

1 **Effect of Various Dietary Fats on Fatty Acid Profile in Duck Liver: Efficient**

2 **Conversion of Short-chain to Long-chain Omega-3 Fatty Acids**

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12 **Abstract**

13 Omega-3 fatty acids, especially long-chain omega-3 fatty acids, have been associated
14 with potential health benefits for chronic disease prevention. Our previous studies found
15 that dietary omega-3 fatty acids could accumulate in the meat and eggs in a duck model.
16 This study was to reveal the effects of various dietary fats on fatty acid profile and
17 conversion of omega-3 fatty acids in duck liver. Female *Shan Partridge Ducks* were
18 randomly assigned to five dietary treatments, each consisting of 6 replicates of 30 birds.
19 The experimental diets substituted the basal diet by 2% of flaxseed oil, rapeseed oil, beef
20 tallow, or fish oil, respectively. In addition, a dose response study was further conducted
21 for flaxseed and fish oil diets at 0.5%, 1%, and 2%, respectively. At the end of the 5-week
22 treatment, fatty acids were extracted from the liver samples and analyzed by GC-FID. As
23 expected, the total omega-3 fatty acids and the ratio of total omega-3/omega-6
24 significantly increased in both flaxseed and fish oil groups when compared with the
25 control diet. No significant change of total saturated fatty acids or omega-3 fatty acids
26 was found in both rapeseed and beef tallow groups. The dose-response study further
27 indicated that 59-81% of the short-chain omega-3 ALA in flaxseed oil-fed group was
28 efficiently converted to long-chain DHA in the duck liver, whereas 1% of dietary flaxseed
29 oil could produce an equivalent level of DHA as 0.5% of dietary fish oil. The more
30 omega-3 fatty acids, the less omega-6 fatty acids in the duck liver. Taken together, this
31 study showed the fatty acid profiling in the duck liver after various dietary fat
32 consumption, provided insight into a dose response change of omega-3 fatty acids,
33 indicated an efficient conversion of short- to long-chain omega-3 fatty acid, and
34 suggested alternative long-chain omega-3 fatty acid-enriched duck products for human

health benefits.

Keywords: omega-3 fatty acid, duck, liver, dietary fat, health benefits

Introduction

Omega-3 polyunsaturated fatty acids (ω -3 PUFAs), in addition as essential nutrients, have been associated with potential health benefits in chronic disease prevention. There are two types of ω -3 PUFAs, known by short-chain ω -3 PUFAs like ALA (alpha-linolenic acids, C18:3n-3) or long-chain ω -3 PUFAs such as EPA (eicosapentaenoic acid, C20:5n-3) and DHA (docosahexaenoic acid, C22:6n-3). Short-chain ω -3 PUFAs are presented in plant oil such as flaxseed and soybean oil, while long-chain ω -3 PUFAs are usually found in marine products such as fish oil. Although short-chain ω -3 PUFAs are more common and less expensive, the potential health benefits of ω -3 PUFAs have been related to long-chain ω -3 PUFAs only.

Compelling data from epidemiological and interventional studies have demonstrated an inverse correlation between long-chain ω -3 PUFAs and risk of some chronic diseases, including cardiovascular diseases,^{1,2} myocardial infarction,^{3,4} psoriasis,⁵ mental illnesses,⁶ cancer,^{7,8} and bronchial asthma.⁹ Although it is not conclusive, some clinical trials also found long-chain ω -3 PUFAs contributed to a lower risk of cancers, such as colon,^{10,11} breast,^{12,13} and prostate cancers.¹⁴ Therefore, the 2015 Dietary Guidelines for Americans recommends the consumption of 8 oz. of seafood per week to provide an average of 250 mg/day of long chain omega-3 Fatty acids for health benefits.¹⁵ Moreover, The American Heart Association's Strategic Impact Goal Through 2020 and Beyond recommends at least two servings with 3.5-oz. fish every week to increase EPA and DHA intakes¹⁶, while

an adequate intake of ω -6 linoleic acid as 17 g/d for men and 12 g/d for women at the age of 19 to 50 years¹⁷.

