science and Technology 15, 427-442.
- Strauss, S.Y., 1991. Direct, indirect, and cumulative effects of three native herbivores on a shared host plant. Ecology 72, 543-558.

- Stuckey, R.L., Forsyth, J.L., 1971. Distribution of naturalized *Carduus nutans* (Compositae) mapped in relation to geology in northwestern Ohio. Ohio Journal of Science 71, 1-15.
- Surles, W.W., Kok, L.T., 1976. Pilot studies on augmentation of *Rhinocyllus conicus* (Col.: Curculionidae) for *Carduus* thistle control. Environmental Entomology 5, 901-904.
- Swope, S.M., Parker, I.M., 2010. Trait-mediated interactions and lifetime fitness of the invasive plant *Centaurea solstitialis*. Ecology 91, 2284–2293.
- Swope, S.M., Stein, I.R., 2012. Soil type mediates indirect interactions between *Centaurea* solstitialis and its biocontrol agents. Biological Invasions 14, 1697-1710.
- Treier, U.A., Broennimann, O., Normand, S., Guisan, A., Schaffner, U., Steinger, T., Müller-Schärer, H., 2009. Shift in cytotype frequency and niche space in the invasive plant *Centaurea maculosa*. Ecology 90, 1366-1377.
- Turner, P.J., Morin, L., Williams, D.G., Kriticos, D.J., 2010. Interactions between a leafhopper and rust fungus on the invasive plant *Asparagus asparagoides* in Australia: a case study of two agents being better than one for biological control. Biological Control 54, 322–330.
- Van Hezewijk, B.H., Bourchier, R.S., 2012. Impact of *Cyphocleonus achates* on diffuse knapweed and its interaction with *Larinus minutus*. Biological Control 62, 113-119.
- Wajnberg, E., Scott, J.K., Quimby, P.C. (Eds.), 2001. Evaluating Indirect Ecological Effects of Biological Control. CABI Publishing, CAB International, Wallingford, UK.
- Wang, Y., Huang, W., Siemann, E., Zou, J., Wheeler, G.S., Carrillo, J., Ding, J., 2011. Lower resistance and higher tolerance of invasive host plants: biocontrol agents reach high densities but exert weak control. Ecological Applications 21, 729–738.
- Wheeler, G.S., Massey, M., Southwell, I.A., 2003. Dietary influences on terpenoids sequestered by the biological control agent *Oxyops vitiosa*: effect of plant volatiles from different

- Melaleuca quinquenervia chemotypes and laboratory host species. Journal of Chemical Ecology 29, 189-208.
- Withers, T.M., Browne, L.B., Stanley, J., 2000. How time-dependent processes can affect the outcome of assays, in: Van Driesche, R., Heard, T., McClay, A., Reardon, R. (Eds.),
  Proceedings of Session: Host-Specificity Testing of Exotic Arthropod Biological Control
  Agents The Biological Basis for Improvement in Safety. X International Symposium on
  Biological Control of Weeds, 8 July 1999, Bozeman, MT. USDA Forest Service, FHTET-991, Morgantown, WV, pp. 27-41.
- Wold, E.N., Marquis, R.J., 1997. Induced defense in white oak: effects on herbivores and consequences for the plant. Ecology 78, 1356-1369.
- Woodburn, T.L., 1997. Establishment in Australia of *Trichosirocalus horridus*, a biological control agent for *Carduus nutans*, and preliminary assessment of its impact on plant growth and reproductive potential. Biocontrol Science and Technology 7, 645–656.
- Wootton, J.T., 1994. The nature and consequences of indirect effects in ecological communities.

  Annual Review of Ecology and Systematics 25, 443-466.
- Zalucki, M.P., van Klinken, R.D., 2006. Predicting population dynamics of weed biological control agents: science or gazing into crystal balls? Australian Journal of Entomology 45, 331-344.

Table 1. Examples of plant-mediated interactions in weed biological control systems (with or without plant impact studies) for agents separated in space and/or time.

