Asymetric Oxidation Reactions Catalyzed by Chiral Substituted Polymers / Nanoclusters; Synthesis of 6-(dimethylamino)-2-phenylisoindolin-1-one Derivative.

by

Bo Hao

B.S., Xiamen University, 2011

AN ABSTRACT OF A DISSERTATION

submitted in partial fulfillment of the requirements for the degree

DOCTOR OF PHILOSOPHY

Department of Chemistry College of Arts and Sciences

KANSAS STATE UNIVERSITY Manhattan, Kansas

2018

Abstract

The discovery of new methodologies to advance the fields of synthetic organic, nanoclusters, and polymer chemistry is critical in the asymmetric synthesis of organic compounds. Particularly, catalytic asymmetric oxidation reactions are economic. The oxidation reactions provide chiral molecules and additional functionality onto the molecules for functional group manipulation. New kinds of polymers, namely chiral-substituted poly-*N*-vinylpyrrolidinones (CSPVPs), stabilize the bimetallic nanoclusters such as Pd/Au or Cu/Au and induce chirality. These chiral polymers wrap around the nanometer-sized (~3 nm) bimetallic nanoclusters and catalyze a number of enantioselective oxidation reactions using oxygen or hydrogen peroxide as the oxidant. Cycloalkanediols were asymmetrically oxidized by 1 atm of oxygen gas to yield the corresponding hydroxyl ketone under the catalysis of Pd/Au (3:1) - CSPVP nanoclusters. Alkenes were oxidized by Pd/Au (3:1)-CSPVP nanoclusters under 2 atmospheric of oxygen in water to give the syn-dihydroxylated products in high chemical and excellent optical yields. Various cycloalkanes underwent regio- and enantio-selective C-H oxidation with Cu/Au (3:1)-CSPVP and 30% hydrogen peroxide to produce the corresponding chiral oxo-molecules in very good to excellent chemical and optical yields. We further discovered an enantioselective desymmetrization of α, α -dialkenyl-alkanols and α, α -dialkenyl-amino acid ethyl esters to give chiral disubstituted lactones and lactams, respectively. A number of medium-sized natural products and drugs were also oxidized regioselectively to give the corresponding mono-oxygenated products. A broad-spectrum predictive C-H oxidation of complex molecules is possible. Chapter 1 mainly discussed the synthesis and characterization of the new classes of chiral substituted PVP and bimetallic nanoclusters. Chapter 2 focus on various kind of oxidation reactions by the catalysis of CSPVP stabilized bimetallic nanoclusters.

Among various bioluminescence assays, firefly luciferase based bioluminescence assays are popular due to their high specific activity, low background noise and ease of use. However, it has been found that some aromatic carboxylic acid substantially inhibited the firefly luciferase reporter enzyme's activity. In order to study firefly luciferase inhibition and the proteins associated with inhibition mechanism, we designed two 6-(dimethylamino)-2-phenylisoindolin-1-one derivatives as probe molecules. The synthesis of one probe molecule is discussed in Chapter 3 and the further investigation of its inhibitory activity on firefly luciferase is being conducted by our collaborator, Dr. Madoka Nakagomi at Itsuu Foundation, Japan.

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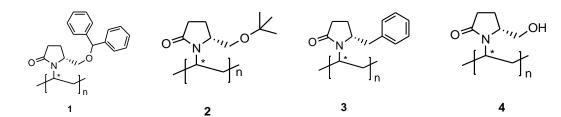
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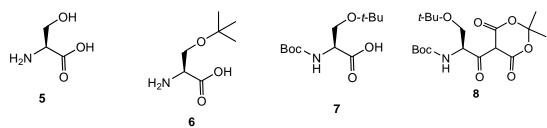
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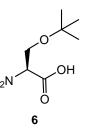
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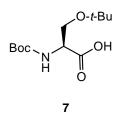
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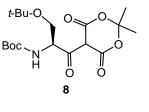
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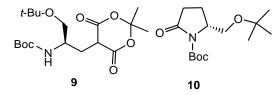


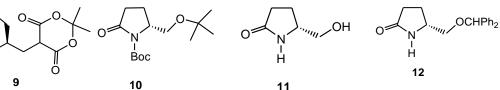


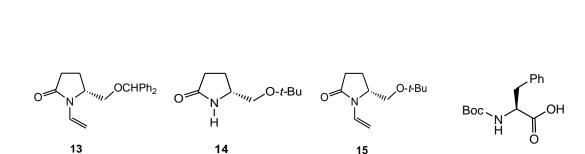




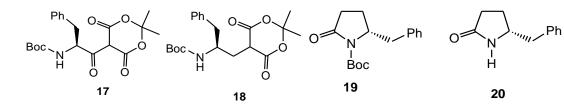


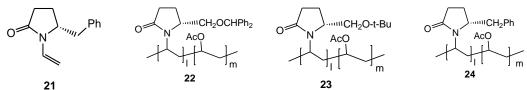


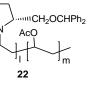


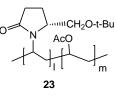


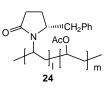
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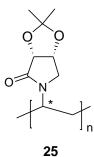


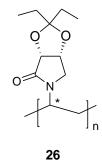


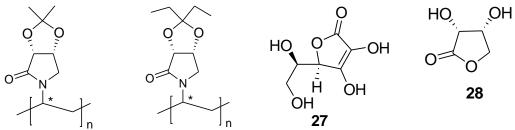


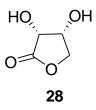


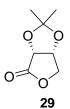


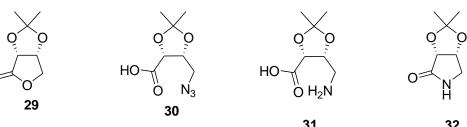


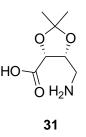


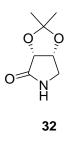


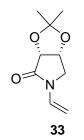


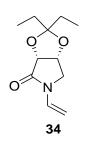


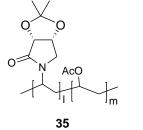


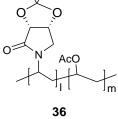


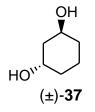


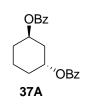


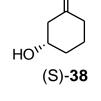




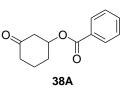




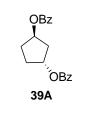


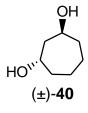


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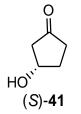


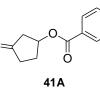
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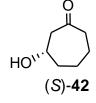


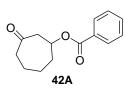


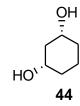


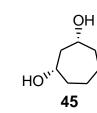


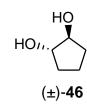


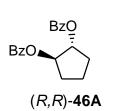


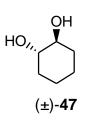


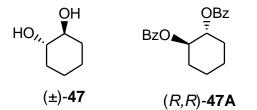


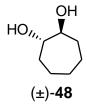




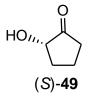


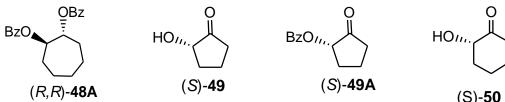


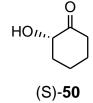


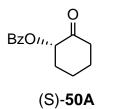


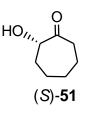
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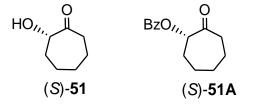


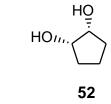


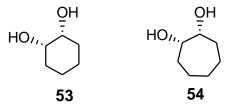


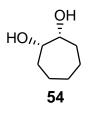


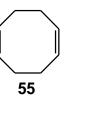


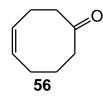




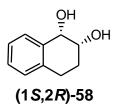


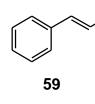


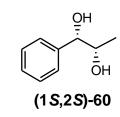


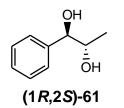


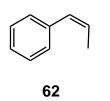


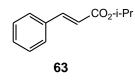


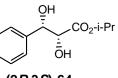




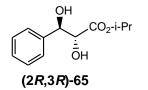






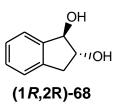


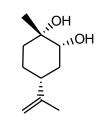
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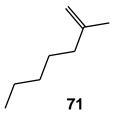
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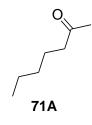
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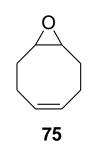


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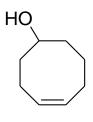
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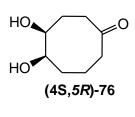
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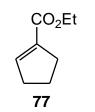


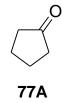
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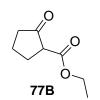


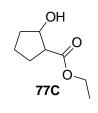


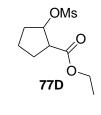


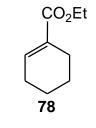


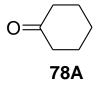
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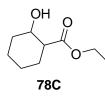


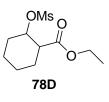


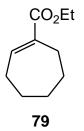


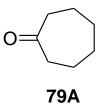


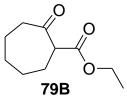


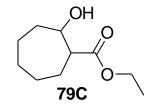




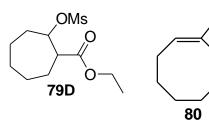


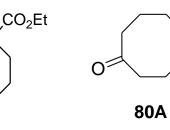


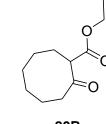




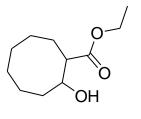


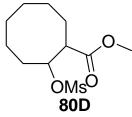


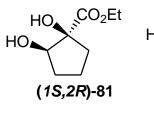


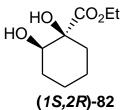


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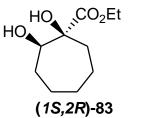


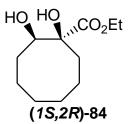


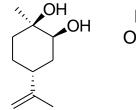


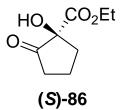
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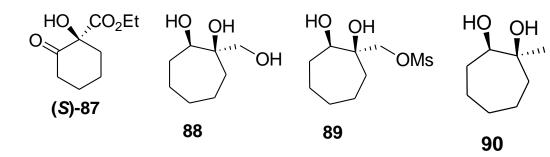


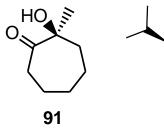


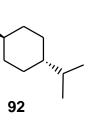


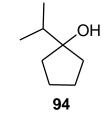


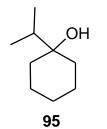
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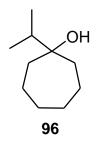


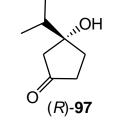






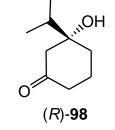


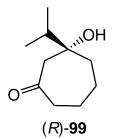


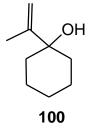


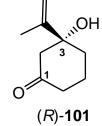
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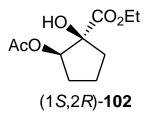
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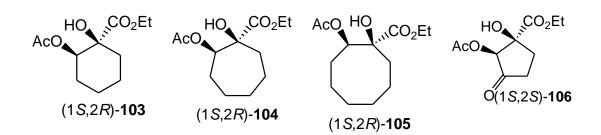


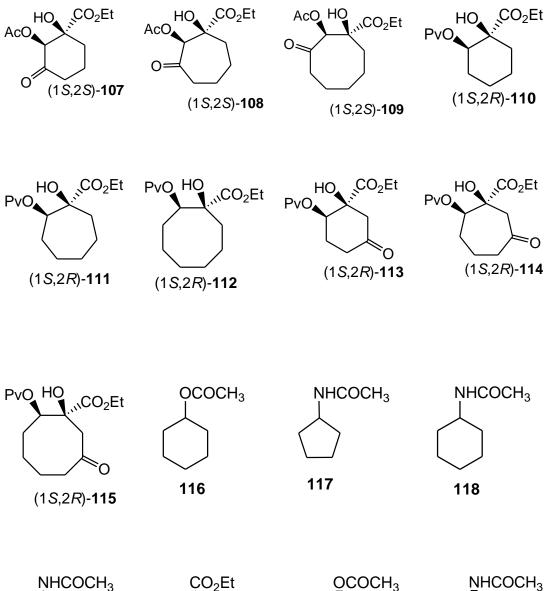


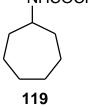


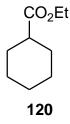


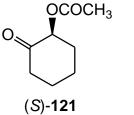


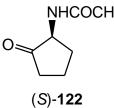


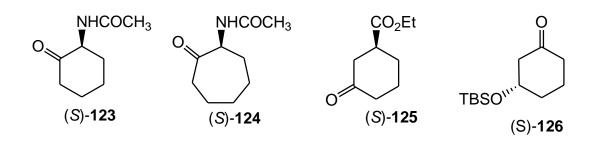


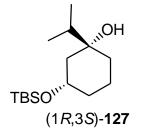


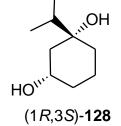


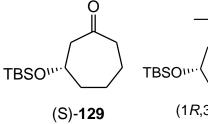


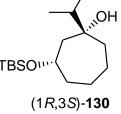


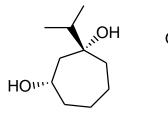




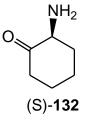


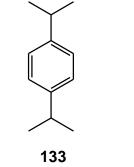


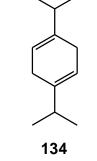


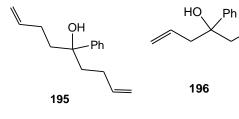


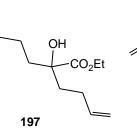
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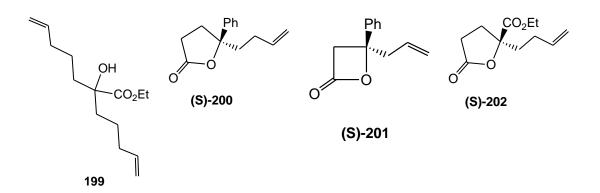


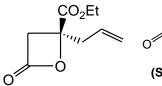


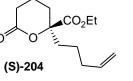


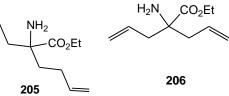
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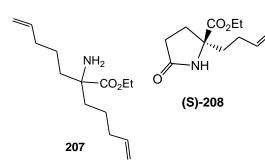


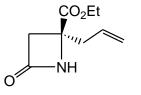




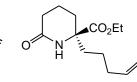


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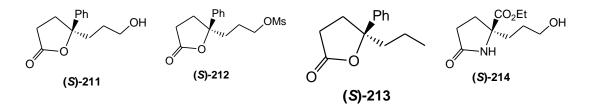


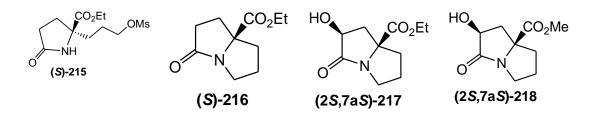


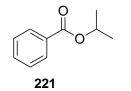
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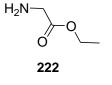


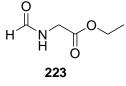
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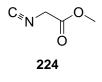


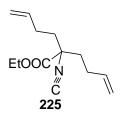


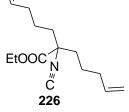


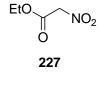


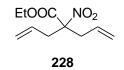


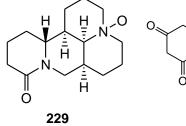


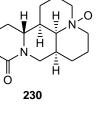




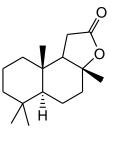




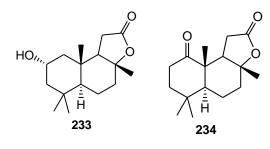


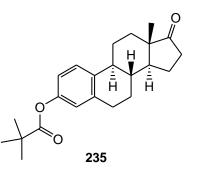


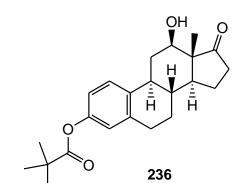
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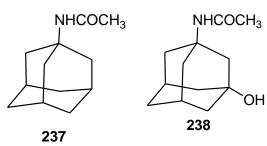


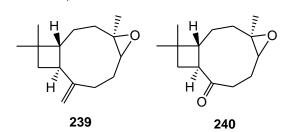
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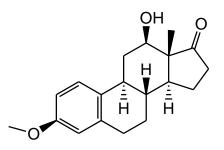


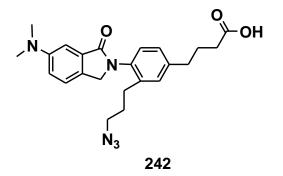






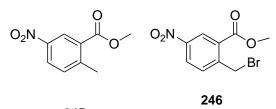






OH O₂N

ОH





245

247

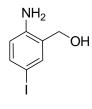


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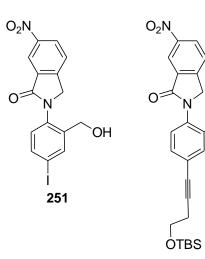
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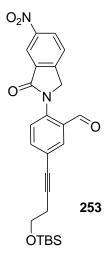
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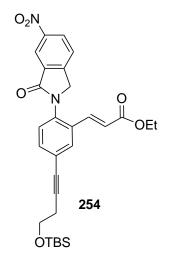


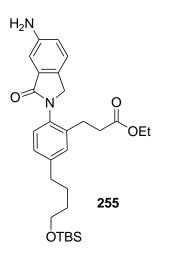
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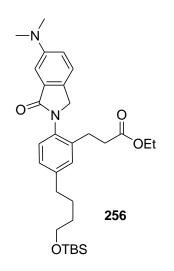


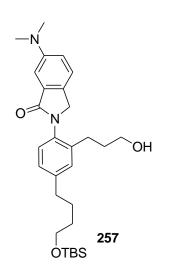


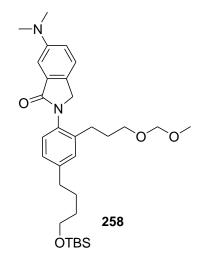


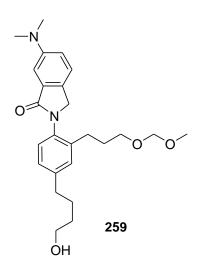


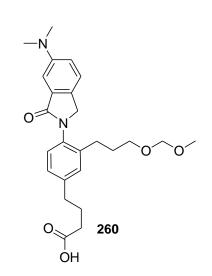


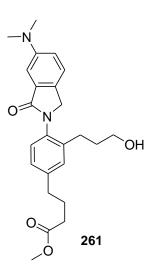


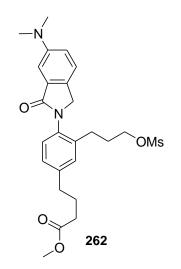


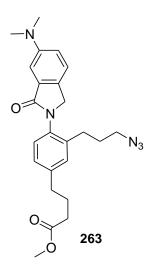












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List of Symbols

 δ
 delta, chemical shift

 ee
 enantiomeric excess

List of Abbreviations

PVP	Poly-N-vinyl-pyrolidinone	
PAMAM	Poly(amidoamine)	
THF	Tetrahydrofuran	
CSPVP	Chiral substituted poly-N-vinyl-pyrolidinone	
AIBN	Azobis(isobutyronitrile)	
NVP	N-vinylpyrrolidone	
DMF	Dimethylfomaide	
DP	Degree of polymerization	
GPC	C Gel Permeation Chromatography	
DLS	Dynamic Light Scattering	
AFM	Atomic Force Microscopy	
NMR	Nuclear magnetic resonance	
TEM	Transition electron microscopy	
XPS	X-ray photoelectron spectroscopy	
CD	Circular dichroism	
TON	Turnover Number	
DCM	Dichloromethane	
EA	A Ethyl Acetate	
HTS	TS High throughput screening	
ATP	Adenosine triphosphate	
D-LH ₂	D-isomer of firefly luciferin	
AMP	Adenylate	
Co-A	Coenzyme A	
NBS	N-Bromosuccinimide	
TBSCI	Tert-Butyldimethylsilyl chloride	
EtOH	Ethanol	
PCC	Pyridinium chlorochromate	
DIBA1-H	Diisobutylaluminium hydride	

MOMCl	Chloromethyl methyl ether
DIPEA	N,N-Diisopropylethylamine
IBX	2-Iodoxybenzoic acid

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Dedication

My beloved wife, Jiawei Gong, and my parents: Mr. Jianmin Hao and Mrs. Lianqing Chu. They are the ones who always stand by my side and encourage me all the time during my Ph.D. study. Thank all my friends in and outside the United States for their companion and friendship in my five years abroad study.

Chapter 1 - Synthesis and Characterization of Chiral Substituted Poly-*N*-vinylpyrrolidones and Bimetallic Nanoclusters

(Partial of the results and discussions in this chapter was published^[1] and adapted with permission of *Journal of the American Chemical Society*. Copyright © 2016 American Chemical Society.)

1.1 Background and Significance

Catalyst is considered as a chemical compound capable of directing and accelerating reactions while remaining unaltered at the end of the reactions. There are many types of catalysts used in the industry or specific reactions, including transition metal salts,^[2] organic molecules,^[3] enzymes,^[4] organometallic compounds,^[5] etc. They can be divided into groups as heterogeneous, homogeneous and biological.^[6] Beside the biological catalyst, the homogeneous catalysts mostly have higher catalytic activity since it can dissolve in the solvent and contact with the substrates directly. However, they often decompose rapidly during the reaction conditions.

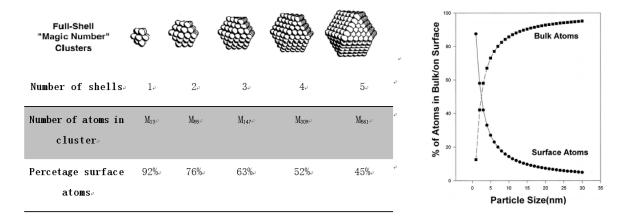


Figure 1. Inverse Relationship Between the Total Number of Atoms in Full Shell Clusters and the Percentage of Surface Atoms.

Transition metal nanoclusters gain great interest recently due to their unique chemical and physical properties result from their nano-dimensions.^[7] Metal clusters have a geometry rule which is designed full-shell or magic number clusters. The number of atoms increases while the percentage of the surface atoms reduced. (**Figure 1**^[8])

Transition metal nanoclusters are nearly monodispersed particles and generally are less than 4 nm in diameter. They have generated great attentions over the past decade. Nanoclusters also have notable potential as new type of catalysts and they also have high activity and selectivity than standard metal catalysts. Without stabilizing material, the metal nanoclusters likely aggregate, due to their high surface area to volume ratio. To overcome aggregation and provide high nanoclusters stability, polymer-stabilized metal nanoclusters have been used.^[9]

Polymer science, which focuses on primarily synthetic polymers, becomes a hot research area in the recent years. Developing methods for the synthesis of polymer with specific functional is critical on both research and industry purpose. Polymer-stabilized chiral catalysts and reagents have received considerable attention in regard to organic synthesis of optically active compounds.^[11] The use of polymer-stabilized catalysts has become one of the essential techniques in organic synthesis. ^[10-12] From the point of view of green chemistry, the polymer-stabilized chiral catalysis method provides a clean and safe alternative method of asymmetric processes. Not only their practical aspect but also the particular microenvironment they may create in a polymer network will make them attractive for utilization in organic reactions, especially in stereoselective synthesis. Chiral polymer synthesis that is directed toward chiral catalysts must be developed. In this case, the polymer, which will be used for the stabilization of bimetallic nanoclusters as asymmetric catalyst, need to induce a stereogenic center on the polymers.

Most polymeric support materials used for the chiral catalyst have been crosslinked polystyrene derivatives, mainly because of their easy preparation and introduction of functional groups on the side chain of the polymer.^[13, 14] Each polymer support would provide a specific microenvironment for the reaction if they can be precisely designed.

Alloying has been serving as an effective method for modifying the catalytic properties of transition-metal surfaces. Supported bimetallic clusters regularly exhibit catalytic properties superior to those of their single-metal counterparts.^[15-17] For this reason, they have attracted great interest in recent years ^[16, 18]. Due to the high ratio of surface area and volume, different kinds of supporting materials are needed, including inorganic oxides, carbon,^[19] organic ligands,^[20] organic polymers,^[10] etc. However, the use of chiral polymers as supporting material and chiral inducers has not been developed. Herein, poly-*N*-vinylpyrrolidinone, which is so called PVP as abbreviate, have been focused and chiral substituted PVP was designed and synthesized. Normally, PVP is a polymeric fine chemical obtained from its monomer, *N*-vinylpyrolidinone (NVP), by free radical polymerization in laboratory. It is soluble in water with a very high solubility, and also soluble in most of the organic solvent, such as alcohols, carboxylic acids, halogen alkanes, esters, ketones and THF. Moreover, PVP is relatively stable under high temperature and no significant changes under 250 °C.

