EFFECTS OF CONTINUED APPLICATION OF NITROGEN AND PHOSPHORUS ON THE CHEMICAL COMPOSITION OF IRRIGATED RICHFIELD SILT LOAM

1050 710

by

GALEN RAY NIEDENTHAL

B. S., Kansas State University, 1969

A MASTER'S THESIS

submitted in partial fulfillment of the

requirements for the degree

MASTER OF SCIENCE

Department of Agronomy

KANSAS STATE UNIVERSITY Manhattan, Kansas

1974

Approved by:

Major Professor

LD 2668 T4 1974 N54 c.2 Document

TABLE OF CONTENTS

ACKN	OWLEDGEMENTS		٠			•	•	•	•		•	٠	•	•	•	٠	•	•	ě	٠	•	•	e •	•	Pag iv
LIST	OF FIGURES		• 1			•	•	•						ě		100	•	•	•	•		800		•	v
LIST	OF TABLES .		•		•	•	٠	•	•	•	•	•	•	•	•	•	•	•	٠	٠		•	•	٠	vii
LIST	OF APPENDIX	TABLE	S .			•	•	•	•		•	100	: • :	•	•		•		•		•		•	•	viii
INTR	ODUCTION		•	• •	٠	•	٠	٠	•	٠	•	•	•	•	٠	•	•	•		•	•	•	•	*	1
REVI	EW OF LITERAT	URE .			•	٠	٠	•	•	•	•	•			•	٠	•					: • :	::•:		3
	Soil Fertilia Organic Car Soil Reaction	rbon n Cha	nges	aı	nd	Se	ecc	ond	lar	· ·	Ef	Te	ect	s	on	. C	Oth	ner	•	٠					3
	Soil Chemic Effects of Co	ontin	uous	s Fe	ert	il	iz	zat	cic	n	or	N	lit	ra	ite	A	cc	eun	ıul	at	ic	on			4
	and Leachin	ng .			::•	٠	٠	•	•	•	•	•	•	•	•	•	•		•	•	•	٠	•		9
	Ammonium-nit:																								12
	Nitrogen Loss Effects of Pl																		•	•	٠	•	•	٠	14
	Properties Effects of Fe		•				•	•					٠					•				•	•		16
	Soils																					•	•	٠	17
METH	DDS AND MATER	IALS	•	h: 0 4	•		•	•	•	•.	•	•	•	•	•		•	•	10.	•	•	•			1 9
	Soil Sampling	g			•		•	•	•	٠		•	•	•	•		•	٠	•		•	•	•	•	19
	Ammonium and																								23
	Total Nitroge																								23
	pH				•	٠		•	•	٠	٠	•	٠	•	•	٠	•	•	•	•	٠	٠	•	•	24
	Available Pho	ospho:	rus		•	•	•	•		•		٠	٠	•		×	•	•	٠	•		•	•	٠	24
	Extractable (Catio	ns .			·			•					٠	•			•	٠			٠	•	٠	24
	Cation Exchar	nge C	apac	eity	y						•			•					•			•			24
	Micronutrien	ts .	-														•	•						•	25
	Carbon				•	•	•	•	•	•	•	•	٠	•	٠	•	٠	٠	•	•	•	٠	•	٠	25
RESU	TS AND DISCU	SSION	• 1	•	ě	•	٠	•	Ē	¥	•	•	٠	•	ĕ	ě	•	٠	•	ě	•	٠	•	•	27
	Ammonium-nit						٠	•	•	•	•	•	•	٠	•	•	•	•	•	•	•	•	•	٠	27
	Nitrate-nitro																					•	•	•	30
	Total Nitroge																					•	٠	•	31
	рН																					•	٠	•	31
	Available Pho																•	•	1.01	٠	ï	•	٠	•	32
	Extractable (_		_	- 50	-		-	_	-	-	37	_	-	-	•	٠	٠	•	•		•	•	32
	Extractable N	Magne:	sium	1.	•		٠	•	٠	•		•	٠		٠		•	•	٠	٠	•	•	•	٠	39
	Extractable S	Sodiw	m.		٠		٠	٠	٠	•	•	•	•		•		•	•	٠		÷		٠	٠	39
	Extractable 1	Potas:	siun	ι.		ï																			39

																												Page
	Cation Exc	:hane	zе	Ca	pa	ic:	Ltj	7	•				•							•		•		•		ï		40
	Extractabl	.e Cc	pp	er		4			•	٠			•		•	٠		•		•		•		•	٠	•	•	40
	Extractabl	e Ir	con		•			•	•	٠	•	٠	•	٠	٠		•		•	٠	٠	٠	٠	٠	٠	•	•	46
	Extractabl	.e Ma	ing	an	es	se			٠	٠				•	٠	٠	٠.		•	•	•		•	•	٠	•	•	46
	Extractabl	.e Zi	inc			•	ï		•	•			•	•	•		•	٠	•	٠		ě	٠	•	٠	•		47
	Total Carb	on		•	•				•				•	•	•		ï		٠	•	٠	•	ı	•	•	•	•	52
	Carbonate-	cart	oon		•				٠	•	•	٠		•	٠			•	•	•	٠	•	•	•	•			52
	Organic Ca	rbor	1	٠	•	•	•	•	•	٠	•	٠	•	•	•	•	•	٠	•	•	•	•	٠	•	•	•	٠	52
SUMMA	ARY AND CON	CLUS	SIO	NS		•	•	•	•	•	ě	•	٠	•	•	•	٠	•	•	•	•	ě	٠	•	•	٠	•	57
LITEF	RATURE CITE	D.	•	•	•	•	•	•	•	•	•	٠	•	•	•	•	•	•	•	•	•	•	•	•	•	•	•	60
APPEN	XTOI			120	-20	21			120	720	20	0	12	100	72	_	2	-	-	-	_	_	·	-	-	2	-	67

ACKNOWLEDGEMENTS

The author would like to express his sincere appreciation to Dr. Larry S. Murphy, major professor, for the suggestion of this problem, and his advice and counseling during the course of study. His assistance in interpreting the data and preparing the manuscript was also greatly appreciated. Thanks is also extended to Dr. William Powers and Dr. David Whitney who served as members of the supervisory committee.

Appreciation is extended to Dr. Roscoe Ellis, Jr. for his advice regarding chemical analysis procedures and to G. W. Wallingford for his suggestions regarding computing techniques and his assistance with carbon analysis procedures.

Sincere thanks are extended to the personnel of the KSU soil fertility laboratory and soil testing laboratory for their assistance with the chemical analysis.

LIST OF FIGURES

Figure	,	Page
1.	Truck-mounted hydraulic soil probe used to obtain soil core samples	22
2.	Method used for sectioning soil cores into desired increments	22
3.	Soil ammonium-N as affected by nitrogen and phosphorus treatments	33
4.	Soil nitrate-N as affected by nitrogen and phosphorus treatments	34
5.	Total soil nitrogen as affected by nitrogen and phosphorus treatments	35
6.	Soil pH values as affected by nitrogen and phosphorus treatments	36
7.	Weak Bray extractable P as affected by nitrogen and phosphorus treatments	37
8.	Extractable soil calcium as affected by nitrogen and phosphorus treatments	41
9.	Extractable soil magnesium as affected by nitrogen and phosphorus treatments	42
10.	Extractable soil sodium as affected by nitrogen and phosphorus treatments	43
11.	Extractable soil potassium as affected by nitrogen and phosphorus treatments	1 μ
12.	Soil cation exchange capacity as affected by nitrogen and phosphorus treatments	45
13.	DTPA-extractable soil copper as affected by nitrogen and phosphorus treatments	48
14.	DTPA-extractable soil iron as affected by nitrogen and phosphorus treatments	49
15.	DTPA-extractable soil manganese as affected by nitrogen and phosphorus treatments	50
16.	DTPA-extractable soil zinc as affected by nitrogen and phosphorus treatments	51

Figure		Page
17.	Total soil carbon as affected by nitrogen and phosphorus treatments	54
18.	Soil carbonate-carbon as affected by nitrogen and phosphorus treatments	55
19.	Soil organic carbon as affected by nitrogen and phosphorus treatments	56

LIST OF TABLES

a	pre	•	Pa	age
	1.	Effects of nitrogen, phosphorus, and potassium on the yield of irrigated corn	•	20
	2.	General soil analysis data from the experimental area prior to the current investigation	•	21
	3.	Mean values for ammonium-N, nitrate-N, total N, pH, and available P as affected by N rate or P application		28
	4.	Mean values for ammonium-N, nitrate-N, total N, pH, and available P as affected by depth		29
	5.	Mean values for calcium, magnesium, sodium, potassium, and CEC as affected by N rate, P application, or depth	•	38
	6.	Mean values for copper, iron, manganese, and zinc as affected by N rate, P application, or depth		47
	7.	Mean values for total carbon, carbonate-carbon, and organic carbon as affected by N rate, P application, or depth.		53

LIST OF APPENDIX TABLES

<u>Table</u>				1	Page
1.	Soil $\mathrm{NH}_{l_{\!$	•	•	•	68
2.	Soil NH4+N (ppm) as affected by treatmentreplication 2	•	•	•	70
3.	Soil NH4+-N (ppm) as affected by treatmentreplication 3	•	٠	•	72
4.	Soil NO_3-N (ppm) as affected by treatmentreplication 1	•	•	•	74
5.	Soil NO3-N (ppm) as affected by treatmentreplication 2	•	٠	•	76
6.	Soil NO3-N (ppm) as affected by treatmentreplication 3	٠	٠	•	78
7.	Total soil N (ppm) as affected by treatmentreplication 1	•	٠	ď	80
8.	Total soil N (ppm) as affected by treatmentreplication 2	•		•	82
9.	Total soil N (ppm) as affected by treatmentreplication 3	•	•	•	84
10.	Soil pH as affected by treatmentreplication 1	•	•	•	86
11.	Soil pH as affected by treatmentreplication 2	•	٠	•	88
12.	Soil pH as affected by treatmentreplication 3	•	•	≅•	90
13.	Weak Bray extractable soil phosphorus (ppm) as affected by treatment	٠	•	•	92
14.	Total soil carbon (%) as affected by treatment	٠	٠	•	93
15.	Soil carbonate-carbon (%) as affected by treatment	8.00	100	•	94
16.	Soil organic carbon (%) as affected by treatment	•		•	95
17.	Soil cation exchange capacity (meq/100 g) as affected by treatmentreplication 1	•	•	•	96
18.	Soil cation exchange capacity (meq/100 g) as affected by treatmentreplication 2	•	::•1	•	97
19.	Soil cation exchange capacity (meq/100 g) as affected by treatmentreplication 3	•	•	•	98
20.	Extractable soil calcium (ppm) as affected by treatment-replication 1	•	•	•	99
21.	Extractable soil calcium (ppm) as affected by treatment replication 2	•	4	•	100

<u> lable</u>				Page
22.			calcium (ppm) as affected by treatment	101
23.	Extractable replication		magnesium (ppm) as affected by treatment	102
24.			magnesium (ppm) as affected by treatment	103
25.			magnesium (ppm) as affected by treatment	104
26.			sodium (ppm) as affected by treatment	105
27.			sodium (ppm) as affected by treatment	106
28.			sodium (ppm) as affected by treatment	107
29.	Extractable replication		potassium (ppm) as affected by treatment	108
30.			potassium (ppm) as affected by treatment	109
31.			potassium (ppm) as affected by treatment	110
32.	DTPA-extract	table	soil copper (ppm) as affected by treatment	111
33.	DTPA-extract	table	soil iron (ppm) as affected by treatment	112
34.	DTPA-extract	table	soil manganese (ppm) as affected by treatment .	113
35	DTPA_extract	-ahla	soil ging (npm) as affected by treatment	114

INTRODUCTION

Substantial increases in agricultural production are necessary to support rapidly growing populations in most areas of the world today. Proper fertilizers, applied at the appropriate rate and time, are essential if needed levels of production are to be achieved and if economic development is to proceed. The inadequate availability of fertilizer today due to the present shortages of natural gas and other energy resources makes proper fertilization techniques even more important.

Nitrogen and phosphorus are two nutrients that have received much concentrated interest in research. Nitrogen supplied to the soil by natural processes is seldom sufficient to produce satisfactory yields of crops.

Accordingly, more and more reliance has been placed on nitrogen fertilizers.

Abundant amounts of phosphorus usually exist in soils, but most of it is present in highly insoluble minerals and is unavailable to plants. Even when available P is applied to the soil, it can be rendered unavailable due to reactions between soil components and phosphorus.

The marked increase in the use of N and P fertilizers in recent years has generated growing interest in the effect of these fertilizers on soil chemical properties, especially when applied at high rates over a period of years. The continued use of single nutrient fertilizers may eventually produce a nutrient imbalance in the soil through excessive removal of certain other nutrients. Nitrogen applied at excessive rates or at improper times may increase the probability of nutrient loss through volatilization or through leaching and possible groundwater contamination.

This investigation was part of a long-range study initiated at the Tribune Branch Experiment Station to evaluate the responses of irrigated corn to various rates of nitrogen, phosphorus, and potassium fertilizers. Beginning in 1961, fertilizer was applied broadcast by hand before planting two corn hybrids, using NH₄NO₃ (34-0-0), triple-superphosphate (0-20-0, or 0-46-0 on a P₂O₅ basis), and KCl (0-0-50, or 0-0-60 on a K₂O basis). Rates of application of each element, expressed in kg/ha, were: nitrogen: 0, 45, 90, 134, 179, and 224; phosphorus: 0 and 19; and potassium: 0 and 37. These trials have been continued with the same amount of nitrogen, phosphorus, and potassium applied to each plot each year. In 1968 and 1969, a uniform application of 11 kg/ha of zinc was applied to the corn test area as ZnSO₄ to offset a low soil zinc condition as determined by soil tests. Since

Recent corn yields at this location have indicated no significant response to the applied potassium or to the additional 19 kg/ha phosphorus application. Nitrogen applications up to 179 kg/ha have increased grain yields considerably, but 224 kg/ha of nitrogen has not produced greater corn yields than the 179 kg/ha rate.

The portion of the study reported in this thesis was concerned with the effect of the continuous applications of N and P on the chemical properties of the Richfield silt loam soil, and to determine the fate of applied nitrogen not utilized by the corn crop.

REVIEW OF LITERATURE

Soil Fertility Treatment Effects on Total Nitrogen and Organic Carbon

Long-term soil fertility treatments have been shown to have dramatic effects on soil chemical properties (28, 32, 45, 46, 62, 68, 69). Dodge and Jones (28) reported that in one long-term study, there was a continual over-all loss of nitrogen and carbon over the entire 30-year cropping period regardless of cropping system or fertilizer treatment. Fertility treatments had relatively little influence on the nitrogen trends in the soil or on the C/N ratio. Plots with the highest nitrogen content at the beginning of the experiment suffered the greatest losses of nitrogen.

Haas and Evans (32) showed that there was a sharp decline in total N and organic C after a 36-year cropping period in another study. Pratt and Chapman (69) found that there were losses of total nitrogen and organic carbon at the 90-120 cm depth after a 20-year lysimeter investigation. They also reported decreases in exchangeable potassium with depth, and decreases in magnesium content with increasing N treatments.

In arguing against the claim that agricultural fertilization is a leading source of nitrates in our water supplies, Stewart pointed out that the total nitrate-N currently available from the soil organic matter and fertilizer is less than the soil organic matter alone furnished at the beginning of cultivation many years ago (79). Since both total N and organic C are reliable estimates of organic matter content and the content of organic matter is an index of the amount of potentially available nitrogen in a soil, it can be seen that these effects of continuous cropping are of considerable importance.

Soil Reaction Changes and Secondary Effects on Other Soil Chemical Properties

Of the many effects long-term applications of nitrogen fertilizers have on the soil, the most important is probably the general tendency of these materials to alter the surface and subsoil pH (1, 2, 12, 23, 46, 62, 66, 67, 88, 89, 90, 91). Nitrogen carriers have a direct, immediate effect on soil acidity and a residual effect which develops more slowly (88). Residual changes in soil pH may be in the same direction as the immediate change at the time of application, as in the case of ammonium salts and calcium cyanamid; or residual effects may completely reverse direct effects, as in the case of anhydrous ammonia, diammonium phosphate, and materials, such as urea, which release ammonia upon hydrolysis. Clevenger and Willis (23) described the immediate decrease in soil pH upon addition of nitrogen fertilizers as due to a "salt effect". Wolcott came to the same conclusions after his work with several N fertilizers (88, 90, 91).

Residual acidifying effects of most common N fertilizers are generally in the following descending order: mono-ammonium phosphate, ammonium sulfate, ammonium chloride, di-ammonium phosphate, ammonium nitrate = anhydrous ammonia = urea = ureaform (66, 88, 90). Calcium nitrate, sodium nitrate, potassium nitrate, and calcium cyanamid possess a residual basic effect on most soils.

The increased acidity of soils due to continuous applications of nitrogen fertilizers has several indirect effects on other soil chemical properties. Abruna, Pearson, and Elkins (1) found that heavy applications of ammonium nitrate and ammonium sulfate caused severe reduction in exchangeable base level after only one year of applications. He reported that exchangeable K was lost from the soil faster than other bases. An indication of a subsoil zone of accumulation of bases leached out of upper horizons was observed.

THIS BOOK CONTAINS NUMEROUS PAGES WITH MULTIPLE PENCIL AND/OR PEN MARKS THROUGHOUT THE TEXT.

THIS IS THE BEST IMAGE AVAILABLE.

110

Adams et al. (2) found that annual applications of N fertilizer exceeding 224 kg/ha reduced soil pH, and that the higher the N rates, the deeper the profile pH was affected. The movement of calcium and magnesium downward to the 30-45 cm zone confirmed the effect of pH on these cations. a In another study, ammonium sulfate applied at a rate of 38 kg N/ha lowered soil pH from 7.4 to 6.0 after 5 years (12). The effect of ammonium nitrate was less pronounced, and calcium nitrate had no influence on pH. The pH changes only had slight effects on exchangeable and soluble cations.

The use of high rates of residually acid nitrogen fertilizer caused appreciable downward movement of calcium and magnesium salts in a study by Hilthold et al. (35). In a Michigan study (91) the effects of ammonium sulfate and calcium nitrate on exchangeable base status were compared. Total exchangeable bases (K + Ca + Mg) declined consistently from year to year at all sampling depths, regardless of treatment.

Nitrogen sources can indirectly effect micronutrient availability Micro because of changes in soil pH. In a study reported by Leo, Odland, and Bell (35), long continued use of ammonium sulfate increased soil acidity and also increased the solubility of toxic aluminum. There was also slight downward movement of organic matter, which increased subsoil organic matter content, CEC, and led to a wider C/N ratio in the subsoil. Viets (84) found that poor aeration (low redox potential) coupled with low pH increased the amount of water-soluble manganese and iron in soils but had little effect on zinc and copper. Copper availability is dependent upon soil pH but does not normally increase appreciably until pH falls below 5.0. Manganese availability increases markedly when the pH falls below 5.5. Field studies with corn in Michigan showed that acidifying nitrogen carriers increased the uptake of manganese, zinc, and boron, while tending to depress the uptake of molybdenum and copper (90).

Owensby et al. (62) observed that high rates of nitrogen applied to bromegrass lowered soil pH in the upper 15 cm of the soil profile. In the 15-30 cm depth, only the 224 kg/ha N rate lowered the pH significantly.

Exchangeable potassium decreased with nitrogen rate initially, but increased at the higher rates in the upper portions of the soil profile. This increased availability was linked to increased acidity at those levels.

Boawn et al. (12) showed that high rates of applied N had no pronounced influence on P availability, but the resultant lowered pH did increase the availability of manganese. Very low soil pH may decrease P availability due to the formation of insoluble iron and aluminum compounds containing phosphorus. Pierre and others (67) noticed that with high nitrogen application rates developed acidity was much less than the theoretical amount expected. They reported that this was due to the low excess base compared with nitrogen in the harvested crop. They also speculated that there were large losses of nitrogen due to denitrification without an equivalent loss of bases. Dancer, Peterson, and Chesters (27) found that as soil pH decreased, the rate of nitrate accumulation decreased and the length of the delay period before nitrate-nitrogen started to accumulate was increased.

In a study by Broadbent and Tyler (19) utilizing ammonium chloride and potassium nitrate, nitrogen immobilized in the soil from the ammonium source increased as pH increased, whereas the reverse was true of the nitrate source. These results appear to be related to the physiological acidity or alkalinity of the nitrogen source. Use of residually acidic fertilizers on a calcareous soil tends to lower pH, but also increases the probability of immobilizing some available nitrogen.

Cornfield (26) found that acid soils tend to accumulate organic nitrogenous residues to a greater extent than do soils of high pH and that the nitrogen in such residues tends to mineralize readily if soil pH is increased. He also found that nitrate accumulation occurred more rapidly in soils having a pH greater than 6.5. Ammonium accumulation was generally high in acid and low in neutral and alkaline soils (25). Generally, soils that fix ammonium under moist conditions have low nitrification values.

While the effect of nitrification on soil acidity has received considerable attention, any effect that nitrogen volatilization might have on acidity has been ignored. Hiltbold and Adams (35) noted that nitrification of an added ammonium salt creates acidity equivalent to the ammonium oxidized plus the anion with which it is associated. Gaseous loss of ammonia or volatilization of nitrogen during nitrification effect a removal of potential acidity equivalent to the nitrogen lost. Denitrification of nitrate results in formation of OHT equivalent to the nitrate reduced and volatilized. The frequent unaccounted losses as well as low recoveries of applied nitrogen by plants indicate that volatilization may appreciably effect the soil acidity resulting from N fertilizers.