Target weed	Herbivore/pathogen	Plant-mediated interaction	Combined agent impact to plant	Reference
			compared to individual impact	(type of study)
Separated in space (two-way interaction)				
Pathogen-Arthropod				
Crofton weed	leaf-spot fungus	Fly on fungus	Equal to fly alone (stem height)	Buccellato et al., 2012
Ageratina adenophora	Passalora ageratinae Crous &	Negative effect (reduced %	Equal to fungus alone (side	(greenhouse,
(Sprengel) King &	A.R. Wood (Mycosphaerellales:	infected leaves on double-	shoot production)	retrospective)
Robinson (Asteraceae)	Mycosphaerellaceae)	galled stems)	Additive (% live leaves)	
	stem gall fly	Fungus on fly	Complementary overall	
	Procecidochares utilis Stone	No effect (gall size, adult fly		
	(Diptera: Tephritidae)	emergence)		
Rush skeletonweed	leaf rust fungus	Fungus on mite	Equal to fungus alone (shoot	Campanella et al., 2009
Chondrilla juncea L.	Puccinia chondrillina Bubak and	Negative effect (reduced shoot	biomass, rosette diameter, rate	(common garden,
(Asteraceae)	Sydow (Uredinales: Pucciniaceae)	growth and gall biomass),	of senescence)	retrospective)
		Slight positive effect (possible	Equal to mite alone (fecundity)	
	leaf gall mite	increased susceptibility to	Additive (shoot length)	

	Eriophyes chondrillae (Canestrini)	galling in one year)		
	(Acari: Eriophyidae)		Complementary overall	
		Mite on fungus not studied		
		Fungus applied first but eventual		
		temporal overlap; results for rust-		
		susceptible plants		
Insect-Insect				
Lantana	leaf-sucking lace bug	Lace bug on flea beetle	Not assessed	Simelane, 2006
Lantana camara L.	Teleonemia scrupulosa Stäl	No effect (adult beetle feeding		(greenhouse,
(Verbenaceae)	(Heteroptera: Tingidae)	and oviposition, preimaginal		retrospective)
		development, adult size),		
	root-feeding flea beetle	Negative effect (reduced larval		
	Longitarsus bethae Savini &	survival),		
	Escalona (Coleoptera:	Also, direct (?) negative effect		
	Chrysomelidae)	of nymphs on adult beetle		
		colonization and oviposition		
		Beetle on lace bug <b>not studied</b>		
Purple loosestrife	leaf-feeding beetle	Leaf beetle on weevil	Equal to leaf beetle alone (shoot	Hunt-Joshi and

Lythrum salicaria L.	Galerucella calmariensis L.	No effect (weevil oviposition,	growth and height, flowering,	Blossey, 2005
(Lythraceae)	(Coleoptera: Chrysomelidae)	larval mass, % plants infested, #	most plant biomass)	(outdoor pot and
		larvae),	Equal to weevil alone (biomass	multi-year field
	rootstock -boring weevil	Negative effect (reduced larval	of dead stems)	cage, retrospective)
	Hylobius transversovittatus Goeze	survival with reduced resources		
	(Coleoptera: Curculionidae)	[pot only])	Complementary? (perhaps if	Hunt-Joshi et al., 2004
			densities of weevil are higher)	(multi-year field
		Weevil on leaf beetle		cage, retrospective)
		No effect (larval mass or		
		survival, insect abundance,		
		oviposition; but very low		
		densities of weevil)		
Broad-leaved paperbark	Early-stage vegetative tips	Early gallers on late galler	Not assessed	Raghu et al., 2012
Melaleuca quinqenervia	puff-ball gall	No relationship (resource		(native range field
(Cav.) S.T. Blake	Sphaerococcus ferrugineus	availability and previous		surveys,
(Myrtaceae)	Froggatt (Homoptera:	abundance of early gallers),		prospective)
	Eriococcidae)	Positive relationship (late galler		
		abundance and previous		
	bud gall	abundance of puff-ball galler		