1.2 Synthesis of Chiral Substituted Poly-N-vinylpyrrolidinone

Poly-*N*-vinylpyrrolidinone, as a chain polymeric material, has two ways to induce the chirality including side-chain chirality and main-chain chirality^[21].(**Figure 2**) Since there are only two carbon the backbone of PVP in each unit and polymerization with the radical mechanism, it is difficult to control the chirality on the main-chain of PVP. Making a chiral substituted

N-vinylpyrrolidinone, as the monomer of chiral substituted PVP, becomes important and sufficient. In particular, with the successful synthesis of CSPVP, we realized the tacticity of CSPVP is not randomly assembled in certain cases. I will discuss it later.^[22]



Figure 2. Polymer Having a Side-chain Chiral Ligand or Main-chain Chiral Ligand.

On the five-membered pyrrolidinone ring, there are three secondary carbons which can induce the stereogenic center. Base on that, we designed and synthesized two classes of CSPVPs. One is the 5-substituted chiral polymer, and the other one is the 3,4-disubstituted chiral polymer. (**Figure 3**)

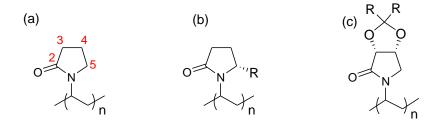


Figure 3. (a) PVP (b) 5-substituted Chiral PVP (c) 3,4-disubstituted Chiral PVP.

1.2.1 Synthesis of 5-Substituted Chiral PVP

We synthesized four 5-substituted chiral PVPs 1 - 4 from cheap optically pure amino acids. (**Table 1**) The synthetic procedure can be divided into two parts, the synthesis of the corresponding monomer and polymerization.

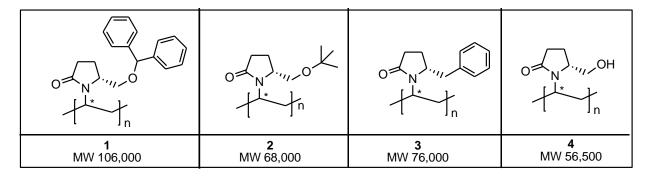
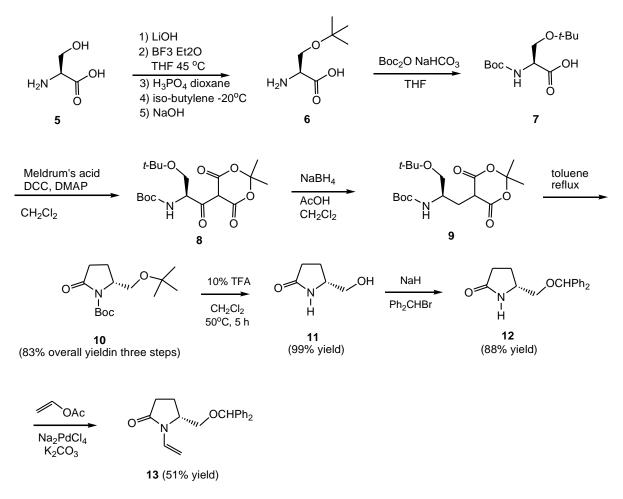


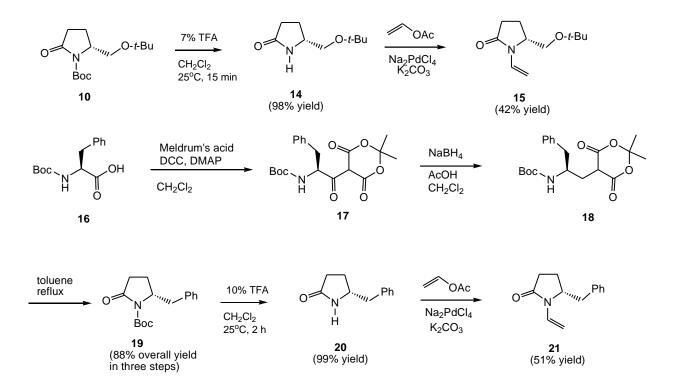
Table 1.5-Substituted Chiral PVP 1 - 4.

The synthetic route for the monomer of CSPVP 1 is shown in Scheme 1. Chiral compounds 11 were made from L-amino acids by following a reported procedure.^[23-26] N-Boc-O-t-butyl L-serine 7 is commercially available material or made from L-serine 5. L-serine was basified with LiOH, complexed with BF₃•EtO₂ to protect both amino group and carboxylic acid, followed with protecting the hydroxyl group by isobutylene and removed boron complex with NaOH to afford *O*-tert-butyl L-serine (6). The amino group in compound 6 was protected by Boc_2O in the presence of base to give compound 7. Compound 7 was coupled with Meldrum's acid with coupling reagent DCC and DMAP, followed by reduction of the ketone function with sodium borohydride, cyclization under heat, and removal of both the Boc group and O-tert-butyl group with trifluoroacetic acid (TFA) in CH₂Cl₂ under 50 °C to give compound 11. Consequently, (R)-5-benzhydryloxymethyl-2-pyrrolidinone (12) was synthesized from the alkylation of compound 11 with sodium hydride and benzhydryl bromide. N-Vinylation of 12 with Na₂PdCl₄, acetate^[27] K_2CO_3 and vinyl under anhydrous condition gave corresponding N-vinylpyrrolidinones 13 with 51% yield.



Scheme 1. Synthesis of N-vinyl-5-benzhydryloxymethylpyrrolidinone (13).

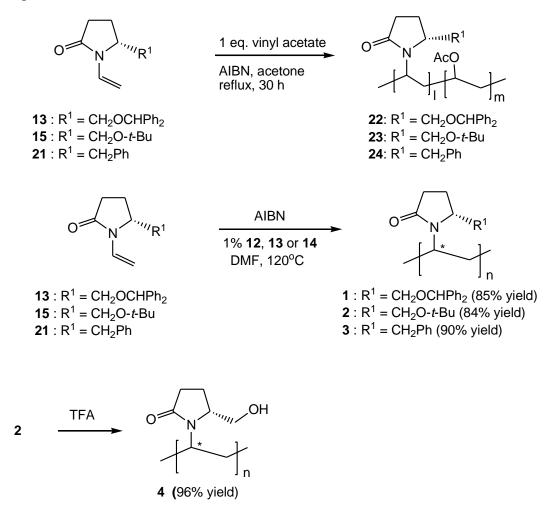
Notably, if compound **10** with 10% TFA in methylene chloride being refluxed under 50 °C for 5 hours, both the Boc group and *O-tert*-butyl group were removed. However, under room temperature for 2 hours, only Boc group in compound **10** was removed by 10% TFA in methylene chloride to give compound **14**. Compound **15** was obtained by the *N*-vinylation of compound **14** with vinyl acetate by the catalysis of Na₂PdCl₄ under base condition.^[27] Moreover, the synthesis of compound **21**, as the monomer of CSPVP **3**, can be made with similar method from Boc-phenylaniline. (**Scheme 2**)



Scheme 2. Synthesis of *N*-vinyl-5-substituted pyrrolidinone 15 & 21.

The preparation of poly-N-vinyl-5-substituted pyrrolidinone 1 - 3 were carried out by dispersion polymerization with N-vinyl-5-substituted pyrrolidinone (CSNVP) 13, 15, 21, as initial monomer, poly(N-vinyl-5-substituted pyrrolidone-co-vinyl acetate) 22 - 24 as dispersant, and 2, 2'-azobisisobutyronitrile (AIBN) as initiator.^[28] (Scheme 3) The copolymerization process of CSNVP 13, 15, 21 and 1 equiv of vinyl acetate with a free radical initiator for both vinyl compounds were refluxed in acetone to give corresponding copolymer. After the formation of copolymer, hexane was added to precipitate the polar polymer from acetone. For the polymerization of CSNVP, Zhai's group prepared polyvinylpyrrolidone (PVP) microspheres in polymerization NVP ethyl acetate by dispersion with initial as monomer, poly(N-vinylpyrrolidone-co-vinyl acetate) (P (NVP-co-VAc)) as dispersant, and AIBN as free radical initiator.^[28] Since the monomer, N-vinyl-5-substituted pyrrolidinone, is much more bulky than normal NVP, the connection of the free radical on the vinyl carbon becomes much more

difficult and needs higher energy to form the backbone of polymer. High temperature and high concentration of the monomer was conducted in the polymerization step. Polymerization of **13**, **15**, **21** separately with a catalytic amount of AIBN and in the presence of 1% of respective copolymer 22 - 24 in DMF at 120 °C for 7 days gave polymers 1 - 3. CSPVP is soluble in DMF and diethyl ether is also needed to isolate CSPVP from solvent. Polymer **4** was prepared through the removal of the *tert*-butyl group of polymer **15** with 10% TFA in dichloromethane. All the four CSPVPs are white solid and gradually become sticky after exposing in the air with the absorption of moisture.



Scheme 3. Polymerization for 5-substituted Chiral PVP.

The modification of CSPVP, such as the step from CSPVP **2** to CSPVP **4**, provides a sufficient method to get new CSPVP containing different functional group with different size and properties. I have also tried to synthesize CSPVP **1** from CSPVP **4** by treatment CSPVP **1** with NaH and benzhydryl bromide. Under a very harsh reaction condition, only 92% of the units were reacted and remained 8% of hydroxyl group on the new polymers. It is because the bulky group is difficult to connect onto the limited space in the polymers in some certain tacticity of CSPVP. It also gives side evidence that the 5-substituted chiral PVP, at least CSPVP **4** is atactic. It can also be confirmed by ¹³C NMR which shows many sets of signal on the similar carbon from different units. Both *R*- and *S*-stereochemistry (labelled * in **Table 1**) are likely presented in the stereogenic center of the polymer alkane backbone, and they are not identifiable.

1.2.2 Synthesis of 3,4-Disubstituted Chiral PVP

With the experience from the successful synthesis of 5-substituted chiral PVP, a new class of CSPVP was designed, due to the hypothesis that in the bimetallic CSPVP nanoclusters, the substituent on the pyrrolidinone ring controls the stereochemical outcome of the reactions, resulting in asymmetric oxidations. Two new polymers **25** & **26** (**Table 2**) are derived from D-isoascorbic acid (**27**), an inexpensive starting material comparing with chiral amino acids especially the unnatural amino acids.

The synthesis of polymer **25** & **26** was achieved from the corresponding monomer **33** & **34**. Chiral compound **32** was made from D-isoascorbic acid by following a reported procedure.^[22, 30, 31] (**Scheme 4**) D-Isoascorbic acid (**27**) was oxidative cleaved by 30% hydrogen peroxide and sodium carbonate. Acetonide formation was carried out by the coupling of compound **28** with acetone under the catalysis of anhydrous CuSO₄. Compound **29** was treated with sodium azide in *N*,*N*-dimethylformamide (DMF) at 100°C to open the lactone ring with S_N 2-type mechanism, followed by the reduction of the azido function with hydrogen and Pd/C in methanol and yielded very polar compound **31**. After heating compound **31** under high vacuum and heat (120 °C), compound **32** is formed and sublimated by using a sublimator. Vinylation of chiral lactam **32** with vinyl acetate (distilled), potassium carbonate and catalytic amount of freshly prepared Na₂PdCl₄ at 50°C gave *N*-vinyllactam **33** in an 80% yield.^[27] Compare with 5-substituted pyrolidinone, the yield of vinylation significantly increased, may due to the steric hinderance effect in **1** - **4**. The nitrogen in compound **33** is on a much opener site for an easier, faster vinylation and avoiding the side products.

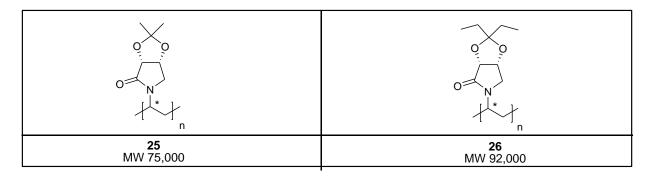
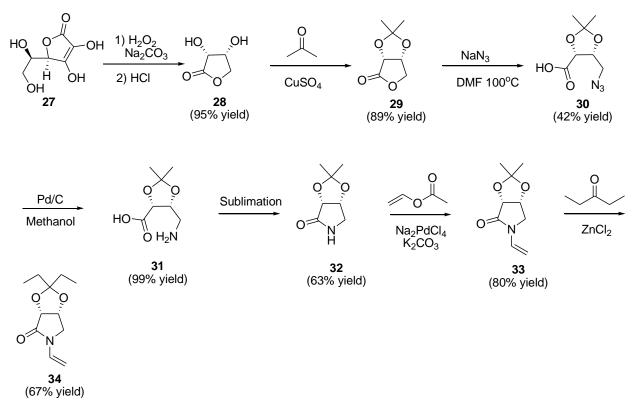


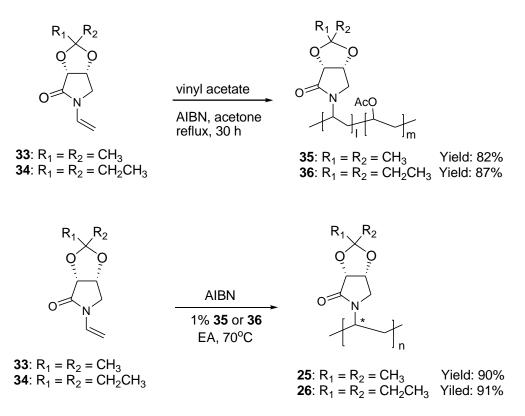
Table 2.3,4-Disubstituted Chiral PVP 25 & 26

To get compound **34**, similar procedure was done by using 3-pentanone instead of acetone to give (3aR, 6aR)-2,2-diethyl-dihydrofuro[3,4-d][1,3]dioxol-4(3aH)-one. After the azidation, reduction and sublimation, a new pyrrolidinone analog with diethyl group was obtained and became difficult for the vinylation without clear reason. Therefore, a new ketal exchange reaction was discovered by dissolving compound **33** in 3-pentanone with adding zinc chloride under 65 °C to distill off the produced acetone for 40 hours to yield 67% of the desired product **34**.



Scheme 4. Synthesis of *N*-vinyl-3,4-disubstituted pyrrolidinone 33 & 34.

Similar to CSPVP **1–4**, CSPVP **25 & 26** were prepared separately in about 90% yield by heating *N*-vinyl pyrrolidinone derivatives **33 & 34**, 1% of respective copolymer **35 & 36** and a catalytic amount of azobisisobutyronitrile (AIBN) in ethyl acetate at 70 °C for 2 days. The polymerization conditions are milder and reactions time are significantly dropped, comparing with CSPVP **1–4**, may also due to the size of the CSNVP.



Scheme 5. Polymerization for 2,3-Disubstituted Chiral PVP 25 & 26.

1.2.3 Characterization of Chiral Substituted PVP

All the CSPVPs are water soluble and their molecular weights were determined by gel permeation chromatography with CSPVP water solution in Dr. Shi's Lab in Grain Science, KSU. (**Figure 4**) TSKgel GMHxl column was used and THF as eluent solvent with a flow rate of 1 mL/min. The molecule weight (Mw) was shown in **Table 1** and **Table 2**. Base on the calculation, the degree of polymerization for CSPVP **1**, **2**, **3**, **25**, **26** are 345, 345, 378, 409, 436, respectively, which means the method for polymerization is constant. Because the relatively difficulty for the polymerization, degree of polymerization (DP) of CSPVP **1–3** are smaller than CSPVP **25** & **26**.

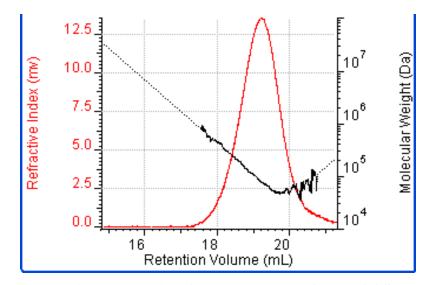


Figure 4. Gel Permeation Chromatography Chart of CSPVP 1.

The polydispersity index is used as a measure of the broadness of a molecular weight distribution of a polymer, and is defined by: Polydispersity index (PI) = Mw / Mn. The number average molecular weight, abbreviate as Mn, is the statistical average molecular weight of all the polymer chains in the sample, and the weight average molecular weight, shown as Mw, is quoted for a molecular weight distribution. There is an equal weight of molecules on either side of Mw in the distribution. The larger the polydispersity index, the broader the molecular weight. A monodisperse polymer where all the chain lengths are equal has an Mw/Mn = 1. Base on the results from GPC, the polydispersity index for CSPVPs are around 1.05 to 1.1.(**Table 3**) The result suggested the size of CSPVPs are uniform from the dispersion polymerization method^[28] and may give constant results on the catalytic activity of the bimetallic-CSPVP nanoclusters.

	CSPVP 1	CSPVP 25	CSPVP 26
Mw	106,184	75,245	91,988
Mn	100,602	68,185	85,232
Polydispersity Index	1.055	1.104	1.079

Table 3.Polydispersity Index of CSPVP 1, 25 & 26.

The average sizes, size distribution were also studied by dynamic light scattering (DLS). In **Figure 5**, Panel A and C are the autocorrelation function over time of **1** & **26** in H₂O (deionized) solution with concentration of 0.12 mol/L, respectively, Panel B and D are size distribution graph of **1** & **26** in H₂O solution with concentration of 0.12 mol/L. DLS graphs of **1** showed a major ensemble of 12–20 nm sizes particles with an average size of 18.6 nm along with a few 152 nm-sized particles, suggesting that different shapes and few large aggregates were presented in the aqueous solution. Similarily, DLS graphs of **26**, taken on ZetaPALS zeta potential analyzer, showed a major ensemble of 11–18 nm sizes particles with an average size of 14.6 nm along with a few 158 nm-sized particles. The size of polymer particle is also in a narrow band which fit the result of polydispersity from GPC analysis.

From the image of atomic force microscopy (AFM) obtained from Nanoscope IIIa SPM atomic force microscope, the size of the polymer can also be measured. (**Figure 6**) In panel A and C, AFM image of CSPVP **1 & 26** are shown and indicated the shapes and sizes of the polymers. Panel B and D measure the height (Y axis) and width (X axis) of polymer **1 & 26**. On the mica, the CSPVPs appeared unshaped with a diameter of ~15–30 nm and a height of 1 nm.

The NMR spectrum is difficult to analysis for the polymers since the signals in ¹H NMR are broad and ¹³C NMR is complex unless the polymer is regularly assembled. The ¹H & ¹³C NMR of commercially available PVP, CSPVP 1 - 3 are all difficult to analyze which means the stereogenic center on the polymers backbone can be isotactic, atactic, and syndiotactic.^[29] Fortunately, it likely has lesser effect on the asymmetric oxidation reactions which will discuss later. However, based on the ¹³C NMR spectra of polymers 25 and 26, they likely adapted syndiotactic or isotactic stereochemistry in the polymer backbone. For example, the ¹³C NMR spectrum of polymer 25 shows only 12 signals having δ values (in ppm) of 160.9 (C=O), 113.5 (O-C-O), 74.7 (C-O), 73.0 (C-O), 48.8 (C-N), 47.7 (C-N), 43.0 (CH₂), 42.7 (CH-N), 29.2 (CH₂), 28.5 (CH₂), 8.4 (CH₃) and 7.0 (CH₃), the simplicity of signal patents suggest the polymer was regularly assembled during polymertization. The exactly tacticity will be studied and confirmed.

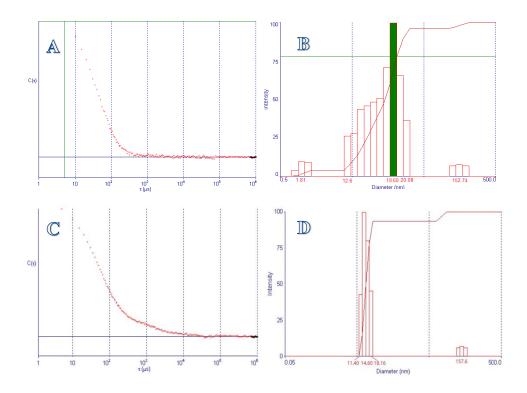


Figure 5. Dynamic Light Scattering Studies of CSPVP 1 (A & B) & 26 (C & D).

Since the purpose of making the different functionalized CSPVP is used for inducing a chiral environment on the asymmetric synthesis, the catalyst, CSPVP stabilized bimetallic nanoclusters was synthesized.

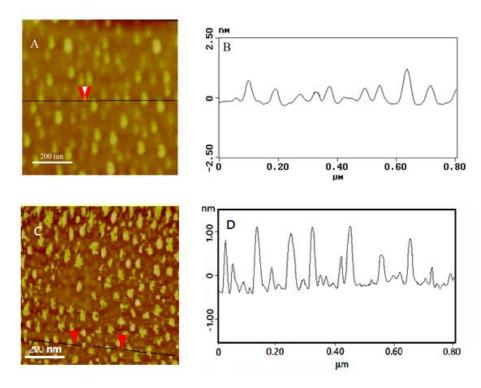


Figure 6. A Representative Atomic Force Microscopy Image and Measurement of Particle Height for CSPVP 1 (A & B) & 26 (C & D).

1.3 Synthesis and Characterization of CSPVP Stabilized Bimetallic Nanoclusters

Various of nanoclusters for different purpose have been made by number of methods including molecular beams, chemical reduction, thermal decomposition, ion implantation, electrochemical synthesis, radiolysis, sonochemical synthesis, and biosynthesis.^[32] We have synthesized various bimetallic nanoclusters including Pd/Au and Cu/Au, using the chemical reduction method^[10] in the presence of various of CSPVPs. We used gold due to its high catalytic activity and synergistic electronic effects, which will enhance the catalytic activity of Pd and Cu.^[10-12, 33-35]

1.3.1 Synthesis of Pd/Au-CSPVP Nanoclusters and Cu/Au-CSPVP Nanoclusters

In the bimetallic nanoclusters, both the valences of two metals are 0, which means the high valence metal salt need to be reduced in the solution. For example^[10], a solution of Na₂PdCl₄ (3 equiv.), HAuCl₄ (1 equiv.), and CSPVP **1** (0.11 equiv.) in deionized water was well mixed to give a yellow solution. To it, 40 equiv. of NaBH₄ were added under ice bath and stirred under room temperature for 30 min (**Scheme 6**) to give a light brown to dark grey solution depending on the concentrations. After adding NaBH₄, Pd(II) in Na₂PdCl₄ and Au(III) in HAuCl₄ were reduced to Pd(0) and Au(0), respectively. The color of the solution changes to black immediately to indicate the formation of Pd/Au-CSPVP nanoclusters. Because the presence of CSPVP as supporting material, Pd(0) and Au(0) do not aggregate due to the metal atom or nanocluster complex with the carbonyl of CSPVP. Different from the bulky metal (0), the high solubility of CSPVP stabilized bimetallic nanoclusters in water makes the nano-size metal as a homogeneous dispersion without precipitate. Without CSPVP, Pd and Au will form black precipitate after adding NaBH₄, suggesting CSPVP is nessecary for the formation of nanoclusters.

Theoritically, only 9 equiv. of hydride are needed for the reduction of Pd (II) (3 equiv.) and Au (III) (1 equiv.). Therefore, for certain reaction, especially C-H oxidation, only 8 equiv. of NaBH₄ was added to avoid the large excess NaBH₄ influencing the reaction outcome.

Similarly, Cu/Au-CSPVP nanoclusters were synthesized using the chemical reduction method. CuCl₂ is a good Cu source which has a high solubility in water, easily storage and relatively stability. However, the Cu/Au-CSPVP nanoclusters made from Cu (II) are not stable due to the precipitation. Therefore, CuCl was used for preparing the Cu/Au-CSPVP nanoclusters. Notably, CuCl can be slowly oxidized by HAuCl₄ to give CuCl₂. To avoid it, CSPVP (0.11 equiv.), HAuCl₄ (1 equiv.) and NaBH₄ (40 equiv.) were added into 0 °C CuCl (3 equiv.) in deionized water solution in sequence to give a dark brown solution. The solution was stirred under room temperature for 30 minutes and Cu/Au-CSPVP nanoclusters solution was ready to use for the asymmetric catalytic reactions. (Scheme 6)

$$3 \operatorname{Na_2PdCl_4} + 1 \operatorname{HAuCl_4:3H_2O} + 0.11 \operatorname{CSPVP} \xrightarrow{H_2O} \operatorname{Pd/Au} (3:1)-\operatorname{CSPVP} \\ \xrightarrow{\operatorname{NaBH_4}} 25^{\circ}\mathrm{C}, \ 0.5 \ \mathrm{h} \\ 3 \operatorname{CuCl} + 1 \operatorname{HAuCl_4:3H_2O} + 0.11 \operatorname{CSPVP} \xrightarrow{H_2O} \operatorname{Cu/Au} (3:1)-\operatorname{CSPVP} \\ \xrightarrow{\operatorname{NaBH_4}} \operatorname{Cu/Au} (3:1)-\operatorname{CSPVP} \\ \xrightarrow{\operatorname{NaBH_4}} 25^{\circ}\mathrm{C}, \ 0.5 \ \mathrm{h} \\ \xrightarrow{\operatorname{NaBH_4}} 25^{\circ}\mathrm{C}, \ 0.5 \ \mathrm{h} \\ \xrightarrow{\operatorname{NaBH_4}} 55^{\circ}\mathrm{C}, \ 0.5 \ \mathrm{h}$$

Scheme 6. The Synthesis of Pd/Au-CSPVP Nanoclusters and Cu/Au-CSPVP Nanoclusters.