As soils become more acidic over a period of time due to nitrogen fertilization, the leachates in the upper horizons tend to show greater acidity than do the soils themselves. As these leachates move downward, the lower depths decrease in pH also. On acid-sensitive crops, low pH may inhibit root growth, thus allowing less utilization of nitrogen deep in the soil profile (2).

Abruna (1) observed that excess soil acidity from long term nitrogen applications may also effect activity of soil microorganisms. The optimum pH of nitrification is about 7.5 to 9.0, so nitrification may be retarded if the soil pH is decreased too much.

Continuous nitrogen applications have been shown to have secondary effects on the availability of soil phosphorus. Grunes (31) found that additions of nitrogen fertilizers to the soil can have both a salt effect and a pH effect on phosphorus availability. In calcareous soils, the salt increased the solubility and hydrolysis of the calcium carbonates to produce a higher calcium ion concentration which in turn reduced the phosphate concentration. Residually-acidic N fertilizers slowed down reversion of soluble phosphorus fertilizers to a more insoluble form and brought more phosphorus into solution from the reaction product of the soil and the P fertilizer.

Lorenz and Johnson (49) found that the physiologically acid ammonium salt, $(NH_4)_2SO_4$, effectively released native soil phosphate better than NH_4NO_3 on a fine sandy loam soil with a pH of 7.8. Olsen et al. (60) discovered that phosphorus solubility in calcareous soils in Colorado increased rapidly as the soil pH decreased. Lowering the pH of alkaline soils was found to increase the phosphorus in solution (21, 68).

Owensby et al. (62) found that high nitrogen rates increased the availability of phosphorus in the surface 15 cm of soil in a study in Kansas. Rennie and Soper (72) found that increased utilization of phosphorus occurred only when the applied nitrogen was in the ammonium form. Nitrate sources of N were relatively ineffective. They attributed this to the fact that the ammonium ion indirectly influences the plants' ability to take up phosphorus, but does not effect the availability of the applied phosphorus fertilizer. At higher soil pH levels (7-9), HPO₁₄-2 is the dominant ionic form of P, and is less readily absorbed by most plants than the H₂PO₄-1 ionic form found within the pH range 5-7.

Effects of Continuous Fertilization on Nitrate Accumulation and Leaching

Oxidation of ammonia to nitrate results in replacement of the basic ion by hydrogen and the conversion of nitrogen to a mobile form in which it may accompany soil bases in leaching (35). Much work has been conducted involving accumulations of nitrate in the soil. Bates and Tisdale (9) found that the form in which nitrates are added to the soil, the nature of the accompanying ion, and the physical placement of the salt in the soil all contribute to the net movement of nitrate-nitrogen in the soil. Upward movement of this ion is influenced by the movement of capillary water resulting from surface evaporation.

Gardner (30) found that correlation of nitrate leaching losses with rainfall is not as direct as might be expected. He hypothesized that the distance which nitrate will move downward depends not so much upon the total rainfall as upon the amount of rainfall which actually passes through the soil. In a comparison of two application methods of ammonium nitrate fertilizer, Nelson (58) observed that there was little movement of ammonium-nitrogen, but nitrate-nitrogen moved downward about 56 cm during the growing season.

Johnston et al. (40) found that large percentages of applied nitrogen were lost in tile drainage effluent from silty clay loam soils. He observed that most nitrogen lost was in the nitrate form, but some was in the form of NH₃, NO₂ and even organic nitrogen. In a study by Stewart et al. (80), more nitrate-nitrogen accumulated under irrigated fields than under dryland cropped conditions in eastern Colorado.

Olsen et al. (59) found that the total amount of nitrate-N in soil profiles was directly related to the rate of nitrogen applied. Other researchers have found that nitrate movement in the soil over time is not as great as would be expected.

Peterson and Attoe (65) found that on well-drained silt loam soils and with moderate rainfall, losses of nitrogen by leaching were small. They observed that most of the nitrate-nitrogen not removed by the crop was found in the soil within the root zone. Moore (55) found that nitrate-nitrogen movement through the soil was considerably slower than movement of the percolating water.

Larson et al. (45) observed an apparent lack of nitrate-N movement and accumulation in fine-textured soils under northern climatic conditions even under high rates of continuous nitrogen fertilization. Boswell and Anderson (13) found little movement of mineral nitrogen in a sandy clay loam and a loamy sand after 5 months. Even with high accumulated rainfall, appreciable amounts of the applied N were evident at 1-2 meter depths 18 months after nitrogen was applied to fallowed plots.

Data obtained by Herron et al. (34) shows that nitrates in large amounts accumulated during a three-year period in Nebraska, but it apparently stayed within the root zone even under irrigated conditions. Accumulation of nitrate-N below 180 cm has been found to be very slight in these soils.

Cornfield (25) found that nitrate-nitrogen accumulation in incubated soils was significantly correlated with total nitrogen and organic carbon contents. Pratt and associates (70) found that in order to ascertain nitrate-N concentrations in the unsaturated zone (the area below the root zone and above the water saturated zone), it is necessary to know (1) the volume of drainage water, (2) the yearly excess of nitrate available for leaching, and (3) an estimate for denitrification.

Since nearly two-thirds of the total annual precipitation occurs in the northern great plains region during the May-September period, leaching during winter months is usually not a problem in this area (34). The usual period when heavy rains are likely to occur and which might cause leaching of nitrates in soils would appear to be spring when cultivated crops are not yet established. Herron et al. (34) summarized that utilization of residual nitrate-nitrogen by plants would appear to be the best and most practical method for preventing build-up of nitrate in the soil profile. Olsen et al. (59) stated that the most effective methods for limiting the amounts of NO₃-N passing through the soil profile include: maintaining a crop cover on the land as much of the time as is feasible, reducing the acreage and frequency of crops that receive fertilizer-N in the rotation, and limiting the rate of N fertilization to approximately that required by the crop.

Nitrates can accumulate near the soil surface after extended dry periods (75, 77, 87). Simpson (75) argued that nitrate-nitrogen is microbiologically assimilated during progressive drying of the topsoil and is protected by the dry conditions from leaching or microbiological reduction. He hypothesized that transport of nitrate ions to the surface in the soil solution by capillary transport is not responsible for the major nitrate accumulation near the soil surface.

Stephens (77) also postulated that most accumulations of nitrate in the topsoil are probably due to the microbiological manufacture of nitrate during nitrification. During dry spells in Uganda, a limited amount of upward movement of soil solution can occur and this gave rise to some accumulation of nitrate in the surface soil.

Wetselaar (87) stated that accumulation of nitrate near the soil surface in Australia must be physical movement, because soil temperatures were too high and soil water content too low for biological nitrification processes to take place. He also rebuked photo-chemical oxidation reactions

of nitrate formation because of shallow penetration of ultra-violet light and the adverse effect of the heat component of radiation on decreasing nitrate content in the surface soil.

Ammonium-Nitrogen Losses by Volatilization

When materials containing or yielding ammonium are applied to the soil in the irrigation water or by broadcast methods followed by irrigation, the greater portion of the ammonium is adsorbed at or near the soil surface (38, 39). The equilibrium reactions between the soil base-exchange compounds and the soil solution adjust so that ammonium compounds are present in the soil solution as long as ammonia exists in the adsorbed state (50). If the soil solution is alkaline, then a part of the ammonia will be present as hydrated ammonia, ammonium hydroxide, ammonium bicarbonate, and ammonium carbonate, depending on the alkalinity, concentration and other factors.

Ray et al. (71) found that migration of ammonium-nitrogen is closely related to the movement of water. The magnitude of migration is dependant upon such soil characteristic as texture, organic matter content, and CEC. There is less retention of ammonium-nitrogen in sandy soils and movement is predominantly downward and lateral. In finer textured soils, such as loams and silt loams, ammonium-N movement is more symmetrical with slightly greater upward and lateral movement than downward movement.

According to Allison et al. (3), subsoils can fix much more ammoniumnitrogen than surface soils. They also found that considerable moist fixation can occur in soil if the predominant clay mineral is illite or vermiculite, and these values are increased by drying and heating.

Nitrogen fertilizer losses as gaseous N compounds must be evaluated before the effects of continuous applications of nitrogen fertilizers on the

soil can be accurately measured. Losses of fertilizer in irrigation or rain water run-off must also be determined. According to Moe et al. (54), applied ammonium nitrate fertilizer, because of its high ionization, is adsorbed and held near the soil surface. A more non-ionized N source, such as urea, is carried further down into the soil with the first increment of rainfall and therefore is less subject to surface run-off loss. Urea is rapidly hydrolyzed to ammonia in the soil and little loss as nitrate-N occurs. Moe further states that run-off losses of surface applied N fertilizers are greatest when the fertilizer is applied to very wet soils or to fallow soils having a surface seal.

There is widespread agreement among investigators that ammonia can be lost readily from the soil by volatilization (39, 48, 50, 86). Allison (4) states that ammonia volatilization losses from the soil, under suitable conditions, may amount to 25% or more of the ammonia added or formed. Kresge and Satchell (44) observed that no ammonia was volatilized from ammonium nitrate as long as the soil pH did not rise above neutral. Losses increased markedly at pH values above 7.0. Losses from alkaline soils containing much NH₃ near the surface increased as soils became dry. There were also high ammonia losses from soils with low CEC.

Jewitt (39) found that substantial quantities of N added in the form of ammonium sulfate to alkaline soils was lost. According to Martin and Chapman (50) evaporation of water was necessary for appreciable volatilization of ammonia from the soil to occur. Losses increased as temperatures became higher.

Meyer et al. (53) also observed that NH₃ losses were greatest when such fertilizers as urea, urea-ammonium nitrate solutions, ammonium sulfate,

and ammonium nitrate were applied to neutral to alkaline soils under conditions of limited rainfall. Losses were magnified by the presence of crop residue on the soil surface and accentuated by cool temperatures which limited nitrification. Fuller (29) observed that volatilization losses from ammonium-containing fertilizers under aerobic conditions are greatly reduced by placing the fertilizers below the surface of the soil. He also stated that losses of volatilized ammonia formed by mineralization of organic compounds in the soil are rarely significant.

Yaalon (92) found that ammonia concentrations in monthly composite rain water samples collected in Israel showed marked dependence on soil temperatures, increasing significantly as soils warmed up in the spring. He attributed this to the release of pedogenic ammonia from calcareous soils at the beginning of the warm spring. Losses from ammonium-containing fertilizers were also considered a contributing factor.

According to Robinson (73), ammonification decreased when soil moisture level decreased below the permanent wilting point (PWP), but still took place quite actively at low soil moisture levels. Since nitrification was retarded at low moisture levels, there was a consequential build up of ammonium-N. After 8 days of incubation at 35°C, little nitrate-N was found in soil that was air dry prior to incubation, whereas nearly 10 ppm ammonium-N were found. At moisture levels below PWP, ammonium-N started to accumulate, while nitrate-N levels remained constant.

Nitrogen Losses Due to Denitrification

Bremner and Shaw (14) described denitrification as a biological process whereby nitrates are reduced to gaseous nitrogen compounds such as nitrous oxide and molecular N. They concluded that denitrification occurs only when the oxygen supply in the soil is restricted. Arnold (8) and Jones (41) support this viewpoint.

Wagner and Smith (85) determined that nitrous oxide (N₂O) may account for a large part of the nitrogen loss under denitrifying conditions. They reported up to 85% of the applied urea nitrogen was lost from a treatment in one study. They also noted that clay soils normally lose more nitrogen due to limited aeration that favors denitrification.

Allison (4), who has stated that nitrogen balance sheets often do not account for all the nitrogen originally present in or added to well-aerated soils, verified that soil nitrogen can be lost as nitric oxide, nitrous oxide, NH₃ and N₂. Soils which are approaching saturation with moisture rapidly release large amounts of their available nitrogen as nitrous oxide. At lower moisture contents, very slow evolution of the gas can take place (8).

Bremner and Shaw (14), Clark et al. (22), Meek et al. (52), and

Patrick and Wyatt (63) all described conditions that enhance denitrification
in the soil. The rate of denitrification increased with a rise in pH,

temperature, and moisture content (14). The instability or reactivity of
nitrous acid in soils is primarily responsible for the large volatile losses
of N commonly observed during the course of the mineralization and nitrification processes in many well-aerated soils (22).

Broadbent (16), Broadbent and Stojanovic (17), and Jannson and Clark (37) all found that appreciable denitrification can occur under apparently aerobic conditions. Kefauver and Allison (43) observed nitrite reduction under aerobic conditions.

Work by Meek and MacKenzie (51) tended to eliminate the accumulation of nitrite as a factor causing large losses of gaseous nitrogen from alkaline soil under aerobic conditions. In another study (74) little N_2O was found under alkaline soil conditions, but N_2O production exceeded N_2

production under acid conditions. Fine textured soils showed a tendency to lose more N_2 than N_2O , and ammonium nitrate treatment favored evolution of nitrous oxide. Smith and Clark (76) found no loss of nitrogen as either nitric oxide or nitrogen dioxide from moist, aerobic soil. Nitrite can accumulate in soil as a lag phase in nitrification under alkaline conditions unfavorable to Nitrobacter spp. (56, 85).

Loewenstein et al. (48) showed that denitrification and nitrification in the soil proceeded simultaneously. They speculated that nitrates produced in the aerobic soil area moved to oxygen-poor regions and became subject to denitrification. Aerobic areas in the soil may have become anaerobic as a result of rapid oxygen consumption or because of concurrent CO₂ evolution by the soil microflora. Broadbent and Clark (18) have noted that the reduction of nitrite under aerobic conditions as a factor of denitrification may be of considerable importance, since nitrite may be formed in soils either from reduction of nitrates when oxygen is lacking or from oxidation of ammonium when oxygen is adequate.

Effects of Phosphorus Application on Soil Chemical Properties

Applications of phosphorus fertilizer over a continuous period of time also have been shown to have an effect on the chemical properties of soils. Several researchers have found that inorganic phosphorus is quite immobile in the soil and does not move far from the point of application (40, 68, 82). Soluble P rarely moves more than 2 or 3 cm from a fertilizer granule before reaction with soil components essentially stops further movement. Repeated applications will result in slow downward movement to 10-15 cm. This limited movement of P in the soil indicates the need for initially placing fertilizer P in the proper position for maximum effectiveness.

According to Taylor (82), essentially no phosphorus moves downward by water percolation because most fertilizer P is converted to water-insoluble forms rapidly after application to the soil. Phosphorus immobility is due to the elements' strong adsorption by finely divided mineral soil particles of the clay fraction (40, 82).

When soluble phosphate fertilizers are added to calcareous soils, they react with CaCO₃ through rapid monolayer sorption on CaCO₃ surfaces, and, at high phosphate concentrations in the vicinity of the fertilizer particles, the precipitation of dicalcium phosphate and tricalcium phosphate or even apatite-like compounds (24).

Phosphorus moves in a calcareous soil primarily in organic forms (33).

Addition of a microbial energy source to the soil increases the movement of organic P significantly, but no increase in the movement of inorganic phosphorus is obtained. Higher plants are unable to utilize the organic phosphorus found in the soil solution (33).

Bingham and Garber (11) found that on alkaline soils heavy phosphorus applications of 1,000 kg/ha resulted in acute copper deficiency, but both copper and zinc solubilities were increased by excessive P fertilization. These heavy treatments reduced the availability of molybdenum in alkaline soils. Zinc uptake was the same regardless of level of phosphorus treatment, whereas manganese, iron, and boron availability was increased with the heavy P treatment.

Effects of Fertilizer on Zinc Availability in Calcareous Soils

Most zinc disorders in plants occur in calcareous soils. Residually acidifying fertilizers applied to calcareous soils have been shown to increase the availability of several micronutrients (11, 84). Zinc availability is often a problem on calcareous soils, owing to the ability

of calcium carbonate to transform zinc to sparingly soluble compounds (57). Zinc may be fixed at the surface of CaCO₃ particles. The greater the level of CaCO₃ of the clay fraction (carbonate clay) the less available the soil zinc. Jurinak and Bauer (42) also described this unavailability of zinc in calcareous soils as being due to zinc adsorption by carbonates or precipitation of zinc hydroxide or carbonate. Some soils retain Zn²⁺ as strongly as zinc silicate, and more strongly than zinc hydroxide or carbonate (83).

METHODS AND MATERIALS

The study consisted of a randomized complete block design with 5 replications of 36 treatments. Only 3 replications of 12 treatments were sampled since previous corn yields indicated no significant response to applied potassium or to an additional application of 19 kg/ha of P to half of each plot (Table 1).

The experimental area was located east of Tribune in Greeley county on the Experiment Station's irrigation site. The soil of the experimental area is a Richfield silt loam, which is a fine, montmorillonitic, mesic Typic Argiustoll. It is a deep, well-drained, nearly level (0-1% slope) soil of the uplands, developed from calcareous loess. The Richfield soil has a dark grayish-brown silt loam surface layer over grayish-brown silty clay loam subsoil, which grades to lighter colored, very friable calcareous silty clay loam at 10-18 inches. Previous soil analysis data from the experimental site are indicated in Table 2.

Soil Sampling. The first replication was sampled on January 23, 1972, and the other two replications were sampled on February 20-21. Soil core samples were taken with a truck-mounted hydraulic soil probe (Figure 1). Soil core samples were taken with a 5 cm diameter, 1.2 m slotted stainless steel sampling tube. The sampling tube was inserted into the soil to a depth of approximately 1 meter and removed with an intact soil core inside. The sampling tube was removed from the drive head and placed on a channel board with a meter stick mounted on one side of the channel. The intact soil core was removed from the sampling tube with a wooden dowel and placed in the channel board for sectioning (Figure 2). The soil core was cut into 5 cm increments for the first meter. The coring process was

Table 1. Effects of nitrogen, phosphorus, and potassium on the yield 1 of irrigated corn.

1961-73 avg.	3,575 6,147 7,715 7,965 8,593 8,655	3,700 6,523 8,216 9,283 9,659	3,763 6,397 8,091 9,722 9,784
1968-73 avg.	4,328 ² /6,962 8,718 9,659 10,349	4,328 7,401 9,032 10,098 10,537 10,725	4,265 7,025 8,844 10,161 10,788 10,662
196	4,140 6,962 8,028 8,028 8,593 8,781	4,265 7,401 9,032 10,035 10,600 10,537	4,265 7,401 8,718 9,910 10,537 10,662
1973	4,328 ² / 6,962 8,718 9,659 10,349	4,328 7,401 9,032 10,098 10,537 10,725	4,265 7,025 8,844 10,161 10,788 10,662
1	3,199 7,276 8,467 8,781 8,718 9,847	3,450 7,087 9,471 11,101 11,854 11,478	3,324 7,589 8,781 10,913 11,478 11,540
1970	5,457 ² / 8,467 10,537 11,227 12,356 12,105	5,457 8,906 10,349 11,227 12,230	5,206 8,906 10,412 11,854 12,230
19	5,080 8,530 9,722 9,533 10,537	5,268 9,094 10,474 11,854 11,917 12,168	5,457 9,345 10,286 11,540 12,293 12,357
1967	2,697 5,394 7,401 7,903 8,404 8,844	3,324 6,084 8,154 9,408 9,408	3,199 5,896 8,028 8,906 9,471 9,533
1964	3,450 5,268 7,715 8,091 8,279 8,216	3,450 5,645 7,401 8,216 8,279 8,844	3,261 5,331 7,213 8,342 8,844 9,157
1961	2,509 5,645 7,726 7,338 8,404	2,572 5,143 7,150 8,174 8,404	2,634 7,025 8,124 8,028 8,028
Treatment N P K kg/ha	000000	119900000000000000000000000000000000000	19 37 19 37 19 37 19 37 19 37
Tre	133 620	133 6 6 0 0 1 3 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5 5	13 13 13 13 13 13 13 13 13 13 13 13 13 1

 $\frac{1}{2}$ Corrected to 12.5% moisture, kg/ha $\frac{2}{2}$ 19 kg/ha phosphorus were added to half the plots

Table 2. General soil analysis date from the experimental area prior to the current investigation.

1 Com	naaita	complos
(COM	DOD'T FE	samples)

		Organic Matter	pН	Avail. P	Exch. K
		%		pp2m	pp2m
Corn:	1961	1.4	7.9	34	550+
Nov.,	1964 N only	1.5	7.8	13	550+
	N + P	1.4	7.8	30	550+
Nov.,	1967 N only	1.4	7.9	11	550+
	N + P	1.5	7.9	37	550+
Dec.,	1970 N only	1.7	7.7	16	550+
	N + P	1.7	7.7	32	550+

repeated until a composite core of 4 meters had been collected. The lower 3 meters were sectioned into 20 cm increments. Two cores were collected from each plot and combined into a single composite sample. Samples were placed in soil sample bags for transportation to the laboratory. Since two 20 cm increments from the duplicate cores (5 cm diameter) would not fit into a sample bag, it was necessary to split these sections along the vertical axis and retain half of each section.

Samples from the first sampling date were returned to the laboratory and placed directly in a forced air oven and dried at 55°C for 4 days. The dried soil samples were then ground with a hammer mill, screened through a 14-mesh screen, mixed and stored in sealed glass bottles. The samples from the second sampling date were frozen for approximately 1 month before being dried and ground.

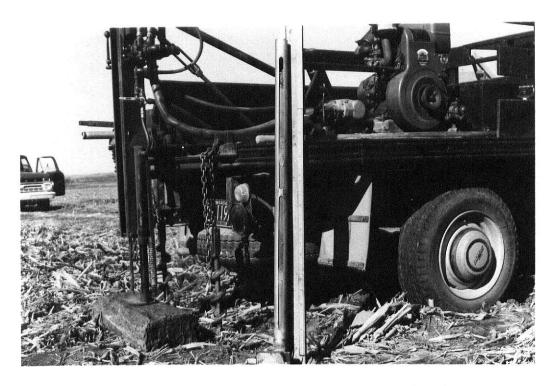


Fig. 1. Truck-mounted hydraulic soil probe used to obtain soil core samples.