Humans can convert short-chain to long-chain omega-3 fatty acids, but the conversion efficiency is limited, usually below 5% in adults¹⁸ or even less than 1% in infants and aging people.¹⁹ When compared with humans, however, waterfowls such as geese have been reported to convert short-chain to long-chain omega-3 fatty acids more efficiently by a series of desaturase and elongase in the liver²⁰ and subsequently excreted into blood circulation to other tissues.²¹ The diverse conversion rates between human and waterfowl have driven scientists to consider that waterfowls may provide an alternate and sustainable source of long-chain ω -3 PUFAs from plant-derived short-chain ALA.²²⁻²⁴

Ducks are aquatic birds which have a high rate of lipogenesis to meet their energetic requirements during ancient migratory flight.²⁵⁻²⁶ It has been reported, for instance, the percent body fat could be reached as high as 37-42% in Peking ducks and 20-30% in Muscovy ducks.²⁷ Although duck products are popular with its unique flavor and juicy texture, the high fat content has raised health concerns. While certain species of lean ducks, such as the *Shan Partridge* contains 7.5% of body fat only have been developed, modification of the fat composition in favor of higher ω -3 PUFAs in replace of ω -6 PUFAs and/or saturated fatty acids may provide promising healthy benefits. It has been noted that supplemented diets with vegetable oil and fish oil effectively enhanced ω -3 PUFAs in the products of pork,²⁸ broilers^{29,30} and broiler eggs.^{31,32} Dietary ALA was also reported to promote EPA and DHA contents in chicken liver.³³ Furthermore, Chen and Hsu reported an increased trend of EPA, DHA, and total ω -3 PUFAs in duck egg yolks by feeding cod liver oil diet.³⁴ In addition to storage and transportation of lipids, liver of

birds has a very high capacity of lipogenesis³⁵. Fatty acids can be synthesis or converted in the liver, and then transported to other tissues such as adipose, cardiac and skeletal muscle. The conversion of fatty acids in the liver and incorporation of them into various tissues are well-established in response to the observed change of fatty acid composition in the relative tissues³⁶. Therefore, fatty acid conversion in the liver may provide impact not only on a varied fatty acid composition but also on the meat quality including flavor and muscle color³⁷. From an aspect of nutrition value, efficient conversion of short- to long-chain ω -3 PUFAs in the liver may boost the levels of long chain ω -3 PUFAs and thus improve the meat quality. Our previous study found that fish oil and sunflower oil could significantly enhance the levels of EPA and DHA in the leg and chest muscles as well as eggs of *Shan Partridge* duck.³⁸ However, the effect of various dietary fats on fatty acid profile and the conversion efficacy of ω -3 PUFAs in duck liver, to our knowledge, has yet to be well studied.

The aim of this study was to assess the modification of fatty acid profiles in the liver of *Shan Partridge* duck after feeding various dietary fats, including ALA-enriched flaxseed oil, ω -6 PUFA-enriched rapeseed oil, saturated fatty acid-enriched beef tallow, and EPA/DHA-enriched fish oil. The conversion efficacy of short-chain to long-chain ω -3 PUFAs was further investigated by a dose-response study for flaxseed and fish oil treatment, respectively.

Materials and methods

Animals

Female *Shan Partridge Ducks* of the same genetic background and of comparable body

weight at the age of 370 days were housed in the same room with incandescent lighting on 15:9 h light-dark cycle. Feed and water were provided for ad libitum consumption.

Experiment design

Shan Partridge Ducks were randomly assigned into 5 dietary treatment groups including a control group (each group with 6 replicates of 30 birds). The experimental diets substituted the basal diet by 2% of flaxseed oil, rapeseed oil, beef tallow, or fish oil, respectively. Control group was feed with the basal diet. In addition, a dose response study was further conducted for flaxseed and fish oil diets only. Total 7 experimental groups fed various substituted basal diet by 0.5%, 1%, and 2% flaxseed or fish oil respectively, and the control group fed with the basal diet. Each group had 6 replicates. The ingredients and calculated nutrient level of the basal control diet was formulated to meet the nutrient requirements of the National Research Council (Table 1). The measured fatty acid values of the experimental diets in the present study is shown in Table 2. Diets were balanced to similar levels of protein, fat, total energy, and fiber. At the end of the 5-week dietary treatment, ducks were sacrificed and fresh duck livers were stored at -20°C for further lipid extraction.

Lipid extraction

One gram of the liver sample was grinded and mixed with 2 mL of chloroform/methanol (1:2, v/v in 0.001% Butylated hydroxytoluene), 1 mL of chloroform, and 1 mL of water. The mixture was then centrifuged at 1,000 rpm for 15 min. The lower layer was then collected. The above procedure was repeated twice. All the three lower layers were combined together and evaporated under a stream of N₂ gas. One mL of chloroform was

124 added to the dried tube before stored at -80°C until further lipid analysis.