For the CSPVP stabilized bimetallic nanocluster catalyzed oxidation reaction, the nanoclusters catalyst solution was used directly into the reaction. But for the characterization of nanoclusters, the crude nanocluster solution was filtered through a Vivaspin 20 centrifugal filter device (3,000 MWCO), washed with deionized water twice to remove low MW inorganic materials, and lyophilized to give a powder. The resulting nanoclusters was dissolved in water to certain concentration and subjected to analysis.

1.3.2 Characterization of Pd/Au-CSPVP Nanoclusters and Cu/Au-CSPVP Nanoclusters

It has been reported the carbonyl in the amide group in PVP was coordinated with the surface of Pd in the nanoclusters.^[36, 37] Because of the chelation, the amide C=O have greater character of $^{\delta-}$ O-C=N^{$\delta+$} which give the pyrolidinone ring at 1648 cm⁻¹ of polymer **26** in IR spectrum shifted to 1643.5 cm⁻¹ for Cu/Au (3:1)-**26** and 1641.9 cm⁻¹ for Pd/Au (3:1)-**26**. The shift of Cu/Au (3:1)-**26** suggests the similar chelation for Cu and CSPVP. (**Figure 7**)

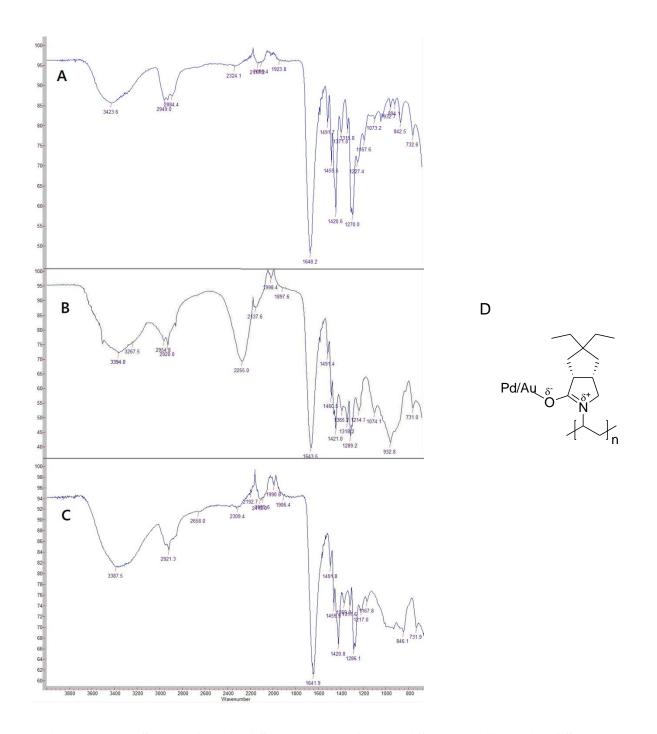


Figure 7. IR Spectra for (A) CSPVP-26, (B) Cu:Au-CSPVP26 (C) Pd:Au-CSPVP26 (D) Pd/Au Complex with CSPVP.

As a nanomaterial, several kinds of microscopy or size distribution analysis were conducted including atomic force microscopy (AFM), transition electron microscopy (TEM) and dynamic

light scattering (DLS). In AFM images obtained from Nanoscope IIIa SPM atomic force microscope, (**Figure 8**) 50-100 nm in diameters and ~ 5 nm in heights of spherical particles were found for both Pd/Au-CSPVP26 and Cu/Au-CSPVP26, as example. And in DLS which was taken on ZetaPALS zeta potential analyzer, (**Figure 9**) 90-120 nm sized nanoparticles were revealed in Pd/Au-CSPVP26 and Cu/Au-CSPVP26 water solutions with a concentration of 4.36 mol/L Pd/Au (3:1) or Cu/Au (3:1) and 0.12 mol/L CSPVP26, suggesting the size of polymer 26 wrapped particles was ~7.5 times larger than polymer 26 alone (14.6 nm shown in **Figure 5**), and the measured sizes of the particles from AFM and DLS were similar. Small amount of 12 - 14 nm size particles shown in DLS may indicate the free CSPVP which was not chelate with metals.

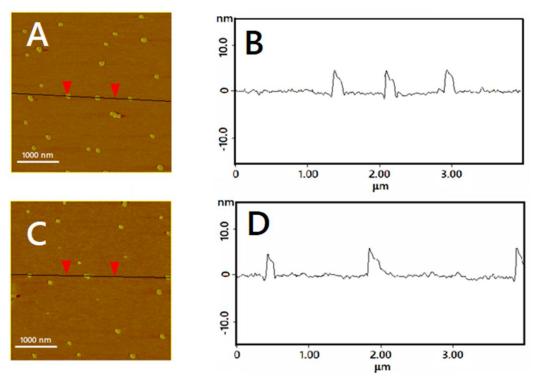


Figure 8. A Representative Atomic Force Microscopy Image and Measurement of Particle Height for Pd:Au-CSPVP26 (Panel A & B) and Cu:Au-CSPVP26 (Panel C & D).

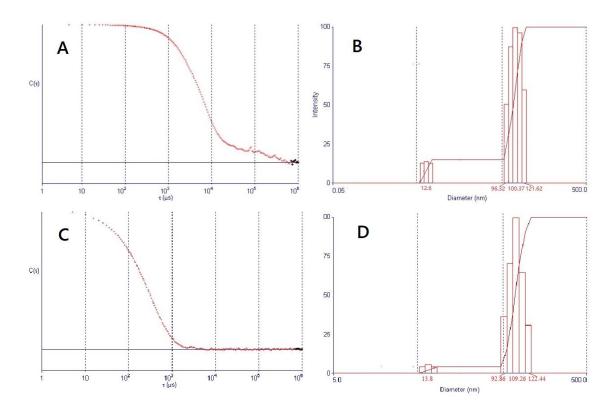


Figure 9. Dynamic Light Scattering Results for Pd:Au-CSPVP26 (Panel A & B), Cu:Au-CSPVP26 (Panel C & D).

TEM images, which is taken on FEI CM100 TEM instrument, revealed average diameters of 3.32 ± 1.08 nm and 3.41 ± 1.13 nm for Pd/Au (3:1)-**26** and Cu/Au (3:1)-**26**, respectively. The nano-sizes (3–4 nm) of the nanoclusters are similar to those reported for Pd/Au using PVP as a stabilizer.^[10, 36] The approximate total numbers of metal atoms and molecules of polymer in a spherical nanocluster can be calculated utilizing the magic-cluster sizes N_{total} = $1/3(10n^3-15n^2+11n-3)$, where n is the number of layers of shell in the metallic nanoparticles with face-centered cubic close-packed structure.^[38] Since the average sizes of Pd/Au (3:1)-**26** and Cu/Au (3:1)-**26** nanoclusters are 3.32 and 3.41 nm, respectively (from TEM), utilizing the aforementioned equation there are respectively ~727 and 923 atoms of Pd/Au (3:1) and Cu/Au

(3:1) in a nanocluster, and each nanoclusters are stabilized by approximately 20 and 25 polymers, respectively. The calculation is shown below:

For Pd/Au (3:1)-2:

(diameters for Au and Pd are 0.288 and 0.275, respectively.) Diameter d = $(3 \ge 0.275 + 0.288) / 4 = 0.278$ Nanocluster diameter: 11.9d = 3.32 nm (the diameter of Pd/Au is obtained from TEM) The shell number: n = 6.5 The number of atoms in a 3.32nm nanocluster: N = 1/3[10n3 - 15n2 + 11n - 3] = 727 atoms Mole of metals in each nanocluster: $1.21 \ge 10^{-21}$ mol Mole of CSPVP in each nanocluster: $1.21 \ge 10^{-21} \ge 0.11 / 4 = 3.3 \ge 10^{-23}$ mol Number of CSPVP in each nanocluster: $3.3 \ge 10^{-23} \le 10^{-23}$ mol Number of CSPVP in each nanocluster: $3.3 \ge 10^{-23} \le 10^{-23}$

For Cu/Au (3:1)-2:

(diameters for Au and Cu are 0.288 and 0.256, respectively.)

Diameter d = $(3 \times 0.256 + 0.288) / 4 = 0.264$

Nanocluster diameter: 12.9d = 3.41 nm (the diameter of Cu/Au is obtained from TEM)

The shell number: n = 7

The number of atoms in a 3.41 nm nanocluster: N = 1/3[10n3 - 15n2 + 11n - 3] = 923 atoms

Mole of metal in each nanocluster: 1.53×10^{-21}

Mole of CSPVP in each nanocluster: $1.53 \times 10-21 \times 0.11 / 4 = 4.22 \times 10^{-23}$

Number of CSPVP in each nanocluster: $5.3 \times 10-23 \times 6.022 \times 1023 = 25.4$

Because of the numbers and extended structure of the chiral polymer, the sizes of the bimetallic nanocluster/polymer found by AFM and DLS are much larger than bimetallic nanoclusters found in TEM (the polymer cannot be observed by TEM).

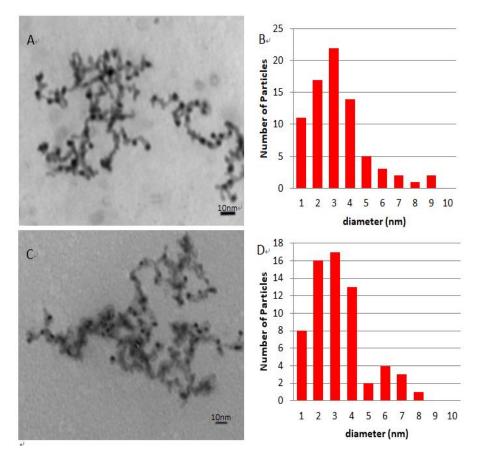


Figure 10. Representative TEM Image and Measurement of the Diameter for Pd:Au-CSPVP26 (Panel A & B), Cu:Au-CSPVP26 (Panel C & D).

Moreover, the elemental analysis including inductively coupled plasma-mass spectrometry (ICP-MS) were carried out. Results of the ICP-MS, taken on NexION® 300D, showed a ratio of 2.985 \pm 0.172 of Pd and Au in the Pd/Au (3:1)-26 sample, and a ratio of 3.148 \pm 0.432 of Cu and Au in the Cu/Au (3:1)-26 sample. This is in agreement of the 3:1 ratio of the two metals used in the preparation of nanoclusters. To digest the metallic samples, the following method is used. To a solution of 10 mg of Pd/Au (3:1)-26 or Cu/Au (3:1)-26 in 100 µL of deionized water

was added 2 mL of aqua regia solution at 70 °C under sonication for 3 h, and the solution was cooled to 25 °C, diluted to 1 mL with a 1% $HNO_3/2\%$ HCl solution. The resulting solution was diluted to certain concentration for the ICP-MS analysis.

In wdie-scan XPS spectrum of the Pd/Au-1 and Cu/Au-1, (Panel A & C) major elements of the polymer, carbon (C 1s at 284.6 eV), oxygen (O 1s at 530.6 eV), and nitrogen (N 1s at 398.6 eV) are shown. Panel B & C showed the binding energy of two maximums at 84.1 eV and 335.1 eV derived from $4f_{7/2}$ Au and $3d_{5/2}$ Pd, respectively in Pd/Au-1 sample. Similarly, 932.7 eV derived from $2p_{3/2}$ Cu is also appeared in Cu/Au-1 sample in Panel F.^[36] X-ray photoelectron spectroscopy (XPS) study was performed on a PHI 5400 XPS system. (**Figure 11**)

The UV-vis spectrum of Pd/Au-26 and Cu-Au-26 were also obtained from Agilent Cary 100 UV-Vis Spectrophotometer. (Figure 12) It is clearly shows the presence of strong signal at 303 nm in the nanoclusters which is very weak in the CSPVP26 alone.

In order to study the chirality of CSPVP stabilized bimetallic nanoclusters, circular dichroism (CD) was taken on Jasco J-815 Circular Dichroism (CD) Spectropolarimeter. (**Figure 13**) For the CSPVP**26** in water solution with a concentration of 0.011 mM alone, there is only a slight negative band on 520 nm. But after forming the bimetallic nanoclusters, they exhibit positive CD band at 280 nm and strong band at 500 nm, which is similar like the reported chiral polymers and chiral Au nanoclusters.^[36, 39, 40] The strong optical absorptions are not from CSPVP**26** or metals. It has been reported the (R)-Au₃₈ and (S)-Au₃₈ nanoclusters presented mirror-imaged CD spectra with similar optical absorption but on opposite sign. It illustrated the chirality of nanoclusters can be performed by CD spectra. The presence of strong optical absorption in Pd/Au-**26** and Cu/Au-**26** indicate the nanoclusters solution is a chiral environment.

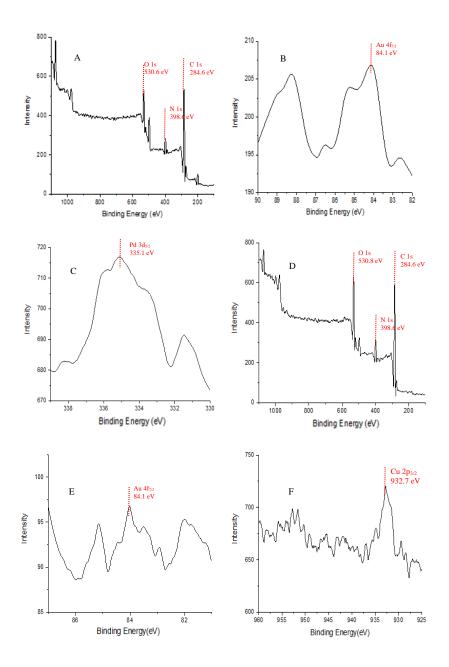


Figure 11. X-ray photoelectron spectroscopy Results for Pd:Au-CSPVP1 (Panel A, B & C), Cu:Au-CSPVP1 (Panel D, E & F).

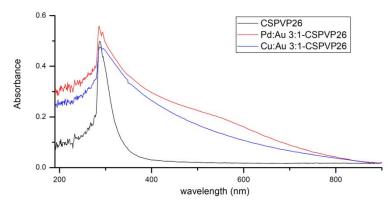


Figure 12 Representative UV-Vis spectra of CSPVP26, Pd:Au-CSPVP26 & Cu:Au-CSPVP26

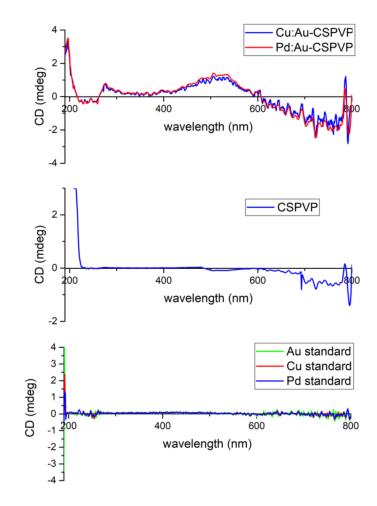


Figure 13. Representative CD spectra of Cu:Au-CSPVP26 & Pd:Au-CSPVP26 (top panel), CSPVP 26 (middle panel), and Au, Cu, and Pd standard solution as controls.

1.4 Conclusion

In this chapter, the synthesis of two classes of new polymers, namely chiral-substituted poly-N-vinylpyrrolidinones (CSPVPs) were discussed. From the characterization results, the CSPVPs are uniform. They were used for the stabilization of the bimetallic nanoclusters such as Pd/Au or Cu/Au and inducted of chirality into the nanomaterial. The size, component and other physical / chemical properties of the bimetallic nanoclusters (~3.5 nm), which are wrapped around by chiral polymers, were also studied and discussed.

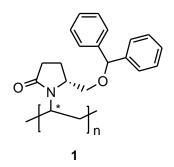
1.5 Synthetic Experimental Procedures

1.5.1 General

¹H NMR spectra (400 MHz) and ¹³C NMR spectra (100 MHz) were measured from a solution in CDCl₃ unless otherwise mentioned. The chemical shift data for each signal on ¹H NMR are given in units of δ relative to TMS ($\delta = 0$) or CHCl₃ ($\delta = 7.26$) in ppm. For ¹³C NMR spectra, the chemical shifts are recorded relative to CDCl₃ ($\delta = 77.0$). Low-resolution mass spectra were taken from an API 2000-triple quadrupole ESI-MS/MS mass spectrometer. High-resolution mass spectra were obtained using a Waters LCT Premier time of flight mass spectrometer. IR spectra were measured directly in solid form from Agilent Cary 630 FTIR. UV-Vis spectra were obtained from Agilent Cary 100 UV-Vis Spectrophotometer. Optical Rotation were measured by Bodenseewerk Perkin-Elmer & Co Gmbh Ueberlingen.

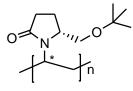
1.5.2 Representative Synthesis

Poly[(5*R*)-5-(benzhydryloxymethyl)-1-vinylpyrrolidin-2-one] (1). (BH7-041)



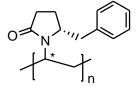
To a hot (120° C) solution of 1.2 mg (1% by weight) of copolymer 24 in 0.5 mL of DMF under argon, were added 120 mg (0.43 mmol) of compound 13 and 0.28 mg (1.7 µmol) of AIBN. The solution was stirred for 6 days at 120°C (until no monomer 13 was found by NMR spectrum of an aliquot from the reaction solution), cooled to 25°C, and diluted with diethyl ether. The precipitate was collected by filtration, dried under vacuum to give 102 mg (84% yield) of polymer 1 as a white solid. The average molecular weight of polymer 1, 106,000, was determined by gel permeation chromatography using TSKgel GMHxl column and THF as solvent with a flow rate of 1 mL/min. ¹H NMR δ 7.3–6.8 (m, 6 H, Ph), 6.6–6.3 (m, 4 H, Ph), 3.8–3.76 (m, 1 H), 3.4–3.0 (m, 3 H), 2.45–1.0 (m, 7 H). ¹³C NMR δ 176.1–175.4, 145.5–144.5, 128.6–124.3, 72.4–69.1, 69.0–67.2, 45.9–41.5, 40.6–39.3, 37.3–34.4, 32.2–34.6, 19.0–18.5. IR (neat) v 3050, 2954, 2890, 1650 (s), 1462, 1441, 1423, 1290.

Poly[(5*R*)-5-(*tert*-butoxymethyl)-1-vinylpyrrolidin-2-one] (2). (BH4-048)



To a solution of 90 mg (0.45 mmol) of monomer **15** and 2 mg of copolymer **22** in 0.2 mL of ethyl acetate under argon was added a solution of 0.3 mg of AIBN in 0.1 mL of ethyl acetate. The solution was heated to reflux for 14 h, cooled to 25° C, diluted with diethyl ether, and filtered to collect the white solid. The solid was dried under vacuum to give 76 mg (83% yield) of polymer **2**. The average molecular weight of polymer **2** was 68,000, which was determined by gel permeation chromatography using TSKgel GMHxl column and THF as solvent with a flow rate of 1 mL/min. ¹H NMR δ 4.0–3.0 (m, 4 H), 2.5–1.4 (bm, 6 H), 1.71 (bs, 9 H, *t*-Bu). The polymer was used in the subsequent catalytic asymmetric oxidation reactions without further purification.

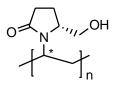
Poly[(5*R*)-5-(benzyl)-1-vinylpyrrolidin-2-one] (3). (BH1-046)



3

To a solution of 0.30 g (1.5 mmol) of monomer **21** and 3 mg of copolymer **23** in 0.6 mL of ethyl acetate under argon was added a solution of 1 mg of AIBN in 0.1 mL of ethyl acetate. The solution was heated to reflux for 6 h, cooled to 25° C, diluted with diethyl ether, and filtered to collect the white solid. The solid was dried under vacuum to give 0.27 g (89% yield) of polymer **3**. The average molecular weight of polymer **3** was 76,000, which was determined by gel permeation chromatography using TSKgel GMHxl column and THF as solvent with a flow rate of 1 mL/min. ¹H NMR δ 7.25–6.25 (m, 5 H, Ph), 4.0–3.0 (m, 4 H), 2.5–1.2 (m, 6 H); ¹³C NMR δ 175.8-174.8, 145.2-144.9, 139.8-135.0, 45.3-41.3, 30.5-30.2, 19.8-18.6. The polymer was used in the subsequent catalytic asymmetric oxidation reactions without further purification.

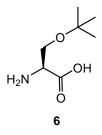
Poly[(5*R*)-5-(hydroxymethyl)-1-vinylpyrrolidin-2-one] (4). (BH4-076)



4

A solution of 0.20 g (3 µmol) of polymer **2** and 34 mg (0.3 mmol) of trifluoroacetic acid in 0.2 mL of CH₂Cl₂ was stirred at 25°C for 2 h, and concentrated to dryness to give 0.14 g (96% yield) of polymer **4**. The average molecular weight of polymer **4** was 56,480, which was determined by gel permeation chromatography using TSKgel GMHxl column and THF as solvent with a flow rate of 1 mL/min. ¹H NMR δ 4.1–2.8 (m, 4 H), 2.5–1.25 (m, 7 H). The polymer was used in the subsequent catalytic asymmetric oxidation reactions without further purification.

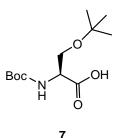
(S)-2-Amino-3-tert-butoxypropanoic acid (6) (BH4-039)



A solution of 0.5 g (4.7 mmol) of L-serine and 0.197 g (4.7 mmol) of LiOH•H₂O IN 3 mL of deionized water was stirred for 1 hour, concentrated to give white solid as serine mono lithium salt and dissolved in 7.05 mL of anhydrous THF. To it, 2.82 mL of BF₃•Et₂O was dropwise added, stirred under Ar at room temperature overnight and 45 °C for 2 hours to give a gray solution. The resulting solution was concentrate under vacuum and the residue was dissolved in 14.7 mL of dioxane in a seal tube. To it, 0.21 mL of 85% H₃PO₄ was added and cooled to -20°C followed by 20 mL of isobutylene (by introducing the gas directly) into the sealed tube reactor and react under room temperature for 2.5 hours, evaporated all the isobutylene, added 1N NaOH solution to pH = 10, concentrated under vacuum to give 3.1g crude product as white

solids. This mixture (containing 6 and other inorganic salts) was used in the following step without further purification.

(S)-3-tert-Butoxy-2-(tert-butoxycarbonyl)propanoic acid (7) (BH4-043)

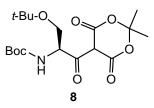


To a solution of 3.1g of crude material *t*-Bu-serine (**6**) in 10 mL of 1:1 THF/H₂O solution, 3 eq of NaHCO₃ and 1.2 eq of Boc₂O were added at 0 °C under ice bath and stirred overnight under room temperature, extracted with 3 x 10 mL diethyl ether. The combined organic layer was acidified with concentrated citric acid to pH = 4-5, extracted with 3 x 10 mL dichloromethane, washed with brine, dried over anhydrous sodium sulfate, concentrate to give desired product. The NMR data is the same as the commercially available compound.

(S)-tert-Butyl

3-tert-butoxy-1-(2,2-dimethyl-4,6-dioxo-1,3-dioxan-5-yl)-1-oxopropan-2-ylcarbamate (8).