Fig. 2. Method used for sectioning soil cores into desired increments.

The soil samples were analyzed for NH_4^+-N , NO_3^-N , total N, pH, available P, extractable Ca, Mg, Na and K, CEC, extractable Cu, Fe, Mn and Zn, total C, $CO_3^{2-}-C$ and organic C.

Ammonium and Nitrate Determinations. A steam distillation procedure outlined by Bremner and Keeney (15) was used for NH_{4}^{+} -N and NO_{3}^{-} -N determinations. A 4 g portion of each sample was weighed and placed in a 300 ml distillation flask along with 20 ml of 2M KCl and 0.2 g of MgO. This mixture was steam distilled and 25 ml of distillate were collected in a 5 ml aliquot of 2.0 percent boric acid-mixed indicator solution. The 30 ml mixture of indicator solution and distillate were then titrated with standard 0.00557 N sulfuric acid from a 5 ml microburet graduated at 0.01 ml intervals. Blanks of deionized water were used to correct all samples. Results were calculated in ppm NH_{1}^{+} -N.

Nitrate-nitrogen was determined on the same soil sample following ammonium-nitrogen determination by adding 0.2 g of ball-milled Devarda's alloy to the distilling flask and immediately distilling another 25 ml of distillate into 5 ml of ${\rm H_3BO_3}$ -mixed indicator solution. The sample was titrated with the standard sulfuric acid and the results calculated in ppm ${\rm NO_3}^-$ -N.

Total Nitrogen. Total soil nitrogen, excluding NO₃-N, was determined on a 1 g soil sample by the macro-Kjeldahl procedure described by Jackson (36).

¹Indicator solution consisted of 20 g H₃BO₃ and 20 ml of mixed indicator dissolved in 700 ml of warm water and brought to a 1 l volume; adjusted with 0.05 N NaOH until 1 ml of mixed indicator solution added to 1 ml of distilled deionized H₂O gave a bluish-green color. Mixed indicator solution made up by dissolving 0.99 g of bromocresol green and 0.66 g of methyl red in 1 l of absolute ethanol.

pH. Soil pH was determined by adding 5 ml of distilled water to 5 g of soil, stirring the mixture, and measuring the pH after 20 minutes by standard potentiometric methods.

Available Phosphorus. Phosphorus was determined as weak acid extractable P using the Bray-1 extracting solution with subsequent colorimetric determination. The extractant used was 0.03 N NH₄F and 0.025 N HCl. The color development of the extracted solution was by reduction of the solution with 1-amino-2-napthol-4-sulfonic acid to allow formation of a phospho-molybdate-complex.

Extractable Cations. Extractable Ca, Mg, Na, and K were determined by using a centrifuge extraction procedure with 0.1 N ammonium acetate as described in Jackson (36). A correction procedure for the solubility of Ca and Mg carbonates in the leachates was also used. The supernatant was evaporated to dryness on a hot plate, then dissolved and brought up to volume in 0.1 N HCl. Calcium and magnesium were determined by the use of a Perkin Elmer model 303 atomic absorption spectrophotomer. Flame photometry was used for determination of Na and K.

Cation Exchange Capacity. Cation exchange capacities were determined by washing the soil sample from the extractable cations procedure with methanol until all excess ammonium acetate was removed. The sample was transferred to a 800 ml Kjeldahl flask with 300 ml deionized water, 10 ml of 50% NaOH and mossy zinc. Ammonia was distilled and the distillate collected in 30 ml of boric acid-methyl purple indicator until 150 ml of distillate were collected. The 175 ml mixture of indicator solution and distillate were then titrated with standard 0.0714 N H₂SO₄. Cation exchange capacities were calculated and expressed as meq/100 g soil.

Micronutrients. Extractable Cu, Fe, Mn, and Zn were determined by using the DTPA-TEA extraction procedure (20, 47). Ten grams of soil were placed in a centrifuge tube and 20 ml of DTPA extracting solution were added. The tubes were covered with plastic stoppers and shaken on a wrist-action shaker for 2 hours. The samples were then centrifuged for 3 minutes and filtered into polyethylene bottles. Extractable Cu, Fe, Mn and Zn were determined by atomic absorption spectrophotometry.

Total carbon was determined on samples from 4 treatments by the use of a Leco carbon analyzer through the courtesy of the Department of Crop and Soil Sciences, Michigan State University (6, 10, 81). One hundred milligrams of finely ground soil (80 mesh) were weighed into a special ceramic crucible and one scoop (approximately 1 gram) each of iron chips and tin accelerators were added. The crucible was then placed in a combustion tube of an induction furnace through which 0, was being passed. The sample was combusted at a temperature of over 1670°C with the carbon in the sample being oxidized to CO2. The gas mixture was passed through (1) a dust trap to filter out the solid tin and iron oxides, (2) a sulfur ${\rm trap\ containing\ MnO}_2\ {\rm to\ absorb\ sulfur\ gases\ which\ may\ have\ been\ oxidized}$ during the combustion of the sample, and (3) a heated catalyst to convert any CO formed to CO2. Moisture was removed from the gas mixture before it entered the analyzer by an anhydrone trap. After combustion and passing through the purification train, the gas mixture was passed into a cylinder housed in a temperature-controlled oven (45°C) in the analyzer. The thermal conductivity of the mixture in the cylinder was measured by a thermistortype thermal conductivity cell. The output of the thermal conductivity cell was read on a special DC digital voltmeter as percent carbon.

Carbonate-carbon was determined using the acid-neutralization method outlined in Black (7). Five grams of soil were placed in a 150 ml beaker and 50 ml of 0.544 N standarized HCl were added. The beaker was then covered with a watchglass, placed on a hot plate, and boiled gently for 5 minutes at 150°C. After cooling, the contents were filtered and washed with 50 ml of deionized water. Five drops of phenolphthalein indicator were added to the filtrate and it was titrated to the endpoint with 0.251 N standardized NaOH. Carbonate-carbon was then calculated and expressed as percent carbonate-carbon.

Approximate organic carbon percentage for each sample analyzed for total carbon was determined by subtracting percent carbonate-carbon from percent total carbon (5).

Statistical analysis was completed on the data using the least squares method. All results were reported at the 5% level of significance.

RESULTS AND DISCUSSION

Detailed soil analysis results are presented in appendix tables 1-47. Tables in the text of this chapter showing mean values are averaged across nitrogen rates, phosphorus rates, or depth, whichever is appropriate concerning the specific treatment or depth effect demonstrated.

Ammonium-nitrogen

Ammonium-nitrogen accumulations in the soil of the experimental area were exceptionally high, especially in samples collected at the later sampling date. Some samples had ammonium-nitrogen concentrations that exceeded 200 ppm. There was a trend for ammonium-N concentration to increase with added nitrogen, but this was not significant at the 0.05 level of significance (Table 3). Ammonium-nitrogen was significantly higher where 19 kg/ha applications of phosphorus had been used. There was a significant decrease in ammonium-N with depth as shown in Figure 3. Little downward movement of ammonium-N occurs since it is tightly adsorbed by the soil colloids and is not susceptible to leaching. However, appreciable concentrations of ammonium-nitrogen were present even at depths of 400 cm (Table 4).

The high concentrations of ammonium-N indicated possible contamination by atmospheric refrigerant ammonia during cold storage. This source of contamination would only have affected samples collected at the later date, since the first set of soil samples were not frozen before being dried. Since adsorption of NH₄ and other cations is related to soil CEC, this particular soil would be expected to have fairly uniform ammonium-N concentrations throughout the profile if contamination was a problem, since CEC did not vary appreciably with depth. Lack of significant variation of CEC with depth raises some question concerning ammonia contamination.

Table 3.	Mean	values f	for	ammonium-N,	nitrate-N,	total	N,	pH,	and	available
	P as	affected	l by	y N rate or	P applicati	on.		4 5 5		

Treatment kg/ha	ammonium-N ppm	$\frac{\texttt{nitrate-N}}{\texttt{ppm}}$	total N ppm	рН	avail. P
0 N	50.90	2.62	737	8.17	8.53
45 N	52.74	2.23	752	8.12	8.54
90 N	54.98	2.72	728	8.14	7.98
134 N	55.15	4.02	751	8.13	5.52
179 N	57.35	4.55	737	8.12	6.55
224 N	56.78	6.96	749	8.11	5.75
LSD	NS	0.45	NS	0.01	1.24
0 P	51.18	3.66	751	8.14	4.68
19 P	58.13	4.04	733	8.12	9.62
LSD .05	2.75	0.26	15	0.01	0.71

Environmental and soil conditions at time of sampling may have been conducive for mineralization of soil N without subsequent formation of nitrate-nitrogen through the nitrification process. Reduced aeration, cold temperatures, and high soil pH all adversely affect nitrification to a greater extent than ammonification.

Since mineralization proceeds most rapidly in well-aerated, warm soils with plenty of basic cations present, this process may have been stimulated while the moist samples were being dried prior to preparation for analysis. Ammonification has been shown to take place at very low soil moisture contents, and at fairly high temperatures (50-70°C). Therefore, ammonification could have been occurring without subsequent nitrification before the soil became too dry, resulting in ammonium-N accumulation.

The increased ammonium-N concentrations accompanying added phosphorus indicate that this increased availability of P may have been utilized by heterotrophic soil organisms to stimulate mineralization of organic nitrogen compounds and increased production of ammonium-N.

Table 4. Mean values for ammonium-N, nitrate-N, total N, pH, and available P as affected by depth.

Depth, cm	ammonium-N	nitrate-N	total N	<u>рН</u>	avail. P
	ppm	ppm	ppm		\mathtt{ppm}
0-5	108.27	9.22	1258	8.01	16.11
5-10	117.20	13.48	1180	7.98	13.10
10-15	110.92	11.52	1141	7.93	8.01
15-20	103.70	7.99	1161	7.93	7.13
20-25	111.37	5.59	1116	7.98	5.72
25-30	106.07	4.17	1052	8.01	3.73
30-35	99.11	3.01	963	8.08	2.22
35-40	91.20	3.11	899	8.08	1.16
40-45	84.57	2.94	844	8.10	
45-50	81.04	3.04	797	8.11	
50-55	73.60	3.23	772	8.13	
55-60	68.03	3.15	748	8.14	
60-65	61.91	2.84	696	8.15	
65-70	57.53	3.13	689	8.16	
70-75	54.51	3.15	694	8.16	
75-80	55.28	2.84	709	8.18	
80-85	51.77	2.75	703	8.17	
85-90	48.62	2.90	672	8.18	
90-95	48.18	2.65	679	8.18	
95-100	44.79	2.84	695	8.18	
100-120	34.77	2.23	575	8.18	
120-140	30.96	2.43	566	8.18	
140-160	27.21	2.56	564	8.18	
160–180	22.37	2.60	541	8.19	
180-200	19.96	2.72	533	8.16	
200-220	20.21	2.72	531	8.16	
220-240	18.60	2.77	510	8.17	
240-260	22.16	3.18	546	8.16	
260-280	20.10	3.06	558	8.17	
280-300	20.73	3.43	586	8.16	
300-320	21.12	2.92	609	8.17	
320-340	19.34	2.92	570	8.19	
340-360	20.24	2.73	624	8.22	
360-380	19.05	2.43	591	8.24	
380-400	18.30	2.49	607	8.25	2 9 2
LSD.05	11.50	1.08	64	0.03	1.43

Some fixed ammonium-N could have been released from the soil organic matter fraction during steam distillation if it were somewhat water soluble. Physically sorbed ammonia also could have been released during the steam distillation procedure. This soil probably had high ammonium-N fixing capacity, since expanding clay minerals like montmorillonite have large

adsorption capabilities. Ammonium-N fixation by the organic fraction of mineral soils increases with pH. Clays saturated with divalent ions also generally have higher ammonium contents than clay saturated with monovalent ions.

Nitrate-nitrogen

Soil nitrate-nitrogen concentrations were much lower than ammoniumnitrogen levels. Individual sample values ranged from none to 27 ppm.

Nitrate-nitrogen decreased with depth as shown in Figure 4, and both nitrogen
and phosphorus treatments had a significant effect on the accumulation of
nitrate-nitrogen (Table 3). The pattern of accumulation indicates nitrates
are moving downward uniformly, as there are no large peaks of nitrate
accumulation in the soil profile. There is an observable increase in
nitrate-N at the 300 cm depth, and then concentration decreases again
(Table 4).

The increased accumulation of nitrate-nitrogen with applied N indicates that at the high rates some nitrogen fertilizer is not being utilized by the corn crop. The effect of P on nitrate-nitrogen may be secondary in that phosphorus has a beneficial effect on soil organisms that enzymatically hydrolyze organic nitrogenous compounds and proteins and subsequently manufacture ammonia which is oxidized to nitrate-nitrogen. If this is the case, then total soil nitrogen and organic carbon, which are the main constituents of organic matter, should decrease with added P. Subsequent data show that this is so.

The effect of phosphorus on nitrate-nitrogen was most prevalent at the low nitrogen rates. The results of available nitrogen analyses indicate that more research is needed on this calcareous soil in regard to immobilization, mineralization, nitrification, and denitrification of soil nitrogen.

Total Nitrogen

Total soil nitrogen, excluding nitrate-nitrogen, decreased dramatically with depth, as illustrated in Figure 5. Concentrations were lowest at the 200-240 cm depth. Nitrogen had no effect on total N (Table 3), while phosphorus decreased the amounts of total nitrogen in the soil at all but the two highest nitrogen rates. This trend was probably explained by the fact that at low N rates, added phosphorus stimulated grain formation in preference to forage production, so that less organic nitrogen was returned to the soil as crop residue after grain removal. As nitrogen fertilization rates increase, grain yields reach a plateau and more phosphorus, as well as other plant nutrients, are incorporated into vegetative material which is returned to the soil following harvest. Total nitrogen content ranged from 130-1680 ppm in individual samples.

pН

The effects of nitrogen and phosphorus fertilizers on soil pH, though small, were significant as shown in Table 3. The effect of the nitrogen fertilizer on decreasing pH can be explained by the residually-acidifying effect of ammonium nitrate. The higher the N rate, the greater the potential acidity. Phosphorus applications at the lower nitrogen rates (0, 45, 90, and 134 kg/ha) decreased soil pH, but had no effect when applied with heavy nitrogen treatments. There was a significant increase in pH with depth, as shown in Figure 6, which implies that surface pH changes have not extended into the lower part of the soil profile. The large amounts of calcium carbonate in this soil indicate that no detrimental decreases in pH can be expected at the present fertilization rates. In fact, a decrease in pH would probably be beneficial as most micronutrients are more readily available at pH levels lower than those present in this soil. Soil pH varied from as low as 7.8 near the surface up to 8.4 at the 400 cm depth.

Available Phosphorus

Weak Bray extractable phosphorus content in the surface 5 cm was quite variable, ranging from 6 ppm with no applied P up to 37 ppm with 19 kg/ha of applied phosphorus as triple-superphosphate. Available phosphorus decreased as nitrogen rates increased. Added nitrogen probably stimulated P uptake by increasing crop growth. Much more available P was present with the P treatment as compared to plots that received no phosphorus fertilizer, as was expected. At low N rates, added P increased soil available P to a much greater extent than at higher N rates, indicating that nitrogen is limiting growth and P uptake at these low nitrogen rates. At higher nitrogen rates, phosphorus levels indicate that it is the limiting nutrient as shown in Figure 7. There was a marked decrease in available phosphorus with depth (Table 4). As phosphorus is relatively immobile, little downward movement of applied P would be expected. Previous data from this study indicate that available phosphorus did not tend to increase with time when 19 kg/ha was added but that the level in the soil remained at a fairly constant, adequate level. When no phosphorus was added, there was a trend for soil phosphorus to decrease with time (Table 2).

Extractable Calcium

Nitrogen applications had a significant effect on soil extractable calcium as shown in Table 5. There was a decrease in calcium with added N when no phosphorus was applied. With applications of nitrogen and phosphorus, no meaningful trends were apparent. Figure 8 indicates the effect of depth on calcium content. The area of lowest concentration (0-20 cm) is probably due to uptake and removal by crops and to loss of part of the calcium pool in the form of CaCO₃ through acidifying effects of organic acids and applied ammonium nitrate. Some calcium also probably leached downward and is

THIS BOOK CONTAINS NUMEROUS PAGES WITH DIAGRAMS THAT ARE CROOKED COMPARED TO THE REST OF THE INFORMATION ON THE PAGE. THIS IS AS RECEIVED FROM CUSTOMER.

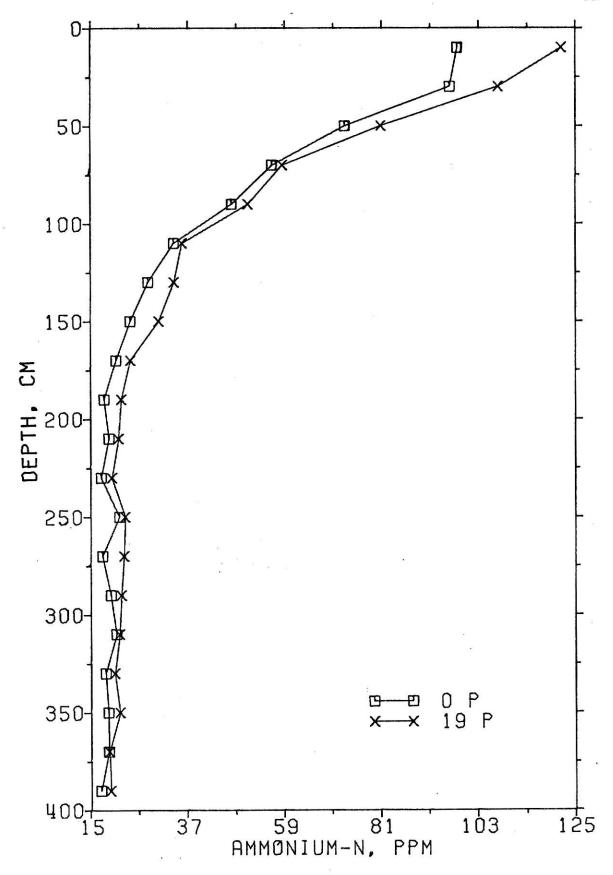


Fig. 3. Soil ammonium—N as affected by nitrogen and phosphorus treatments.

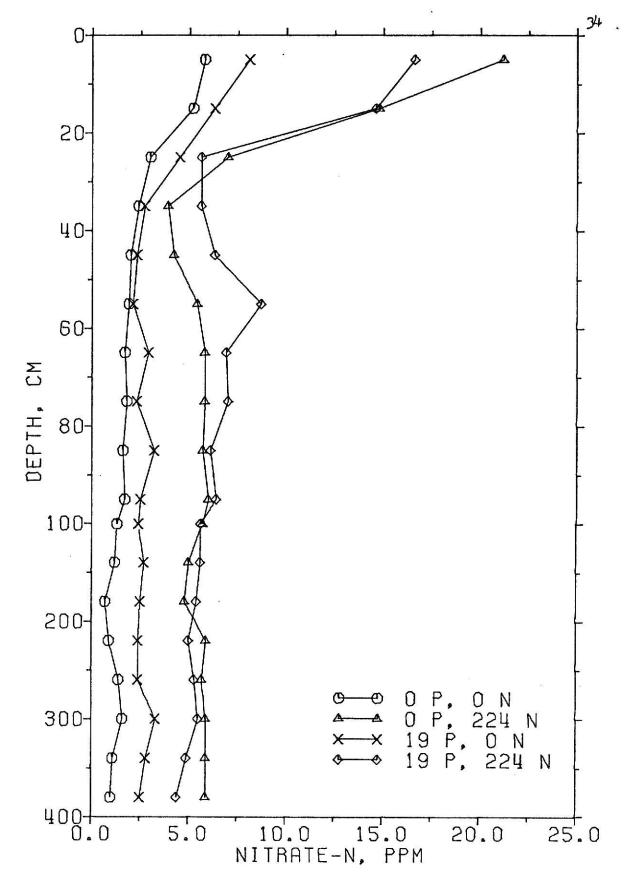


Fig. 4. Soil nitrate-N as affected by nitrogen and phosphorus treatments.

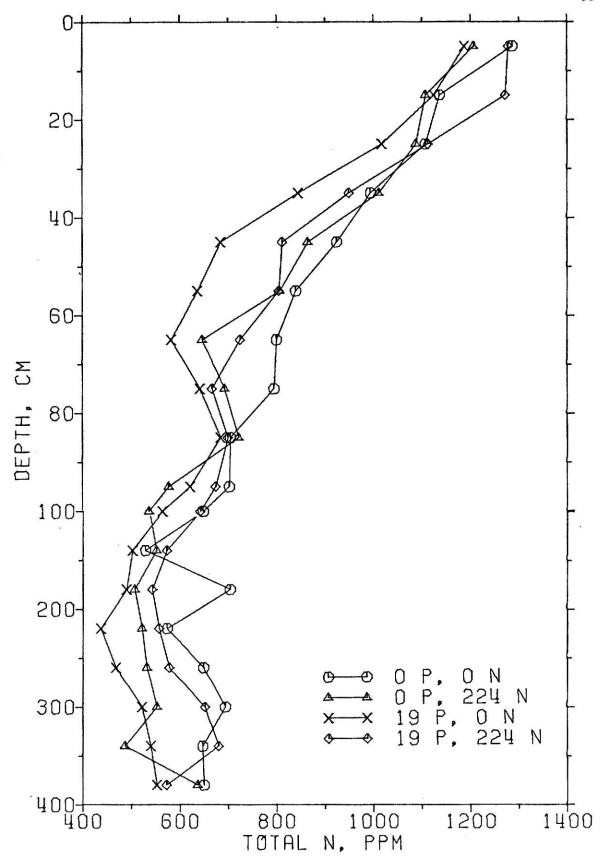


Fig. 5. Total soil nitrogen as affected by nitrogen and phosphorus treatments.