125 Fatty acid analysis

126 Fatty acid methyl esters were synthesized according to the protocols of the Kansas
127 Lipidomics Research Center. Briefly, each lipid extracted sample was transferred to
128 Teflon-lined screw cap tube. An internal standard, pentadecanoic acid (C15:0), was added
129 to each sample. Derivatization was performed with 1mL of 3 M methanolic hydrochloric
130 acid at 78°C for 30 min. Then 2 mL of water and 2 mL of hexan:chloroform (4:1, v/v)
131 were added to each tube. The upper phase was collected after vortex and centrifuge. After
132 the above procedure was repeated twice, three upper phase were combined and dried
133 under nitrogen gas, re-dissolved in 200 µL of hexane and transferred into a GC vial with
134 insert.

135 Fatty acid methyl esters were analyzed using an Agilent 6890N gas chromatography
136 equipped with a programmed temperature vaporization injector, an Agilent 7683
137 autosampler, and Agilent flame ionization detector (Santa Clara, CA). The GC was fitted
138 with a HP-88 capillary column (100m × 0.25mm × 0.2µm, Agilent, Santa Clara, CA). The
139 injector temperature was operated at 275°C with an injection volume of 1 µL. The
140 detector temperature was set at 260°C. Helium was used as the carrier gas at a flow rate
141 of 1.6 mL/min. The flow rate of air and hydrogen were 400 mL/min and 30 mL/min,
142 respectively. The oven temperature ramp was programmed from an initial value of
143 150 °C for 1 min to 175 °C at 10 °C/min for 10 min, and then to 210°C at 5 °C/min for 5
144 min hold, finally to 230 °C at the same speed for 11 min. The total run time per sample is
145 40.5 min and the sampling rate of the FID was 20 Hz. Fatty acid peaks were identified by

comparison of the relative retention times with the Supelco[®]37 component fatty acid methyl ester mix standards. The content of each fatty acid was calculated based upon the area of each identified peak.

Statistical analysis

Data are expressed as mean \pm SD. All the data were analyzed by two-way analysis of variance (ANOVA) and followed by pairwise comparison with Tukey adjustment using SAS 9.2 (SAS Institute Inc., Cary, NC, USA). A value of $P < 0.05$ was considered to be statistically significant.

Results

Fatty acid profile in duck liver

A representative gas chromatography selected from each treatment group was showed in Figure 1. Total 23 fatty acids including the internal standard (peak 2, pentadecanoic acid, 15:0) were identified and analyzed in the duck liver samples, including saturated fatty acids (SFA 14:0, 15:0, 16:0, 17:0, 18:0, 22:0, and 24:0), monounsaturated fatty acids (MUFA 16:1n-10, 16:1n-7, 18:1n-9, 18:1n-7, and 20:1), ω -3 PUFAs (18:3, 18:4, 20:5, 22:5, and 22:6), and ω -6 PUFAs (18:2, 20:2, 20:3, 20:4, 22:4, and 22:5). As shown in Table 3, the contents of fatty acids in duck liver fed various dietary fats for 5 weeks varied. No significant difference of total SFA, total MUFA, total ω -6 PUFAs, or individual ω -6 PUFA 18:2 and 20:2 was found among the treatment groups. Both short-chain ω -3 PUFA ALA (C18:3) and long-chain ω -3 PUFA DHA (C22:6) were significantly abundant in flaxseed oil group, while long-chain ω -3 PUFA

DHA only were considerably found in fish oil group. The highest content of total ω -3 fatty acids was detected in fish oil-fed group, followed by flaxseed oil-fed and rapeseed oil-fed group. The content of arachidonic acid, one of the ω -6 PUFAs (20:4), was significantly lower in both flaxseed oil and fish oil groups when compared with the control. The ratios of total PUFA/SFA and $\Sigma n3/\Sigma n6$ were significantly higher in the flaxseed oil, rapeseed oil, and fish oil groups than that in the beef tallow or the control groups.