(BH7-030)



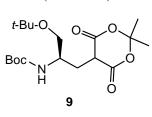
A solution of 4.0 g (15.3 mmol) of *N*-Boc-*O*-tert-butyl-L-serine (**7**), 3.3 g (23 mmol) of Meldrum's acid (2,2-dimethyl-1,3-dioxane-4,6-dione), and 2.8 g (22.9 mmol) of 4-(dimethylamino)pyridine (DMAP) in 80 mL of dichloromethane under argon was cooled to 0° C over an ice-water bath. To it, a solution of 3.47 g (16.8 mmol) of

N,*N*'-dicyclohexylcarbodiimide (DCC) in 20 mL of dichloromethane was added dropwise. The mixture was stirred for 12 hours under argon and the precipitated (*N*,*N*'-dicyclohexylurea) was removed by filtration. The filtrate was washed with 50 mL of 5% HCl, and then water, dried (anhydrous Na₂SO₄), and concentrated to dryness leaving 5.83 g of compound **8** as a white solid. This compound was used in the following step without further purification. ¹H NMR δ 5.67–5.63 (m, 1 H), 5.49–5.47 (m, 1 H), 3.74–3.70 (m, 2 H), 3.62 (s, 1 H, NH), 1.76 (s, 3 H), 1.73 (s, 3 H), 1.46 (s, 9 H), 1.26 (s, 9 H). ¹³C NMR δ 206.8, 172.7, 152.3, 105.9, 79.0, 75.0, 62.0, 60.6, 59.9, 30.0 (3 C), 28.4 (3 C), 25.6 (2 C). MS (ESI, MeOH): *m/z* = 388.7 ([M + H]⁺).

(R)-tert-Butyl

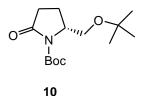
3-tert-butoxy-1-(2,2-dimethyl-4,6-dioxo-1,3-dioxan-5-yl)propan-2-ylcarbamate (9).

(BH7-031)



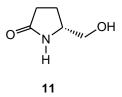
To a cold (0°C) solution of 5.80 g (15.2 mmol) of compound **8** in 150 mL of dichloromethane under argon were added 5.51 g (91.8 mmol) of acetic acid and 1.51 g (39.8 mmol) of NaBH₄. The solution was stirred at 0°C for three hours and at 25°C for 12 hours, diluted with 30 mL of water, the dichloromethane layer was separated, and the water layer was extracted with dichloromethane three times. The combined organic layers were washed with water and brine, dried (anhydrous Na₂SO₄), concentrated to give a yellow oil, which was crystallized from diethyl ether to give 5.27 g of compound **9** as a white solid. ¹H NMR δ 5.08–5.09 (m, 1 H), 4.08–4.00 (m, 1 H), 3.62 (s, 1 H, NH), 3.45–3.41 (m, 2 H), 2.5–2.45 (m, 1 H), 2.16–2.01 (m, 1 H), 1.80 (s, 3 H), 1.72 (s, 3 H), 1.40 (s, 9 H), 1.05 (s, 9 H); ¹³C NMR δ 208.0, 174.6, 151.0, 105.7, 79.3, 74.7, 69.8, 49.6, 47.1, 31.9, 28.4, 25.7. MS (ESI, MeOH): m/z = 374.0 ([M + H]⁺), 299.9.

(*R*)-N-(*tert*-Butoxycarbonyl)-5-(*tert*-butoxymethyl)-pyrrolidin-2-one (10). (BH7-032)



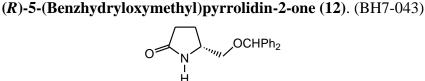
A solution of 5.26 g (14.10 mmol) of compound **9** in 40 mL of toluene under argon was heated to reflux for 6 hours, cooled to 25°C, concentrated under vacuum, and column chromatographed on silica gel using hexane and ethyl acetate (4:1) as eluent to give 3.44 g (83% overall yield in three steps) of compound **10** as a yellow oil. $[\alpha]_D^{22} = +80.7$ (c 1.0, CHCl₃); ¹H NMR δ 4.09–4.07 (m, 1 H), 3.52 (dd, J = 10, 4 Hz, 1 H), 3.31 (dd, J = 10, 1.6 Hz, 1 H), 2.56 (dt, J = 17.6, 9.6 Hz, 1 H), 2.20 (dd, J = 17.6, 9.2 Hz, 1 H), 2.05–1.84 (m, 2 H), 1.40 (s, 9 H), 1.02 (s, 9 H). ¹³C NMR δ 175.3, 150.1, 82.6, 73.2, 62.9, 58.0, 32.5, 28.2, 27.5, 21.7. MS (ESI, MeOH): m/z = 294.1 ([M + Na]⁺), 272.0 ([M+ H]⁺), 194.1, 172.1, 116.0. HRMS-ESI: m/z [M + H]⁺ calcd for C₁₄H₂₆NO₄: 272.1862; found: 272.1887.

(R)-5-(Hydroxymethyl)pyrrolidin-2-one (11). (BH7-033)

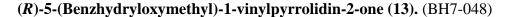


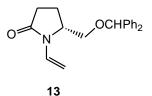
A solution of 1.0 g (3.6 mmol) of compound **10** in 5 mL of 10% trifluoroacetic acid in dichloromethane was stirred at 50°C for 5 hours, cooled to 25°C, and diluted with dichloromethane. The solution was washed with an aqueous solution of NaHCO₃, then water and brine, dried (MgSO₄), concentrated to give 0.42 g (99% yield) of compound **11** as a brown oil. This material was used in the subsequent step without further purification. $[\alpha]_D^{22} = -60.2$ (c 1.0, CHCl₃); ¹H NMR δ 6.7–6.6 (bs, 1 H, NH), 4.46 (dd, J = 11.2, 4 Hz, 1 H), 4.21 (dd, J = 11.2, 7.2 Hz, 1 H), 4.15–4.04 (m, 1 H), 2.54–2.32 (m, 4 H, CH₂ and OH), 1.98–1.87 (m, 1 H);

¹³C NMR δ 181.6, 65.2, 57.5, 30.3, 22.3; MS (ESI, MeOH): m/z = 116.0 ([M + H]⁺, 100%), 98.1. HRMS-ESI: m/z [M + Na]⁺ calcd for C₅H₉NO₂Na⁺: 138.0531 found: 138.0537.



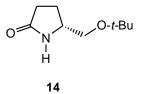
To a cold (0°C) solution of 0.43 g (3.74 mmol) of compound **11** in 1 mL distilled THF under argon, was added 0.15 g (3.74 mmol) of NaH (60% in mineral oil; NaH was washed twice with dried diethyl ether). The mixture was stirred at 0°C for 30 minutes and added 1.11 g (4.48 mmol) of bromodiphenylmethane. The mixture was stirred at 50°C for 12 hours, diluted with 1 mL of H₂O, and the organic THF layer was separated and concentrated under vacuum and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as an eluent to give 0.93 g (88% yield) of compound **12** as a brown oil. $[\alpha]_D^{22} = -63.2$ (c 1.0, CHCl₃); ¹H NMR δ 7.4–7.2 (m, 10 H, Ar), 6.06–5.98 (bs, 1 H, NH), 5.35 (s, 1 H), 3.95–3.88 (m, 1 H, CHN), 3.48 (dd, J = 9.6, 3.6 Hz, 1 H, CHO), 3.36 (dd, J = 9.6, 8 Hz, 1 H, CHO), 2.41–2.27 (m, 2 H, CH₂C=O), 2.24–2.14 (m, 1 H), 1.79–1.68 (m, 1 H); ¹³C NMR δ 178.1, 141.8 (2 C), 128.7 (4 C), 127.9 (2 C), 127.1 (4 C), 84.1, 73.0, 54.1, 29.9, 23.4; MS (ESI, MeOH): *m/z* = 304.1 ([M + Na]⁺), 194.0, 167.1. HRMS-ESI: *m/z* [M + Na]⁺ calcd for C₁₈H₁₉NO₂Na⁺: 304.1313; found: 304.1320.





To a solution of 1.20 g (4.27 mmol) of compound **12** in 50 mL of vinyl acetate under argon, were added 4.0 g of 3 Å molecular sieves, 27 mg (0.34 mmol) of Na₂PdCl₄, and 1.06 g (7.69 mmol) of K₂CO₃. The mixture was stirred at 50°C under argon for 28 hours, cooled to 25°C, filtered, concentrated under vacuum, and column chromatographed on silica gel using a mixture of hexane and diethyl ether (1:1) as an eluent to give 0.67 g (51% yield) of pure compound **13** as a brown oil. $[\alpha]_D^{22} = +24.6$ (c 1.0, CHCl₃); ¹H NMR δ 7.35–7.20 (m, 10 H), 6.98 (dd, J = 16.4, 9.2 Hz, 1 H, CH=), 5.31 (s, 1 H), 4.38 (d, J = 9.2 Hz, 1 H, =CH₂), 4.34 (d, J = 16.4 Hz, 1 H, =CH₂), 4.13–4.08 (m, 1 H), 3.68 (dd, J = 9.2, 4.8 Hz, 1 H), 3.52 (dd, J = 9.2, 2.8 Hz, 1 H), 2.70 (dt, J = 17, 10 Hz, 1 H), 2.42–2.34 (m, 1 H), 2.20–2.11 (m, 2 H); ¹³C NMR δ 178.8, 142.5, 138.1, 128.7 (4 C), 127.9 (2 C), 127.1 (4 C), 96.8, 84.2, 73.6, 54.0, 30.2, 23.7. MS (ESI, MeOH): m/z = 330.1 ([M + Na]⁺). HRMS-ESI: m/z [M + Na]⁺ calcd for C₂₀H₂₁NO₂Na⁺: 330.1470; found: 330.1490.

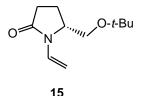




A solution of 10.2 g (37.5 mmol) of compound **10** in 100 mL of 7% trifluoroacetic acid (TFA) in dichloromethane was stirred at 25°C for 15 minutes and then diluted with 30 mL of saturated NaHCO₃. The mixture was separated, and the aqueous layer was extracted with dichloromethane. The combined organic layer was washed with water, and brine, dried (MgSO₄), concentrated to give 6.28 g (98% yield) of compound **14** as a brown oil. This material was used in the subsequent step without further purification. $[\alpha]_D^{22} = -10.8$ (c 1.0, CHCl₃); ¹H NMR δ 7.8–7.72 (bs, 1 H, NH), 3.94–3.86 (m, 1 H, CHN), 3.43 (dd, J = 9.0, 3.2 Hz, 1 H, CH₂O), 3.25 (dd, J = 9.0, 8.0 Hz, 1 H, CH₂O), 2.61–2.45 (m, 2 H, CH₂C=O), 2.28–2.18 (m,

1 H), 1.86–1.75 (m, 1 H), 1.18 (s, 9 H); ¹³C NMR δ 175.5, 74.4, 65.2, 59.7, 32.4, 28.0 (3 C), 22.5. MS (ESI, MeOH): m/z = 194.1 ([M + Na]⁺; 100%), 172.3. HRMS-ESI: m/z [M + Na]⁺ calcd for C₉H₁₇NO₂Na⁺: 194.1157; found: 194.1172.

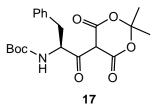
(*R*)-5-(*t*-Butyloxymethyl)-1-vinylpyrrolidin-2-one (15). (BH4-058)



To a solution of 2.0 g (11.6 mmol) of compound **14** in 35 mL of vinyl acetate under argon, were added 4.0 g of 3 Å molecular sieves, 0.27 g (0.9 mmol) of Na₂PdCl₄, and 2.25 g (16.3 mmol) of K₂CO₃. The mixture was stirred at 50°C under argon for 14 hours, cooled to 25°C, filtered, concentrated under vacuum, and column chromatographed on silica gel using a mixture of hexane and ethyl acetate (1:2) as an eluent to give 0.96 g (42% yield) of pure compound **15** as a yellow oil. $[\alpha]_D^{22} = +9.6$ (c 1.0, CHCl₃); ¹H NMR δ 7.0 (dd, J = 16, 9 Hz, 1H, CH=), 4.45 (d, J = 9 Hz, 1 H, =CH₂), 4.44 (d, J = 16 Hz, 1 H, =CH₂), 4.06–4.00 (m, 1 H, CHN), 3.52 (dd, J = 9.2, 4.8 Hz, 1 H, CH₂O), 3.41 (dd, J = 9.2, 2.8 Hz, 1 H, CH₂O), 2.64 (dt, J = 17.2, 10.8 Hz, 1 H), 2.35–2.26 (m, 1 H), 2.14–2.05 (m, 2 H), 1.13 (s, 9 H); ¹³C NMR δ 175.3, 139.7, 93.1, 74.6, 63.5, 58.8, 32.4, 28.2 (3 X), 20.8. MS (ESI, MeOH): *m*/*z* = 220.2 ([M + Na]⁺), 198.2 (M+H⁺), 187.1. HRMS-ESI: *m*/*z* [M + Na]⁺ calcd for C₁₁H₁₉NO₂Na⁺: 220.1313; found: 220.1333.

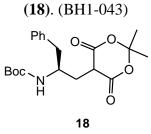
(S)-tert-Butyl

1-(2,2-dimethyl-4,6-dioxo-1,3-dioxan-5-yl)-1-oxo-3-phenylpropan-2-ylcarbamate (17). (BH1-042)



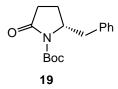
A solution of 1.33 g (5.0 mmol) of Boc-L-phenylalanine, 1.08 g (7.5 mmol) of Meldrum's acid, and 0.95 g (7.5 mmol) of 4-(dimethylamino)pyridine (DMAP) in 70 mL of dichloromethane under argon was cooled to 0°C over an ice-water bath. To it, a solution of 1.13 (5.5 mmol) g of *N*,*N*'-dicyclohexylcarbodiimide (DCC) in 15 mL of dichloromethane was added dropwise. The mixture was stirred for 12 hours under argon and the precipitated (*N*,*N*'-dicyclohexylurea) was removed by filtration. The filtrate was washed with 5% HCl twice, and then water, dried (anhydrous Na₂SO₄), and concentrated to dryness leaving 1.90 g of **17** as white solid. This compound was used in the following step without further purification. ¹H NMR δ 7.3 -7.15 (m, 5 H), 4.5–4.4 (m, 1 H), 4.3–4.2 (m, 1 H), 3.9–3.8 (m, 1 H), 2.8–2.7 (m, 2 H), 1.7 (s, 3 H), 1.65 (s, 3 H); ¹³C NMR δ 207.5, 170.8, 153.1, 135.8, 129.6 (2 C), 128.9 (2 C), 127.4, 105.7, 80.3, 62.7, 60.3, 33.8, 28.4, 26.8.

(R)-tert-Butyl 1-(2,2-dimethyl-4,6-dioxo-1,3-dioxan-5-yl)-3-phenylpropan-2-ylcarbamate

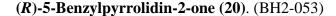


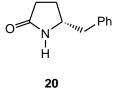
To a cold (0° C) solution of 1.90 g (4.85 mmol) of compound **17** in 25 mL of dichloromethane under argon were added 1.75 g (29.1 mmol) of acetic acid and 0.48 g (12.6 mmol) of NaBH₄. The solution was stirred at 0° C for three hours and at 25°C for 12 hours, diluted with 30 mL of water, the dichloromethane layer was separated, and the water layer was extracted with dichloromethane three times. The combined organic layers were washed with water and brine, dried (MgSO₄), concentrated to give 1.78 g of compound **18** as a white solid. This compound was used in the subsequent step without further purification. ¹H NMR δ 7.35–7.19 (m, 5 H), 4.48–4.43 (m, 1 H), 4.24–4.20 (m, 1 H), 3.92–3.90 (m, 1 H), 2.88–2.83 (m, 2 H), 2.31–2.11 (m, 2 H), 1.81 (s, 3 H), 1.77 (s, 3 H), 1.25 (s, 9 H); ¹³C NMR δ 174.7, 150.2, 137.3, 129.6 (2 C), 129.0 (2 C), 127.1, 83.2, 59.2, 39.7, 31.3, 28.3 (3 C), 21.8.^[41] MS (ESI, MeOH): *m/z* = 400.2 ([M + Na]⁺).

(*R*)-*N*-(*tert*-Butoxycarbonyl)-5-benzylpyrrolidin-2-one (19). (BH1-029)



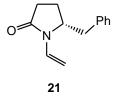
A solution of 0.35 g (0.93 mmol) of compound **18** in 20 mL of toluene under argon was heated to reflux for 4 hours, cooled to 25°C, concentrated, and column chromatographed on silica gel using hexane and ethyl acetate (4:1) as eluent to give 0.23 g (88% overall yield in three steps) of compound **19** as a yellow oil. The material was used in the subsequent step without purification. $[\alpha]_D^{22} = +48.7$ (c 1.5, CHCl₃); Lit.^[41] +48.5 (c 1.50, CHCl₃). ¹H NMR δ 7.34–7.29 (m, 2 H), 7.28–7.24 (m, 1 H), 7.20–7.17 (m, 2 H), 4.39–4.34 (m, 1 H, CHN), 3.13 (dd, J = 13.5, 4 Hz, 1 H), 2.73 (dd, J = 13.5, 9.6 Hz, 1 H), 2.35–2.27 (m, 2 H), 2.20–1.90 (m, 1 H), 1.85–1.76 (m, 1 H), 1.57 (s, 9 H); ¹³C NMR δ 174.8, 150.1, 137.3, 129.6 (2 C), 129.0 (2 C), 127.1, 83.3, 59.3, 39.7, 31.3, 28.4 (3 C), 21.8. MS (ESI, MeOH): m/z = 298.1 ([M + Na]⁺).





A solution of 1.0 g (12.9 mmol) of compound **19** in 5 mL of 10% trifluoroacetic acid in dichloromethane was stirred at 25°C for 2 hours, diluted with dichloromethane, and washed with an 10% aqueous solution of NaHCO₃, then water and brine, dried (MgSO₄), concentrated to give 0.61 g (96% yield) of compound **20** as a brown oil. This material was used in the subsequent step without further purification. $[\alpha]_D^{22} = +39.7$ (c 1.19, EtOH); Lit.^[41] +39.6 (c 1.19, EtOH). ¹H NMR δ 7.33–7.12 (m, 5 H), 5.95–5.90 (bs, 1 H, NH), 3.92–3.83 (m, 1 H, CHN), 2.83 (dd, J = 13.4, 5.6 Hz, 1 H), 2.72 (dd, J = 13.4, 7.6 Hz, 1 H), 2.34–2.20 (m, 3 H), 1.89–1.80 (m, 1 H); ¹³C NMR δ 178.1, 137.8, 129.3 (2 C), 129.2 (2 C), 127.1, 55.9, 43.2, 30.3, 27.2; MS (ESI, MeOH): m/z = 198.1 ([M + Na]⁺).

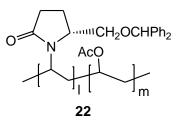
(*R*)-5-(Benzyl)-1-vinylpyrrolidin-2-one (21). (BH2-059)



To a solution of 0.60 g (3.42 mmol) of compound **20** in 30 mL of vinyl acetate under argon, were added 2.0 g of 3 Å molecular sieves, 88 mg (0.30 mmol) of Na₂PdCl₄, and 0.86 g (6.2 mmol) of K₂CO₃. The mixture was stirred at 50°C under argon for 28 hours, cooled to 25°C, filtered, concentrated under vacuum, and column chromatographed on silica gel using a mixture of hexane and diethyl ether (1:1) as an eluent to give 0.46 g (67% yield) of pure compound **21** as a yellow oil. $[\alpha]_D^{22} = +38.3$ (c 1.0, CHCl₃); ¹H NMR δ 7.34–7.23 (m, 3 H), 7.18–7.14 (m, 2 H), 7.08 (dd, J = 16.4, 9.6 Hz, 1 H, =CH), 4.63 (d, J = 16.4 Hz, 1 H, =CH₂), 4.60 (d, J = 9.6 Hz, 1 H, =CH₂), 4.22- 4.17 (m, 1 H, CHN), 3.00 (dd, J = 14, 3 Hz, 1 H), 2.81 (dd, J = 14, 8 Hz, 1 H), 2.24–2.17 (m, 1 H), 2.10–1.91 (m, 3 H); ¹³C NMR δ 173.7, 137.1, 129.7, 128.9 (2 C), 128.4, 127.1 (2 C), 95.3, 57.0, 36.6, 30.1, 22.8; MS (ESI, MeOH): m/z = 224.1 ([M + Na]⁺), 202.0 (M+H⁺; 100%). HRMS-ESI: m/z [M + Na]⁺ calcd for C₁₃H₁₅NONa: 224.1051; found: 224.1076.

Copolymer 22 from (R)-5-(benzhydryloxymethyl)-1-vinylpyrrolidin-2-one (13) and

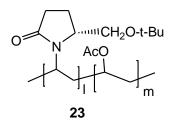
vinylacetate. (BH7-040)



To a solution of 93 mg (0.33 mmol) of compound **13** and 28 mg (0.33 mmol) of vinyl acetate in 0.15 mL of acetone under argon, was added 10 mg (0.066 mmol) of AIBN, and the solution was stirred under reflux for 30 h. The solution was cooled to 25° C, diluted with hexane, and stirred for 10 min. The white precipitate was collected by filtration, dried under vacuum to give 0.106 g (88% yield) of copolymer **22** as a white solid. ¹H NMR δ 7.50–6.3 (m, 11 H, Ph & CH), 3.6–1.0 (m, 16 H).

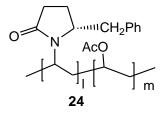
Copolymer 23 from (R)-5-(tert-butoxymethyl)-1-vinylpyrrolidin-2-one (15) and vinylacetate.

(BH3-035)



From 30 mg (0.15 mmol) of **15**, 13 mg (0.15 mmol) of vinyl acetate and 0.25 mg of AIBN, copolymer **23**, 39 mg (90% yield), was obtained. ¹H NMR δ 4.90–2.75 (m, 5 H), 2.70–0.80 (m, 20 H).

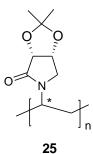
Copolymer 24 from (*R***)-5-benzyl-1-vinylpyrrolidin-2-one (21) and vinylacetate**. (BH1-045)



From 50 mg (0.25 mmol) of monomer **21**, 21.4 mg (0.25 mmol) of vinyl acetate and 0.7 mg of AIBN, copolymer **24**, 65 mg (91% yield), was obtained. ¹H NMR δ 7.20–6.80 (m, 3 H), 6.75–6.30 (m, 2 H), 3.50–3.00 (m, 3 H), 2.5–1.1 (m, 13 H).

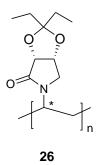
Poly[(3aR,6aR)-2,2-dimethyl-5-vinyl-dihydro-3aH-[1,3]dioxolo[4,5-c]pyrrol-4(5H)-one] (25).

(BH9-043)



To a solution of 0.3 mg (0.2% by weight) of copolymer **35** in 0.5 mL of ethyl acetate under argon, were added 70 mg (0.38 mmol) of compound **33** and 0.3 mg (2 µmol) of AIBN. The solution was stirred for 2 days at 70°C (until no monomer **33** was detected by NMR spectrum of an aliquot from the reaction solution), cooled to 25°C, and diluted with hexane. The precipitate was collected by filtration, dried under vacuum to give 63 mg (90% yield) of polymer **25** as a white solid. The average molecular weight of polymer **25**, 75,000 (n = 410), was determined by gel permeation chromatography using TSKgel GMHxl column and water as solvent with a flow rate of 1 mL/min. $[\alpha]_D^{22} = -74.3$ (c 1.6, H₂O); ¹H NMR δ 4.7–4.3 (m, 2 H, CHO), 3.8–3.0 (m, 3 H, CHN, CH₂N), 1.6–1.1 (8 H, CH₂, 2 CH₃); ¹³C NMR δ 161.0, 113.0, 74.7, 72.1, 48.7, 47.8, 46.1, 45.6, 27.2, 25.9. IR (neat) v 3020, 2940, 2143, 1648 (s), 1420, 1285, 1169, 1024, 830.

Poly[(3a*R*,6a*R*)-2,2-diethyl-5-vinyl-dihydro-3a*H*-[1,3]dioxolo[4,5-c]pyrrol-4(5*H*)-one] (26). (BH10-016)



To a solution of 0.4 mg (0.2% by weight) of copolymer **36** in 0.8 mL of ethyl acetate under argon, were added 0.18 g (0.99 mmol) of compound **34** and 0.72 mg (4 µmol) of AIBN. The solution was stirred for 2 days at 70°C (until no monomer **34** was detected by NMR spectrum of an aliquot from the reaction solution), cooled to 25°C, and diluted with hexane. The precipitate was collected by filtration, dried under vacuum to give 0.16 g (91% yield) of polymer **26** as a white solid. The average molecular weight of polymer **26**, 92,000 (n = 436), was determined by gel permeation chromatography using TSKgel GMHxl column and water as solvent with a flow rate of 1 mL/min. $[\alpha]_D^{22} = -138.9$ (c 1.8, H₂O); ¹H NMR δ 4.7–4.6 (m, 2 H, CHO), 3.7–3.0 (m, 3 H, CHN, CH₂N), 1.6–1.3 (6 H, CH₂), 0.9–0.6 (m, 6 H, CH₃); ¹³C NMR δ 160.9, 113.5, 74.7, 73.0, 48.8, 47.7, 43.0, 42.7, 29.2, 28.5, 8.4, 7.0. IR (neat) v 3423, 2949, 2884, 1648 (s), 1460, 1420, 1270, 842; UV-Vis 299 nm (ϵ = 45090 L mol⁻¹cm⁻¹).