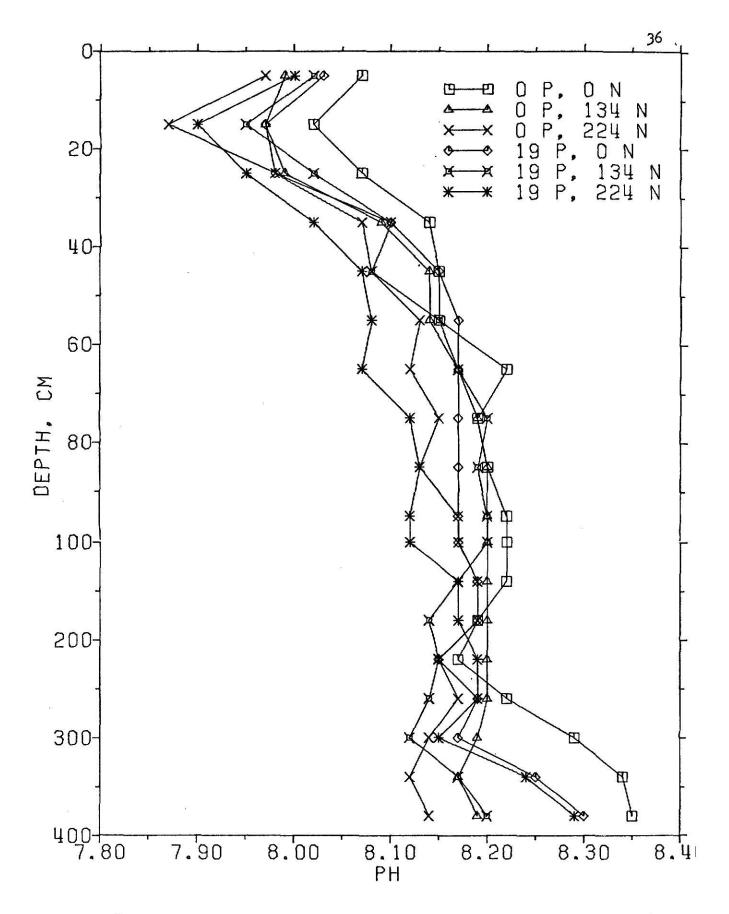


Fig. 6. Soil pH values as affected by nitrogen and phosphorus treatments.

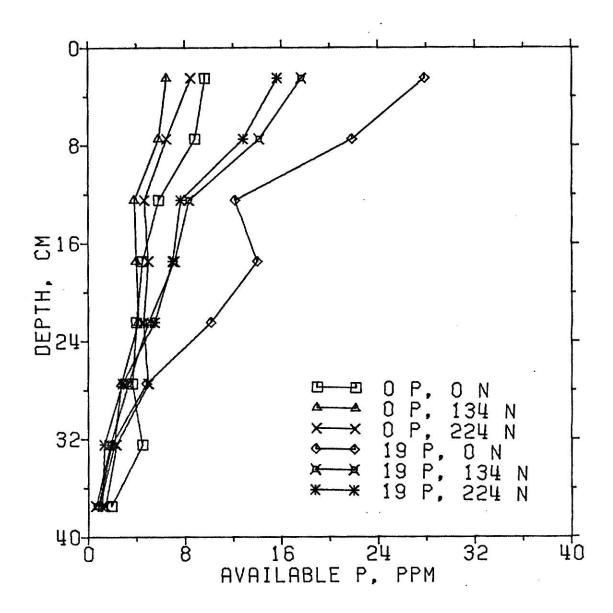


Fig. 7. Weak Bray extractable P as affected by nitrogen and phosphorus treatments.

Table 5. Mean values for calcium, magnesium, sodium, potassium, and CEC as affected by N rate, P application, or soil depth.

Treatment	calcium	magnesium	sodium	potassium	$\frac{\text{CEC}}{\text{meq}/100}$ g
kg/ha	ppm	ppm	ppm	ppm	
0 N	15341	736	88.7	570	22.05
134 N	16039	711	83.3	557	21.75
224 N	14711	672	95.4	517	22.17
LSD .05	912	16	7.3	17	0.19
0 P	15106	705	92.9	540	22.05
19 P	15621	708	85.4	556	21.93
LSD _{.05}	NS	NS	6.0	14	NS
Depth, cm					
0-5 5-10 10-15 15-20 20-25 25-30 30-35 35-40 40-45 45-50 50-55 55-60 60-65 65-70 70-75 75-80 80-85 85-90 90-95 95-100 LSD .05	8755 8749 8698 8865 11706 15836 17834 16594 16459 17855 18495 18583 17916 17905 17854 17928 17525 17045	539 526 519 509 565 624 660 649 652 685 712 753 788 806 825 854 857 860 877 867	84.9 91.4 79.0 81.3 92.1 99.1 99.1 92.9 91.8 85.4 90.8 85.4 91.8 NS	594 563 525 510 585 464 435 4475 475 507 553 655 684 764 764	23.22 23.30 23.60 23.27 23.56 23.67 22.83 22.12 21.48 21.25 21.08 21.15 21.06 21.28 21.01 21.25 21.01 21.25 21.26 0.49

responsible for the zone of increased extractable calcium accumulation at the 60-70 cm depth. A correction procedure for free calcium and magnesium carbonates was used but extractable calcium concentrations still exceeded 20,000 ppm lower in the profile. The pattern for extractable calcium concentration is similar to that for carbonate carbon, indicating the close relationship between these two ions in this calcareous soil.

Extractable Magnesium

There was a significant decrease in soil extractable magnesium with applied nitrogen as shown in Table 5. This was possibly due to removal by the growing crops and to some downward leaching of soluble MgCO₃ though no zone of accumulation was apparent (Figure 9). Magnesium concentration increased with depth. Phosphorus application had no effect on soil extractable magnesium. Magnesium concentrations ranged from 450-900 ppm.

Extractable Sodium

Extractable sodium was significantly affected by both nitrogen and phosphorus fertilization. When 19 kg/ha of phosphorus was added, sodium content increased as N rates increased as shown in Figure 10. At the 0-N rate, the addition of phosphorus led to a decrease in extractable sodium content. Depth had no effect on sodium, indicating that plant uptake of this element is low and that it is associated with the mineral fraction of the soil and not the organic fraction. Sodium concentration in the soil was erratic and showed no pattern (Table 5).

Extractable Potassium

Extractable soil potassium was affected by nitrogen, phosphorus, and depth (Table 5). Potassium concentrations decreased with applied nitrogen, indicating increased plant uptake as yields increased. Extractable soil potassium increased when phosphorus was applied. This may have been due to the slight decrease in pH due to applied phosphorus. The decrease in alkalinity may have increased the availability of soil K. The addition of phosphorus also tended to increase soil extractable calcium, though the increase was non-significant (0.05). Since Ca⁺⁺ tends to replace K⁺ on the soil exchange complex, this increase in extractable Ca⁺⁺ would therefore result in an increase in extractable potassium.

The general decrease in extractable K at the 30-60 cm depth is likely due to uptake by plant roots. Since much potassium is found in the vegetative portions of corn plants, considerable amounts of plant K are returned to the soil upon incorporation of plant residues. This explains the accumulation of extractable potassium near the soil surface as shown in Figure 11. Corn yield data (Table 1) show that the addition of 37 kg K/ha does not influence any significant increase in grain yield. However, there is a slight increase at the two highest N rates when comparing the N + P treatments with the N + P + K treatments. This observation, in conjunction with the significant decrease in extractable potassium with increased N applications, implies that continuous heavy applications of N and P may ultimately result in the necessity to apply fertilizer K in the future. The grand mean for extractable soil potassium was 548 ppm.

Cation Exchange Capacity

Soil cation exchange capacity was affected by nitrogen treatment, but no obvious trend was apparent (Table 5). Phosphorus tended to decrease the soil CEC, but the effect was insignificant at the 0.05 level. There was a definite decrease in CEC with depth, which was due to less organic matter below the surface 25 cm as shown in Figure 12. Soil cation exchange capacity varied from approximately 20 meg/100 g to 25 meg/100g soil.

Extractable Copper

Soil DTPA-extractable copper was not significantly affected by either nitrogen or phosphorus treatment as shown in Table 6. There was a trend for soil copper to decrease with added phosphorus at the two lower nitrogen rates. Regardless of phosphorus treatment, DTPA-extractable copper was highest when 134 kg N was applied. Perhaps additional nitrogen increased extractability but at the highest nitrogen rate higher crop uptake may have reduced the

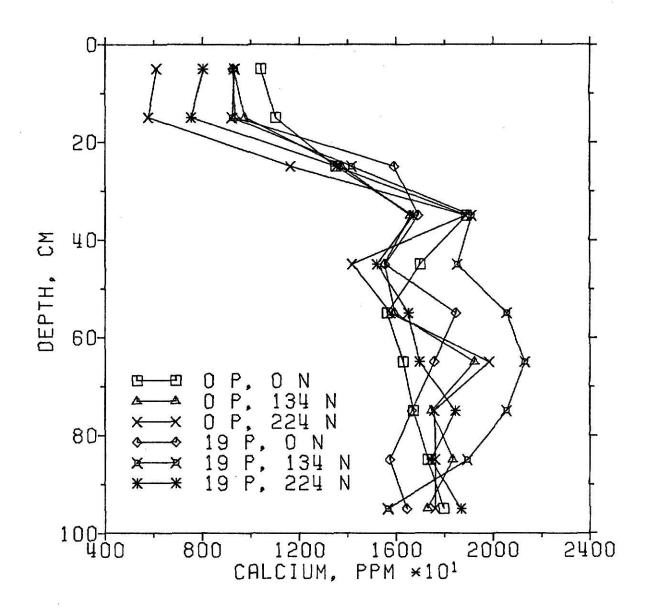


Fig. 8. Extractable soil calcium as affected by nitrogen and phosphorus treatments.

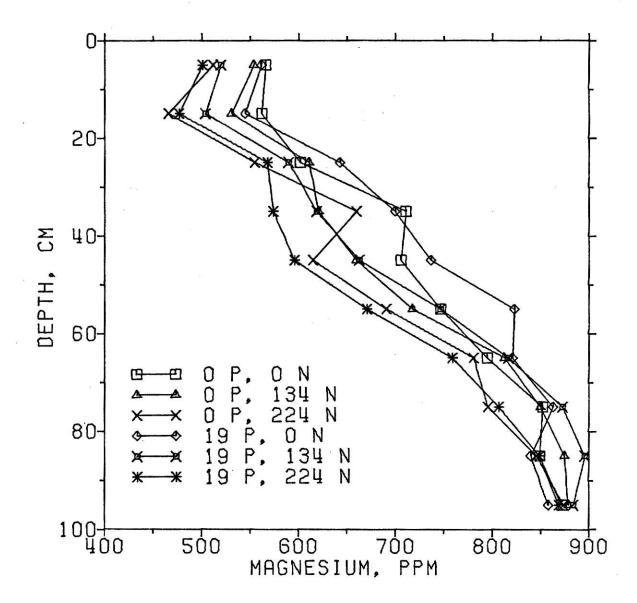


Fig. 9. Extractable soil magnesium as affected by nitrogen and phosphorus treatments.

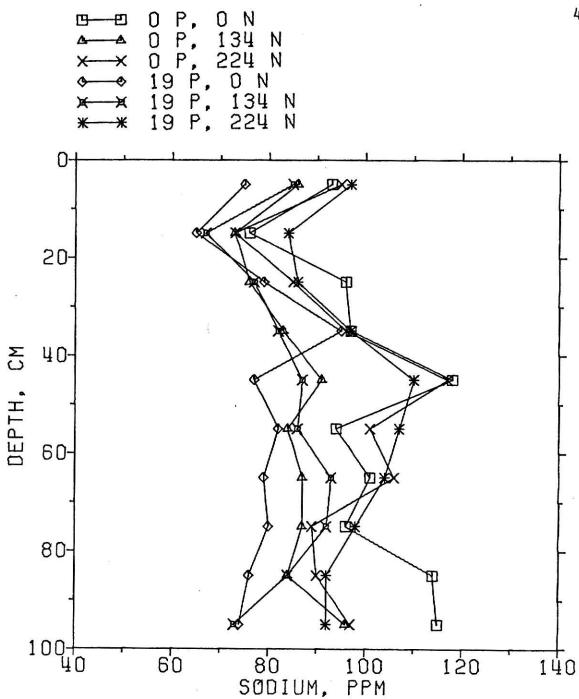


Fig. 10. Extractable soil sodium as affected by nitrogen and phosphorus treatments.

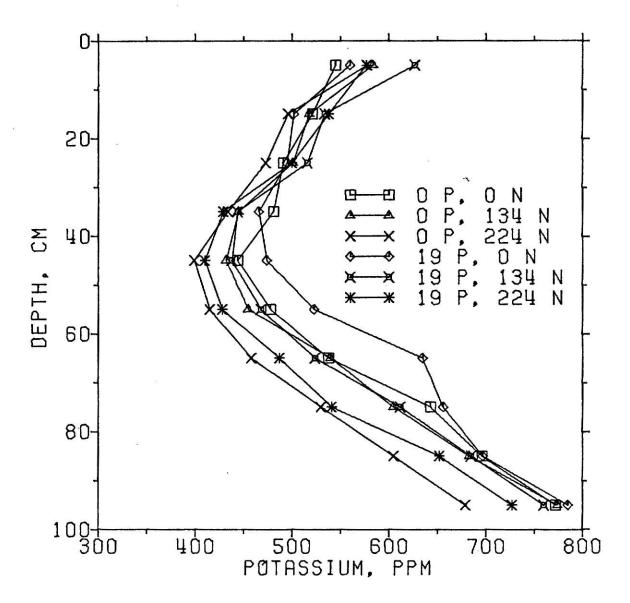


Fig. 11. Extractable soil potassium as affected by nitrogen and phosphorus treatments.

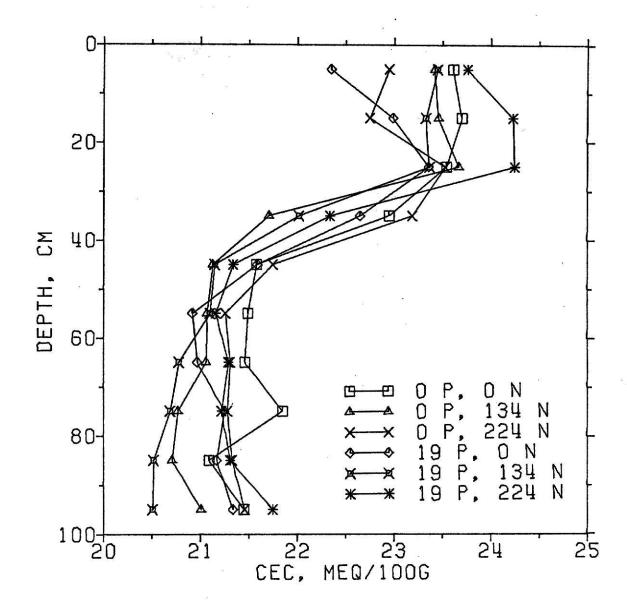


Fig. 12. Soil cation exchange capacity as affected by nitrogen and phosphorus treatments.

amount of available copper in the soil. Copper concentrations were highest at the 25-35 cm depth. At the lower depths (35-50 cm) added phosphorus resulted in decreased amounts of available copper as shown in Figure 13. Low copper concentrations in the surface 10 cm are probably due to crop uptake and removal. Soil DTPA-extractable copper was considered to be adequate with a grand mean of 0.67 ppm.

Extractable Iron

Regardless of treatment, DTPA-extractable iron decreased with depth as shown in Figure 14. Iron availability is often a problem in calcareous soils in Kansas, especially with sorghum, soybeans, pinto beans, and corm, and the decreased extractability with depth is probably due to increasing pH and soil calcium carbonate content. Nitrogen treatments had significant effects on extractable iron, and the same pattern of concentration developed as was noted with copper. The effect of phosphorus was insignificant, but some suggestion existed of an increase in extractable iron with P applications (Table 6). Extractable iron concentrations ranged from 3-7 ppm, which ranks in the medium to high category in Kansas.

Extractable Manganese L

DTPA-extractable manganese levels were high enough that no deficiency problems should exist. Manganese concentrations varied from approximately 3-18 ppm. Manganese was affected by nitrogen and phosphorus treatments as well as depth, as shown in Table 6. The increase in extractable manganese with nitrogen was probably due to secondary effects of high nitrogen rates on soil pH and CaCO₃ content near the soil surface. Phosphorus addition decreased available manganese throughout the profile. This may have been due more to increased uptake and removal by the crop than to any other factor. There was a decrease in available manganese with depth regardless of treatment as shown in Figure 15.

Table 6. Mean values for copper, iron, manganese, and zinc as affected by N rate, P application, or soil depth.

Treatment kg/ha	copper	iron	manganese	zinc
	ppm	ppm	ppm	ppm
0 N	.671	3.99	7.51	1.03
134 N	.697	5.07	7.56	1.08
224 N	.652	4.37	8.49	0.84
LSD .05	NS	0.39	0.69	NS
0 P	.679	4.36	8.40	0.99
19 P	.668	4.59	7.31	0.97
LSD _{.05}	NS	NS	0.57	NS
Depth, cm 0 0-10 10-20 20-30 30-40 40-50 LSD	.637 .661 .713 .704 .652	5.48 5.30 4.52 3.66 3.42 0.50	13.03 10.16 7.19 4.80 4.08 0.89	1.84 2.07 0.74 0.14 0.13 0.45

Extractable Zinc

DTPA-extractable zinc concentrations varied from nearly none up to 2.5 ppm. The reduction in soil zinc levels was quite pronounced below 30 cm. This is a prime reason why zinc deficiency symptoms commonly develop in crops growing on newly leveled land where the subsoil is exposed at the surface. Eleven kg/ha of zinc were applied to all plots in 1968 and 1969 to offset a low soil zinc condition. The decrease in zinc in the upper 15 cm is probably due to crop removal since that period of time (Figure 16). Available soil zinc concentration with depth seemed closely correlated with expected pH effect on zinc availability. No clear differences due to treatment are evident in data presented in Table 6.

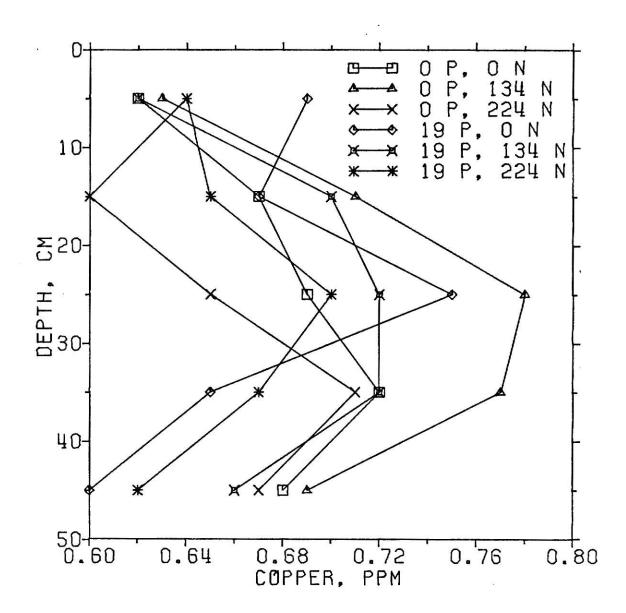


Fig. 13. DTPA-extractable soil copper as affected by nitrogen and phosphorus treatments.

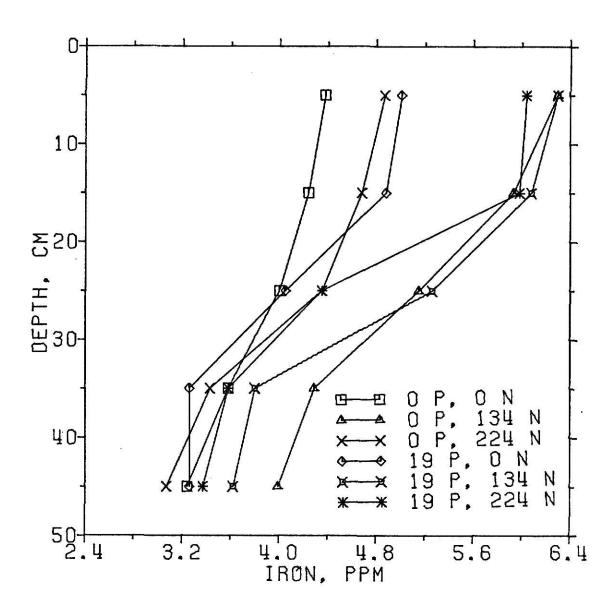


Fig. 14. DTPA-extractable soil iron as affected by nitrogen and phosphorus treatments.