	Fergusonina turneri Taylor	only)		
	(Diptera: Fergusoninidae)			
		Late galler on early gallers		
	Late-stage vegetative tips	Negative relationship (resource		
	blister gall	availability and previous		
	Lophodiplosis indentata Gagne'	abundance of late galler),		
	(Diptera: Cecidomyiidae)	Positive relationship (only puff-		
		ball galler abundance and		
		previous abundance of late		
		galler)		
Dalmation toadflax	root-boring moth	Moth on weevil	Not fully reported but stem-	Saner et al., 1994
Linaria dalmatica (L.)	Eteobalea intermediella Riedl	No effect (weevil oviposition,	mining weevil reduced stem	(native range,
Mill.	(Lepidoptera: Cosmopterigidae)	preimaginal survival; but only	biomass	greenhouse,
(Scrophulariaceae)		one of two generations of moth		retrospective)
	stem-mining weevil	included in this study)		
	Mecinus janthinus Germar			
	(Coleoptera: Curculionidae)	Weevil on moth		

No effect (larval survival)

## Separated in time but not space (one-way interaction)

## No examples found

Separated in space and time (one-way interaction of 1<sup>st</sup> agent on 2<sup>nd</sup> agent)

Pathogen-Arthropod				
Yellow starthistle	1 <sup>st</sup> to colonize: leaf rust fungus	No effect (number of seed heads	Not assessed	O'Brien et al., 2010
Centaurea solstitialis L.	Puccinia jaceae Otth var.	attacked by insects)		(field, retrospective)
(Asteraceae)	solstitialis (Uredinales:			
	Pucciniaceae)			
	2 <sup>nd</sup> to colonize: seed-head insects  Chaetorellia succinea (Costa)  (Diptera: Tephritidae)  Eustenopus villosus (Boheman)  (Coleoptera: Curculionidae)			
	1 <sup>st</sup> : leaf rust fungus	No effect (adult feeding or	Equal to seed-head weevil alone	Swope and Parker,
	Puccinia jaceae	oviposition/plant, larval	(seed production, partial	2010
		survival),	assessment only)	(field, retrospective)
	2 <sup>nd</sup> : seed-head weevil	Negative/positive effect (lesser		
	Eustenopus villosus	proportion of seeds consumed		

## by larvae)

	1 <sup>st</sup> : leaf rust fungus	Non-serpentine soils	Non-serpentine soils	Swope and Stein, 2012
	Puccinia jaceae	No effect (larval survival),	Equal to seed-head insects alone	(field, retrospective)
		Negative/positive effect (lesser	(seed production, partial	
	2 <sup>nd</sup> : seed-head insects	proportion of seeds consumed	assessment only)	
	Chaetorellia australis Hering	by larvae)		
	(Diptera: Tephritidae)		Serpentine soils	
	Chaetorellia succinea	Serpentine soils	Greater (seed production, partial	
	Eustenopus villosus	Positive effect (larval survival),	assessment only)	
	Urophora sirunaseva (Hering)	Negative/positive effect (lesser		
	(Diptera: Tephritidae)	proportion of seeds consumed		
		by larvae)		
Insect-Insect				
Garlic mustard	1 <sup>st</sup> : root-crown weevil	Stem level	Equal to root-crown weevil	Gerber et al., 2007
Alliaria petiolata (M.	Ceutorhynchus scrobicollis	Negative (reduced attack rate	alone (plant survival, biomass,	(native range,
Bieb.) Cavara &	Nerensheimer & Wagner	due to reduced stem height and	seed production)	common garden,
Grande (Brassicaceae)	(Coleoptera: Curculionidae)	diameter)	Additive (stem height)	prospective)
	2 <sup>nd</sup> : shoot-mining weevil	Plant level (increased stem #		
	Ceutorhynchus alliariae Brisout	canceled out reduced stem		