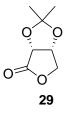
(3R,4R)-3,4-Dihydroxy-dihydrofuran-2(3H)-one (28). (BH11-084)



To a cold (0 $^{\circ}$ C) solution of 17.6 g (0.1 mol) of *D*-isoascorbic acid (**27**) in 250 mL of H₂O, 21.2 g (0.2 mol) of Na₂CO₃ and 22.7 g (0.2 mol) of 30% H₂O₂ were added dropwise, the solution was stirred at 0 $^{\circ}$ C for 10 minutes and 42 $^{\circ}$ C for 30 minutes. To the reaction solution, 4.0 g of

activated charcoal was added in portion over 10 minutes, and the mixture was stirred at 80 °C for 30 minutes, filtered through Celite during hot and washed the filter cake with 50 mL of H₂O. The filtrate was carefully acidified with 6 *N* HCl until pH 1, concentrated on a rotary evaporator and then under vacuum to give 36.9 g of solid. The crude products were purified by adding 200 mL of ethyl acetate and heating to reflux for 10 minutes. The hot solution was filtered and the filtrate containing the desired product was saved. The extraction of the product from the solid was repeated by refluxing in 200 mL of ethyl acetate and filtered. Both filtrates were combined, cool over an ice-water bath, and the crystallized white solid was collected by filtration to give 11.0 g (94 % yield) of the titled compound. $[\alpha]_D^{22} = -73.2$ (c 0.5, H₂O), Lit^[22] $[\alpha]_D^{22} = -72.8$ (c 0.498, H₂O); ¹H NMR (DMSO-d6) δ 5.75 (dd, *J* = 7.2, 3.2 Hz, 1 H, OH), 5.34 (s, 1 H, OH), 4.39–4.34 (m, 1 H), 4.26 (dt, *J* = 10, 3.2 Hz, 1 H), 4.24–4.19 (m, 1 H), 4.03 (dd, *J* = 10, 3.2 Hz 1 H); ¹³ C NMR (DMSO-d6) δ 177.1, 72.5, 70.1, 69.0; MS (ESI, MeOH): m/z = 141.4 ([M + Na]⁺), 72.4.

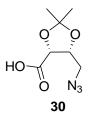
(2R,3R)-O-Isopropylidene-D-erythronolactone (29). (BH9-145)



To a solution of 1.6 g (13.6 mmol) of **28** in 25 mL of anhydrous acetone under argon, 4.5 g (28.5 mmol) of CuSO₄ was added. The mixture was stirred for 36 hours and the precipitated inorganic salt was removed by filtration over a pad of Celite. The filtrate was concentrated to dryness leaving 2.32 g of yellow solid. The crude product was column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as an eluent to give 1.90 g (89% yield) of the titled compound as a pale yellow solid. $[\alpha]_D^{22} = -114.64$ (c 1.5, acetone), Lit^[42]. $[\alpha]_D^{20} = +116.3$ (c 1.5, acetone); ¹H NMR δ 4.85 (bs, 1 H), 4.72 (bs, 1 H), 4.39 (bs, 2 H), 1.42

(s, 3 H), 1.34 (s, 3 H); ¹³C NMR δ 174.8, 113.9, 75.8, 74.8, 70.5, 26.9, 25.6; MS (ESI, MeOH): m/z = 181.3 ([M + Na]⁺), 95.6.

(2R,3R)-4-Azido-4-deoxy-2,3-O-isopropylidene-D-erythronic acid (30). (BH9-147)



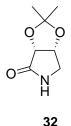
A solution of 1.90 g (12.0 mmol) of **29** and 2.73 g (42.1 mmol) of NaN₃ in 12 mL of distilled DMF was heated to 110 °C under argon for 24 hours. The reaction solution was diluted with 500 mL of diethyl ether, 10 mL of H₂O and 20% aqueous HCl to pH 2, and extracted with 40 mL of ethyl acetate four times. The combined extract was washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as an eluent to give 1.03 g (42% yield) of the titled compound as a pale yellow solid. The major by-product was azido((4R,5R)-5-(hydroxymethyl)-2,2-dimethyl-1,3-dioxolan-4-yl)methanone. $[\alpha]^{22}_{D}$ (c 0.47, acetone); Lit.^[30] +72 (c 0.47, acetone); ¹H NMR δ 9.52–9.0 (bs, 1 H, OH), 4.67 (d, *J* = 7.2 Hz, 1 H), 4.60–4.56 (m, 1 H), 3.60 (dd, *J* = 13.2, 3.2 Hz, 1 H), 3.40 (dd, *J* = 13.2, 6 Hz, 1 H), 1.63 (s, 3 H), 1.42 (s, 3 H); ¹³C NMR δ 172.2, 112.2, 73.8, 70.8, 52.0, 27.1, 26.2; MS (ESI, MeOH): m/z = 224.1 ([M + Na]⁺), 143.6, 85.6.

(2R,3R)-4-Amino-4-deoxy-2,3-O-isopropylidene-D-erythronic acid (31). (BH9-151)



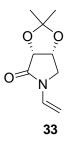
To a solution of 1.01 g (5.0 mmol) of **30** in 100 mL of methanol, 50 mg of 10% Pd/C was added, and the mixture was shake under 2 atm. of hydrogen gas for 4 hours in a hydrogenator. The mixture was removed from the hydrogenator, filtered through a pad of Celite, the filtrated was concentrated to dryness leaving 0.86 g (99% yield) of the titled compound as a pale yellow solid. $[\alpha]_D^{22} = +91.6$ (c 1.0, 60% aqueous acetone), $\text{Lit}^{[30]} [\alpha]_D^{20} = +92$ (c 1.02, 60% aqueous acetone); ¹H NMR (DMSO-d6) δ 4.39 (d, J = 7.2 Hz, 1 H), 4.31–4.24 (m, 1 H), 2.76 (t, J = 8 Hz, 2 H), 1.39 (s, 3 H), 1.23 (s, 3 H); MS (ESI, MeOH): m/z = 176.1 ([M + H]⁺), 198.1 ([M + Na]⁺).

(2R,3R)-4-Amino-4-deoxy-2,3-O-isopropylidene-D-erythronolactam (32). (BH9-031)



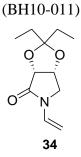
An amount of 0.16 g (0.93 mmol) of **31** was placed in a sublimator and heated to 160–170 °C/0.5 mm Hg under vacuum, and 92 mg (63% yield) of **32** was obtained as white solid. This material was used in the subsequent step without further purification. $[\alpha]^{22}{}_{D} = -60.3(c \ 0.78, MeOH);$ Lit^[30]. -59 (c 0.78, MeOH); ¹H NMR δ 6.31 (bs, 1 H, NH), 4.79 (t, J = 5.2 Hz, 1 H), 4.58 (d, J = 6 Hz, 1 H), 3.59 (dd, J = 11, 4.8 Hz, 1 H), 3.49 (d, J = 11 Hz, 1 H), 1.49 (s, 3 H), 1.39 (s, 3 H); ¹³C NMR δ 174.4, 113.0, 76.7, 74.9, 46.0, 27.2, 25.8; MS (ESI, MeOH): m/z = 158.4 ([M + H]⁺).

(2R,3R)-4-Amino-4-deoxy-2,3-O-isopropylidene-N-vinyl-D-erythronolactam (33). (BH9-035)



To a solution of 90 mg (0.57 mmol) of compound **32** in 10 mL of vinyl acetate under argon, were added 1.0 g of 3 Å molecular sieves, 13 mg (0.046 mmol) of Na₂PdCl₄, and 0.14 g (1.03 mmol) of K₂CO₃. The mixture was stirred at 50°C for 14 hours, cooled to 25°C, filtered through Celite, concentrated under vacuum, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as an eluent to give 84 mg (80% yield) of **33** as a yellow oil. $[\alpha]_D$ -20.1(c 1.0, CHCl₃); ¹H NMR δ 7.05 (dd, *J* = 16.0, 9.2 Hz, 1 H, CH=), 4.82–4.79 (m, 1 H), 4.70 (d, *J* = 6.4 Hz, 1 H), 4.53 (d, *J* = 9.2 Hz, 1 H, =CH₂), 4.48 (d, *J* = 16 Hz, 1 H, =CH₂), 3.62 (d, *J* = 2.8 Hz, 2 H), 1.39 (s, 3 H), 1.35 (s, 3 H); ¹³C NMR δ 169.4, 129.2, 113.0, 96.7, 78.1, 72.1, 48.7, 27.2, 25.9; MS (ESI, MeOH): m/z = 184.0 ([M + H]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₉H₁₄NO₃⁺: 184.0974 found: 184.0952.

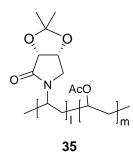
(3aR,6aR)-5-Vinyl-2,2-diethyl-dihydro-3aH-[1,3]dioxolo[4,5-c]pyrrol-4(5H)-one (34).



A mixture of 0.25 g (1.37 mmol) of compound **33**, 5 mg of $ZnCl_2$ (0.04 mmol) and 7 mL of 3-pentanone was placed in a distillation apparatus equipped with a short distill head and heated to 65 °C to remove the by-product acetone over 40 hours. The resulting mixture was diethyl ether, filtered through Celite, and washed with a small amount of diethyl ether. The filtrate was

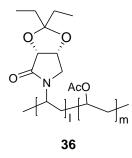
concentrated under vacuum and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as an eluent to give 192 mg (67% yield) of **34** as a yellow oil. $[\alpha]_D$ -43.2 (c 1.0, CHCl₃); ¹H NMR δ 7.07 (dd, J = 16.0, 9.2 Hz, 1 H, CH=), 4.84–4.81 (m, 1 H), 4.72 (d, J = 6.4 Hz, 1 H), 4.55 (d, J = 9.2 Hz, 1 H, =CH₂), 4.49 (d, J = 16 Hz, 1 H, =CH₂), 3.64 (d, J = 2.8 Hz, 2 H), 1.70 (q, J = 7.2 Hz, 2 H), 1.65 (q, J = 7.2 Hz, 2 H), 0.91 (t, J = 6 Hz, 6 H); ¹³C NMR δ 169.3, 128.8, 115.5, 96.6, 78.0, 71.9, 48.6, 30.0, 29.6, 8.4, 7.6. MS (ESI, MeOH): m/z = 212.2 ([M + H]⁺) 234.3 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₁H₁₈NO₃⁺: 212.1287 found: 212.1269.

Poly(3,4-disubstituted *N*-vinyl pyrrolidinone-co-vinyl acetate) [P(DVP-co-VAc)] 35 from compound 33 and vinyl acetate. (BH9-042)



To a solution of 10 mg (55 μ mol) of *N*-vinylpyrrolidinone 33 and 4.7 mg (55 μ mol) of vinyl acetate in 20 μ L of acetone under argon, was added 0.1 mg (0.6 μ mol) of azobisisobutyronitrile (AIBN), and the solution was stirred under reflux for 30 h. The solution was cooled to 25°C, diluted with hexane, and stirred for 10 min. The white precipitate was collected by filtration, dried under vacuum to give 12 mg (82% yield) of copolymer **35** as a white solid. ¹H NMR δ 4.90–2.75 (m, 5 H), 2.70–0.80 (m, 22 H).

Poly(3,4-disubstituted *N*-vinyl pyrrolidinone-co-vinyl acetate) [P(DVP-co-VAc)] 36 from compound 34 and vinyl acetate. (BH10-15)



To a solution of 15 mg (55 μ mol) of compound **34** and 4.7 mg (55 μ mol) of vinyl acetate in 20 μ L of acetone under argon, was added 0.1 mg (0.6 μ mol) of AIBN, and the solution was stirred under reflux for 30 h. The solution was cooled to 25°C, diluted with hexane, and stirred for 10 min. The white precipitate was collected by filtration, dried under vacuum to give 17.2 mg (87% yield) of copolymer **36** as a white solid. ¹H NMR δ 4.90–2.75 (m, 5 H), 2.70–0.80 (m, 26 H).

Chapter 2 - Asymmetric Oxidation Reactions and Late Stage Oxidation Reactions Catalyzed by Bimetallic Nanoclusters

(Partial of the results and discussions in this chapter was published^[1] and adapted with permission of *Journal of the American Chemical Society*. Copyright © 2016 American Chemical Society.)

Organic transformation reactions using bimetallic nanoclusters^[15] in the oxidation of alcohols,^[10, 43, 44] formic acid oxidation,^[45] aldehyde oxidation,^[46] C-H oxidation,^[36] Ullmann coupling,^[47] Suzuki coupling reaction,^[48] tandem oxidation-Michael addition reaction,^[49] have been reported. However, these reports do not include studies of enantio- or stereo-selectivity.

Catalytic asymmetric oxidation is challenging and the results can have a great impact to the chemistry community. With the catalysis of CSPVP stabilized bimetallic nanoclusters, several kinds of the oxidation reaction, including alcohol oxidations, alkene dihydroxylations, alkane C-H oxidations, oxidative ring closing of dienes and late-stage oxidations of complex molecules are all investigated and discussed in this chapter.

2.1 Selective Asymmetric Oxidation Reactions of Alcohols.

2.1.1 Background and Significance

Oxidation of alcohols to their respective aldehyde or ketone is a useful and fundamental organic reaction. Many methods have been developed by inducing stoichiometric amount of oxidants, which may produce large amount of highly polluted and toxic waste, such as bromate^[50], permanganate^[51] or chromate^[52]. Recently, transition metal nanoparticle–catalyzed aerobic alcohol oxidation with high catalytic activity and selectivity has been investigated. ^[10, 44]

Alcohol can be selectively oxidized to ketone or aldehyde in the presence of other functional groups.

Although desymmetrization of meso diols and diol derivatives using various organic reagents and enzymes have appeared,^[53-55] reports on the enantioselective oxidation of a mixture of racemic diols *via* kinetic resolution are limited, with the exception of oxidation of racemic benzylic vicinal diols.^[56] Hence, asymmetric oxidation of racemic diols by kinetic resolution or meso diols using bimetallic nanoclusters with chiral stabilizer may give high catalytic activity, regio- and enantioselectivity.

2.1.2 Results and Discussions

With the general idea, (\pm) -trans-1,3-cyclohexanediol (37) was used to study the reaction (\pm) -trans-1,3-cyclohexanediol was prepared by the conditions and possible products. seperation of mixture of cis & *trans*-1,3-cyclohexanediol by adding CuCl₂. cis-1,3-Cyclohexanediol was precipitated in diethyl ether due to the formation of complex with Cu²⁺ and *trans*-1,3-cyclohexanediol remained in diethyl ether. Ammonia acetone solution was *cis*-1,3-cyclohexanediol/Cu²⁺ destroyed the complex used to and yield pure cis-1,3-cyclohexanediol (44).

For instance, treatment of (\pm)-**37** with 0.2 mol% Pd/Au–CSPVP **3** (the amount of catalyst is based on the total moles of Pd and Au) and 0.3 equiv of K₂CO₃ in water under 1 atmospheric oxygen to yield (*S*)-3-hydroxycyclohexanone (**38**). After screening different ratios of Pd versus Au such as 1:0, 9:1, 3:1, 1:1, 0:1 of the bimetallic nanoclusters, Pd/Au (3:1)-CSPVP appears to provide the highest catalytic activity and chemical yields in the aerobic catalytic asymmetric oxidation of (\pm)-*trans*-cyclohexanediol (**37**).(**Table 4**, entries 1–5) Higher temperature was

needed when using palladium without gold and the reaction gave lower enantioselectivity. However, gold nanaclusters cannot catalyze the oxidation reaction even under 100 $^{\circ}$ C which indicate palladium is the real catalyst in the reaction and gold can improve the catalytic reactivity. Another control experiment was conducted by using normal PVP (MW = 40,000) instead of CSPVP, which gave achiral product. It is also an evidence which shows the chiral environment was induced from CSPVP.(**Table 4**, entry 6)

	ſ	OH 0.15	mol% Pd/Au (ratio	o)-(CS)PVP ↓		
	НО ^{,,,}	O ₂ (1	atm.), K ₂ CO ₃ , H ₂ C	D, HO ^{\\.}		
	(:	±)- 37		(S)- 38		
entry	PVP	Pd:Au	reaction time	Temperature(°C)	% yield	% ee
1	3	1:0	14 days	80	42	64
2	3	9:1	14 days	60	44	83
3	3	3:1	7 days	60	47	88
4	3	1:1	7 days	60	38	82
5	3	0:1		no reaction under 1	00 °C	
6	achiral	3:1	7 days	60	46	0

 Table 4.
 Reaction Condition Studies for the Oxidation of (±)-trans-cyclohexanediol(37).

After understanding the best ratio of Pd/Au, the reaction condition was consistently used for the oxidation of (\pm) -*trans*-cycloalkanediols to give the similar and comparable results. The first generation of polymer 1 - 4 were tested in the same reaction condition. 0.2 mol% of Pd/Au (3:1)-CSPVP 1 - 4 and 0.3 equiv. of K₂CO₃ in water under 1 atmospheric oxygen at 60 °C for 7 days was used for the oxidation of (\pm) -*trans*-1,3-cyclohexanediols (**37**) and gave 46–49% chemical yields and 70–99% ee of (*S*)-3-hydroxycyclohexanone (**38**) along with the recovered

(R,R)-37 (**Table 5**, entries 1–4). It appears CSPVP1 gives the best enantioselectivity due to the bulkiest C5-substituent group. It is also another provement thet the enantioselective oxidation was because of the appearance of CSPVP. Therefore, Pd:Au(3:1)-CSPVP1 was locked to be the catalyst of diol oxidation reactions.

Through kinetic resolution, two enantiomers react with different reaction rates in a chemical reaction with a chiral catalyst, resulting in an enatioencriched sample of the less reactive enantiomer. In the oxidation of (\pm) -*trans*-1,3-cyclohexanediols (**37**), 50% of diols was recovered with 92% ee, which means at least 96% of (R,R)-**37** was not oxidized in 7 days. With the kinetic resolution of Pd:Au(3:1)-CSPVP1, (S,S)-**37** was oxidaized much faster than (R,R)-**37**. Therefore, stopping the reaction before half of the starting material reacted was needed to get optical pure (S)-**38**. Since the reactions were difficult to stop at exactly half point and partial of (S,S)-**37** remained in the recovered enriched (R,R)-**37**, the enantiomeric excess of the recovered (R,R)-**37** was related to the experiment operation and have potential to increase. It also provides an effective way to separate the racemic **37** to (R,R)-**37** from (S,S)-**37**. Moreover, hydroxyl ketone (S)-**38** did not undergo oxidation, revealing an electron-withdrawing ketone group impeded further oxidation.

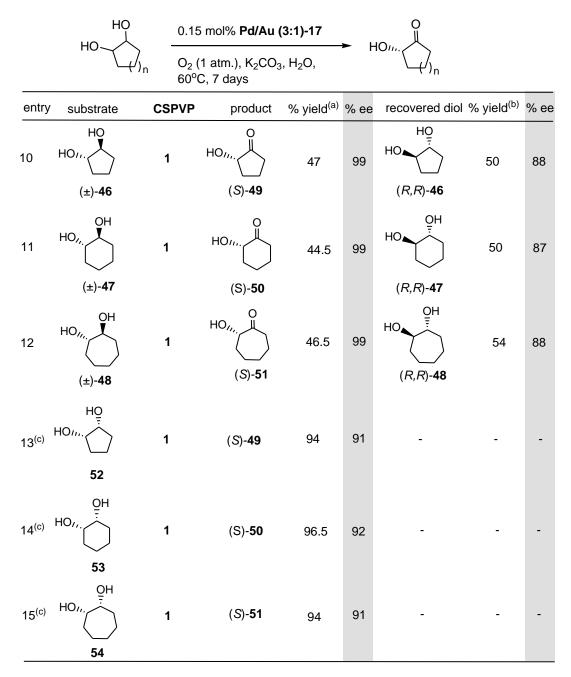
Using 0.2 mol% of Pd/Au (3:1)-1, other five- and seven-membered cycloalkanes, such as 1,3-*trans*-cycloalkanediols (\pm)-**39**, (\pm)-**40** and *trans*-1,2-cycloalkanediols (\pm)-**46**–(\pm)-**48**, were similarly oxidized to give excellent chemical and optical yields. Results are presented in **Table 5** (entries 5 and 6, and entries 10–12). *trans*-(\pm)-**39** and *trans*-(\pm)-**40** were obtained with Cu²⁺ complex method similar like *trans*-(\pm)-**37**. *trans*-(\pm)-**46**–(\pm)-**48** were separated from mixture of *cis* and *trans* mixture by column chromatography on silica gel. Meanwhile, *cis*-**43**–**45** and **52**–**54** were isolated for oxidation study.

When the amounts of catalysts were increased to 0.40 mol% (double) or 0.80 mol% (quadruple) of Pd/Au (3:1)-1, the oxidation of (\pm) -37 were respectively completed in 6 and 5 days, with chemical yields of 47% (99% ee) and 49% (99% ee) of (*S*)-38. Hence, an increase of the amounts of catalyst shortens the reaction time. The recyclability of the Pd/Au (3:1)-1 catalyst was also examined. Hence, the catalyst, recovered from the oxidation reaction of (\pm) -37, was reused under similar reaction conditions for a second time, and 39% yield (99% ee) of (S)-38 was isolated. The catalyst was further recycled for a third time but only 18% yield (98% ee) of (S)-38 was obtained, showing the catalyst can be reused albeit with lower catalytic activities. The turnover number (TON) of the catalytic oxidation reaction after three cycles is > 706.

The meso 1,3 and 1,2-*cis*-cycloalkanediols do not undergo oxidation reaction under similar reaction conditions as those of *trans*-diols. It may due to the fact that *cis*-diols form stable complexes with the nanoparticles.^[57] However, under elevated pressure of oxygen (30 psi.) and temperature (120°C), the reactions took place in 3 days to give 89–97% yields in 90–92% ee of the oxidized (*S*)-hydroxyl ketones (**Table 5**, entries 7–9 and entries 13–15). The higher reaction temperature (120°C) lowered slightly the optical yields comparing with those obtained from *trans*-cycloalkanediols. This oxidative desymmetrization of meso-diols is synthetically useful, since it provides the chiral hydroxyketones without the recovery of starting diols. Importantly, both *trans*- and *cis*-diols gave only the (S)-hydroxy ketones. No changes in reaction rate and chemical and optical yields in the oxidation reaction of meso-**53** were found when doubling the amount of **1** but maintaining the amount of Pd/Au (3:1).

		но	0.15 mol% Pd/Au (3:1)-CSPVP		o L			
	нс	∫_() _n	O ₂ (1 atm.), K 60ºC, 7 days	₂ CO ₃ , H ₂ O,		HO		
entry	substrate	CSPVP	product	% yield ^(a)	% ee	recovered diol	% yield ^(b)	% ee
1	OH HO ^{\\'} (±)- 37	1	HO ¹¹ (S)- 38	49	99	OH 	50	92
2	(±)- 37	2	(S)- 38	46	83	(<i>R</i> , <i>R</i>)- 37	52	79
3	(±)- 37	3	(S)- 38	46.5	88	(<i>R</i> , <i>R</i>)- 37	52	81
4	(±)- 37	4	(S)- 38	46	70	(<i>R</i> , <i>R</i>)- 37	51	65
5	HO HO ^{''} (±)- 39	1	HO ^{''} (S)- 41	48	99	HO HO (<i>R</i> , <i>R</i>)- 39	53	91
6	ОН НО ^{\\'} (±)- 40	1	HO ¹¹ .(S)- 42	46	99	HO (<i>R</i> , <i>R</i>)- 40	53	85
7 ^(c)	HO HO HO	1	HOX. (S)- 41	97	90	-	-	-
8 ^(c)	OH 	1	HO ^{VV} (S)- 38	89	91	-	-	
9 ^(c)	ОН НО ^{\''} 45	1	HO''' (S)-42	94.5	92	-	-	-

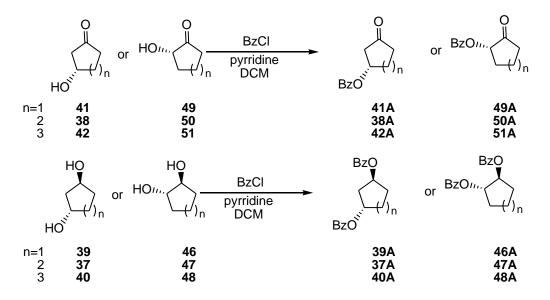
(to be continued)



(a) Isolated yields were based on starting racemic or meso-diols.
 (b) Percent recoveries were based on recovered unreacted starting diols.
 (c) Reactions were carried out at 120°C under 30 psi. of oxygen for 3 days.