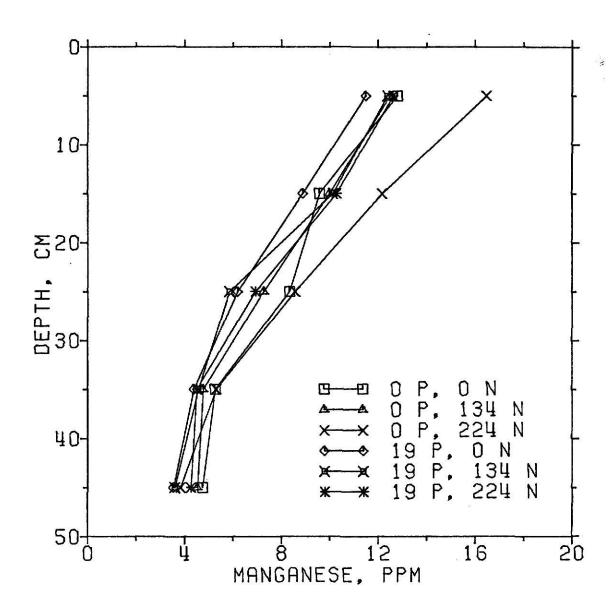


Fig. 15. DTPA-extractable soil manganese as affected by nitrogen and phosphorus treatments.

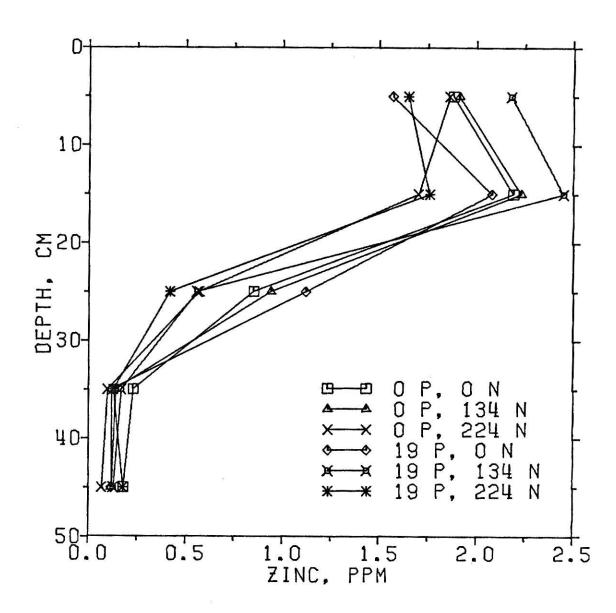


Fig. 16. DTPA-extractable soil zinc as affected by nitrogen and phosphorus treatments.

Total Carbon

In analysis for total soil carbon, carbonate-carbon, and organic carbon, only the four extreme treatments were included (Figure 17).

Applications of 224 kg N/ha decreased total soil carbon, while addition of 19 kg P/ha increased amounts of total soil carbon. However, the effects of these treatments were not significant as shown in Table 7. Total carbon concentration increased with depth, reaching a maximum near the 35-55 cm depth, and then declined. Total carbon varied from 1.0 to nearly 2.2%. Carbonate-carbon

Soil carbonate-carbon varied from 0.3-1.8%. There were large concentrations of carbonate-carbon at the 40-60 cm depth in the profile as shown in Figure 18. This accumulation is probably due to downward movement of soluble calcium and magnesium carbonates through the leaching process. No consistent differences due to treatment were apparent, but there was a general trend for carbonate-carbon to decrease with nitrogen treatment, especially in the upper 25 cm. Depth means, as shown in Table 7, indicate significant differences in carbonate-carbon concentration with profile depth. Organic Carbon

Organic carbon decreased with depth as shown in Figure 19. This was due to decreased organic matter content with depth. There was a general trend for the nitrogen applications to increase soil organic carbon, especially in the upper 20 cm, but this effect was not significant (Table 7). Organic carbon decreased with the addition of 19 kg/ha of phosphorus, but this trend was evident only under the 224 kg/ha nitrogen application.

Table 7. Mean values for total carbon, carbonate-carbon, and organic carbon as affected by N rate, P application, or soil depth.

Treatment kg/ha	total carbon	carbonate-carbon %	organic carbon
0 N 224 N LSD .05	1.54 1.51 NS	1.22 1.17 NS	0.32 0.34 NS
0 P 19 P LSD _{.05}	1.52 1.54 NS	1.17 1.22 NS	0.35 0.32 0.02
Depth, cm			
0-5 5-10 10-15 15-20 20-25 25-30 30-35 35-40 40-45 45-50 50-55 55-60 60-65 65-70 70-75 75-80 80-85 85-90 90-95 95-100 LSD	1.20 1.14 1.15 1.08 1.21 1.46 1.68 1.96 2.06 1.93 1.90 1.79 1.81 1.66 1.52 1.46 1.41 1.44 1.34	0.46 0.48 0.47 0.67 1.01 1.28 1.56 1.67 1.71 1.65 1.60 1.56 1.46 1.40 1.34 1.29 1.29 1.26 1.23 0.17	0.74 0.66 0.67 0.61 0.54 0.46 0.41 0.39 0.39 0.22 0.25 0.20 0.25 0.21 0.12 0.13 0.10 0.16 0.09 0.11 0.08

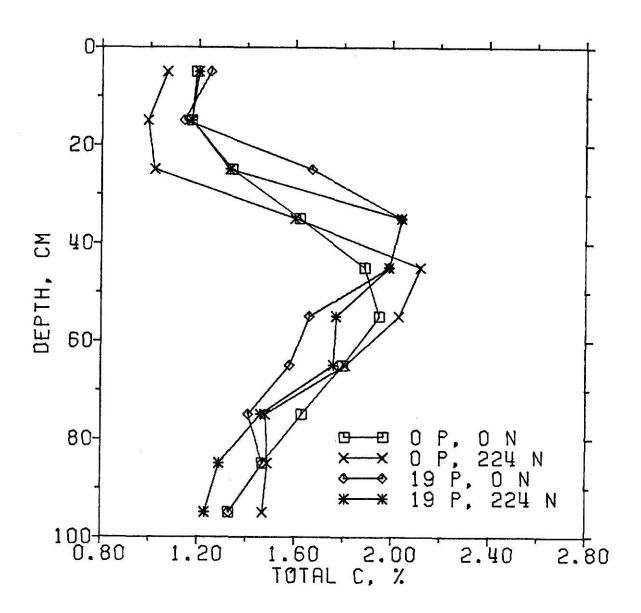


Fig. 17. Total soil carbon as affected by nitrogen and phosphorus treatments.

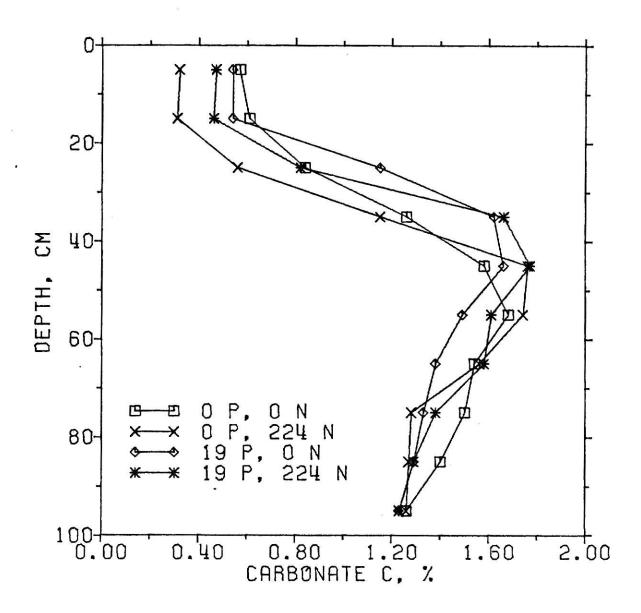


Fig. 18. Soil carbonate-carbon as affected by nitrogen and phosphorus treatments.

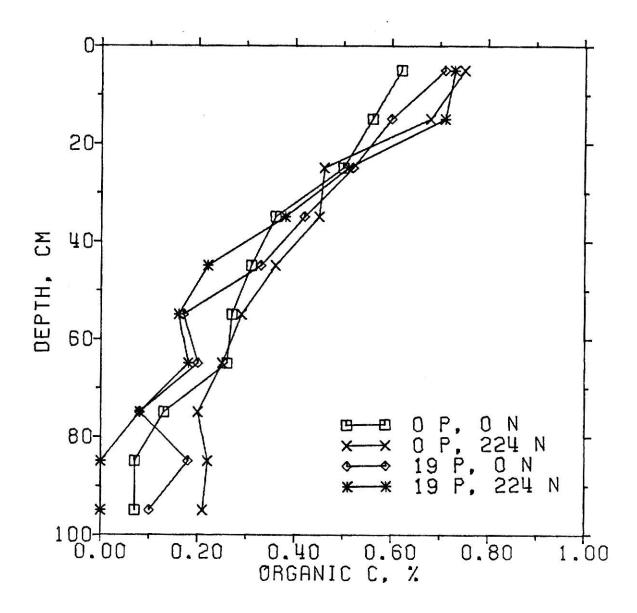


Fig. 19. Soil organic carbon as affected by nitrogen and phosphorus treatments.

SUMMARY AND CONCLUSION

The results of this study indicate that continuous applications of nitrogen and phosphorus fertilizer have had measurable effects on chemical properties of the Richfield soil.

Unaccountably high concentrations of ammonium-nitrogen indicate either possible ammonium-N contamination of the samples or build-up of ammonium-N in the samples through a microbiological process such as mineralization without subsequent nitrification. Depending upon the source of the ammonium-N, many questions concerning the nitrogen status of this soil remain unanswered. More research in this area is needed since ammonium-N is included in the Kansas test for available soil nitrogen.

Nitrate-nitrogen levels in the profile indicate little accumulation or leaching of this form of nitrogen. Nitrogen and phosphorus treatments increased the amount of nitrate-nitrogen in the soil as fertilizer rates increased. Fertilization with nitrogen certainly has not produced accumulations of NO₃-N in the soil which could pose a threat to groundwater quality. This demonstrates that adherance to recommended rates of nitrogen fertilization tends to eliminate the problem of soil nitrate-N loss due to leaching.

Nitrogen applications had no measurable effect on total soil nitrogen, but the addition of 19 kg/ha of phosphorus significantly decreased total soil nitrogen.

Both nitrogen and phosphorus applications significantly decreased soil pH, even though measured differences were very small. Since this soil contains such large amounts of calcium carbonate, no forseeable drastic change in pH should take place using the present rates of fertilization. Soil pH slowly increased with depth and reached a maximum of about 8.4 at the 400 cm depth due to the presence of free calcium carbonate.

Weak Bray extractable phosphorus decreased as nitrogen rates increased, and increased markedly when 19 kg P/ha were applied. Little movement of fertilizer phosphorus is indicated due to sharp decreases in extractable phosphorus with depth.

An interesting trend in this study was that the application of 19 kg P/ha decreased total soil nitrogen and organic carbon. This would lead one to speculate that soil organic matter content also decreased. Though this was not measured, soil CEC did decrease slightly, and surface CEC is closely related to organic matter content.

Heavy rates of nitrogen fertilizer tended to decrease extractable calcium and magnesium levels in the soil. Phosphorus had no effect on extractable calcium and magnesium concentrations. Both calcium and magnesium concentrations increased considerably with depth. The Ca:Mg ratio in this soil is quite high, exceeding 16:1 near the soil surface and approaching 27:1 lower in the profile. The soil K:Mg ratio varies from 1:1 at the surface to 1:1.5 at the 50 cm depth. This high magnesium content may explain the lack of response of corn to applied magnesium that researchers have found recently in Kansas.²

Extractable soil potassium content decreased when increasing nitrogen rates were applied, which was probably due to increased plant removal of soil K. Addition of 19 kg P/ha produced an increase in extractable soil potassium, which was probably due to the effect of phosphorus on lowering soil pH and thereby increasing availability of soil K.

²Whitney, D. A. and R. Ellis, Jr. 1973. Kansas Fertilizer Research Report of Progress. 202:124.

Cation exchange capacity, which varied from 20-25 meq/100 g soil, decreased with depth. Soil CEC was affected by nitrogen treatment, but no trend was apparent.

Higher applications of nitrogen seemed to generally increase the availability of iron and manganese, while phosphorus only affected the availability of soil manganese. Levels of all micronutrients were adequate in the upper 15 cm of soil. Zinc levels below 30 cm were quite low.

Nitrogen and phosphorus had inverse effects on all forms of soil carbon, but measurable effects were quite small and usually insignificant. Carbonate-carbon comprised from 75-80% of the total soil carbon when averaged across all depths.

The results of this study indicate that applied phosphorus effects on soil chemical properties are more significant to date than the effects of nitrogen. When high rates of nitrogen are used, decreases in soil potassium are quite apparent. This would indicate that potassium fertilization may eventually be necessary on areas where high nitrogen rates have been applied for some time, especially when large amounts of forage along with grain are removed, such as is the case with silage production.

LITERATURE CITED

- 1. Abruna, F., R. W. Pearson, and C. B. Elkins. 1958. Quantitative evaluation of soil reaction and base status changes resulting from field applications of residually acid-forming nitrogen fertilizers. Soil Sci. Soc. Amer. Proc. 22:539-542.
- 2. Adams, W. E., R. W. Pearson, W. A. Jackson, and R. A. McCreery. 1967. Influence of limestone and nitrogen on soil pH and coastal bermudagrass yield. Agron. J. 59:450-453.
- 3. Allison, F. E., M. Kefauver, and E. M. Roller. 1953. Ammonium fixation in soils. Soil Sci. Soc. Amer. Proc. 17:107-110.
- 4. Allison, F. E. 1955. The engima of soil nitrogen balance sheets. Adv. Agron. 7:213-250.
- 5. Allison, L. E. 1965. Organic carbon. <u>In</u> C. A. Black (ed.) Methods of soil analysis. II. Chemical and microbiological properties. Agronomy 9:1367-1378.
- 6. Allison, L. E., W. B. Bollen, and C. D. Moodie. 1965. Total carbon. In C. A. Black (ed.) Methods of soil analysis. II. Chemical and microbiological properties. Agronomy 9:1346-1366.
- 7. Allison, L. E. and C. D. Moodie. 1965. Carbonate. In C. A. Black (ed.) Methods of soil analysis. II. Chemical and microbiological properites. Agronomy 9:1379-1396.
- 8. Arnold, P. W. 1954. Losses of nitrous oxide from soil. J. Soil Sci. 5:116-128.
- 9. Bates, T. E. and S. L. Tisdale. 1957. The movement of nitratenitrogen through columns of coarse-textured soil materials. Soil Sci. Soc. Amer. Proc. 21:525-528.
- 10. Belo, J. A. O. 1970. Determination of total carbon by dry combustion and its relation to forms of soil nitrogen as measured in the laboratory and in the greenhouse. Ph.D. thesis. Michigan State University.
- 11. Bingham, F. T. and M. J. Garber. 1960. Solubility and availability of micronutrients in relation to P fertilization. Soil Sci. Soc. Amer. Proc. 24:209-213.
- 12. Boawn, L. C., C. E. Nelson, F. G. Viets, Jr., and C. L. Crawford. 1960. Nitrogen carrier and nitrogen rate influence on soil properties and nutrient uptake by crops. Washington Ag. Exp. Sta. Bulletin 614.
- 13. Boswell, F. C. and O. E. Anderson. 1964. Nitrogen movement in undisturbed profiles of fallowed soils. Agron. J. 56:278-281.

- 14. Bremner, J. M. and K. Shaw. 1958. Denitrification in soil. II. Factors affecting denitrification. J. Agr. Sci. 51:40-52.
- 15. Bremner, J. M. and D. R. Keeney. 1965. Steam distillation methods for determination of ammonium, nitrate, and nitrite. Anal. Chem. Acta. 32:485-495.
- 16. Broadbent, F. E. 1951. Denitrification in some California soils. Soil Sci. 72:129-137.
- 17. Broadbent, F. E. and B. F. Stojanovic. 1952. The effect of partial pressure of oxygen in some soil nitrogen transformations. Soil Sci. Soc. Amer. Proc. 16:359-363.
- 18. Broadbent, F. E. and F. E. Clark. 1965. Denitrification. <u>In</u> W. V. Bartholomew and F. E. Clark (ed.) Soil nitrogen. Agronomy 10:357-362.
- 19. Broadbent, F. E. and K. B. Tyler. 1965. Effect of pH on N immobilization in two California soils. Plant and Soil 23:314-322.
- 20. Brown, A. A., James Quick, and J. L. Eddings. 1971. A comparison of analytical methods for soil zinc. Soil Sci. Soc. Amer. Proc. 35:105-107.
- 21. Burd, J. S. 1948. Chemistry of the phosphate ion in soil systems. Soil Sci. 65:227-247.
- 22. Clark, F. E., W. E. Beard, and D. H. Smith. 1960. Dissimilar nitrifying capacities of soils in relation to losses of applied nitrogen. Soil Sci. Soc. Amer. Proc. 24:50-54.
- 23. Clevenger, C. B and L. G. Willis. 1935. Immediate effects of fertilization upon soil reaction. Agron. J. 27:833-846.
- 24. Cole, C. V., S. R. Olsen, and C. O. Scott. 1957. The nature of phosphate sorption by calcium carbonate. Soil Sci. Soc. Amer. Proc. 17:352-356.
- 25. Cornfield, A. H. 1952. The mineralization of the nitrogen of soils during incubation: Influence of pH, total nitrogen, and organic carbon content. J. Sci. Food Agric. 3:343-349.
- 26. Cornfield, A. H. 1959. Mineralization, during incubation, of the organic nitrogen compounds in soils as related to soil pH. J. Sci. Food Agric. 10:27-28.
- 27. Dancer, W. S., L. A. Peterson, and G. Chesters. 1973. Ammonification and nitrification of N as influenced by soil pH and previous N treatments. Soil Sci. Soc. Amer. Proc. 37:67-69.
- 28. Dodge, D. A. and H. E. Jones. 1947. The effect of long-time fertility treatments on the nitrogen and carbon content of a prairie soil. Soil Sci. Soc. Amer. Proc. 12:294.

- 29. Fuller, W. H. 1963. Reactions of nitrogenous fertilizers in calcareous soils. Agr. Food Chem. 11:188-193.
- 30. Gardner, W. R. 1965. Movement of nitrogen in soil. <u>In</u> W. V. Bartholomew and F. E. Clark (ed.) Soil nitrogen. Agronomy 10:550-572.
- 31. Grunes, D. L. 1959. Effect of nitrogen on the availability of soil and fertilizer phosphorus to plants. Adv. in Agron. 11:361-396.
- 32. Haas, H. J., C. E. Evans, and E. F. Miles. 1957. Nitrogen and carbon changes in great plains soils as influenced by cropping and soil treatments. U. S. D. A. Tech. Bulletin 1164.
- 33. Hannapel, R. J., W. H. Fuller, and R. H. Fox. 1964. Phosphorus movement in a calcareous soil. II. Soil microbial activity and organic phosphorus movement. Soil Sci. 97:421-427.
- 34. Herron, G. M., G. L. Terman, A. F. Drier, and R. A. Olson. 1968. Residual nitrate-N in fertilized deep loess-derived soils. Agron. J. 60:477-482.
- 35. Hiltbold, A. E. and F. Adams. 1960. Effect of nitrogen volatilization on soil acidity changes due to applied nitrogen. Soil Sci. Soc. Amer. Proc. 24:45-47.
- 36. Jackson, M. L. 1958. Soil chemical analysis. Prentice-Hall Inc. Sixth printing, 1970, by the author, Department of Soil Science, University of Wisconsin, Madison, Wisconsin.
- 37. Jannson, S. L. and F. E. Clark. 1952. Losses of nitrogen during decomposition of plant material in the presence of inorganic nitrogen. Soil Sci. Soc. Amer. Proc. 16:330-334.
- 38. Jenny, J., A. D. Ayers, and J. S. Hosking. 1945. Comparative behavior of ammonia and ammonium salts in soils. Hilgardia 16:429-457.
- 39. Jewitt, T. N. 1942. Loss of ammonia from ammonium sulfate applied to alkaline soils. Soil Sci. 54:401-409.
- 40. Johnston, W. R., F. Ittihadieh, R. M. Daum, and A. F. Pillsbury. 1965. Nitrogen and phosphorus in tile drainage effluent. Soil Sci. Soc. Amer. Proc. 29:287-289.
- 41. Jones, E. J. 1951. Loss of elemental nitrogen from soils under anaerobic conditions. Soil Sci. 71:193-196.
- 42. Jurinak, J. J. and N. Bauer. 1956. Thermodynamics of zinc adsorption on calcite, dolomite, and magnesite type minerals. Soil Sci. Soc. Amer. Proc. 20:466-471.
- 43. Kefauver, M. and F. E. Allison. 1957. Nitrite reduction by <u>Bacterium denitrificans</u> in relation to oxidation-reduction potential and oxygen tension. J. Bact. 73:8-14.