	(Coleoptera: Curculionidae)	height)  No effect (attack rate, larval survival despite increased N-content, new adult recruitment)		
Marelediale	1 SL	,	Forelts and hard warn't alone	Milhorth and Nachala
Musk thistle	1 <sup>st</sup> : rosette (crown) weevil	Neutral to negative effect	Equal to seed-head weevil alone	Milbrath and Nechols,
Carduus nutans L.	Trichosirocalus horridus (Panzer)	(density-dependent, see Table	to greater (seed production)	2004a
(Asteraceae)	(Coleoptera: Curculionidae)	2)	(see Table 2)	(greenhouse and
				common garden,
	2 <sup>nd</sup> : seed-head weevil			retrospective)
	Rhinocyllus conicus Froelich			Milbrath and Nechols,
	(Coleoptera: Curculionidae)			2004b
				(common garden,
				retrospective)
Diffuse knapweed	1 <sup>st</sup> : root-boring weevil	Negative effect (proportion of	Not assessed	Van Hezewijk and
Centaurea diffusa	Cyphocleonus achates (Fahr.)	heads attacked)		Bourchier, 2012
Lamarck (Asteraceae)	(Coleoptera: Curculionidae)			(field, retrospective)
	2 <sup>nd</sup> : seed-head weevil			
	Larinus minutus Gyll. (Coleoptera:			

Table 2. Plant-mediated interaction of *Trichosirocallus horridus* on *Rhinocyllus conicus*, two biological control agents of musk thistle (from Milbrath and Nechols, 2004a,b).

	Outcome of interaction due to prior infestation by <i>T. horridus</i>			
Mechanism of plant-mediated	Low T. horridus larval	High T. horridus larval		
interaction	density	density		
Phenology: synchrony	No	Yes, 1 week delay		
between R. conicus adults and				
flower heads altered?				
Behavior: host plant selection	No	Yes, adults/plant reduced 57%		
(adult presence, oviposition)		and eggs/plant reduced 64%		
by adult <i>R conicus</i> altered?				
Resource limitation: change in	No	Yes, 34% less receptacle dry		
quantity of food for R.		mass/plant		
conicus larvae?				
Physiology: change in plant	Not tested	Yes, but only observed when		
quality for R. conicus larvae?		R. conicus larval densities low		
Combined effect on	No effect	63% reduction		
recruitment of new R.				
conicus adults				
	Percentage reduction in musk	thistle seed production		
	(compared to control)			
T. horridus alone	0%	Not tested but less than effect		
		of R. conicus		

R. conicus alone	45%	Assumed ~45%
Combined	45%	Estimated 59%

Table 3. Recommendations for agent release based on different outcomes of plant-mediated impact studies for agents involved in negative (competitive), neutral, or positive (facilitative) one-way interactions, where the agents are separated in time, or in space and time. Agent 1 attacks plant before agent 2 within the life cycle of the plant (annuals and biennials) or within a growing season (perennials); between-season effects involving perennial plants are not considered.

	Experimental out	tcome		
Agent 1 impact on plant	IF plant-	AND combined agent impact	THEN recommend-	Literature example
relative to Agent 2	mediated	to plant compared to highest	ation for agent	
	interaction <sup>a</sup>	individual impact	release is <sup>b</sup>	
Agent 1 greater than	+ or 0 or -	Greater (than Agent 1 alone)	Both species <sup>c</sup>	Neutral: Gerber et al., 2007
agent 2 (and agent 2 not				
zero)				
	-	Same as Agent 1	Agent 1	
		Less	n/a	
Agent 1 has impact; agent	+ or 0 or -	Same as Agent 1	Agent 1	
2 has NO impact				