A dose response study

In order to investigate the conversion efficacy of short-chain to long-chain ω -3 fatty acids in the liver, a dose response study using flaxseed oil diet at 0.5%, 1%, and 2% doses versus fish oil diet was conducted. As shown in Table 4, the contents of ALA in the livers of ducks fed various doses of flaxseed oil increased from the basal line of 0.06 to 0.16, 0.36, and 0.65 mg/g fresh weight gradually. Meanwhile, DHA content in flaxseed oil-fed group also increased from 0.29 to 0.72, 1.06, and 1.01 mg/g fresh weight. In fish oil-fed groups, DHA but not EPA content increased significantly from 0.29 to 1.11, 1.76, and 2.08 mg/g fresh weight. On the contrary, the content of ω -6 arachidonic acid (AA, 20:4) decreased in 2% of flaxseed oil- and 1-2% of fish oil-fed groups significantly.

Conversion between ω -3 fatty acids

The effect of short-chain ω -3 ALA-enriched flaxseed oil diet and long-chain ω -3 fatty acids-rich fish oil diet on liver DHA content is shown in Figure 2. DHA in duck liver became predominant in both fish oil and flaxseed oil groups. Among of ω -3 fatty acids, 91%, 92%, and 85% were DHA in the liver of ducks fed various fish oil doses at 0.5%,

1%, and 2%, respectively. Meanwhile, 81%, 73%, and 59% of total ω -3 fatty acids were converted to DHA in the duck liver fed flaxseed oil at 0.5%, 1%, and 2%, respectively. When compared with fish oil group, 1% of flaxseed oil produced an equivalent level of DHA as 0.5% of dietary fish oil.

Ratio of total ω -3/ ω -6 fatty acids

As shown in Figure 3, the ratios of $\Sigma\omega$ 3/ $\Sigma\omega$ 6 in duck liver gradually increased as the doses of flaxseed oil or fish oil increased. Fish oil group possessed a higher $\Sigma\omega$ 3/ $\Sigma\omega$ 6 value than flaxseed oil group at each dose, while a comparable value was observed between 1% of flaxseed oil and 0.5% of fish oil treatment.

Discussion

Fatty acid manipulation via dietary means may provide an effective method to obtain healthy animal products for humans. Our previous studies investigated the effect of dietary fat on fatty acid composition showing that different dietary fats could change ω -3 fatty acid composition in the duck eggs and muscle tissues. However, little information is available about ω -3 fatty acid profile in the liver modified by various dietary fats. Therefore, this present study, to our knowledge, is the first time to examine the modulation of different dietary fats on fatty acid profile and contents in the duck liver.

After 5-week's dietary treatment, all the dietary fats except for beef tallow showed significant modifications of the fatty acid profile and content in the duck liver. Although beef tallow provided more SFA than the control diet, no significant difference was found in 2% beef tallow-fed group, suggesting an effective transport and storage of SFA into

non-hepatic tissues such as adipose tissues. Furthermore, the MUFA-enriched rapeseed oil treatment did not affect any fatty acids except for ALA that significantly increased in hepatic tissues.

The most significant modification was observed in the groups fed with either flaxseed oil or fish oil. Ducks fed with flaxseed oil and fish oil diets were found to have much higher total ω -3 PUFA and ω -3/ ω -6 ratio than other groups. The ratio of ω -3/ ω -6 was achieved as high as 0.28 for flaxseed oil-fed group and 0.36 for fish oil-fed group. Such ratio is much higher than the modern Western diet and is compatible with that of our ancestors about 100-150 years ago.³⁹ The increase of both ratios in flaxseed oil and fish oil groups directly not only due to the increase of ω -3 fatty acids but also due to the decrease of ω -6 fatty acids, especially for AA. AA is a precursor of the derived eicosanoids such as PGE₂, TXA₂ and LTB₄. The decrease of ω -6 fatty acids like AA may thus reduce risk of platelet aggregation, hemorrhage, and vasoconstriction.⁴⁰⁻⁴¹ Some studies also suggested that a lower ratio of ω -3/ ω -6 diets suppress inflammation in patients with rheumatoid arthritis,^{42,43} and have a beneficial effect on patients with asthma.⁴⁴ The ω -3/ ω -6 ratio maybe a useful indicator to evaluate the healthy benefits of the functional food products.