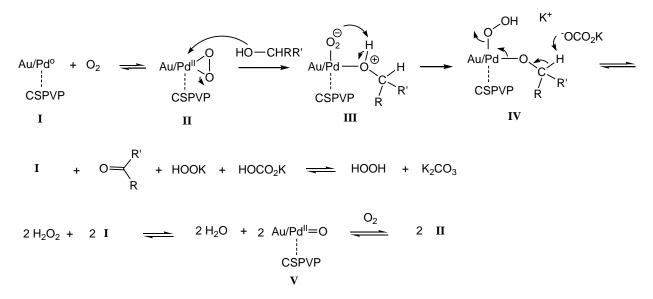
The absolute configurations of (*S*)-**38**, (*S*)-**41** and (*S*)-**49**–(*S*)-**51** were determined by comparison of the sign of the reported specific rotations.^[58-61] The specific rotation of (*S*)-**42** has not been reported previously and its absolute configuration was assumed based on the retention time of its benzoate derivative in HPLC/chiral column. Optical purities were measured through HPLC/chiral column of their benzoyl derivatives, which were synthesized by the treatment of the hydroxyl ketones with benzoyl chloride and pyridine. In all cases, PVP was used in place of CSPVP under similar reaction conditions to provide the racemic products for HPLC/chiral column analyses. The absolute configurations and optical purities of the unreacted diols (*R*,*R*)-**37**, (*R*,*R*)-**39**, (*R*,*R*)-**40** and (*R*,*R*)-**46**–(*R*,*R*)-**48** were similarly determined by comparison of the sign of the reported specific rotations^[62, 63] and through HPLC/chiral column of their dibenzoate derivatives obtained from the reactions of the diols with benzoyl chloride and pyridine. (Scheme 7)



Scheme 7. Benzoylation of (S)-Hydroxycycloalkanones and Dibenzoylation of Recovered (*R*,*R*)-Cycloalkanediols.

Dr. Hua has proposed the mechanism for the oxidation of diols.^[1] (**Scheme 8**) It has been reported a η^2 -peroxido Pd^{II} (peroxopalladium) species **II**^[64] formed with the oxidation of Pd(0).^[65] Then the hydroxyl group in diols attacks Pd(II), open the three membered ring and gives complex **III**, which undergoes proton transfer to give **IV**. Finally, the base, K₂CO₃, removes the C-1 proton and gives the corresponding ketone, **I** and H₂O₂. Oxidation of **I** with H₂O₂ gives **II** and go into next catalytic circle.^[66] The exact mechanism for the enantioselectivity is not clear.

Nanoclusters Pd/Au-CSPVP provides excellent enantioselectivities (91–99% ee) in the oxidation of diols, and results of the enantioselectivity from the oxidation of *trans*-diols are comparable to those reported using enzymes.^[67, 68]



Scheme 8. Proposed Mechanism for the Asymmetric Catalytic Oxidation of Racemic and Meso 1,3-Cycloalkanediols and 1,2-Cycloalkanediols.

2.2 Asymmetric Dihydroxylation of Alkenes.

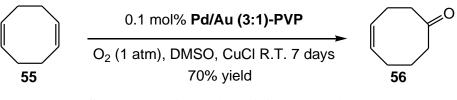
2.2.1 Background and Significance

The development of catalytic reaction of alkene transformed the chemical industry and gave various ways for the organic synthesis. For example, the Nobel Prize in chemistry 2001 was divided to Professor Knowles, Noyori and Sharpless for their work on chirally catalyzed hydrogenation or dihydroxylation reactions. Dihydroxylation is the process by which an alkene is converted into a vicinal diol. Although there are many routes to accomplish this oxidation, the most common and direct processes use a high-oxidation-state transition metal, such as *anti*-Dihydroxylation^[69], Upjohn *syn*-Dihydroxylation^[70] and Prevost syn-Woodward Dihydroxylation^[71], catalytic asymmetric *syn*-dihydroxylation reactions were majorly conducted by the Sharpless asymmetric dihydroxylation process.^[72] However, the use of toxic and volatile osmium tetraoxide catalyst makes the reaction purification difficult. Moreover, the enantioselective dihydroxylation of some Z-1,2-disubstituted alkenes and alkenes containing stereogenic centers using Sharpless method gave lower enantioselectivity.^[72] Therefore, Other catalysts^[73] with osmium-free asymmetric dihydroxylation reactions including palladium-catalyzed difunctionalization of alkenes,^[74] RuO₄,^[75] KMnO₄,^[76] and iron complexes^[77] have also been investigated. Until now, Asymmetric dihydroxylation of alkenes bimetallic nanoparticles and chiral polymers have not been using discovered. Pd/Au(3:1)-CSPVP catalyst without osmium dihydroxylated alkenes with similar or higher enantioaselectivity compare with sharpless method.

2.2.2 Results and Discussions

Although no dihydroxylation of alkenes took place at 1 atmospheric of oxygen with Pd:Au(3:1)-PVP nanoclusters, but with the co-catalyst CuCl, (no reaction without CuCl) the

corresponding ketone (**56**) was isolated from the oxidation of cyclooctadiene (**55**) with 70% yield in DMSO. The dihydroxylation product was not detected. (**Scheme 9**)

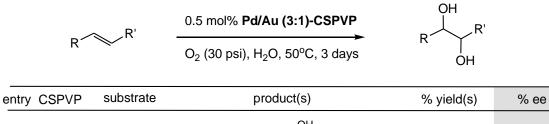


Scheme 9. Oxidation of Cyclooctadiene.

However, various alkenes were readily oxidized at 30 psi. of oxygen in excellent to good chemical yields and excellent optical yields. Results of the oxidation are summarized in Table 6. For example, treatment of 1,2-dihydronaphthalene (57) with 0.5 mol% of Pd/Au (3:1)-1 in water at 25°C under 30 psi. of oxygen for 3 days gave (15,2R)-58 in 86% yield and 99% ee (Table 6, entry 1). Only a single product was isolated in the reaction. The Sharpless dihydroxylation of 57 using DP-PHAL gave 56% ee of (1R,2S)- 58.^[78] Other disubstituted alkenes including *trans*- and *cis*- β -methylstyrene (59 and 62, respectively; Table 6, entries 2 and 3) also underwent oxidation reactions at 25°C. In the oxidation of trans-59, the syn-adduct (15,25)-60 was isolated in 87% yield and 99% ee along with a small amount of anti-adduct (1R,2S)-61 in 6% yield and 97% ee. Similarly, in the oxidation of cis-62, the syn-adduct (1S,2R)-61 was isolated in 90% yield and 98% ee as well as 8% yield of anti-adduct (1R,2R)-60 in 98% ee. However, only syn-addut (1S,2R)-58 was isolated when 0.3 equiv of K₂CO₃ was added to the oxidation reaction of 57 at 25°C. The disubstituted alkenes without aromatic ring were also examined, for example, cyclohex-2-enone (73) and cyclooct-4-enone (75), were oxidized under similar condition except at 50 °C to (2R,3R)-74 (86% yield) and (4S,5R)-76 (85% yield), respectively. (**Table 6**, entries 9 and 10) The increase of reaction temperature indicates the aryl group, as a electron-donating group, increased the activity for the oxidation reaction. The higher ee value (99%), the oxidation of 73 and 75 gave, indicate the enantioselectivity of CSPVP26 is better than CSPVP1 even the reaction temperature are higher. Notably, the dihydroxylation takes place from the *re* face of the alkenes (in regarding to C1 of 57, 59, 62, 73 and 75).

The oxidation of alkenes containing an electron-withdrawing group such as *trans*-cinnamic acid ester **63** and indene (**66**) only proceeded at 50°C giving the *syn*-adducts (2*R*,3*S*)-**64** (82% yield; 99% ee) and (1*S*,2*R*)-**67** (67% yield; 93% ee), respectively, as the major products. The respective *anti*-adducts, (2*R*,3*R*)-**65** (3% yield; 97% ee) and (1*R*,2*R*)-**68** (11% yield; 94% ee) were also isolated as minor products. (**Table 6**, entries 4 and 5) Since (1*R*,2*R*)-**68** was obtained in 11% yield, *anti*-adduct was tried to improve as a major product. As expected, in the presence of 0.3 equiv of K₂CO₃ at 70°C for 5 h, (1*R*,2*R*)-**68** was isolated in 59% yield and 91% ee. (**Table 6**, entries 6)

Trisubstituted alkenes such as (R)-(+)-limonene (**69**), containing a stereogenic center, also underwent stereoselective oxidation at 25°C to give a single stereoisomer (1*S*,2*R*,4*R*)-**70**. (**Table 6**, entries 7) The formation of (1*S*,2*R*,4*R*)-**70** from the dihydroxylation reaction of limonene is remarkable, since the dihydroxyl functions were delivered from the same side of the bulky isopropenyl group. The cyclic trisubstituted alkene reacted preferentially compared to the acyclic terminal disubstituted olefin group, suggesting electron-donating groups such as alkyl and aryl enhance the reactivity towards the nanoclusters.



			,		···) ··· (·)	
1 ^(a)	1	57	(1 <i>S</i> ,2 <i>R</i>)	,"ОН	86	99
2 ^(a)	1	59	<u>ОН</u> <u>-</u> ОН (1 S,2 S)-60		87 , 6	99 , 97
3 ^(a)	1	62	(13,23)-00	(1 <i>R</i> ,2 <i>S</i>)-61 ОН ОН (1 <i>R</i> ,2 <i>R</i>)-60	90 , 8	98 , 98
4	1	63	i-Pr OH CO ₂ -i-Pr OH (2 <i>R</i> ,3 <i>S</i>)-64	ОН со, 	₂ -i-Pr 82,3	99 , 97
5	1	66	ОН (1 <i>S</i> ,2 <i>R</i>)-67	он он (1 <i>R</i> ,2 R)-68	67,11	93 , 94
6 ^(b)	1	66	(1 <i>S</i> ,2 <i>R</i>)-67	(1 <i>R</i> ,2R)-68	5 , 59	89 , 91
7	1			он он }	92	-
8 ^(c)	26	(<i>R</i>)-69	(1 <i>S</i> ,2 <i>R</i> ,4 <i>R</i>)	-70 	73	97

(To be continued)

entry	CSPVP	substrate	product(s)	% yield(s)	% ee
9	26	73 0	OH HO (2 <i>R</i> ,3 <i>R</i>)-74	86	99
10	26	56	HO HO (4S, <i>5R</i>)-76	85	99
11	26	CO ₂ Et 77 CO ₂ Et	HO (<i>1S,2R</i>)-81	77	99
12	26	78	HO (1 <i>S</i> ,2 <i>R</i>)-82	83	99
13	26	79 CO ₂ Et	HO, CO ₂ Et (1 <i>S</i> ,2 <i>R</i>)-83	85	99
14	26	CO ₂ Et	HO HO ,CO ₂ Et (1S,2R)-84	81	99

(a) Reactions were conducted at 25° C for 3 days; (b) reactions were conducted at 70° C for 5 h in the presence of 0.3 equiv K₂CO₃; (c) 5% mol of Pd:Au-CSPVP nanoclusters was used.

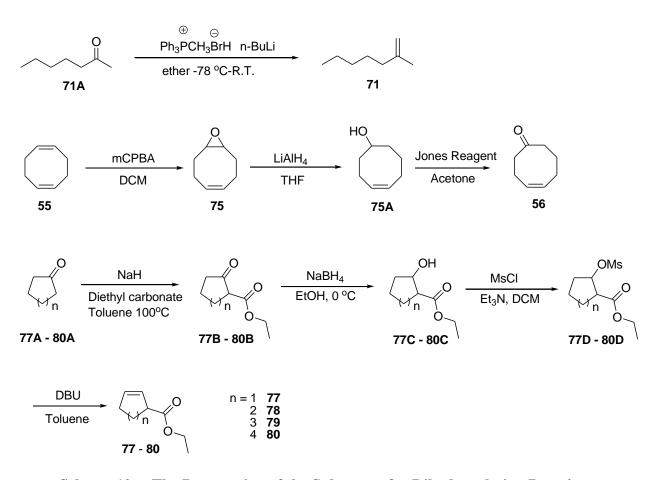
Table 6. Results of Catalytic Asymmetric Dihydroxylation of Alkenes.

However, the terminal disubstituted olefin group can also be oxidized by increasing the amount of the catalyst. With 5 % of Pd/Au(3:1)-CSPVP**26**, 2-methylhept-1-ene (**71**) were oxidized to (*S*)-2-methylheptane-1,2-diol (**72**) with 73% chemical and 97% optical yield. (**Table 6**, entries 8)

Molecule (+)-(R)-**71** has previously been obtained from an improved Sharpless oxidation in a 85% yield and 86% ee.^[79, 80] The successful oxidation of **71** expanded the scope of the oxidation method.

The oxidation of trisubstituted alkenes containing an electron-withdrawing group, ethyl cycloalk-1-enecarboxylate (77 - 80) were also oxidized at 50°C and gave the *syn*-adducts (1S,2R)-81 - 84 with 77 - 81% yield and ~99% ee. (Table 6, entries 11–14)

The substrates were commercially available if not mentioned below. Compound **71** was made by Wittig Reaction from 2-heptanone (**71A**). With the epoxidation of cyclooctadiene (**55**) with mCPBA in methylene chloride, reduction with LiAlH₄ in THF and oxidation of alcohol with Jones Reagent, cyclooct-4-enone (**56**) was obtained. Compound **77–80** were made from cycloalkanone **77A–80A**, respectively. The compounds **77A–80A** were treated with NaH and diethyl carbonate in toluene, reduced the ketone with NaBH₄, protected the resulting alcohol with MsCl-Et₃N and eliminate to give **77–80** with 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU). The preparation of the substrates was conducted by Dr. Man Zhang and Mr. Kyle Apley. (**Scheme 10**)

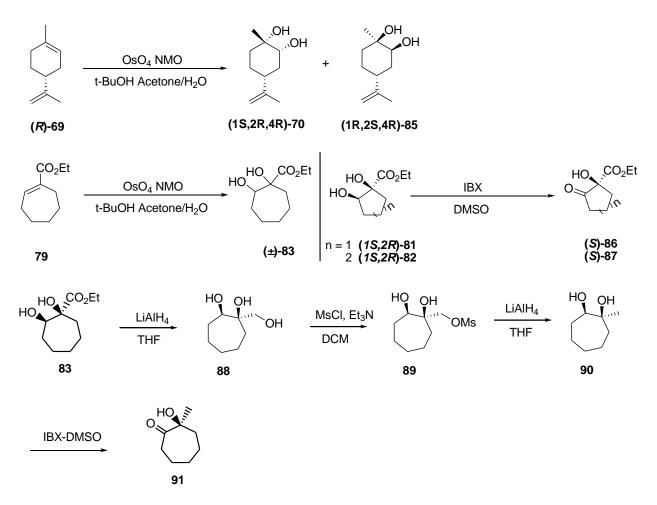


Scheme 10. The Preparation of the Substrates for Dihydroxylation Reactions.

The absolute configurations were determined by comparing the sign and specific rotations of the respective reported molecules.^[67, 68, 81, 82] To determine the stereochemistry of **70** (which has not been reported previously), we independently prepared (1S,2R,4R)-**70** and its (1R,2S,4R)-diastereomer (**85**)^[83] by the reaction of (*R*)-**69** with OsO₄ (catalytic amount)-NMO in t-BuOH-H₂O followed by silica gel column chromatography. The *syn*-stereochemistry of the products were confirmed independently by oxidizing **79** with OsO₄-NMO in t-BuOH-H₂O to give (±)-**83**, whose ¹H NMR spectrum was identical to that of (-)-**83**. The absolute configuration of **81** and **82** were deduced by oxidation with IBX-DMSO to the corresponding (-)-(1*S*)-1-hydroxy-2-oxocycloalkanecarboxylic ethyl esters (**86** and **87**, respectively), whose specific rotations are similar to those reported (+)-(*1R*)-isomers but possessing opposite sign.^[84]

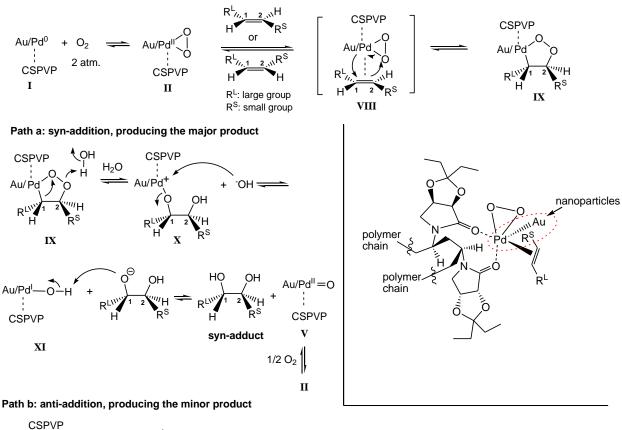
The C1-absolute stereochemistry of (-)-**83** was established by converting it to the known compound, (2*S*)-2-hydroxy-2-methylcycloheptanone (**91**). ^[85] Molecule **83** was reduced by LiAlH₄ to give corresponding triol and mesylated the primary alcohol with MsCl/Et₃N to give **89**. Molecule **89** was reduced with LiAlH₄ and oxidized the secondary alcohol with IBX/DMSO to give **91**. The highest reported % ee's of the dihydroxylation products of **59**, **62**, **63** (the ethyl ester analog), **66**, **71** and **73** from Sharpless asymmetric dihydroxylation reactions are 98,^[86] 81,^[87] 99,^[82], 89,^[88] 86^[79] and 99,^[89] respectively. Optical purities of all dihydroxylated molecules were measured using HPLC/chiral column. Hence, the Pd/Au-CSPVP provided higher or similar enantiomeric selectivities in the catalytic asymmetric dihydroxylation reactions.

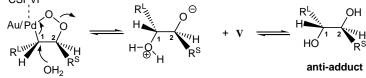
Dr. Hua has also proposed the mechanism for the dihydroxylation of alkenes.^[1] Similar to that described in **Scheme 8**, Pd(0) was oxidized to η^2 -peroxido Pd(II) (peroxopalladium) species **II** and complexed with C=C in alkenes. Through intermediate **VIII**, metallo-1,2-dioxolane **IX**^[65] was formed. Carbon-1 of **IX** undergoes a 1,2-alkyl shift from Pd to oxygen (path a) leading to intermediate **X**, which hydrolyzes to give the *syn*-adducts. In some case, the 1,2-alkyl shift takes place rapidly and only the *syn*-adducts were found. In other investigated alkenes, in addition to major path a, water attacks C1 of intermediate **IX** from the backside, path b, providing *anti*-addition diols as the minor products. (**Scheme 12**)



Scheme 11. Confirmation of the Absolutely Configurations of the Dihydroxylation Products.

The asymmetric aerobic oxidation of alkenes by Pd/Au-CSPVP at ambient temperature or 50° C provided excellent chemical and optical yields of the *syn*-dihydroxylated products and in certain cases where higher temperature (such as 70° C) is required, an addition of a weak base such as K₂CO₃ afforded *anti*-dihydroxylated molecules as the major products.





Scheme 12. Proposed Mechanism and Stereo- and Regio-chemistry of the Enantioselective Oxidation of Alkenes.

2.3 Selective C-H Asymmetric Oxidation Reactions of Cycloalkanes.

2.1.1 Background and Significance

C-H oxidation has a long history and an ongoing presence in research at the forefront of chemistry and interrelated fields. In a single molecule, even medium-size organic molecules, so many different C-H bond can be oxidized. The active C-H oxidative sites normally have weaker

C-H bond dissociation energy and other effect, such as electronic effect, steric effect, conjugation and hyperconjugation, etc. ^[90] Therefore, the active C-H oxidation sites are normally on aromatic ring,^[91] benzylic^[92] or allylic^[93] position. Regio- and enantioselective C-H oxidation of cycloalkanes is one of the most challenging transformations in organic synthesis. In general, directing groups such as hydroxyl group, ester or amide are needed for the regioselectivity. Although various of different catalyst such as iron complexes,^[94-96] gold reagents,^[97] and others^[90, 98-100] in C-H oxidation have been reported, no research on the use of bimetallic nanoclusters and chiral supports in the catalytic enantio- and region-selective C-H oxidation appears. C-H oxidation is a highly atom-economic process, and regioselective catalytic asymmetric C-H oxidation would provide a powerful tool for organic synthesis. To explore the scope of the oxidation reactions, a more challenging catalytic asymmetric C-H oxidation was investigated.

2.1.2 Results and Discussions

Since the Pd:Au(3:1)-CSPVP did not work well for the C-H oxidation, different bimetallic nanoclusters have been investigate for the C-H oxidation including Fe:Au(3:1)-CSPVP and Cu:Au(3:1)-CSPVP. Both Fe:Au(3:1)-CSPVP and Cu:Au(3:1)-CSPVP have the catalytic activity on the C-H oxidation of isopropylcyclohexane to yield 3-isopropylcyclohexanone. However, Fe:Au(3:1)-CSPVP nanoclusters are not stable at high temperature, forming the black precipitate which is magnetic. Therefore, instead of using Pd:Au (3:1)-CSPVP nanoclusters, Cu:Au (3:1)-CSPVP nanoclusters was used for the C-H oxidation of the cycloalkanes.

Under oxygen atmosphere (30 psi.), no C-H oxidation product was detected, making the stronger oxidizer (hydrogen peroxide) necessary for the oxidation reactions. After making the crude Cu:Au(3:1)-CSPVP solution, the substrate was added into it followed with 30% H_2O_2 . The

remaining NaBH₄ was also quanched by H_2O_2 with the formation of hydrogen gas. Since the organic substrates are difficult to dissolve in water, acetonitrile was added slowly into the reaction system until the entire substrate dissolving after heating to 50 °C. Too much acetonitrile (>1/2 by volumn by experience) may produce black precipitate after heating for several hours which means the nanoclusters decomposition.

Results of the catalytic asymmetric C-H oxidation reactions of various substituted achiral cycloalkanes using 1 mol% Cu/Au (3:1)-**CSPVP** and H_2O_2 at 50°C are summarized in **Table 7**. The products are chiral molecule obtained from the desymmetrization process of the achiral substrates. For example, oxidation of meso-*trans*-1,4-diisopropylcyclohexane (**92**) gave an 87% yield of (2*R*,5*S*)-**93**. (**Table 7**, entry 1) There are four C-H bond positions which has potenssial to be oxidized. The four methyls on isopropyl group, two methine C-H's on isopropyl group, two methine C-H's on cyclohexanone ring and four methylene groups on the ring. It appears the four methyle methyls were not oxidized. Moreover, the four methylene group was selectively oxidize two of them to give a chiral product with 81% ee. The Cu:Au(3:1)-CSPVP1 performed both region- and enantioselectivity on the catalytic oxidation reaction of **92**. The specific rotation and ¹H and ¹³C NMR spectral data of (2*R*,5*S*)-**93** were in agreement with those of the reported compound synthesized from (*S*)-perillaldehyde.^[101]

Similarily, 1-isopropylcycloalkanols **94–96** and 1-isopropenylcyclohexanol (**100**) were conducted to be oxidized with the catalysis of Cu:Au(3:1)-CSPVP1 and yield corresponding ketone (*R*)-**97–**(*R*)-**99**, and (*R*)-**101**, respectively (**Table 7**, entries 2–5). The hydroxyl group is the directing group in the reaction and only γ position away from hydroxyl group were oxidized to give both region- and enantioselectivity. Notebly, the -C=CH₂ of **100**, which is more active

than the alkane moiety, was not oxidized at 50° C under H₂O₂ with the Cu:Au (3:1)-CSPVP nanoclusters,

The regiochemistry of (*R*)-**97**–(*R*)-**99**, and (*R*)-**101** were analyzed by 2D COSY spectroscopy. In the COSY spectrum of (*R*)-**97**, the signal at δ 2.41–2.30 ppm assigned for C2-hydrogens has no correlation with other protons, while the signals at ~2.25 and 2.05 ppm assigned for C5 hydrogens show correlation with C4 hydrogens at 2.01 and 1.60 ppm. Similar correlations were found in the 2D COSY spectra of (*R*)-**98**, (*R*)-**99**, and (*R*)-**101**. Moreover, (*R*)-**98** and (*R*)-**99** were independently synthesized from (*S*)-**38** and (*S*)-**42**, respectively, by silylation of the C3-hydroxyl group followed by addition reaction with isopropylmagnesium bromide,^[102] removal of the silicon protecting group, and oxidation with IBX-DMSO.(Scheme 13) Their NMR spectra and specific rotations were similar to those obtained from the C-H oxidations. The products indicated the chelation of nanoclusters and the *tert*-hydroxy group directed asymmetric oxidation at γ -carbon away from C-OH group of cycloalkanes.