Agent 1 and 2 same	+ or 0 or -	Greater	Both <sup>c</sup>	
	-	Same as Agent 1 or 2	Agent 1 or Agent 2 <sup>c</sup>	
		Less	n/a	
Agent 2 greater than	+ or 0 or -	Greater (than Agent 2 alone)	Positive or Neutral	
agent 1 (and agent 1 not			Both <sup>c</sup>	
zero)			Negative	Milbrath and Nechols, 2004a,
			Both or Agent 2? <sup>d</sup>	b
	-	Same as Agent 2	Agent 2	
	-	Less	Agent 2	
Agent 2 has impact;	+ or 0 or -	Same as Agent 2 or Less	Agent 2	Milbrath and Nechols, 2004a,
agent 1 has NO impact				b

a + = positive interaction, 0 = neutral interaction, - = negative interaction

<sup>&</sup>lt;sup>b</sup> Assuming appropriate host-specificity. If no specific plant-mediated interaction noted (positive, neutral, negative), then recommendation applies to all scenarios. n/a = specific combined impact outcome not possible by definition of a one-way interaction.

<sup>c</sup> For recommendations of Both, may choose to import only Agent 1 at first. For recommendations of Both or a choice between agents, may modify choice using plant demography model-derived recommendations.

<sup>&</sup>lt;sup>d</sup> Further risk assessment needed.

Table 4. Recommendations for agent release based on different outcomes of plant-mediated impact studies for agents involved in negative (competitive), neutral, or positive (facilitative) two-way interactions. Agents attack plant simultaneously within a growing season but are spatially separated; between-season effects are not considered.

	Experin	nental outcome			
Agent 1 impact on plant	IF plant-medi	ated	AND combined agent impact to	THEN recommend-	Literature
relative to agent 2	interaction <sup>a</sup>		plant compared to highest	ation for agent release	example
	Agent 1 on 2	Agent 2 on 1	individual impact	is <sup>b</sup>	
Agent 1 greater than	+ or 0 or -	+ or 0 or -	Greater (than Agent 1 alone)	Agent 2 on 1= negative	
agent 2 (and agent 2 not				Both or Agent 1? <sup>d</sup>	
zero)				All other cases	
				Both species <sup>c</sup>	
	+ or 0 or -	-	Same as Agent 1 or Less	Agent 1	
	-	+ or 0	Same as Agent 1 (Less n/a)	Agent 1	Buccellatto et
					al., 2012
Agent 1 has impact; agent	+ or 0 or -	+ or 0 or -	Same as Agent 1 or Less	Agent 1	

## 2 has NO impact

Agent 1 and 2 same	+ or 0 or -	+ or 0 or -	Greater	Both <sup>c</sup>	Buccellatto et
					al., 2012
	-	-	Same as Agent 1 or 2, or Less	Agent 1 or Agent 2 c	
	-	+ or 0	Same as Agent 1 or 2 (Less n/a)	Agent 1 or Agent 2 c	
	+ or 0	-	Same as Agent 1 or 2 (Less n/a)	Agent 1 or Agent 2 c	
Agent 2 greater than	+ or 0 or -	+ or 0 or -	Greater (than Agent 2 alone)	Agent 1 on 2= negative	
agent 1 (and agent 1 not				Both or Agent 2? <sup>d</sup>	
zero)				All other cases	
				Both species <sup>c</sup>	
	-	+ or 0 or -	Same as Agent 2 or Less	Agent 2	Buccellatto et
					al., 2012
	+ or 0	-	Same as Agent 2 (Less n/a)	Agent 2	

Agent 2 has impact; + or 0 or - + or 0 or - Same as Agent 2 or Less Agent 2 agent 1 has NO impact

 $<sup>^{</sup>a}$  + = positive interaction, 0 = neutral interaction, - = negative interaction

<sup>&</sup>lt;sup>b</sup> Assuming appropriate host-specificity. If no specific plant-mediated interaction noted (positive, neutral, negative), then recommendation applies to all scenarios. n/a = specific combined impact outcome not considered possible.

<sup>&</sup>lt;sup>c</sup> For recommendations of Both, may choose to import only one agent at first. For recommendations of Both or a choice between agents, may modify choice using plant demography model-derived recommendations.

<sup>&</sup>lt;sup>d</sup> Further risk assessment needed.