It is interesting that duck liver possesses an efficient conversion of all the short-chain ω -3 fatty acids into long-chain DHA. About 60% of ALA in the flaxseed oil was converted to DHA in the duck liver, while 85% of total ω -3 fatty acids, mostly EPA and DHA, in the fish oil was converted to DHA. Such high conversion efficiency may be related to the broad substrate specificity of the duck elongase enzymes that convert the short-chain ω -3 PUFAs to final DHA exceptionally.²³

The results of dose response study showed that the total ω -3 fatty acids, specifically DHA, and the ratio of ω -3/ ω -6 increased as the dose increased in both flaxseed oil and fish oil treatments. It should be noted that 60-81% of the short-chain omega-3 ALA in flaxseed oil-fed group was efficiently converted to long-chain DHA in the duck liver and 1% of dietary flaxseed oil produced an equivalent level of DHA or ω -3/ ω -6 ratio as 0.5% of dietary fish oil. Therefore, the ducks fed flaxseed oil could be an alternative source of fish DHA. Considering that the cost of flaxseed oil is much less expensive than fish oil, it appears commercially applicable for flaxseed oil-enriched diet to be used by waterfowl to provide healthy products.

Taken together, this study investigated the effects of various dietary fats on fatty acid profile and contents of ω -3 fatty acids in duck liver. Total ω -3 fatty acids and the ratio of total ω -3/ ω -6 significantly increased in both flaxseed oil- and fish oil-fed groups. About 60-81% of the short-chain ω -3 ALA in flaxseed oil-fed group was efficiently converted to long-chain DHA in the duck liver, whereas 1% of dietary flaxseed oil could produce an equivalent level of DHA as 0.5% of dietary fish oil. It is significant that the short-chain ALA was efficiently converted to long-chain DHA in the duck liver, which may provide an alternative DHA-enriched duck products for human health benefits.

Authors' contributions: All authors participated in the review of the manuscript; JS and LL designed the experiments, XC and XD conducted the experiments and performed analysis, and WW and XC wrote the manuscript.

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Table 1. Composition and nutrient levels of the basal diet

Ingredients	Content (g/kg)	Nutrient	Content (g/kg)
Maize grain	400	Metabolizable energy	11.2 ^b
Wheat	290	Crude protein	16.5
Soybean meal	120	Total phosphorus	0.70
Wheat bran	90	Total calcium	3.35
Calcium hydrophosphate	12	Total lysine	0.79
Stone powder	80	Total methionine	0.40
Salt	3	Ether extract	29.0
Premix ^a	5		

^a Supplied per kg of diet: vitamin A 1,500 U, cholecalciferol 200 U, vitamin E (DL- α -tocopheryl acetate) 10 U, riboflavin 3.5 mg, pantothenic acid 10 mg, niacin 30 mg, cobalamin 10 μ g, choline chloride 1,000 mg, biotin 0.15 mg, folic acid 0.5 mg, thiamine 1.5 mg, pyridoxine 3.0 mg, Fe 80 mg, Zn 40 mg, Mn 60 mg, I 0.18 mg, Cu 8 mg, Se 0.3 mg; ^b Unit: MJ/kg.

Table 2. Measured fatty acids in the experimental diets

Fatty acid*	Content (g/100g total fatty acids)				
	Control	Flaxseed oil	Rapeseed oil	Beef tallow	Fish oil
SFA	32.61	25.50	26.22	41.94	30.36
MUFA	37.75	33.03	48.50	36.90	45.65
PUFA	29.84	41.59	25.40	20.25	35.12
Total ω3	7.53	20.54	5.66	4.74	15.57
18:3ω3	6.22	19.69	4.83	4.21	4.42
20:5ω3	0.98	0.64	0.63	0.35	6.14
22:6ω3	0.33	0.21	0.20	0.18	5.01
Total ω6	19.34	19.12	17.83	13.25	14.20
18:2ω6	18.77	18.67	17.42	12.93	12.73
20:2ω6	0.11	0.07	0.12	0.07	0.78
20:4ω6	0.46	0.38	0.29	0.25	0.69
PUFA/SFA	0.92	1.63	0.97	0.48	1.16
$\Sigma\omega$3/$\Sigma\omega$6	0.39	1.07	0.32	0.36	0.77

* SFA: Saturated fatty acids; MUFA: Monounsaturated fatty acids; PUFA: Polyunsaturated fatty acids.