More directing group effect, such as alcohol, ester, and amide, were studied. Compounds 102 - 105 were designed and investigated due to the presence of acetoxy, hydroxyl and ethyl ester group. Oxidation of 102 - 105 with 1 mol% Cu/Au (3:1)-26 and H₂O₂ at 50°C gave exclusively C3-oxo products 106 - 109 (76–83% yields), respectively.(Table 7, entries 6 – 9) Compare with the oxidation results from 94 - 96, the active C-H bonds for the Cu/Au (3:1)-CSPVP catalytic oxidation of 102 - 105 should be γ '- carbon which are not in agreement with the real results. The oxidation took place at the γ -carbon away from C-OH group of cycloalkanes. It indicated the directing group in 102 - 105 were acetoxy group, which directed the oxidation on the γ -carbon away from C=O of acetoxy group. With the appearance of C=O, the directing effect of hydroxyl group was override, indicated the directing effect of C=O was stronger than hydroxyl group.

5 mol% Cu/Au (3:1)-CSPVP

Cyclic alkanes

Chiral oxidized cyclic alkanes

30% H_2O_2 , CH_3CN , 50°C, 7 days

entry	CSPVP	substrate	product	% yield	% ee
1 ^(a)	1	92	(2 <i>R</i> ,5 <i>S</i>)- 93	87	81
2 ^(a)	1	Э2 ОН 94	(2A,53)- 93	91	92
3 ^(a)	1	ОН 95	(<i>R</i>)-98	98	91
4 ^(a)	1	ОН 96	(<i>R</i>)-99	89	93
5 ^(a)	1	ОН 100	(R)-101	98	93
6 ^(a)	26	HO AcO (1 <i>S</i> ,2 <i>R</i>)- 102	HO CO_2Et AcO $\gamma \gamma$ O(1S,2S)-106	83	-

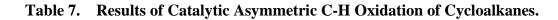
(To be continued)

entry	CSPVP	substrate	product	% yield	% ee
7 ^(a)	26	HO CO ₂ Et AcO (1 <i>S</i> ,2 <i>R</i>)- 103	HO Aco (1S,2S)- 107	83%	-
8 ^(a)	26	HO, CO ₂ Et ACO (1 <i>S</i> ,2 <i>R</i>)- 104	HO CO_2Et $O = \gamma \gamma$ (1S,2S)-108	76%	
9 ^(a)	26	AcO HOCO ₂ Et	AcO HO , CO ₂ Et	83%	
10 ^(a)	26	HO PvO (1 <i>S</i> ,2 <i>R</i>)- 81A	No product detec	cted	
11	26	HO CO ₂ Et PvO (1 <i>S</i> ,2 <i>R</i>)- 110	HO PvO y y (1 <i>S</i> ,2 <i>R</i>)- 113	83%	-
12	26	HO, CO ₂ Et PVO (1 <i>S</i> ,2 <i>R</i>)-111	HO, CO ₂ Et PvO, y - 0 (1 <i>S</i> ,2 <i>R</i>)- 114	88%	-
13	26	PvO HO	PvO HO ,,,CO ₂ Et	88%	-

(To be continued)

entry	CSPVP	substrate	product	% yield	% ee
14	26		ОСОСН ₃ 0 у (S)-121	94	94
15	26	NHCOCH ₃	(S) 121 NHCOCH ₃ (S)-122	76	92
16	26	NHCOCH ₃	NHCOCH ₃ Ο γ (<i>S</i>)-123	72	93
17	26	NHCOCH ₃	NHCOCH ₃ γ (<i>S</i>)-124	84	93
18	26	CO ₂ Et	CO ₂ Et γ (<i>S</i>)- 125	88	92

(a) 1% mol of Cu:Au-CSPVP nanoclusters was used.

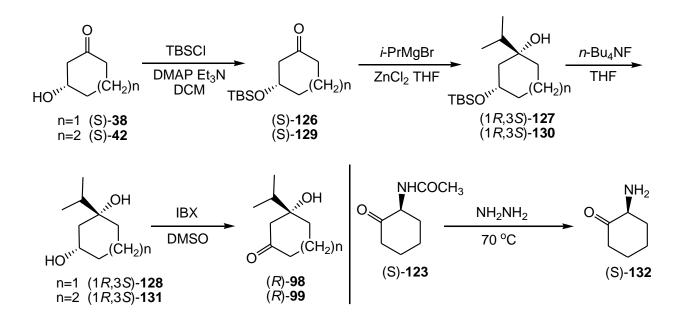


After changing the acetoxy group to pivaloyl group, the oxidation took place at the γ '- carbon of the C1-hydroxyl group, which was in agreement of the hydroxyl directing effect. For example, oxidation of compounds **110–112** gave only **113–115**, respectively, in 83–90% yields.(**Table 7**, entries 11–13) Because the bulky *tert*-butyl moiety blocks the chelation of C=O in PvO with

nanoclusters, allowing the OH group to chelate and direct the oxidation. The regiochemistry of **113–115** were revealed by their 1H and 2D COSY NMR spectra. Notably, no desired oxidation product was isolate from the treatment of monopivaloyl (PvO) analog of (*1S,2R*)-Ethyl 1,2-dihydroxycyclopentanecarboxylate (**81A**) even heating to 100 $^{\circ}$ C with seal tube. (**Table 7**, entries 10) It may due to the crowed environment on the five-membered ring which prevented the active C-H bond oxidation.

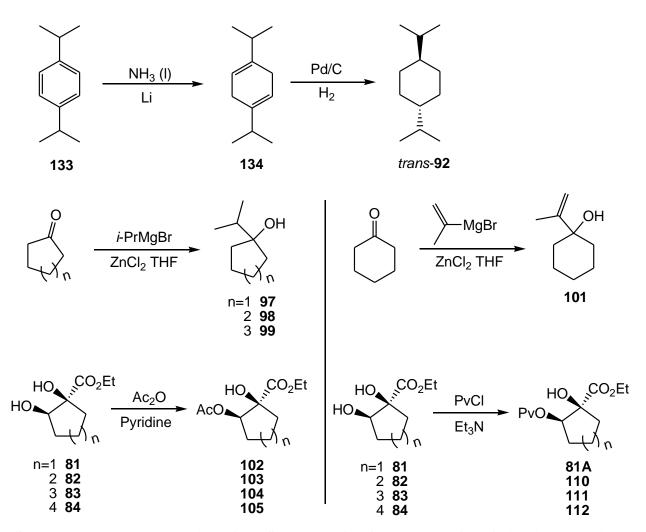
This functional-group directing effect provides guidance for desymmetrized C-H oxidation. Hence, monofunctionalized cyclohexanes 116 - 120 were oxidized readily to give respective (S)-121 - (S)-125 in excellent chemical (72–94%) and optical (92–94% ee) yields (Table 7, entries 14–18). Notebly, in order to study the effect of borane in the catalyst solution, molecular weight cut off membrane was used to remove all the inorganic small molecules including borane. The remaining solution was used directly to conduct the C-H oxidation of 120 to 125 and gave similar results as the crude catalyst solution with 87% yield and 92% ee. The acetoxy, acetamide, and ethyl ester functions apparently directed the oxidation at the γ -carbon away from C=O group on the ring. From the above results, the aptitude of directive oxidation can be summarized as followed: CH₃CO₂ ~ CH₃CONH > OH > CO₂Et.

The NMR spectra of **121**, **123–125** were identical to those reported.^[103-105] The sign of specific rotation of (*S*)-**121**, is opposite to that reported (*R*)-**121**.^[103] Compound (*S*)-**123** was converted to 2-(*S*)-aminocyclohexanone (**132**) by the treatment with 85% hydrazine at 70°C, (**Scheme 13**) and the sign of specific rotation of the resulting amine, $[\alpha]_D^{22} = -71.3$ (c 0.5, CHCl₃), is opposite to that of reported 2-(*R*)-aminocyclohexanone.^[103] The absolutely configuration of **122** and **124** were assumed based on the oxidation result of **118** to (*S*)-**123**. Absolute configuration of (*S*)-**125**, $[\alpha]_D^{22} = +2.62$ (c 1.61, MeOH), was similar to that reported.^[105]



Scheme 13. The Synthesized Molecules for Confirming the Absolutely Configurations of the Cycloalkanes Oxidation Products.

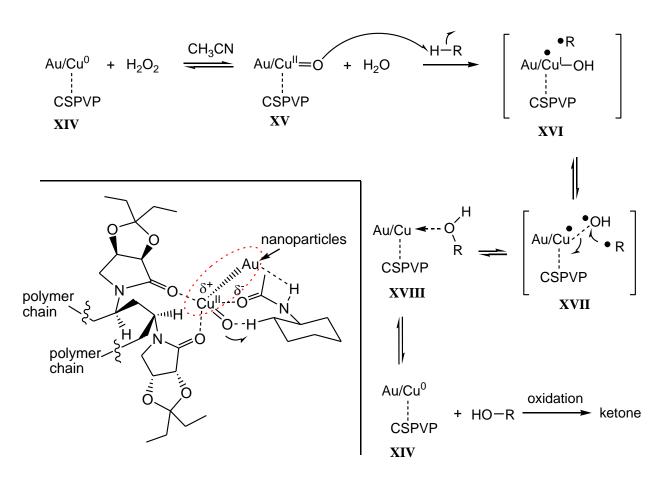
Trans-92 was made by Birch Reduction by Li-NH₃ from 1,4-diisopropylbenzene (133). Compounds 94 – 96 and 100 were made from the addition reactions of isopropylmagnesium bromide or isopropenylmagnesium bromide with cycloalkanones in the presence of 0.2 equiv of ZnCl₂ in THF at 0°C, respectively.^[102] Alcohols 81 – 84 were mono-acetylated with acetic anhydride (Ac₂O) and pyridine to give 89–95% yields of 102 – 105 and mono-pivaloylated with pivaloyl chloride and Et₃N to give 85–88% yields of 110 – 112. Compounds 116 – 120 were simply esterificated or acetylated from cyclic alcohol or amine, respectively.(Scheme 14) Dr. Man Zhang and Mr. Kyle Apley helped to prepare the substrates 94 – 96 and 116 – 120.



Scheme 14. The Preparation of the Substrates for Cycloalkane C-H Oxidation Reactions

Dr. Hua has also proposed the mechanism for the Cu/Au-CSPVP catalytic C-H oxidation reactions based on the previously suggested mechanism of copper complexes bearing trispyrazolylborate ligands.^[1, 106] (Scheme 15) Cu⁰/Au XIV first reacts with H₂O₂ to give copper(II) oxo complex XV, which activated the C-H bond by pulling a hydrogen atom from the substrate to produce hydroxyl copper(I) radical and cycloalkyl radical complex XVI. Cycloalkyl radical and hydroxyl radical then form a C-O bond to yield alcohol and regenerates XIV. The alcohols were further oxidized under the reaction conditions to give ketones. Cu(II) complexed with the directing group and O in Cu(II)=O formed hydrogen bonding with the H on

the active site, constructed a six (hydroxyl group as directing group) or seven-membered ring (carbonyl as directing group) transition state.



Scheme 15. Proposed Mechanism of the Enantioselective Catalytic Oxidation of Alkanes.

Various substituted cycloalkanes were oxidized by Cu/Au-CSPVP and H_2O_2 to furnish chiral cycloalkanones. With the presence of various directing group, the regio-selective C-H oxidation was achieved with high chemical yield. Moreover, the desymmetrization of 1,4-disubstituted cyclohexane or monosubstituted cycloalkanes expanded the scope of the asymmetric oxidation catalyzed by CSPVP stabilized bimetallic nanoclusters and gave application prospects for the oxidation of complex compounds.

2.4 Asymmetric Oxidative Ring-Closing Reaction of Dienes.

2.1.1 Background and Significance

After the discovery of C-H oxidation on the cyclic compounds, acyclic substrates have also been investigated. Unfortunately, the acyclic compounds, such as heptan-4-yl acetate or N-(heptan-4-yl)acetamide, cannot be oxidized with the similar condition. However, based on the directing effect of OH group, 5-methylnona-1,8-dien-5-ol was conduct for the oxidation reaction and the desymmetrized ring-closing product γ -Lactone was isolated without other C-H oxidation products.(discovered by Dr. Man Zhang in Hua's laboratory) γ -Lactones or lactams are part of the important core structural in many research fields such as pharmaceutical chemistry, natural products, perfumes and food additives.^[107, 108] Different kinds of catalysts with different catalytic mechanisms have been reported for the formation of chiral lactone or lactam, such as gold,^[109] rhodium,^[110] titanium,^[111] palladium,^[112] and chiral phosphoric acid^[113]. Various copper catalyzed oxidative ring-closing reaction has been reported, such as arylation, ^[114-116] lactone formation^[107] or lactam formation^[117]. However, the use of bimetallic NCs and chiral supports in the asymmetric oxidative ring-closing reaction for making lactone or lactam has not been reported. The development of the new methodology gives an efficient and fast route for the synthesis and functionalization of lactone or lactam with a stereogenic center on the γ position.

2.1.2 Results and Discussions

Base on Dr. Man Zhang's discovery of the γ -Lactone formation, the reaction condition was optimized for getting desirable results with better chemical and optical yield. For example, compound **197** was added into 4 mol% of Cu/Au(3:1)-**26** in water, (crude nanocluster solution),

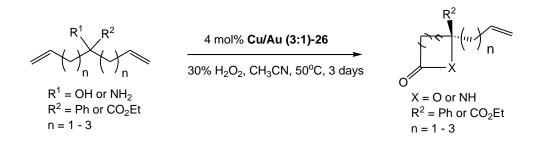
followed with adding 30% H_2O_2 , heating to 50 °C and adding mimimum amount of CH_3CN to dissolve the substrate. Notebly, because of the appearance of ester group in the substrates, 40 equiv. of NaBH₄ (compare with 1 equiv. of HAuCl₄, 100 equiv. of substrates) will be a significant amount which will hydrolysis the ester group. Therefore, the amount of NaBH₄ was cutted to 10 equiv. (compare with 1 equiv. of HAuCl₄, 100 equiv. of substrates) for the oxidative ring-closing reaction.

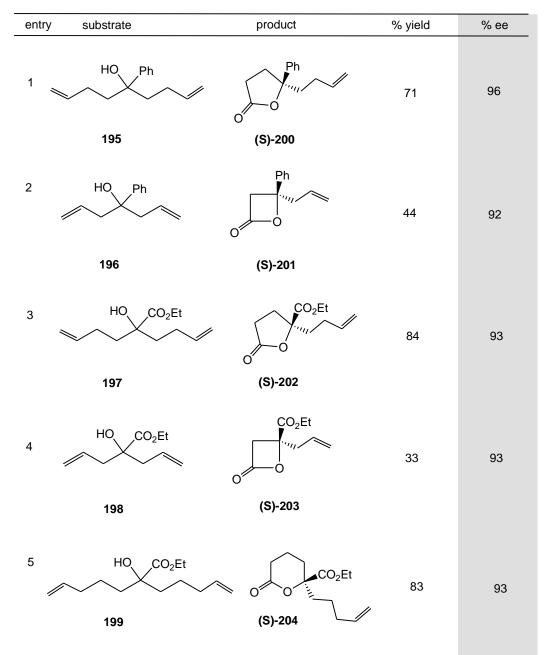
The results for the oxidative ring-closing reaction were summarized in **Table 8**. For example, treatment of **195** with 4 mol% Cu/Au (3:1)-**26** and 30% H₂O₂ in CH₃CN-H₂O at 50°C gave (*S*)-**200** in 71% yield and 96% ee. (**Table 8**, entry 1) Instead of using CSPVP**26**, CSPVP**25** and CSPVP1 were also used to investigate the enantioselectivity of 5-monosubstituted polymers and 3,4-disubstituted polymers. With CSPVP **25** or CSPVP1, 95% or 90% ee product were obtained respectively, indicating CSPVP26 gave the highest enantioselectivity among the three polymers. Other analogs with phenyl group or ethyl ester group such as **196** – **199** were conducted with similar condition to yield similarly catalytic asymmetric oxidative ring closing reaction product **201–204**, respectively. (**Table 8**, entries 2 – 5) With this method, highly stained four-membered ring lactone **201** and **203** were produced although the yield is lower because of the formation of polymers.

Not only γ -lactone, γ -lactam **208** – **210** were also produced with similar reaction condition from the amino ester analog **205** – **207**, respectively, in good chemical (64-93%) and excellent optical (92-95% ee) yields. (**Table 8**, entries 6 – 8)

The absolute configuration of (*S*)-**200** was determined by converting it to (-)-(*S*)-5phenyl-5-propyl-dihydrofuran-2-one (**213**) with $[\alpha]_D^{22} = -42.1$ (c 0.02, MeOH), and the optical rotation agrees to that reported.^[118] Molecule (*S*)-**200** was ozonolysized with O₃ in methylene chloride and reduced with NaBH₄ to give corresponding alcohol **211**. Then the alcohol **211** was mesylated by MsCl in the presence of Et₃N and reduced to remove OMs with NaBH₄ and gave known compound (*S*)-**213**.^[119] The absolute configuration of (*S*)-**208** was determined by conversion into a previously reported pyrrolizidinone (**218**) with similar reported optical rotation.^[120] Molecule (*S*)-**208** was ozonolysized with O₃ in methylene chloride and educed with NaBH₄ to give corresponding alcohol **214**. The alcohol **214** was mesylated by MsCl in the presence of Et₃N to give a good leaving group OMs in compound **215**. After the treatment of **215** with NaH, N⁻ in the lactam ring attacked the CH₂OMs and gave cyclized product **216**. Molecule **216** was treated with LDA to form enolate and hydroxylated by MoOPH^[121] (57% yield along with 8% of the α -OH isomer). Finally, ethyl ester (**217**) was tranfered to methyl ester **218**. (Scheme **16**)

Compounds **195–199** were maily prepared by Kyle Apley and they were made by the Grignard Reaction from isopropyl benzoate or diethyl oxalate with corresponding alkenyl magnesium bromide, respectively. Compounds **205** and **207** were prepared from glycine ethyl ester (**222**) by the sequence: (i) treatment with ethyl formate to give **223**,^[122] (ii) isocyano product (**224**) formation with POCl₃-Et₃N,^[122] (iii) alkylation with NaH and corresponding alkenyl bromide, (iv) hydrolysis of isocyano group to give desired products. Compound **206** were synthesized by treatment of ethyl 2-nitroacetate (**227**) with 1-propenyl acetate followed the reduction with Zn/AcOH. (**Scheme 17**)





(To be continued)

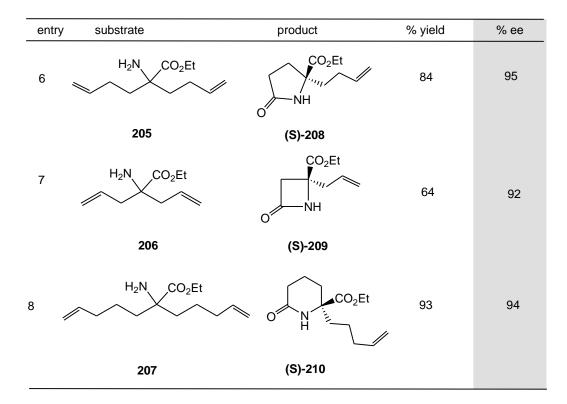
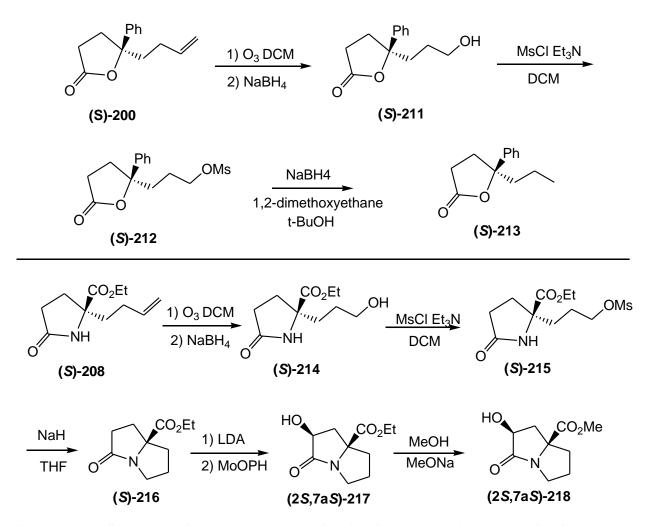
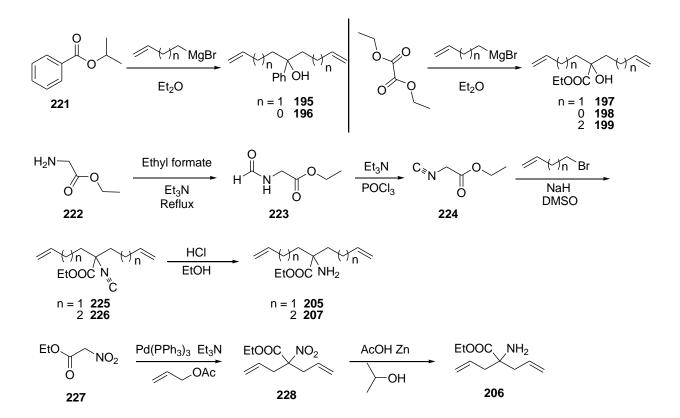


 Table 8.
 Results of Catalytic Oxidative Ring-Closing Reactions.



Scheme 16. Syntheses of Known Molecules for Confirming the Absolutely Configurations of the Oxidative Ring-Closing Products.

A mechanism for the formation of lactones and lactams is proposed (**Scheme 19**), Similarily, Cu^0/Au **XIV** first reacts with H_2O_2 to give copper(II) oxo complex **XV**, which subtracts a hydrogen atom from OH or NH₂ in the substrates, gives hydroxyl copper(I) radical and a oxygen or nitrogen radical intermediate **XIX**. The two radicals then form a Cu-O(N) bond and the C=C in substrates is oxidized by hydroxyl copper to give corresponding alcohol **XXI** and Cu⁰/Au **XIV**. After removing a molecule of formaldehyde from **XXI**, intermediate **XXII** is attacked by water and give **XXIII** which is further oxidized to corresponding lactone or lactam.



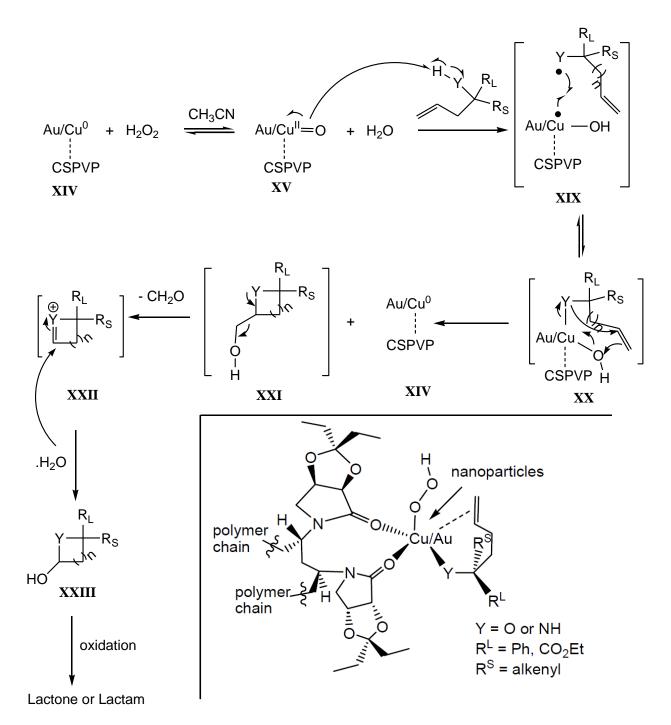
Scheme 17. Preparations of the Substrates for Oxidative Ring-Closing Reactions

2.5 Selective Late-stage C-H Oxidation of Complex Molecules.

2.1.1 Background and Significance

Among the frontier challenges in chemistry in the twenty-first century are the interconnected goals of increasing synthetic efficiency and diversity in the construction of complex molecules. The selective and efficient modification of complex molecules, such as nature products and bioactive compounds, with disparate functional groups is a long-standing challenge that has been solved gradually by using catalysts to perform relatively pure transformations. The straightforward step(s), which is so-called "late-stage" functionalization, provide various analogs with similar scaffold without doing intricate total synthesis. Due to the size of the molecule,

more active sites can be converted in single molecule and selective transformation is important.





Dienes.

However, most of the reported C-H oxidation^[123] and halogenation^[124] often take place at the allylic C-H, or benzylic C-H bond due to a lower bond dissociation energy comparing with that of unactivated CH₂ and CH₃ bonds. The "late-stage" aliphatic C-H alkylation,^[125, 126] C-H azidation,^[127] and C-H oxidation^[128-130] have been investigated in recent years and newly installed functional groups can be further converted into various functionalities, building up a molecule library for testing the bioactivity or drug mechanism study.