- 44. Kresge, C. B. and D. P. Satchell. 1960. Gaseous loss of ammonia from nitrogen fertilizers applied to soils. Agron. J. 52:104-107.
- 45. Larson, K. L., J. F. Carter, and E. H. Vasey. 1971. Nitrate-nitrogen accumulation under bromegrass sod fertilized annually at six levels of nitrogen for 15 years. Agron. J. 63:527-528.
- 46. Leo, W. M., T. E. Odland, and R. S. Bell. 1959. Effect on soils and crops of long continued use of sulfate of ammonia and nitrate of soda with and without lime. Rhode Island Agr. Exp. Sta. Bulletin 344.
- 47. Lindsay, W. L. and W. A. Norvell. 1969. Development of a DTPA micronutrient soil test. Agron. Abst. p. 84.
- 48. Loewenstein, H., L. E. Engelbert, O. J. Attoe, and O. N. Allen. 1957. Nitrogen loss in gaseous form from soils as influenced by fertilizers and management. Soil Sci. Soc. Amer. Proc. 21:397-400.
- 49. Lorenz, O. A. and C. M. Johnson. 1953. Nitrogen fertilization as related to the availability of phosphorus in certain California soils. Soil Sci. 75:119-129.
- 50. Martin, J. P. and H. D. Chapman. 1951. Volatilization of ammonia from surface-fertilized soils. Soil Sci. 71:25-34.
- 51. Meek, B. D. and A. J. MacKenzie. 1965. The effect of nitrite and organic matter on aerobic gaseous losses of nitrogen from calcareous soil. Soil Sci. Soc. Amer. Proc. 29:176-178.
- 52. Meek, B. D., L. B. Grass, and A. J. MacKenzie. 1969. Applied nitrogen losses in relation to oxygen status of soils. Soil Sci. Soc. Amer. Proc. 33:575-578.
- 53. Meyer, R. D., R. A. Olson, and H. G. Rhoades. 1961. Ammonia losses from fertilized Nebraska soils. Agron. J. 53:241-244.
- 54. Moe, P. G., J. V. Mannering, and C. B. Johnson. 1968. A comparison of nitrogen losses from urea and ammonium nitrate in surface runoff water. Soil Sci. 105:428-433.
- 55. Moore, D. G. 1960. Nitrogen movement in soil as affected by time of application, form of nitrogen, and fixation. Diss. Abstracts 21:414-415.
- 56. Morrill, L. G. and J. E. Dawson. 1967. Patterns observed for the oxidation of ammonium to nitrate by soil organisms. Soil Sci. Soc. Amer. Proc. 31:757-760.
- 57. Navrot, J. and S. Ravikovitch. 1969. Zinc availability in calcareous soils: III. The level and properties of calcium in soils and its influence on zinc availability. Soil Sci. 108:30-37.
- 58. Nelson, C. E. 1953. Methods of applying ammonium nitrate fertilizer on field corn, and a study of the movement of NH, and NO₃ nitrogen in the soil under irrigation. Agron. J. 45:154-157.

- 59. Olsen, R. J., R. F. Hensler, O. J. Attoe, S. A. Witzel, and L. A. Peterson. 1970. Fertilizer nitrogen and crop rotation in relation to movement of nitrate-nitrogen through soil profiles. Soil Sci. Soc. Amer. Proc. 34:448-452.
- 60. Olsen, S. R., C. V. Cole, F. S. Watanabe, and L. A. Dean. 1954. Estimation of available phosphorus in soils by extraction with sodium bicarbonate. U. S. D. A. Circular 939.
- 61. Olsen, S. R., F. S. Watanabe, H. R. Cosper, W. E. Larson, and L. B. Nelson. 1954. Residual phosphorus availability in long-time rotation on calcareous soils. Soil Sci. 78:141-151.
- 62. Owensby, C. E., K. L. Anderson, and D. A. Whitney. 1969. Some chemical properties of a silt loam soil after 20 years' nitrogen and phosphorus fertilization of smooth bromegrass. Soil Sci. 108:24-29.
- 63. Patrick, W. H. Jr. and R. Wyatt. 1964. Soil nitrogen loss as a result of alternate submergence and drying. Soil Sci. Soc. Amer. Proc. 28:647-653.
- 64. Pearson, R. W., F. Abruna, and J. Vicente-chandler. 1962. Effect of lime and nitrogen applications on downward movement of calcium and magnesium in two humid tropical soils of Puerto Rico. Soil Sci. 93: 77-82.
- 65. Peterson, L. A. and O. J. Attoe. 1965. Importance of soil nitrates in determination of need and recovery of fertilizer nitrogen. Agron. J. 57:572-574.
- 66. Pierre, W. H. 1928. Nitrogenous fertilizers and soil acidity: I. Effects of various nitrogeneous fertilizers on soil reaction. Agron. J. 20:254-269.
- 67. Pierre, W. H., J. R. Webb, and W. D. Shrader. 1971. Quantitative effects of nitrogen fertilizer on the development and downward movement of soil acidity in relation to level of fertilization and crop removal in a continuous corn cropping system. Agron. J. 63:291-297.
- 68. Pratt, P. F., W. W. Jones, and H. D. Chapman. 1956. Changes in phosphorus in an irrigated soil during 28 years of differential fertilization. Soil Sci. 82:295-306.
- 69. Pratt, P. F. and J. D. Chapman. 1961. Gains and losses of mineral elements in an irrigated soil during a 20-year lysimeter investigation. Hilgardia 30:445-467.
- 70. Pratt, P. F., W. W. Jones, and V. E. Hunsaker. 1972. Nitrate in deep soil profiles in relation to fertilizer rates and leaching volume. J. Environmental Quality 1:97-102.
- 71. Ray, H. E., J. M. MacGregor, and E. L. Schmidt. 1957. Movement of ammonium nitrogen in soils. Soil Sci. Soc. Amer. Proc. 21:309-312.

- 72. Rennie, D. A. and R. J. Soper. 1958. The effect of nitrogen additions on fertilizer-phosphorus availability. II. J. Soil Sci. 9:155-167.
- 73. Robinson, J. B. D. 1957. The critical relationship between soil moisture content in the region of wilting point and the mineralization of natural soil nitrogen. J. Agr. Sci. 49:100-105.
- 74. Schwartzbeck, R. A., J. M. MacGregor, and E. L. Schmidt. 1961. Gaseous nitrogen losses from nitrogen fertilized soils measured with infrared and mass spectroscopy. Soil Sci. Soc. Amer. Proc. 25:186-189.
- 75. Simpson, J. R. 1960. The mechanism of surface nitrate accumulation on a bare fallow soil in Uganda. J. Soil Sci. 11:45-60.
- 76. Smith, D. H. and F. E. Clark. 1960. Volatile losses of nitrogen from acid or neutral soils or solutions containing nitrate and ammonium ions. Soil Sci. 90:86-92.
- 77. Stephens, D. 1962. Upward movement of nitrate in a bare soil in Uganda. J. Soil Sci. 13:52-59.
- 78. Stewart, B. A., L. K. Porter, and D. D. Johnson. 1963. Immobilization and mineralization of nitrogen in several organic fractions of soil. Soil Sci. Soc. Amer. Proc. 27:302-304.
- 79. Stewart, B. A. 1970. A look at agricultural practices in relation to nitrate accumulations. <u>In</u> O. P. Engelstad (ed.) Nutrient mobility in soils: Accumulation and losses. Soil Sci. Soc. Amer. Pub. No. 4, Soil Sci. Soc. Amer., Madison, Wisconsin, pp 47-60.
- 80. Stewart, B. A., F. G. Viets, Jr., G. L. Hutchinson, and W. D. Kemper. 1967. Nitrate and other water pollutants under fields and feedlots. Environ. Sci. and Tech. 1:736-739.
- 81. Tabatabai, M. A. and J. M. Bremner. 1970. Use of the Leco automatic 70-second carbon analyzer for total carbon analysis of soils. Soil Sci. Soc. Amer. Proc. 34:608-610.
- 82. Taylor, A. W. 1967. Phosphorus and water pollution. J. Soil Water Conservation 22:228-231.
- 83. Udo, E. J., H. L. Bohn, and T. C. Tucker. 1970. Zinc adsorption by calcareous soils. Soil Sci. Soc. Amer. Proc. 34:405-407.
- 84. Viets, F. G., Jr. 1962. Chemistry and availability of micronutrients in soils. J. Agr. Food Chem. 10:174.
- 85. Viets, F. G., Jr. and R. C. Hageman. 1971. Factors affecting the accumulation of nitrate in soil, water, and plants. Agriculture Handbook No. 413. Agricultural Research Service, U. S. D. A. 54 pp.
- 86. Wagner, G. H. and G. E. Smith. 1958. Nitrogen losses from soils fertilized with different nitrogen carriers. Soil Sci. 85:125-129.

- 87. Wetselaar, R. 1961. Nitrate distribution in tropical soils. I. Possible causes of nitrate accumulation near the surface after a long dry period. Plant and Soil 15:110-120.
- 88. Wolcott, A. R. 1964. The acidifying effects of nitrogen carriers. Ag. Ammonia News, July-August ed.
- 89. Wolcott, A. R. and H. D. Foth. 1964. Lime takes the sting out of acid-forming N fertilizers. Crops and Soils, Nov. ed., p. 9.
- 90. Wolcott, A. R., H. D. Foth, J. C. Schickluna, and J. F. Davis. 1964. Soil reaction key helps select best N fertilizer. Crops and Soils, Jan.-Feb. ed., p. 21.
- 91. Wolcott, A. R., H. D. Foth, J. F. Davis, and J. C. Schickluna. 1965. Nitrogen carriers: I. Soil effects. Soil Sci. Soc. Amer. Proc. 29:405-410.
- 92. Yaalon, D. H. 1964. The concentration of ammonia and nitrate in rain water over Israel in relation to environmental factors. Tellus 16:200-204.

APPENDIX

Table 1. Soil NH_{4}^{+} -N (ppm) as affected by treatment.

£			N, kg/r	ıa		
Depth,	0	45	90	134	179	224_
0-5 5-10 10-25 25-30 35-45 25-35 30-45 30-45 30-45 45-55 65-75 85-95 95-120 140-160 180-220 240-240 240-240 240-240 240-340 240-340 240-340 340-360	8.6922070204656688919306204903 46233333322222160313098721474-4-1	74.9 38.0 46.8 25.8 30.6 21.3 22.4 15.8 61.8 11.5 2.6 21.5 2.7 2.7 2.7 2.7 2.7 2.7 2.7 2.7	77.6 54.7 96.2 6 2 6 2 7 7 5 7 5 2 2 9 6 2 2 1 7 2 2 5 5 2 2 1 7 2 2 2 5 5 2 2 1 7 2 2 2 2 2 1 7 2 2 2 2 2 2 1 7 2 2 2 2	95.887247056464491880924461305547317 91388865766646791880924461305547317 1189.7	97.76.1.90.2.0.7.1.93.3.7.98.8.7.3.4.5.6.7.1.3.2.8.1.7.6.5.1.1.2.7.4.6.8.7.3.4.5.6.7.1.3.2.8.1.7.6.5.1.1.2.1.7.4.2.1.2.1.7.4.2.1.2.1.2.1.2.1.2.1.2.1.2.1.2.1.2.1.2	79368338832509851258377276438993165 65633644342934877276438993165 64786677234834272218468317793165
360-380 380-400	-	- 5.5	21.5 19.3	17.8 22.0	13.7 10.0	20.5 18.9

Table 1. Soil NH_{4}^{+} -N (ppm) as affected by treatment (Cont.)

	Market Market Advantage of the State of the		N, kg/ha	l		
Depth,	0	45	90	134	179	224
0-5 5-10 10-15 15-20 20-25 20-35 30-35 30-45 45-50 50-65 65-75 60-75 75-80 85-90 95-100 120-140 140-160 180-200 240-280 240-280 280-300 300-320	744.6.76.2.9.9.9.4.0.7.8.5.8.4.5.0.3.0.9.9.7.8.0.2.3.9.4.4.9.3.7.5.5.1.2.2.2.1.2.2.1.1.1.2.5.7.0.8.2.3.9.1.2.1.2.1.1.2.5.7.0.8.2.3.9.1.2.1.2.1.2.1.2.1.2.1.2.1.2.1.2.1.2.1	136.00 6.00 6.00 7.00	976865678555553445447712473409976782635283019767826352	131.8 770.255.4102183651842596797193040 897286.02183651842596797193040 101.02183651842596797193040	61.29390021854475473876288708644 61.293900218544754876288708644 55557434876288708644 157.4	75.4.3.3.8.8.8.0.9.2.4.4.2.0.4.9.8.0.2.1.5.3.9.0.1.8.2.6.4.5.2.3.2.6.1.5.3.9.0.1.8.2.6.4.5.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.3.2.6.4.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2
260-280 280-300	8.2 12.3	18.7 20.0	19.3 19.5	16.0 23.4	15.6 17.4	19.2 26.6

Table 2. Soil NH_{4}^{+} -N (ppm) as affected by treatment.

		N, kg/h	າສ		
0	45	90	134	179	224
63.7 143.1 129.0 143.1 129.0 148.8 149.7 149.1 129.0 148.8 149.7 159.8 169.0 169	16946474236697323049806364319387 824.42.36697323049806364319387 777548091277297466.4319387 16946474236697323049806364319387	84.62.27.034.59.42.234.024.936.57.35.10.55.6 84.62.27.034.59.42.234.02.49.36.57.35.10.55.6 84.62.27.034.59.42.234.02.49.36.57.35.10.55.6	1028.1.5.5.6.8.1.4.6.8.8.2.1.5.1.4.8.8.4.5.7.0.3.2.3.0.4.2.1.5.1.4.8.8.4.5.7.0.3.2.3.0.4.2.4.3.3.3.3.3.3.3.3.3.3.3.3.3.3.3.3.3	1034.72 124.23 127.23 1	174.579.27761642245982461659679 1751.1730.459.1642245982461659679 1106.798785.649.6982461659679 18887649615.461659679 18887649615.461659679
29.8 16.6	31.5 19.4	14.0 19.2	18.3 27.5	15.2 15.9	29.9 24.6
	63.4 113.1 63.4 113.1 63.4 113.1 63.4 113.1 63.4 113.1 63.4 113.1 63.4 13.4 13.4 13.4 13.4 13.4 13.4 13.4 1	63.7 142.4 113.1 129.1 63.7 56.2 91.0 88.4 81.8 92.0 73.4 84.8 81.8 92.0 73.6 69.7 73.6 69.7 73.6 69.7 73.6 69.7 73.6 69.7 73.6 73.	0 45 90 63.7 38.1 84.9 142.4 52.6 111.6 113.1 74.9 100.2 129.1 82.4 79.2 63.7 98.6 92.7 56.2 101.4 118.7 91.0 54.7 72.0 88.4 73.4 109.3 84.8 73.2 84.4 81.8 71.3 46.5 92.0 57.6 77.9 73.7 44.6 52.9 49.5 38.9 69.4 69.7 50.7 60.2 73.0 59.3 41.2 71.3 42.4 37.2 40.8 52.3 42.4 50.7 37.0 33.0 58.3 47.4 37.2 48.0 52.9 37.4 43.3 39.8 27.9 26.1 27.0 39.3 35.3 24.6 15.7 25.4 19.4 18.3 19.6 13.3 <td< td=""><td>63.7 38.1 84.9 41.7 142.4 52.6 111.6 102.4 113.1 74.9 100.2 78.5 129.1 82.4 79.2 151.9 63.7 98.6 92.7 84.6 56.2 101.4 118.7 98.1 91.0 54.7 72.0 100.5 88.4 73.4 109.3 81.5 84.8 73.2 84.4 111.6 81.8 71.3 46.5 99.8 92.0 57.6 77.9 84.1 73.7 44.6 52.9 48.4 49.5 38.9 69.4 52.6 69.7 50.7 60.2 79.8 73.0 59.3 41.2 75.8 52.6 41.2 71.3 65.2 40.8 52.3 42.4 49.1 50.7 37.0 33.0 54.5 52.6 41.2 71.3 65.2 47.1 68.0 52.9 37.4 48.4 50.7 58.3 47.4 37.2 47.1 68.0 52.9 37.4 48.4 49.1 50.7 37.0 39.3 30.5 54.5 52.6 16.6 37.4 16.3 13.5 30.5 14.0 83.3 24.6 16.6 37.4 18.3 24.0 19.6 13.3 18.5 21.8 16.6 15.7 19.7 25.4 19.4 18.3 24.0 19.6 13.3 18.5 21.3 20.9 21.1 20.1 20.1 20.1 20.1 20.1 20.1 20.1</td><td>0 45 90 134 179 63.7 38.1 84.9 41.7 103.5 142.4 52.6 111.6 102.4 124.7 113.1 74.9 100.2 78.5 151.2 129.1 82.4 79.2 151.9 127.3 63.7 98.6 92.7 84.6 170.2 56.2 101.4 118.7 98.1 114.7 91.0 54.7 72.0 100.5 117.8 84.8 73.4 109.3 81.5 96.4 84.8 73.2 84.4 111.6 88.8 81.8 71.3 46.5 99.8 64.2 92.0 57.6 77.9 84.1 73.5 73.7 44.6 52.9 48.4 73.0 49.5 38.9 69.4 52.6 50.2 69.7 50.7 60.2 79.8 61.2 73.0 59.3 41.2 75.8</td></td<>	63.7 38.1 84.9 41.7 142.4 52.6 111.6 102.4 113.1 74.9 100.2 78.5 129.1 82.4 79.2 151.9 63.7 98.6 92.7 84.6 56.2 101.4 118.7 98.1 91.0 54.7 72.0 100.5 88.4 73.4 109.3 81.5 84.8 73.2 84.4 111.6 81.8 71.3 46.5 99.8 92.0 57.6 77.9 84.1 73.7 44.6 52.9 48.4 49.5 38.9 69.4 52.6 69.7 50.7 60.2 79.8 73.0 59.3 41.2 75.8 52.6 41.2 71.3 65.2 40.8 52.3 42.4 49.1 50.7 37.0 33.0 54.5 52.6 41.2 71.3 65.2 47.1 68.0 52.9 37.4 48.4 50.7 58.3 47.4 37.2 47.1 68.0 52.9 37.4 48.4 49.1 50.7 37.0 39.3 30.5 54.5 52.6 16.6 37.4 16.3 13.5 30.5 14.0 83.3 24.6 16.6 37.4 18.3 24.0 19.6 13.3 18.5 21.8 16.6 15.7 19.7 25.4 19.4 18.3 24.0 19.6 13.3 18.5 21.3 20.9 21.1 20.1 20.1 20.1 20.1 20.1 20.1 20.1	0 45 90 134 179 63.7 38.1 84.9 41.7 103.5 142.4 52.6 111.6 102.4 124.7 113.1 74.9 100.2 78.5 151.2 129.1 82.4 79.2 151.9 127.3 63.7 98.6 92.7 84.6 170.2 56.2 101.4 118.7 98.1 114.7 91.0 54.7 72.0 100.5 117.8 84.8 73.4 109.3 81.5 96.4 84.8 73.2 84.4 111.6 88.8 81.8 71.3 46.5 99.8 64.2 92.0 57.6 77.9 84.1 73.5 73.7 44.6 52.9 48.4 73.0 49.5 38.9 69.4 52.6 50.2 69.7 50.7 60.2 79.8 61.2 73.0 59.3 41.2 75.8

Table 2. Soil NH_{μ}^+ -N (ppm) as affected by treatment (Cont.)

			N, kg/ha	a	· · · · · · · · · · · · · · · · · · ·	
Deoth,	0	45	90	134	179	224_
0-5 5-10 10-15 15-20 20-25 230-35 35-45 20-35 35-45 45-55 65-75 85-95 90-120-140 140-180 95-120-140 140-220 240-240 280-320 280-320 320-320	137.75.10 137.75.10 137.20 137	165.725.2001769846669431269298998600 1314595.0017698885529112695.600 1964476329122222218	0 0 6 2 7 4 2 7 2 2 2 7 6 8 9 5 1 2 8 8 4 8 5 5 9 4 5 4 5 9 6 2 1 2 1 2 2 1 1 5 8 1 7 0 8 8 7 5 5 5 4 5 3 2 2 1 1 2 2 2 1 1 2 1 2 2 2 1 1 2 1 2	1734.04.78.97.04.206.41.194.51.15.20.97.9 1275.404.78.97.04.206.41.194.51.15.20.97.9 127968684825.77.55.74.424.22.22.18.93.39.9	92446633775244448024750418674663 1242218867.5244448024750418674663 12922222222222222222222222222222222222	139.6.4 139.1.3 139
340-360 360-380 380-400	40.1 33.7 28.0	19.6 20.8 22.5	17.5 17.1 15.4	16.1 21.8 24.6	23.0 15.4 22.5	23.4 19.0 18.2

Table 3. Soil NH_4^+ -N (ppm) as affected by treatment.

			N, kg/l	na		
Depth,	0	45	90	134	179	224
0-5 10-15 15-20 25-30 35-45 45-55 65-75 85-95 100-140 140-180 120-140 140-180 120-240 140-180 120-240 140-220 240-230 240-230 240-330 320-340	1221438.0706208138659744409147464153 12214137474141498853.65977444050.53 17777776390722484050.53	1049.74.28.21.1.25.70.09.21.88.1.50.39.78.24.4.3.2.3 149.84.1.25.70.09.21.88.1.50.39.78.24.4.3.2.3 149.74.28.21.1.25.70.09.21.88.1.50.39.78.24.4.3.2.3 149.74.28.21.1.25.70.09.21.88.1.50.39.78.24.4.3.2.3	196.1.2.8.59.1.8.1.94.8.7.9.2.96.7.7.2.4.6.4.7.1.4.6.1.0.1.0.1.0.1.0.1.0.1.0.1.0.1.0.1.0.1	698.98.20.28.17.56.87.73.38.72.63.44.70.86.21 698.98.20.28.17.56.87.73.38.72.63.44.70.86.21	107.3 122.8 1316.6 1330.1 134.9 134.9 134.9 134.9 134.9 134.9 135.1 136.1 137.	59.56.90.28.20.36.16.57.4.54.6.1.16.8.37.77.0.59.0 105.8.20.36.16.57.4.54.6.1.16.8.37.77.0.59.0 104.3.16.5.7.4.54.6.1.16.8.37.77.0.59.0
340-360 360-380 380-400	19.9 17.5 14.7	14.4 19.6 16.1	18.3 20.4 14.7	27.2 37.2 18.2	17.1 17.0 18.5	19.7 17.5 28.2

Table 3. Soil NH_{μ}^+ -N (ppm) as affected by treatment (Cont.)