Table 3. Fatty acid contents in the duck liver fed various dietary fats for 5 weeks*

Fatty acid**	Content (mg/g fresh weight)				
	Control	Flaxseed oil	Rapeseed oil	Beef tallow	Fish oil
SFA	8.45±1.94	8.90±0.67	9.99±2.45	9.78±0.97	11.13±3.70
MUFA	6.97±2.29	7.88±1.35	12.20±4.90	9.39±1.32	11.34±5.99
PUFA	5.68±0.93 ^a	7.8±0.5 ^{ab}	8.02±2.22 ^{ab}	6.33±0.64 ^a	9.62±3.34 ^b
Total ω3	0.36±0.02 ^a	1.69±0.09 ^c	0.95±0.25 ^b	0.33±0.03 ^a	2.45±0.67 ^d
18:3ω3	0.06±0.01 ^a	0.65±0.15 ^c	0.35±0.25 ^b	0.08±0.02 ^a	0.19±0.12 ^{ab}
20:5ω3	UD	0.04±0.01 ^a	0.03±0.02 ^a	UD	0.18±0.02 ^b
22:6ω3	0.29±0.03 ^a	1.01±0.13 ^b	0.58±0.07 ^a	0.25±0.02 ^a	2.08±0.51 ^c
Total ω6	5.32±0.92	6.08±0.43	7.07±1.98	6.00±0.62	7.16±2.78
18:2ω6	2.50±0.70	3.88±0.61	4.55±2.14	3.29±0.66	5.35±2.67
20:2ω6	0.08±0.02	0.09±0.01	0.11±0.03	0.09±0.02	0.09±0.04
20:4ω6	2.74±0.29 ^a	2.11±0.30 ^b	2.41±0.25 ^{ab}	2.62±0.21 ^a	1.72±0.26 ^c
PUFA/SFA	0.68±0.06 ^a	0.88±0.07 ^b	0.80±0.09 ^b	0.65±0.02 ^a	0.86±0.05 ^b
Σω3/ Σω6	0.07±0.01 ^a	0.28±0.02 ^c	0.14±0.01 ^b	0.05±0.00 ^a	0.36±0.09 ^d

*Values are expressed as mean ± SD (n=3-6). Means in a row without a common letter differ, $p < 0.05$.

**SFA: Saturated fatty acids; MUFA: Monounsaturated fatty acids; PUFA: Polyunsaturated fatty acids. UD: undetectable.

Table 4. Fatty acid contents of duck liver fed various dose of dietary fats for 5 weeks*

Fatty acid**	Content (mg/g fresh weight)						
	Control	Flaxseed oil			Fish oil		
		0.5%	1%	2%	0.5%	1%	2%
SFA	8.45±1.94	8.38±1.26	9.20±0.73	8.90±0.67	10.28±2.04	9.83±1.10	11.13±3.70
MUFA	6.97±2.29	6.35±2.80	6.89±1.20	7.88±1.35	11.15±3.62	9.21±2.45	11.34±5.99
PUFA	5.68±0.93 ^a	6.70±0.63 ^a	7.82±0.66 ^{ab}	7.8±0.5 ^{ab}	7.44±1.46 ^{ab}	7.96±0.39 ^{ab}	9.62±3.34 ^b
Total ω3	0.36±0.02 ^a	0.90±0.05 ^b	1.45±0.24 ^{cd}	1.69±0.09 ^{cd}	1.22±0.17 ^{bc}	1.91±0.01 ^d	2.45±0.67 ^e
18:3ω3	0.06±0.01 ^a	0.16±0.05 ^a	0.36±0.15 ^b	0.65±0.15 ^c	0.09±0.03 ^a	0.11±0.02 ^a	0.19±0.12 ^a
20:5ω3	0.00±0.01 ^a	0.01±0.00 ^a	0.03±0.01 ^a	0.04±0.01 ^a	0.01±0.01 ^a	0.05±0.01 ^a	0.18±0.12 ^b
22:6ω3	0.29±0.03 ^a	0.72±0.09 ^b	1.06±0.08 ^b	1.01±0.13 ^b	1.11±0.15 ^b	1.76±0.05 ^c	2.08±0.51 ^c
Total ω6	5.32±0.92	5.81±0.65	6.37±0.43	6.08±0.43	6.22±1.34	6.05±0.40	7.16±2.78
18:2ω6	2.50±0.70 ^a	2.64±1.06 ^a	3.44±0.52 ^{ab}	3.88±0.61 ^{ab}	3.71±1.21 ^{ab}	3.72±0.55 ^{ab}	5.35±2.67 ^b
20:2ω6	0.08±0.02	0.07±0.01	0.09±0.01	0.09±0.01	0.10±0.03	0.09±0.03	0.09±0.04
20:4ω6	2.74±0.29 ^{ab}	3.10±0.42 ^a	2.83±0.16 ^{ab}	2.11±0.30 ^{cd}	2.42±0.15 ^{bc}	2.24±0.16 ^c	1.72±0.26 ^d