			19 P, kg/	/ha		
<	8		N, kg/ha	1		
Depth,	0	45	90	134	179	224
0-5 10-15 15-25 20-35 15-25 20-35 15-25 20-35 15-25 20-35 15-25 20-35 15-25 20-35 15-2	78.4 78.4 78.9	2512.3.0.9.0.3.6.9.3.6.4.8.3.2.8.7.2.9.1.4.9.5.3.6.0.3.6.6.1.9.0.2.1.5.6.4.8.3.2.8.7.2.9.1.4.9.5.3.6.0.3.6.6.1.9.0.2.1.2.3.6.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2	76.392307189166437034400083800257742 5071.3923.07189166437034400083800257742 18844750.222111118844750.257742	47.8 130.0 134.8 100.1 134.0 134.0 134.0 131.3 1631	214.34.7262.731.938.75.912.975.92.502.78.32.29 214.51.434.7262.731.938.75.912.975.92.502.78.32.29 215.434.7262.731.938.75.912.975.92.502.78.32.29 215.434.7262.731.938.75.912.975.92.502.78.32.29 215.434.7262.731.938.75.912.975.92.502.78.32.29	170.431.434.94.94.3522.26.70.996.506.96.810.28.80 1731.56.8.78.8.8.35.55.5.996.506.96.810.28.80 1731.66.8.78.8.8.35.55.5.996.506.96.810.28.80 1731.66.8.78.8.8.35.55.5.996.506.96.810.28.80 1731.66.8.78.8.8.35.55.5.996.506.96.810.28.80 1731.66.8.78.8.8.35.55.5.9996.506.96.810.28.80 1731.66.8.78.8.8.35.55.5.9996.506.96.810.28.80 1731.66.8.78.8.8.35.55.5.9996.506.96.810.28.80 1731.66.8.96.8.96.810.28.80 1731.66.8.96.810.28.80 1731.66.810.80 1731.66.80 1731.66.80 1

Table 4. Soil NO3 -N (ppm) as affected by treatment.

		N, kg/h	а		
0	45	90	134	179	224
3.5 3.5 4.1 3.7 2.0 0.8 0.6 1.6 0.2	2.5	8.2 7.6 8.5 7.0 7.0 7.0 7.0 7.0 7.0 7.0 7.0 7.0 7.0	78.427345416542532243644563544 78.683427345416542532243644563544	4796509420626043666 22197537563 4209420626043666 22197537563	3333854819460031200892693395335513 12739561346799001970907855454434454
-	=	1.6 0.8	3.1 4.1	2.0 0.4	5.5 4.7
	3.5 3.5 4.1 3.7 2.0 0.8 0.6 1.6	3.5 2.5 3.5 3.9 4.1 - 3.7 - 2.0 - 0.8 - 0.6 - 1.6 - 0.6 -	0 45 90 3.5 2.5 8.2 3.5 3.9 7.6 4.1 - 8.0 3.7 - 5.5 2.0 - 5.1 0.8 - 9.2 0.6	3.5	0 45 90 134 179 3.5 2.5 8.2 7.4 14.4 14.4 3.5 3.9 7.6 18.1 27.7 2.0 4.1 - 8.0 16.4 20.9 20.9 3.7 2.0 6.6 20.9 2.0 <td< td=""></td<>

Table 4. Soil NO3 -N (ppm) as affected by treatment (Cont.)

Depth, om 0 45 90 134 179 0-5 5.1 7.6 6.4 9.6 2.9	7.6 6.4 9.6 2.9 10.0
	7.6 6.4 9.6 2.9 10.0 10.3 9.2 10.7 15.6 15.9 6.8 8.6 11.7 12.1 16.1
5-10 6.8 10.3 9.2 10.7 15.6 10-15 5.9 6.8 8.6 11.7 12.1 15-20 7.8 4.5 5.3 5.7 8.4 20-25 3.7 3.3 2.9 5.1 3.9 25-30 4.5 3.7 1.4 3.1 1.8 30-35 1.0 1.4 2.2 3.5 1.2 35-40 2.5 2.2 2.0 3.3 2.0 40-45 2.0 0.8 1.6 4.5 2.2 45-50 3.5 1.0 1.2 3.9 2.0 45-50 3.5 1.0 1.2 3.9 2.0 55-60 0.6 1.8 2.0 3.3 1.4 60-65 2.3 2.2 2.0 4.7 0.4 75-80 - 1.2 1.6 3.7 0.8 80-85 2.5 1.0 2.0 3.3 1.8 85-90 2.7 2.2 - 2.7 0.2	4.5 5.3 5.7 8.4 12.3 3.3 2.9 5.1 3.9 5.2 3.7 1.4 2.2 3.5 1.2 7.8 2.2 2.0 3.3 2.0 10.0 0.8 1.6 4.5 2.2 12.3 1.0 1.2 3.9 2.0 13.8 1.0 1.2 3.9 2.0 13.8 1.2 2.5 4.1 1.0 17.0 1.8 2.0 3.3 1.4 20.6 2.2 2.0 4.7 0.4 16.8 3.3 0.2 4.1 1.4 15.2 1.0 2.0 3.3 1.8 16.3 13.0 1.0 2.0 3.3 0.4 12.5 11.8 1.0 2.0 3.3 0.4 12.5 11.8 1.0 2.2 2.7 0.2 11.8 10.9 2.7 2.0 3.1 0.4 6.8 0.9 2.7 2.0 3.1 0.4

Table 5. Soil NO3 -N (ppm) as affected by treatment.

×			N, kg/h	а		
Depth,	0	45	90	134	179	224
0-5 5-10 10-15 15-20 20-30 30-35 40-25 20-30 30-45 50-65 60-75 80-95 80-95 100-120 140-180 180-220 240-240 240-240 240-240 240-240 240-240 240-240 240-3	2376949949355437914842422632161393	2794694027977499454215 049 52172712 1001120 110 0121121	6.7 6.7 6.1 6.1 6.1 6.1 6.1 6.1 6.1 6.1	8.1 9.0 9.0 9.8 3.6 6.6 9.4 6.8 1.9 7.7 9.1 6.7 1.9 9.9 6.2 8.2 8.8 8.9 1.0 1.0 1.0 1.0 1.0 1.0 1.0 1.0 1.0 1.0	5.9.7.2.3.3.8.6.9.8.1.8.8.9.9.1.8.7.9.7.7.6.7.8.8.8.7.9.7.1.7.6.5.0.1.0.0.0.0.0.0.0.0.0.0.0.0.0.0.0.0.0	39072126731531355856092999433262686 2493086343332344443211122222344343333

Table 5. Soil NO3 -N (ppm) as affected by treatment (Cont.)

100 VI V			N, kg/ha			
Depth,	0	45	90	134	179	224
0-5 10-15 15-25 15	25736744394348872388554481945284	898643544322234344452131222113211 898643544322234344452131222113211	62493671585425131014254708199886 13503747444643233432001012211221	14.605699990036699878 66396398222 11111302 33222222102	13640763434543333233442233334244543 11640763434543333233442233334244543	122963226578585975844102999914788
340-360 360-380 380-400	2.9 2.6 1.7	1.7	1.9 2.8 1.6	0.2 0.7 2.6	3.8 5.9 3.5 5.4	7.3 7.6 7.1

Table 6. Soil NO3-N (ppm) as affected by treatment.

5200			
0	Ρ.	kg/	-
•	Ρ.	K O' I	n_H

			N, kg/h	a		
Depth,	0	45	90	134	179	224
0-5 5-10 10-20 20-35 20-35 35-45 20-35 35-45 45-55 65-75 85-95 100-180 120-220 120-180 120-220 120-220 120-220 120-220 120-30 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-220 120-30	808664353433243332222111021101100 81086643534332433322221110211101100	8.7.7.4.8.8.4.0.4.2.2.7.9.9.7.6.4.8.6.7.9.4.1.9.5.9.9.1.2.4.9.2.7.0.0.0.0.2.1.0.0.1.0.1.0.1.0.1.0.1.0.1.0	905428069856710387482416171367717	11.4.8.5.2.9.6.6.5.1.9.7.1.4.1.4.5.3.2.1.3.7.2.1.8.1.8.0.4.8.1.4.7.5.5.1.2.3.3.2.2.1.3.2.3.2.2.1.3.2.2.0.1.3.3.2.3.3.3.4.2.2.2.2	015941662868339212327471803152860 17015941662868339212327471803152860	504645996359656275583474830075375 460274522335632443447655790999989
360-380 380-400	1.6 1.0	0.5	0.7 0.5	5.7	4.5 3.6	6.8 10.7

Table 6. Soil NO3-N (ppm) as affected by treatment (Cont.)

10			/ 1
19	Р.	kg/	na
1 7		11 5	1104

		11	N, kg/ha			
Depth,	0	45	90	134	179	224
0-5 10-15 15-20 20-25 20-30 10-25 20-30 10-20 20-25 20-35 20-35 20-35 450-55 650-75 850-95 100-140 140-180 140-220 140-240 140-220 140-230 140-2	6.1 1.2 1.2 1.2 1.2 1.2 1.2 1.3 1.3 1.3 1.3 1.3 1.3 1.3 1.3	698.1703035571531373549479934444446 698.1155243341332221332011002221011	5553373604786044009624 92 5402667 133952212121102111111 00 01114321	6.2.7.6.2.4.8.9.9.4.4.4.1.3.9.4.9.6.4.9.0.2.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.9.8.8.0.6.5.1.6.2.1.4.4.4.4.4.4.4.4.4.4.4.4.4.4.4.4.4.4	223614580193935616331130225868816 90818546531313333332322345433333332	48733421971429384515186356832019633 142775422130113223342433333345432332
360-380 380-400	1.0 0.5	1.9 1.2	2.8 2.8	1.4 2.1	2.1	3.3 2.3

Table 7. Total soil N (ppm) as affected by treatment.

			Ur, AS/	па		
			N, kg/h	a		
Depth,	0	45	90	134	179	224
0-5 5-10 10-15 15-20 20-25 25-30 30-35 35-40 40-45 45-50 50-55 55-60	1170 1100 1080 1070 1000 1030 870 730 750 630 810 840	1190 1050 1100 1070 1010 960 950 810 770 730 670 500	1070 1100 1050 1010 1100 1060 970 880 860 790 1100	880 1020 810 840 820 800 560 510 540 540 560	1260 1060 1020 960 970 840 880 820 760 640 640 630	1140 1100 970 970 880 830 810 860 790 740 690 710
60-65 65-70 70-75 75-80 80-85 85-90 90-95 95-100 100-120 120-140	790 710 680 830 800 450 480 570 440	530 580 480 580 580 500 550 420	760 660 640 610 780 690 790 640	640 650 600 730 620 680 650 560 630	750 620 650 600 520 560 550 480 530	570 570 570 570 570 570 570 470 470
140-160 160-180 180-200 200-220 220-240 240-260 260-280 280-300 300-320 320-340 340-360 360-380 380-400	470 650 680 640 640 670 610 780 730 640 770	340 470 3750 3500 570 580 460 350 580	430 550 550 4390 400 400 400 550 400 500 400	520 400 420 550 540 540 640 640 750 710	570 570 570 570 410 530 530 530 470	470 470 470 470 470 470 470 470 470 470

Table 7. Total soil N (ppm) as affected by treatment (Cont.)

			N, kg/ha			
Depth,	0	45	90	134	179	224
0-5 5-10 10-15 15-20 20-25 20-25 20-35 35-40 45-50 45-55 50-65 65-70 75-80 85-95 95-100 120-140 140-160 160-180	1060 1060 1060 1000 1000 1000 1000 1000	980 930 930 830 820 620 620 670 630 630 570 530 530 530 530 530 530 530 53	90 1160 1050 960 960 950 830 770 670 670 680 570 650 550 650 650 650 650 650 650	1250 1070 1090 1070 980 830 760 730 760 760 760 760 760 760 760 760 760 76	1220 11200 1220 1220 1220 1220 1220 122	1060 1180 1010 1220 1060 920 850 780 780 780 580 670 580 670 580 580 580 580
180-200 200-220 220-240 240-260	190 130 140 170	520 480 570 500	550 650 630 550	550 570 520 620	420 380 300 460	600 560 680 610
260-280 280-300 300-320 320-340 340-360 360-380 380-400	210 350 210 300 360 310 270	630 610 520 610 850 510 410	560 560 630 610 710 630 670	500 670 600 680 730 650 560	420 500 390 480 480 420 480	510 600 600 820 1120 600 500

Table 8. Total soil N (ppm) as affected by treatment.

			N, kg/h	a		
Deoth,	•	1. ~	00	104	1.00	204
<u>cm</u>	0	45	90	134	179	224
0-5	1060	1020	1150	1280	1140	1450
5-10	1160	1000	1120	1300	1160	1260
10-15	1180	1090	1130	1040	1220	1160
15-20	940	1080	990	1280	1100	1290
20-25	960	1120	1180	1190	1220	1390
25-30	950	1040	1250	960	970	1310
30-35	900	790	1010	970	1060	1080
35-40	840	940	900	76 0	870	1080
40-45 45-50	860 840	1000 1150	630 660	820 920	820 540	1000 1000 820
50-55	720	880	540	600	750	860
55 - 60	580	1200	780	600	760	840
60-65 65-70	560 780	740 960 1080	840 730	520 700 690	720 680 670	780 730
70-75 75-80 80-85	900 600 520	900 960	690 820 610	680 650	760 790	900 1080 1080
85-90	650	740	590	720	850	960
90-95	610	980	600	820	680	740
95-100	770	1210	470	660	660	700
100-120	600	1020	430	660	600	500
120-140	450	900	470	500	580	750
140-160	600	1110	480	470	560	630
160-180	520	530	710	530	500	650
180-200	600	690	580	490	460	580
200-220	480	450	450	370	400	900
220-240	460	630	430	400	540	650
240-260	560	530	750	660	720	590
260-280	740	590	660	520	700	680
280-300	590	510	460	600	630	560
300-320	740	710	670	580	680	680
320-340	620	610	580	800	660	550
340-360	560	790	580	680	620	580
360-380	680	710	580	500	570	840
380-400	570	410	920	640	600	700

Table 8. Total soil N (ppm) as affected by treatment (Cont.)

			N, kg/ha			
Depth,	0	45	90	134	179	224_
0-5 5-10 10-20 20-25 20-35 30-45 20-35 35-45 45-55 60-75 85-95 95-120 140-160 180-220 240-280 180-220 240-280 280-320	1280 1180 13400 13400 14800 14	1260 1140 1210 1150 1150 1100 1100 1100 1100 110	1370 1470 14550 14	1380 1240 1240 1450 1180 1180 11920 1180 11920 1180 11920 1180 11920 119	1430 1480 1480 1480 1480 1000 1000 1000 100	1460 1280 1280 1280 1280 1280 1280 1280 128
340-360 360-380 380-400	830 870 790	560 650 610	640 540 610	7 20 790 740	730 580 720	690 740 720

Table 9. Total soil N (ppm) as affected by treatment.

7	****		N, kg/h	ıa		
Depth,	0	45	90	134	179	224
Depth, cm 0-5 5-10 10-15 15-20 20-25 25-30 30-45 45-50 50-65 60-65 65-70 75-80 80-95	1560 1670 1300 1350 1390 1320 1330 1330 1140 1330 1010 950 1010 890 870 930 880 890	1280 1110 1240 1450 1210 1130 1080 950 930 760 820 930 690 940 680 890 760 770 900	90 1480 1310 1180 1140 1180 1020 1000 840 750 900 820 650 640 600 720 880 720 810	1340 1280 1260 1260 1200 1160 1140 1090 1030 920 930 720 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 900 870 870 870 870 870 870 870 870 870 8	1410 1300 1030 1020 1060 1200 1020 1120 1180 900 850 840 830 540 680 600 900	1210 1080 1040 1220 1080 1040 1160 1080 940 940 1000 760 520 530 520
95-100 100-120 120-140 140-160 160-180 180-200 200-220 240-260 240-260 260-280 280-300 300-320 320-340 340-360 360-380 360-380 380-400	900 610 600 1030 740 600 640 680 750 680 550	860 640 5780 5820 5660 5810 5840 5840 5840 5840	760 600 520 580 620 510 500 560 570 600 560 560 560	720 860 800 760 820 760 710 810 880 690 780 770 530 780 980 990	900 640 510 470 560 460 500 580 530 530 570 560	540 540 540 550 500 500 500 500 500 500

Table 9. Total soil N (ppm) as affected by treatment (Cont.)

		19	N, kg/ha			
Depth,	0	45	90	134	179	224
0-5 5-10 10-15 15-20 25-35 30-45 25-35 35-45 45-55 65-75 65-75 85-95 95-120 120-140	1380 1230 1090 1060 1090 910 800 780 560 560 560 520 510 730 980 620 540 620 640	1620 1430 1240 1240 1440 1270 1480 1210 1020 1020 1020 1020 7700 750 650 7690 630 530	1090 1310 1150 1270 1060 9860 9810 9810 7800 7800 7800 7800 7800 7800 7800 7	1160 1170 1060 1210 1230 1090 1100 940 750 670 660 660 680 680 570 460	1680 1350 1360 1360 1270 1110 1110 1090 880 750 670 650 8690 780 780 780 780	1370 1320 1330 1370 1070 1070 1070 670 6800 640 630 640 630 660 510 660 520 640 640
140-160 160-180 180-200 200-220 220-240 240-260	630 560 500 560 480	530 770 470 480 490 630	530 420 230 400 440 420	470 490 580 580 540	520 650 530 590 450 550 540	440 490 460 460 520 510 630
260-280 280-300 300-320 320-340 340-360 360-380 380-400	480 500 540 600 510 480 600	620 530 510 470 540 540	430 460 550 510 480 440 580	450 580 640 580 630 660 620	550 580 490 550 530 450	400 580 450 400 480

Table 10. Soil pH as affected by treatment.

		N, kg/h	a		
0	45	90	134	179	224
8.1000011111122122222221111223334	888778888888888888888888888888888888888	8.0099001111111222222222122111222222 8.8888888888	109000101121111222222222222222222222222	09890112202221222222222222112134	8.7.8.9.9.0.1.0.1.1.1.2.1.2.1.2.2.2.2.2.2.2.2.2.2
8.4 8.4	8.3 8.2	8.2 8.2	8.2 8.3	8.4	8.2 8.2
	8.1 8.0 8.1 1.1 1.1 1.1 1.1 1.1 1.2 1.2 1	8.1 8.0 8.0 8.0 7.9 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1	0 45 90 8.1 8.0 8.0 8.0 8.0 7.9 8.0 7.9 7.9 8.0 7.9 8.0 8.1 8.0 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.2 8.1 8.2 8.2 8.2 8.1 8.2 8.2 8.1 8.2 8.2 8.1 8.2 8.2 8.1 8.2 8.2 8.1 8.2 8.2 8.1 8.2 8.2 8.1 8.2 8.2 8.1 8.2 8.2 8.2 8.1 8.1 8.1 8.1 8.2 8.2 8.2 8.1 8.1 8.2 8.2 8.2 8.2 8.3 8.2 8.2 </td <td>8.1 8.0 8.0 8.0 8.0 8.0 8.0 8.0 8.0 8.0 7.9 7.9 8.0 7.9 8.0 8.0 8.1 8.0 8.0 8.0 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.2 8.1 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.1 8.2 8.2 8.2 8.1 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.1 8.1 8.2 8.2 8.2 8</td> <td>0 45 90 134 179 8.1 8.0 8.0 8.1 8.0 8.1 8.0 8.0 7.9 7.9 8.0 8.0 7.9 7.9 7.9 8.0 7.9 8.0 8.0 8.0 8.1 8.0 8.0 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.2 8.1 8.2 8.2 8.2 8.2 8.2 8</td>	8.1 8.0 8.0 8.0 8.0 8.0 8.0 8.0 8.0 8.0 7.9 7.9 8.0 7.9 8.0 8.0 8.1 8.0 8.0 8.0 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.2 8.1 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.1 8.2 8.2 8.2 8.1 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.2 8.1 8.1 8.2 8.2 8.2 8	0 45 90 134 179 8.1 8.0 8.0 8.1 8.0 8.1 8.0 8.0 7.9 7.9 8.0 8.0 7.9 7.9 7.9 8.0 7.9 8.0 8.0 8.0 8.1 8.0 8.0 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.1 8.1 8.1 8.2 8.2 8.2 8.1 8.2 8.2 8.2 8.2 8.2 8

Table 10. Soil pH as affected by treatment (Cont.)

19 P. kg/ha

Table 11. Soil pH as affected by treatment.

			,
0	Ρ.	kg/	1
£ 1		v cr /	ng

			N, kg/h	a		
Depth,	0	45	90	134	179	224
0-5 10-15 15-25 15	8.01000112323333333222322223444	7,7,7,8,8,8,8,8,8,8,8,8,8,8,8,8,8,8,8,8	7.00 7.00 7.00 7.00 7.00 7.00 7.00 7.00	777787888888888888888888888888888888888	8,77,78,88,88,88,88,88,88,88,88,88,88,88	8.0989900001111121111222221111111111111111
280-300	8.4	8.1	8.0	8.2	8.1	8.1

Table 11. Soil pH as affected by treatment (Cont.)

			N, kg/ha			
Depth,	0	45	90	134	179	224
0-5 5-10 10-15 15-20 20-25 25-30 30-45 25-30 30-45 45-50 45-55 60-65 75-80 85-95 100-180 140-180 140-180 140-180 140-220 240-220 240-220 240-230 240-320 240-320 240-320 240-320 320-340	8.0 9.9 9.9 9.0 8.1 1.1 1.1 1.1 1.1 1.1 1.1 1.1	8.0 7.9 7.9 7.9 7.9 8.0 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1 8.1	8.8.9.9.9.9.0.1.1.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2.2	8.00 8.00 9.00 112222221112111211122 8.10 8.10 8.10 8.10 8.11 8	8.000910100001021212212222222222222222222	8.0 0 9 9 9 9 9 0 0 1 1 1 1 1 1 1 1 1 2 1 2 1 2 2 2 2 2
320-340 340-360 360-380 380-400	8.1 8.2 8.2 8.3	8.1 8.1 8.1 8.2	8.1 8.3 8.3 8.3	8.2 8.1 8.2 8.0	8.2 8.2 8.2 8.1	8.2 8.2 8.2 8.2

Table 12. Soil pH as affected by treatment.

0 P, kg/haN, kg/ha Depth, 45 134 179 224 0 90 cm 0-5 8.0 8.1 8.0 8.1 7.9 8.0 7.9 5-10 8.1 8.0 8.0 8.0 8.0 10-15 8.1 8.0 7.9 7.9 8.0 8.1 15-20 8.0 7.9 7.9 7.9 8.0 8.0 20-25 8.1 8.1 8.1 8.0 8.0 8.0 25-30 8.1 8.1 8.1 8.0 8.0 8.1 30 - 358.2 8.1 8.2 8.2 8.1 8.0 35-40 8.1 8.1 8.2 8.1 8.0 8.1 40-45 8.3 8.1 8.1 8.1 8.0 8.1 45-50 8.2 8.1 8.2 8.1 8.1 8.0 8.3 50-55 8.1 8.1 8.1 8.0 8.1 8.3 55-60 8.0 8.1 8.1 8.1 8.2 8.2 60-65 8.2 8.1 8.3 8.2 8.1 8.3 8.2 8.1 8.1 65-70 8.2 8.1 70-75 8.2 8.2 8.1 8.1 8.1 8.1 75-80 8.1 8.2 8.2 8.2 8.1 8.2 8.2 8.2 8.1 80-85 8.2 8.2 8.1 8.3 8.2 85-90 8.2 8.1 8.2 8.2 8.2 8.2 8.1 8.2 8.2 90-95 8.2 8.3 8.1 8.2 8.2 8.2 8.2 95-100 8.2 8.2 100-120 8.2 8.2 8.3 8.1 8.2 8.1 8.2 8.2 8.1 8.2 120-140 8.3 8.3 8.2 8.2 8.2 8.2 8.2 8.1 140-160 8.2 160-180 8.2 8.2 8.1 8.2 8.2 8.2 8.1 8.2 180-200 8.2 8.2 8.2 8.2 8.1 8.2 200-220 8.2 8.3 8.2 8.2 8.2 8.2 8.1 220-240 8.3 8.2 8.2 8.2 8.1 8.2 240-260 8.2 8.2 8.2 8.2 8.2 8.2 260-280 8.2 8.1 8.1 280-300 8.2 8.2 8.2 300-320 8.2 8.2 8.2 8.2 8.2 8.1 320-340 8.3 8.2 8.2 8.2 8.1 8.1 8.3 8.3 8.3 8.2 8.1 8.2 340-360 8.4 8.2 8.1 8.1 360-380 8.3 8.3 8.2 380-400 8.1 8.1 8.3

Table 12. Soil pH as affected by treatment (Cont.)

			N, kg/ha			
Depth,	0	45	90	134	179	224
0-5 5-10 10-15 15-20 20-25 20-25 20-35 30-45 45-50 50-65 65-70 75-80 85-90 90-120 140-160 120-140 140-180 120-240 140-180 120-240 140-280 140-280 140-280 180-280 180-280 180-280 180-32	888888888888888888888888888888888888888	8.1.9.0.0.0.2.1.0.1.1.1.1.2.2.2.2.2.2.1.1.1.1	88.78.88.88.88.88.88.88.88.88.88.88.88.8	8.0.0.0.1.1.1.2.2.3.2.2.2.2.2.2.2.2.2.2.2.2.2.2	8.00001101111222222222222222222222222222	887778888888888888888888888888888888888
320-340 340-360 360-380 380-400	8.2 8.2 8.2 8.3	8.1 8.1 8.2 8.2	8.2 8.2 8.3 8.3	8.2 8.2 8.2 8.2	8.2 8.2 8.3 8.2	8.3 8.4 8.4 8.4

Weak Bray extractable soil phosphorus (ppm) as affected by treatment. Table 13.

	224	0000000	121 127 2000000 2000000	747 747 768 7040 7040 7040
	179	00011 0001 0001 0000 000 000 000 000 00	268 208 208 208 208 208 208 208 208 208 20	200 200 200 200 200 200 200 200 200 200
P, kg/ha kg/ha	134	401 408 404 600 600 600 600 600 600 600 600 600	2000 2000 1300 1000	110 100 100 100 100 100 100 100 100 100
19 P, kg/ N, kg/ha	90	#4 #0 #8 #4 #4 #4 #4 #4 #4 #4 #4 #4 #4 #4 #4 #4	233 1893 200 200 200 200 200 200 200 200 200 20	427 427 500 500 100 100 100 100 100 100 100 100
	45	02400400 02400400	30.23.05 12.00 12.00 1.00	23 24 25 20 20 20 20 20 20 20 20 20 20 20 20 20
	0	0.01 88.00 0.00 0.00 0.00 0.00 0.00 0.00	1222500 1432500 1432500	844 844 846 846 846 846 846 846 846 846
	224	000440000 0000000000000000000000000000	00000000000000000000000000000000000000	00000000000000000000000000000000000000
	179	000000000000000000000000000000000000000	80 04 00 44 0000000	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~
g/ha /ha	134	0404000 04440000	000000000	~~~~~~~~~~ ~~~~~~~~~~~~~~~~~~~~~~~~~~~
0 P, kg/k N, kg/hs	90	11 202444441 22200220	8 N N 4 4 4 4 4 N N O N N O N O	00000000
	4.5	00000 0000 0000 0000 0000 0000 0000 0000	00000000000000000000000000000000000000	2 2 2 2 2 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3
	0	8 6 7 7 7 7 7 8 9 9 9 9 9 9 9 9 9 9 9 9 9 9	00000000000000000000000000000000000000	00000000000000000000000000000000000000
0	Depth,	Heb. 1 0-5 0-5 10-15 15-20 20-25 30-35 35-40	Rep. 2 0-5 0-5 10-15 15-20 25-30 30-35 35-40	Reb. 3 0-5 0-5 10-15 15-20 20-25 25-30 30-35

Total soil carbon (%) as affected by fertilizer treatment (kg/ha). Table 14.

	19 P 224 N	10111111111111111111111111111111111111
cation 3	19 P 0 N	2 4 2 3 2 4 2 3 4 2 4 3 8 0 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
Replica	0 P 224 N	229229323232323 229229323232323232323232
	0 0 N	11121122221111111111111111111111111111
	19 P 224 N	11111111111111111111111111111111111111
tion 2	19 P 0 N	11111111111111111111111111111111111111
Replication	0 P 224 N	223244242424242424242424242424242424242
	0 0 N	010 000 000 000 000 000 000 000 000 000
	19 P 224 N	00000000000000000000000000000000000000
tion 1	19 P 0 N	00000000000000000000000000000000000000
Replication	0 P 224 N	10000000000000000000000000000000000000
Sec. 50	0 0 N	3336998649346003 333698649346603 333698649346603
	Depth,	988705505050 9505050505050505050505050505050

42 19 224 PZ Replication 00 AZ (kg/ha) 224 treatment NP 19 224 fertilizer N AZ Replication 19 Ъу AZ 224 affected 155565 155565 155565 155565 155565 155565 155565 155565 155565 155565 155565 155565 155565 15566 155665 156665 15666 156665 1566 AZ ឧ (% AZ 122244 12224 19 224 carbonate-carbon 100000 10000 AZ Replication 00 NA 0 54 Soil AZ 15 Depth, 0 S Tabl.

Soil organic carbon (%) as affected by fertilizer treatment (kg/ha). Table 16.

	19 P 224 N	000000000000000000000000000000000000000	
tion 3	19 P 0 N	000000000000000000000000000000000000000	
Replica	0 P 224 N	000000000000000000000000000000000000000	
	0 O		
	19 P 224 N	00000000000000000000000000000000000000	00000
tion 2	19 P 0 N	00000000000000000000000000000000000000	
Replica	0 P 224 N	00000000000000000000000000000000000000	الم الم الم
	0 N N	00000000000000000000000000000000000000	-00-0
	19 P 224 N	000000000000000000000000000000000000000	00400
tion 1	19 P 0 N	00000000000000000000000000000000000000	
Replication	0 P 224 N	00000000000000000000000000000000000000	0.4.0.04
	0 0 P4 N	00000000000000000000000000000000000000	
	Depth,	70000000000000000000000000000000000000	00000000000000000000000000000000000000

Table 17. Soil cation exchange capacity (meq/100 g) as affected by treatment.

			224	٠,	, (, 4	, v	, ~	· · ·	-	, ,	, q	, . , 	· -	· -	-	-	1	· -	1	1	2	22.0
	19 P, kg/ha	N, kg/ha	134	ς.	, (C	·	~	3	21.8		-		-	;	-	0	0	7	.		· -	+	+
tion 1			0	2	N	3	2	2	21.4	-	Ţ	7	0	0	+	0	0	‡	ij	+	7	7	21.6
Replication		o (224	2	2	2	2	2	3	2	2	-	.	ij	ö	+	-	2	 1	.			21.5
	0 P, kg/ha	N, kg/ha	134	2	8	3	2	-	21.2	0	_ .	H	0	0	0	ċ	0	0	H	÷	+	<u>ਂ</u>	2
			0	8	2	*	c'	-	21.4	0	+	0	0	ċ	0	0	.	7	8	0	0	0	
		, , ,	cm cm	15	5-1	0-1	5-2	0-2	25-30	0-3	5-4	7-0	5-5	0-5	5-6	9-0	5-2	0-2	2-8	0-8	5-9	6-0	5-1

Soil cation exchange capacity (meq/100 g) as affected by treatment. Table 18.

			Replication	ation 2		
		0 P, kg/ha			19 P, kg/ha	
		N, kg/ha			N. kg/ha	
	0	134	727	0	134	524
54	ů		ω,	22.6	· ·	23.6
25	0		6	ď	~	· ·
25	.2		å	3	÷	٠.
25	4 .		3.	3	÷	÷
-+	9.		å	3	~	Š
25	.2	23.7	25.4	25.0	24.3	25.0
56			• -	3	o.	÷
3	9.		ď	7	-	-
N	ń		÷	1.	-	÷
N	0		1.	÷	•	-
N	τ.		4	0	÷	;
N			·	•		÷
N	⊣.		1.	0	<u>, ;</u>	-
N			o	•		-
\vdash	⊅.		ö	7	0	0
2			0	Ţ	o.	-
N					ं	+
21	9.		4	÷	o	.i
22			ä	÷	÷	÷
22			÷	÷	ċ	÷

by treatment. 224 P, kg/ha kg/ha 134 as affected z. 19 0 (g Replication cation exchange capacity (meq/100 224 kg/ha kg/ha 134 <u>م</u> ż 0 0 Soil 19. Depth, 0 E Tabl

1. 12122 123122 123311 123411 13311 13014 14136 14136 17488 17488 16544 16544 16544 224 as affected by treatment -- replication kg/ha kg/ha 15106 15279 16824 16824 17704 17704 18666 18666 18709 17104 16179 16089 17068 134 ď ż 19 16547 173366 173366 173366 173366 17337 17372 17372 17372 17372 17372 17372 17372 17372 17372 17372 17372 17372 6029 0 Extractable soil calcium (ppm) 06352 052237 052237 052237 120523 130521 14680 182107 18355 18355 164174 164174 164174 224 kg/ha kg/ha 66079 60199 60 134 ф ф Z 0 17947 173889 173889 173889 198864 174168 174168 174168 174168 17378 17378 17378 17378 17378 17378 17378 17378 0 20. Depth, Table CH

20174 19629 17934 18612 16947 05960 05960 04869 04869 04752 14591 16736 16724 17324 18043 19705 N 224 treatment -- replication kg/ha kg/ha 0.5620 0.5620 0.5523 0.55680 0.5680 0 134 19 P. N. 0.5400 0.5400 0.65330 0.65330 1.65536 0 affected by 8 calcium (ppm) 04987 04555 045555 046555 046551 18558 118638 116528 116528 118345 118345 224 kg/ha kg/ha 06000 06530 06530 06530 06530 05276 17264 18704 17804 18049 19244 19244 19462 134 soil 4 ž O Extractable 06100 05903 05159 05159 05159 15284 16607 16607 18291 17799 17799 17799 06230 07282 0 21. 9887765 905110 905110 905110 905110 905110 905110 905110 905110 905110 905110 905110 905110 905110 905110 905110 905110 Depth, Table G E

05642 07724 07724 07724 09435 19009 17650 16260 16260 17963 17964 17964 17964 17964 17964 17964 17964 17964 as affected by treatment -- replication 224 kg/ha kg/ha 07638 076363 066363 066363 114339 117339 12248 20272 202841 20841 20843 20843 20843 20843 20843 20843 134 ď ż 19 06374 0058454 0054945 1054945 105663 105663 116663 119668 119668 119668 119668 119668 5909 0 Extractable soil calcium (ppm) 007209 0067401 0067401 008355 115953 117342 117342 117359 117359 117374 118374 118374 118374 224 P, kg/ha kg/ha 06075 05050 05050 05050 10197 11632 12400 12659 15391 16179 14919 14919 14919 14919 14919 14919 14919 14919 14919 06002 134 N 0 00743 008855 008855 108855 110885 110 0 22. 20-70 20-70 20-70 20-80 20-80 20-90 100 Depth, Table 日

ij treatment--replication 224 kg/ha kg/ha 134 ь, z 19 by 0 affected 8 magnesium (ppm) 224 kg/ha kg/ha 8888330 888830 88880 888830 88880 88800 88800 88800 88800 88800 88800 88800 88800 88800 88800 88800 88800 8 134 soil ď N. 0 Extractable 0 23 Depth, rable CE

2 magnesium (ppm) as affected by treatment--replication 224 kg/ha kg/ha 888887778777780878 88787877787878 88787877787878 888887778787878 134 ь. С ż 19 0 224 kg/ha kg/ha 88476644019076777 88476640190788077 88476640190788077 134 Extractable soil **Д** Z 0 7.78 8.73 0 24. Depth, Table ន ប

3 treatment--replication 224 P, kg/ha N. kg/ha 134 19 рy 8878888877688778877 8878888887788778877 88778871788717887 0 affected as magnesium (ppm) 224 kg/ha kg/ha 134 8888777 87277 87277 88887 8887 8888 8777 8778 soil Р, N. 0 Extractable 0 25. Depth, rable CE

ij 224 Extractable soil sodium (ppm) as affected by treatment--replication kg/ha kg/ha 134 ż 19 0 224 P, kg/ha kg/ha 134 N. 0 0 26. Depth, Table CH

N Extractable soil sodium (ppm) as affected by treatment--replication 19 P, kg/ha N, kg/ha 134 0 224 P, kg/ha N, kg/ha 134 0 0 27. 9887765 9887765 9887765 1007 100 Depth, Table 5

3 treatment--replication kg/ha kg/ha 134 ž 19 0 ЪУ affected ន (mdd) 224 sodium kg/ha kg/ha 134 soil 4 z 0 Extractable 77888887878700002200 72888887878700002200 28 Depth, Φ Tabl E E

as affected by treatment -- replication 1. 224 19 P, kg/ha kg/ha 134 , N 8887769065776577669 731838617386657767669 731838776773806673767007 0 potassium (ppm) 224 0 P, kg/ha N, kg/ha 134 Extractable soil 0 29. Depth, Table 日

2 treatment--replication 224 kg/ha kg/ha 134 ᆄ 19 ż Ъу 0 affected ಜ potassium (ppm) 224 kg/ha kg/ha 134 soil ď ž 0 Extractable 010000 to 0000 to to 0000 to 0 0 30. Depth, Table 日

by treatment--replication 224 874790178877808718881 87478877887788881 87478877887788888 kg/ha kg/ha 134 凸 Z 19 0 affected S (mdd) 224 potassium kg/ha kg/ha 134 8877678 80406760 8040676 80406 soil **д** ż 0 Extractable 0 31 9880 901-150 1001-100 1001-110 Depth, Table S

628 67 67 67 67 67 67 67 224 88488 26262 treatment. 19 P, kg/ha kg/ha 38228 669629 134 26238 ż 240 480 80 80 80 80 80 98820 64833 64883 0 as copper (ppm) 224 99955 92522 99996 26668 P, kg/ha DTPA-extractable soil N, kg/ha 28882 62 62 73 67 134 \$288 \$28 \$28 0 668659 62869 22225 32355 0 Rep. 2 Depth, 0-10 10-20 20-30 40-50 0-10 10-20 20-30 30-40 40-50 0-10 10-20 20-30 30-40 40-50 Table Rep. C E Rep.

affected by 32.

00460 00000 00000 33.58 500 to 000 to 00 224 19 P, kg/ha DTPA-extractable soil iron (ppm) as affected by treatment. kg/ha 134 ż 22.4.4.9 24.4.99 200000 0 2000 2000 2000 2000 2000 333 FF 4.82 3.33 2.787 61 224 kg/ha kg/ha 4.55.00 1.29.20 1.39.20 25659 25669 25669 4.57 6.07 6.57 6.57 134 <u>ц</u> z 0 20002 20002 0 33. Rep. 3 Depth, 0-10 10-20 20-30 30-40 40-50 0-10 10-20 20-30 30-40 40-50 0-10 10-20 20-30 30-40 40-50 Table Rep. Rep. E O

Table 34. DTPA-extractable soil manganese (ppm) as affected by treatment. ă l ă

		0 P, kg/ha			19 P, kg/ha		
		N, kg/ha			N, kg/ha		
Depth,	0	134	224	0	134	224	
Rep. 1					٠		
0-1	0.4.0	400	100	4.4.	20,	00.	
30-40 40-50	44. 25. 25. 25.	8.70 6.02 6.40 6.40	4.50 0.10 0.00	3.86	4.11	4.33 4.33	
Rep. 2							
0-10	16.12	14.37	12.96	14.12 12.52 8.83	14.02	15.28 12.57	
140	5.57	in	400	1000	$i\omega \omega$	icn	
Rep. 3	¥2						
0-1	, ·	6.7	2.1	∞ Q	wσ	0.0	
20-30	6.79	3.75	6.24	3.68	7.4.5 2.8.5 2.5.5	2.90	
0-5	٦.	0	7.	٦.	٦.	ż	

by

GALEN RAY NIEDENTHAL

B. S., Kansas State University, 1969

AN ABSTRACT OF A MASTER'S THESIS

submitted in partial fulfillment of the

requirements for the degree

MASTER OF SCIENCE

Department of Agronomy

KANSAS STATE UNIVERSITY

This investigation, part of a long-range study initiated at the Tribune Branch Experiment Station in 1961 to evaluate the response of irrigated corn to various rates of fertilizers, was concerned with the effect of continuous applications of nitrogen and phosphorus on the chemical properties of the Richfield silt loam soil, and to determine the fate of applied nitrogen not utilized by the corn crop. Soil core samples were taken in January and February of 1972 to a depth of 4 meters and analyzed for several soil constituents.

Ammonium-nitrogen accumulation in the soil was high, exceeding 200 ppm in some samples near the soil surface. There was a significant (0.05) decrease in ammonium-nitrogen with depth. Ammonium-nitrogen was significantly increased by the addition of 19 kg P/ha, but the increase due to applied nitrogen was not significant.

Soil nitrate-nitrogen concentrations were much lower than ammoniumnitrogen levels, rarely exceeding 25 ppm. Nitrate-nitrogen decreased with depth, and there were no noticeable peaks of nitrate accumulation in the profile. Nitrogen and phosphorus treatments increased the amount of nitratenitrogen in the soil as fertilizer rates increased.

Total soil nitrogen, excluding nitrate-nitrogen, decreased with depth. Nitrogen applications had no measurable effect on total soil nitrogen, but the addition of 19 kg P/ha significantly decreased total soil nitrogen.

Both nitrogen and phosphorus significantly lowered soil pH, even though measured differences were small. Soil pH increased with depth, ranging from approximately 7.9 near the surface to 8.3 at the 400 cm depth.

Weak Bray extractable phosphorus decreased as nitrogen rates increased, and increased markedly when 19 kg P/ha were applied. There was a marked decrease in available phosphorus with depth.

High rates of nitrogen fertilizer tend to decrease extractable calcium and magnesium in this soil. Phosphorus had no significant effect on calcium and magnesium. Extractable sodium content was increased by nitrogen fertilization when phosphorus was applied. Sodium concentration decreased when phosphorus was applied by itself. Sodium concentration in the profile varied quite erratically and was not affected by depth.

Extractable soil potassium content decreased when increasing nitrogen rates were applied. Addition of 19 kg P/ha produced an increase in extractable soil potassium. Potassium levels were quite high, explaining the lack of crop response to applied potassium at the experimental site.

Soil cation exchange capacity, ranging from 20-25 meq/100 g, was not significantly affected by nitrogen or phosphorus treatments. Soil cation exchange capacity decreased with depth.

Levels of all micronutrients seemed to be adequate in the upper 15 cm of soil. Iron, manganese, and zinc decreased with depth, but copper had a zone of highest concentration at the 25-35 cm depth. Phosphorus had a significant effect only on manganese concentration, causing a decrease in DTPA-extractable manganese. Conversely, nitrogen applications led to an increase in soil extractable manganese.

Nitrogen and phosphorus had inverse effects on all forms of soil carbon, but measurable effects were quite small and usually insignificant. Carbonate-carbon comprised from 75-80% of the total soil carbon when averaged across all depths.