PUFA/SFA	0.68±0.06 ^a	0.81±0.05 ^{bc}	0.85±0.03 ^c	0.88±0.07 ^c	0.73±0.06 ^{ab}	0.82±0.10 ^{bc}	0.86±0.05 ^c
Σω3/ Σω6	0.07±0.01 ^a	0.16±0.02 ^b	0.23±0.02 ^{bc}	0.28±0.02 ^{cd}	0.20±0.03 ^b	0.32±0.02 ^{de}	0.36±0.09 ^e

* Values are expressed as mean ± SD (n=6). Means in a row without a common letter differ, $p < 0.05$.

** SFA: Saturated fatty acids; MUFA: Monounsaturated fatty acids; PUFA: Polyunsaturated fatty acids. Values are expressed as mean ± SD (n=6).

Figure legends:

Figure 1. Representative Gas Chromatography of fatty acid profile in the liver of ducks fed with various dietary fats for 5 weeks. *Shan Partridge Ducks* were randomly assigned into 5 dietary treatments: either the basal diet or 2% of flaxseed oil, rapeseed oil, beef tallow, or fish oil, respectively. At the end of the 5-week treatment, fatty acids in duck liver were analyzed by GC-FID. Totally 23 fatty acids were identified and detected as follows. 1. myristic acid, C14:0; 2. pentadecanoic acid, C15:0 (internal standard); 3. palmitic acid, C16:0; 4. cis-6-hexadecenoic acid, C16:1n-10; 5. palmitoleic acid, C16:1n-7; 6. margaric acid, C17:0; 7. steric acid, C18:0; 8. oleic acid, C18:1n-9; 9. vaccenic acid, C18:1n-7; 10. linoleic acid, C18:2n-6; 11. arachidic acid, C20:0; 12. α -linolenic acid, C18:3n-3; 13. stearidonic acid, C18:4n-3; 14. gondoic acid, C20:1n-9; 15. eicosadienoic acid, C20:2n-6; 16. dihomogamma-linolenic acid, C20:3n-6; 17. arachidonic acid, C20:4n-6; 18. eicosapentaenoic acid, C20:5n-3; 19. lignoceric acid, C24:0; 20. adrenic acid, C22:4n-6; 21. docosapentaenoic acid, C22:5n-6 (Osbond acid or all-cis-4,7,10,13,16-docosapentaenoic acid); 22. docosapentaenoic acid, C22:5n-3 (clupanodonic acid or all-cis-7,10,13,16,19-docosapentaenoic acid); 23. docosahexaenoic acid, C22:6n-3.

Figure 2. Dose response of ω -3 fatty acids in the liver of ducks fed with various doses of flaxseed oil or fish oil for 5 weeks. *Shan Partridge Ducks* were randomly assigned into a dose response study by feeding either flaxseed oil or fish oil diets at 0.5%, 1%, and 2%, respectively. At the end of the 5-week treatment, fatty acids in duck liver were analyzed by GC-FID. About 59-81% and 85-92% of total ω -3 fatty acids were converted to DHA in the duck liver fed various doses of flaxseed oil and fish oil, respectively. The dose of 1% flaxseed oil produced an equivalent level of DHA as 0.5% fish oil. Values are expressed as mean \pm SD (n=6). Means in a group without a common letter differ, $p < 0.05$.

Figure 3. Dose response of total ω -3/ ω -6 ratio in the liver of ducks fed with various disease of flaxseed oil or fish oil for 5 weeks. *Shan Partridge Ducks* were randomly assigned into a dose response study by feeding either flaxseed oil or fish oil diets at 0.5%, 1%, and 2%, respectively. At the end of the 5-week treatment, fatty acids in duck liver were analyzed by GC-FID. The ratios of ω 3/ ω 6 in duck liver gradually increased as the doses of flaxseed oil or fish oil increased. Fish oil group possessed a higher ω 3/ ω 6 value than flaxseed oil group, but a comparable value was observed between 1% of flaxseed oil and 0.5% of fish oil treatment. Values are expressed as mean \pm SD (n=6). Means in a group without a common letter differ, $p < 0.05$.