2.1.2 Results and Discussions

With the successful C-H oxidations of small size cycloalkanes, medium to large size molecule or nature products were also oxidized to their corresponding analogs. Due to the oxidation difficulty and valuable products, more catalyst (5 mol % – 30 mol %) were used and several substrates gave positive results. For example, oxymatrine (**229**) is one of quinolizidine alkaloid compounds extracted from a Chinese herb called *Sophora Flavescens* and can be used for the protection of cardiovascular system.^[131] With the catalysis of 30 mol % Cu/Au(3:1)-**26** in CH₃CN and H₂O₂, single oxidative product **230** was isolate with 60% yield and recover 33% of **229**. (**Table 9**, Entry 1) Cu/Au-**26** catalyzed reaction has highly selectivity with the directing effect by the carbonyl on C-15, β -carbon away from C=O. The structure was determined by comparing the ¹³C NMR with reported compound.^[132] No C-H oxidation of oxymetrine has been reported previously except microorganism catalyzed oxidation of **230**, giving 10 products with hydroxyl or oxo group on C-12, C-13, C-14.^[132] COSY study has been taken. Base on the assignment of oxymatrine which was reported^[133] and the COSY of the product, the ¹H NMR assignment was shown below. (**Figure 14**)

	δ	intergration	assignment
	5.15-5.09	1	11CH
8 9 10	4.41	1	17CH
$\begin{array}{c} 12 H 7 H 1 0 \\ 0 13 N = 6 N 2 \end{array}$	4.20	1	17CH
	3.50-3.25	2	14CH ₂
16 H $4 15 N 17 5 3$	3.22-3.02	5	2CH ₂ , 6CH, 10CH ₂
	2.82-2.60	2	<mark>3</mark> СН, <mark>9</mark> СН
O H	2.48-2.37	1	12CH
<u> </u>	2.32	1	12CH
		1	8CH
	1.89-1.48	7	3CH, 4CH ₂ , 5CH, 7CH, 8CH, 9CH
$ \begin{array}{c} 12 \\ H \\ 7 \\ H \\ 13 \\ 16 \\ H \\ 15 \\ N \\ 17 \\ H \\ 4 \\ 0 \end{array} $	5.15-5.09 4.41 4.20 3.50-3.25 3.22-3.02 2.82-2.60 2.48-2.37 2.32 2.10-2.04	1 1 1 2 5 2	11CH 17CH 17CH 14CH ₂ 2CH ₂ , 6CH, 10CH 3CH, 9CH 12CH 12CH 12CH 8CH 3CH, 4CH ₂ , 5CH,

Figure 14 COSY Study and ¹H NMR Assignment for 13-Oxo-oxymatrine *N*-oxide (230)

A medium-sized natural product, ambroxide (231), also underwent regioselective oxidation with 5 mol % of Cu/Au(3:1)-26 to give sclareolide (232) with 74% yield and (2*S*)-2-hydroxysclareolide (233) with 6% yield. (Table 9, Entry 2) The oxygen in 231 activated α -C for C-H oxidation and gave lactone, as reported.^[134, 135] However, unlike reported iron complex,^[134] sclareolide (232) was further oxidized to (2*S*)-2-hydroxysclareolide (233) in 38% yield and 3% yield of 1-oxosclareolide (234) along with 46% recovery of 232, in a separate experiment. (Table 9, Entry 3) Notably, the C2 – equatorial hydroxyl function of 233 does not undergo further oxidation, likely due to steric hindrance (C2-axial-H is shielded by C15- β - and C16-methyls).^[136] ¹H and ¹³C NMR spectral data of 233 and 234 are identical to those reported.^[96, 136]

	complex molecule -			
		0% H ₂ O ₂ , CH ₃ CN, 50°C, 7 days		
entry	substrate	product(s)	% yield(s)	% recover
1		$ \begin{array}{c} 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 230 \end{array} $	O] 60	33
2			74	
	231	232 0		3
3			6	46
	232	HO _{1,1} HO _{1,1} HO ₁ HO	38	
			3	

Cu/Au (3:1)-CSPVP

(To be continued)

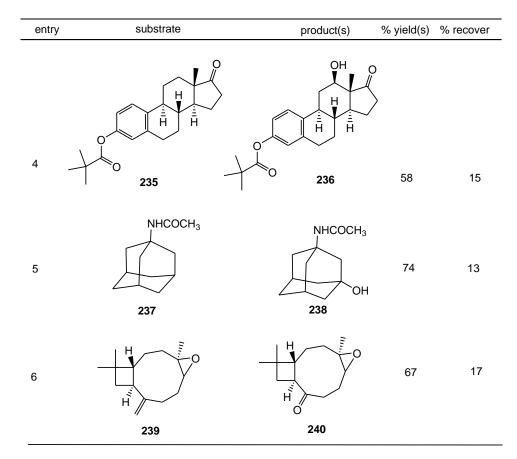
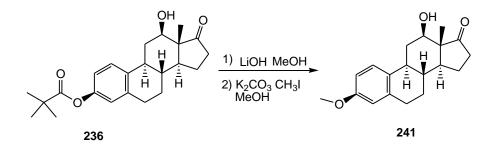


 Table 9.
 Results of Late-stage C-H Oxidation for Complex Molecules.

Steroid type of compound was also selectively oxidized. For example, estrone is an estrogenic hormone and has been discontinued for medical use with the superseding by its derivatives.^[137] With 5 mol % of Cu/Au(3:1)-26 in CH₃CN/H₂O₂, 58% yield of 12 β -hydroxyl estrone derivative (236) was isolated from pivaloyl ester of estrone (235) with 15% recovery yield. (Table 9, Entry 4) The activated C-12 was directed by the carbonyl group which is β -carbon away. Copper catalyzed C-H oxidation of estrone derivatives with 12 β -hydroxyl product has been reported and the conversion from ketone to imine as a directing group for copper is necessary.^[138] The structure of 236 was determined by comparison of the reported 12 β -hydroxyl estrone derivative derivative 241^[138] after the convertion of pivaloyl ester to methoxy group by hydrolysis with LiOH/MeOH and methylation with CH₃I. (Scheme 20)

Amantadine has been approved for use as an anti-viral and an anti-parkinsonian medical by FDA. *N*-Acetylamantadine (**237**) was oxidized at the δ -carbon (away from C=O) to give **238** in 74% yield with the catalysis of 5 mol % Cu/Au(3:1)-**26**. (**Table 9**, Entry 5) The oxidation took place at C3 (δ -carbon) of **237**, which may due to ring strain of adamantane structure.^[139] (**Table 9**, Entry 5) The ¹H and ¹³C NMR data of **238** are identical to those reported.^[140]

To my surprise, caryophyllene oxide (**239**) was also oxidized, but not similar C-H oxidation. The terminal C=C was oxidized to C=O and gave kobusone (**240**) with 67% yield by the catalysis of 5 mol % Cu/Au(3:1)-**26**. The ¹H and ¹³C NMR data of **240** are identical to those reported.^[141] The synthesis of kobusone has been reported from caryophyllene oxide in 2 steps (47% overall yield), as reported.^[141]



Scheme 19. The Synthetic Route for the Confirmation of the C-H Oxidation Product.

In summary, the oxidation produced ketone function (in some cases, the hydroxyl group), which can be converted into amines, amides, or oxime for possible enhancement of bioactivity or for biological mechanistic study. Importantly, there are few methods in the literature can produce the proposed oxidized products in merely one step. Even though the oxidations of complex molecules with the catalysis of Cu/Au-CSPVP are not predictable for now, but it still gives a chance for the late-stage modification and provides various value products.

2.6 Conclusion

In this chapter, several of oxidation reactions were discussed. Cycloalkanediols were asymmetric oxidized by 1 atm of oxygen gas to yield corresponding hydroxyl ketone under the catalysis of Pd/Au(3:1)–CSPVP nanoclusters. Alkenes were oxidized by Pd/Au(3:1)-CSPVP nanoclusters under 2 atmospheric of oxygen in water to give the syn-dihydroxylated products in high chemical and excellent optical yields. Various cycloalkanes underwent regio- and enantio-selective C-H oxidation with Cu/Au(3:1)-CSPVP and 30% hydrogen peroxide to produce the corresponding chiral oxo-molecules in very good to excellent chemical and optical yields. An enantioselective desymmetrization of α , α -dialkenyl-alkanols and α , α -dialkenyl – amino acid ethyl esters to give chiral disubstituted lactones and lactams, respectively. A number of medium-sized natural products. A broad-spectrum predictive C-H oxidation of complex molecules is possible.

2.7 Synthetic Experimental Procedures

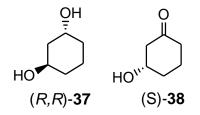
2.6.1 General

¹H NMR spectra (400 MHz) and ¹³C NMR spectra (100 MHz) were measured from a solution in CDCl₃ unless otherwise mentioned. The chemical shift data for each signal on ¹H NMR are given in units of δ relative to TMS ($\delta = 0$ ppm) or CHCl₃ ($\delta = 7.26$ ppm). For ¹³C NMR spectra, the chemical shifts are recorded relative to CDCl₃ ($\delta = 77.0$ ppm). Low-resolution mass spectra were taken from an API 2000-triple quadrupole ESI-MS/MS mass spectrometer. High-resolution mass spectra were obtained using a LCT Premier time of flight mass spectrometer from Waters. IR spectra were measured directly in solid form from Agilent Cary

630 FTIR. Optical Rotation were measured by Bodenseewerk Perkin-Elmer & Co Gmbh Ueberlingen. Optical purities of various oxidized products were determined by HPLC using chiral column, Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries, Exton, PA.

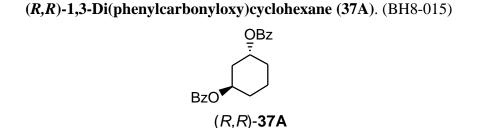
2.6.2 Procedures and Analysis Results for Oxidation of *trans*-Cycloalkanediols.

(*R*,*R*)- *trans*-1,3-Cyclohexanediol (37) & (S)-3-hydroxycyclohexanone (38) (BH7-056)

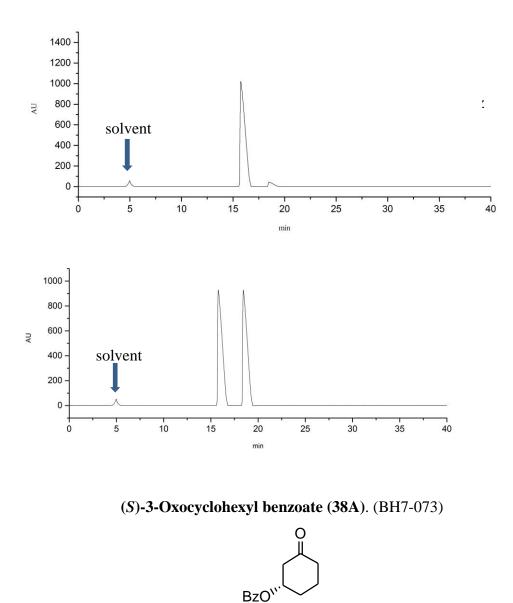


To a solution of 4.8 mL of Pd/Au (3:1)-1 (2.1 µmol of Pd/0.7 µmol of Au-0.08 µmol of 1) in H₂O, were added 0.16 g (1.39 mmol) of racemic *trans*-1,3-cyclohexanediol (**37**) and 68 mg (0.42 mmol) K₂CO₃ under 1 atmospheric pressure of O₂ (a balloon was used). The solution was stirred at 60 °C for 7 days and the reaction progress was monitored by ¹H NMR until about a half of the diol was consumed. The reaction solution was cooled to 25°C, diluted with water, and extracted three times with dichloromethane (20 mL each). The combined extracts were dried (anhydrous Na₂SO₄), concentrated, and column chromatographed on silica gel using a mixture of ethyl acetate and diethyl ether (3:7) as an eluent to give 78 mg [97.5% yield based on reacted (*S*,*S*)-**37**] of (*S*)-3-hydroxycyclohexanone (**38**) in 99% ee and 81 mg (50% recovery based on racemic diol) of (*R*,*R*)-**37** in 92% ee. (*S*)-**38**: $[\alpha]_D^{22} = +45.2$ (c 0.8, CHCl₃); Lit.^[58] +37.3 (c 0.80, CHCl₃; 82% ee). ¹H NMR δ 4.13–4.07 (m, 1 H, CHO), 3.30 (bs, 1 H, OH), 2.56 (dd, J = 14, 4.4 Hz, 1 H, CH₂C=O), 2.33 (dd, J = 14, 8 Hz, 1 H, CH₂C=O), 2.24 (t, J = 7 Hz, 2 H, CH₂C=O), 2.05–1.87 (m, 2 H), 1.75–1.56 (m, 2 H); ¹³C NMR δ 210.9, 69.8, 50.6, 41.1, 32.9, 20.9. MS (ESI, MeOH): *m*/*z* = 137.1 ([M + Na]⁺). The % ee's of the hydroxyketones and diols were determined by subjecting the benzoate derivatives to HPLC using chiral column

[Chiralpak AD(-H) column, size: 0.46 cm x 25 cm]. (*R*,*R*)-**37**: NMR spectra were identical to the authentic *trans*-1,3-cyclohexanediol. $[\alpha]_D^{22} = +3.18$ (c 2.0, CH₂Cl₂); Lit.^[63] -3.5 (c 2.0, CH₂Cl₂). The % ee was determined from the HPLC/chiral column of the benzoate derivative (*R*,*R*)-**37A &** (*S*)-**38A**.



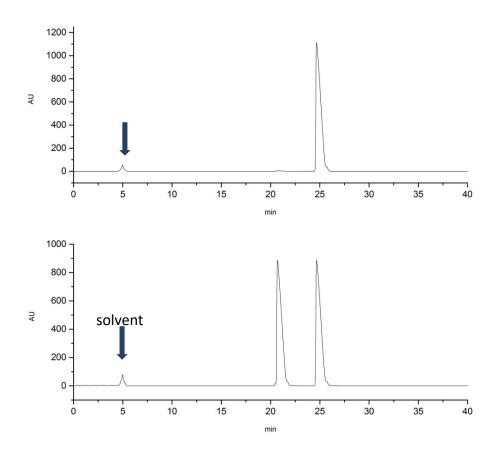
A solution of 10 mg (86 µmol) of (*R*,*R*)-**37**, 24 mg (0.17 mmol) of benzoyl chloride in 0.1 mL of pyridine and 0.3 mL of dichloromethane was stirred at 25°C for 8 h under argon. The solution was diluted with 1 mL of water, concentrated to dryness, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 17 mg (61% yield) of (*R*,*R*)-**37A** in 92% ee. ¹H NMR δ 8.06 (d, J = 8 Hz, 4 H), 7.56 (t, J = 8 Hz, 2 H), 7.45 (t, J = 8 Hz, 4 H), 5.47–5.45 (m, 2 H), 2.15–2.12 (m, 2 H), 1.94–1.91 (m, 2 H), 1.90–1.60 (m, 4 H); ¹³C NMR δ 166.0 (2 C), 133.1 (2 C), 130.8 (2 C), 129.8 (4 C), 128.6 (4 C), 70.8 (2 C), 36.1, 30.6 (2 C), 19.7. MS (ESI, MeOH): *m*/*z* = 347.2 ([M + Na]⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane:*i*-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 15.7 min (*R*,*R* enantiomer, major), t_R = 18.6 min (*S*,*S* enantiomer, minor). The racemic *trans*-dibenzoate **37A** were similarly prepared for HPLC/chiral column study from the corresponding racemic *trans*-diols.



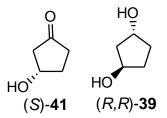
(S)-**38A**

To a solution of 50 mg (0.44 mmol) of (*S*)-**38** in 1.5 mL of dichloromethane under argon at 0°C, was added 0.1 mL of pyridine and 0.12 g (0.85 mmol) of benzoyl chloride. The reaction solution was stirred at 0°C for 1 h, diluted with water, and extracted with CH₂Cl₂ twice. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a mixture of ethyl acetate and hexane (1:2) as an eluent to give 88 mg (92% yield) of (*S*)-**38A** in 99% ee. ¹H NMR δ 8.00 (d, J = 7.6 Hz, 2 H), 7.56 (t, J = 7.6 Hz 1 H), 7.43 (t, J = 7.6 Hz, 2 H), 5.55–5.50 (m, 1 H, CHO), 2.75 (dd, J = 14.8, 4 Hz, 1 H),

2.66 (dd, J = 14.8, 5.6 Hz, 1 H), 2.50–2.40 (m, 2 H), 2.17–2.2.0 (m, 3 H), 1.97–1.85 (m, 1 H); ¹³C NMR δ 208.4, 165.7, 133.4, 129.8, 128.7, 128.6, 72.4, 46.8, 41.3, 29.6, 21.1. MS (ESI, MeOH): m/z = 241.1 ([M + Na]⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane-i-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 21 min (*R* enantiomer, minor), t_R = 25 min (*S* enantiomer, major).

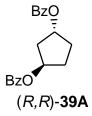


(S)-3-Hydroxycyclopentanone (41) & (R,R)-cyclopentanediol (39) (BH7-067)

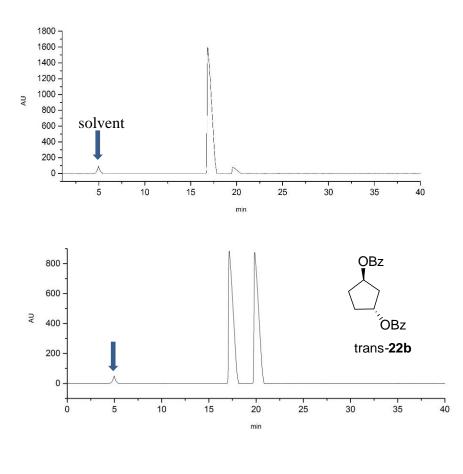


From 15 mg (0.147 mmol) of (±)-cyclopentanediol (**39**), 0.48 mL of Pd/Au (3:1)-**1** (0.22 µmol of Pd/0.07 µmol of Au-0.008 µmol of **1**) in H₂O, 6.8 mg (96% yield based on reacted diols **39**; 99% ee) of (*S*)-**41** and 8.0 mg (53% recovery of (*R*,*R*)-**39** in 91% ee. Compound (*S*)-**41**: $[\alpha]_D^{22} =$ -33.8 (c 0.6, CH₂Cl₂); Lit.^[59] -18.3 (c 0.50, CH₂Cl₂; for S-configuration, 83% ee). ¹H NMR δ 4.63–4.60 (m, 1 H, CHO), 2.52–2.36 (m, 2 H), 2.28–2.12 (m, 3 H), 2.10–2.00 (m, 1 H), 1.85–1.87 (bs, 1 H, OH); ¹³C NMR δ 218.0, 69.9, 47.9, 35.8, 32.3. MS (ESI, MeOH): *m/z* = 123.0 ([M + Na]⁺). (*R*,*R*)-**39**: The NMR spectra were identical to those of the authentic 1,3-*trans*-cyclopentanediol. $[\alpha]_D^{22} = +17.3$ (c 1.0, CHCl₃). The % ee was determined from the HPLC/chiral column of the benzoate derivative (*R*,*R*)-**39A &** (*S*)-**41A**.

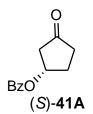
(*R*,*R*)-1,3-Diphenylcarbonyloxycyclopentane (39A). (BH8-021)



From 15 mg (0.147 mmol) of (R,R)-19 and 82 mg (0.59 mmol) of benzoyl chloride in 0.2 mL of pyridine and 0.5 mL of dichloromethane, 26 mg (57% yield) of (*R*,*R*)-**39A** in 91% ee. ¹H NMR δ 8.06 (d, J = 8.5 Hz, 4 H), 7.56 (t, J = 8.5 Hz, 2 H), 7.45 (t, J = 8.5 Hz, 4 H), 5.49–5.44 (m, 2 H), 2.16–2.13 (m, 2 H), 1.94–1.86 (m, 4 H); ¹³C NMR δ 166.6 (2 C), 133.1 (2 C), 130.8 (2 C), 129.8 (4 C), 128.6 (4 C), 71.5 (2 C), 39.7, 24.5 (2 C). MS (ESI, MeOH): *m/z* = 332.8 ([M + Na]⁺),. The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane:*i*-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 16.8 min (*R*,*R* enantiomer, major), t_R = 19.6 min (*S*,*S* enantiomer, minor). The racemic *trans*-dibenzoate **39A** were similarly prepared for HPLC/chiral column study from the corresponding racemic *trans*-diols.

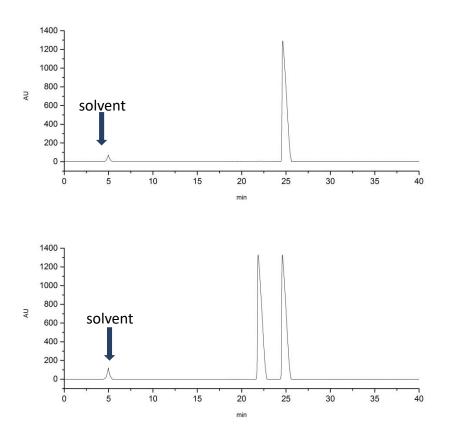


(S)-3-Oxocyclopentyl benzoate (41A). (BH7-075)

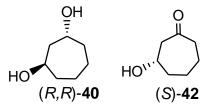


From 6.8 mg (68 µmol) of (*S*)-**41** and 19 mg (0.14 mmol) of benzoyl chloride and 0.1 mL of pyridine in 1 mL of dichloromethane, 13 mg (94% yield) of (*S*)-**41A** in 99% ee. ¹H NMR δ 8.09–8.07 (m, 2 H), 7.59–7.46 (m, 1 H), 7.45–7.42 (m, 2 H), 5.49–5.44 (m, 1 H), 2.73–2.68 (m, 1 H), 2.55–2.51 (m, 1 H), 2.37–2.25 (m, 4 H); ¹³C NMR δ 215.8, 165.7, 133.4, 129.8, 128.7, 128.6, 72.5, 44.4, 35.6, 29.2. MS (ESI, MeOH): m/z = 227.2 ([M + Na]⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column,

n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 22 \text{ min}$ (*R* enantiomer, minor), $t_R = 24.5 \text{ min}$ (*S* enantiomer, major).



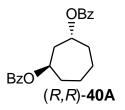
(*R*,*R*)-Cycloheptanediol (40) & (*S*)-3-Hydroxycycloheptanone (42). (BH7-104)



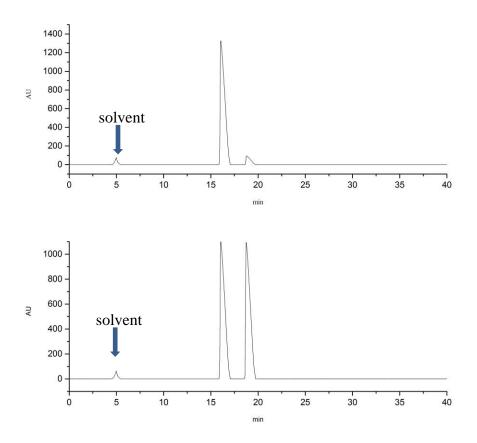
From 30 mg (0.23 mmol) of (±)-cycloheptanediol (**40**), 0.8 mL of Pd/Au (3:1)-**1** (0.35 µmol of Pd/0.12 µmol of Au-0.013 µmol of **1**) in H₂O, 13.4 mg (92% yield based on reacted diols **40**; 99% ee) of (*S*)-**42** and 15.8 mg (53% recovery) of (*R*,*R*)-**40**. Compound (*S*)-**42**: $[\alpha]_D^{22} = +16.4$ (c 0.8, CH₂Cl₂). ¹H NMR^[60] δ 4.13–4.06 (m, 1 H, CHO), 2.84–2.74 (m, 2 H), 2.54–2.40 (m, 2 H),

1.94–1.70 (m, 6 H), 1.65–1.55 (m, 1 H); ¹³C NMR δ 212.4, 67.7, 51.8, 44.5, 39.0, 24.5, 23.9. MS (ESI, MeOH): m/z = 151.1 ([M + Na]⁺). (*R*,*R*)-40: The NMR spectra were identical to those of authentic 1,3-*trans*-cycloheptanediol. $[\alpha]_D^{22} = +16.9$ (c 1.0, CHCl₃). The % ee was determined from the HPLC/chiral column of the benzoate derivative (*R*,*R*)-40A & (*S*)-42A.

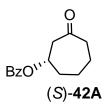
(*R*,*R*)-1,3-(Diphenylcarbonyloxy)cycloheptane (40A). (BH8-020)



From 15 mg (0.12 mmol) of (*R*,*R*)-**40** and 64 mg (0.46 mmol) of benzoyl chloride in 0.2 mL of pyridine and 0.5 mL of dichloromethane, 22 mg (56% yield) of (*R*,*R*)-**40A** in 85% ee. ¹H NMR δ 8.06 (d, J = 8 Hz, 4 H), 7.56 (t, J = 8 Hz, 2 H), 7.45 (t, J = 8 Hz, 4 H), 5.49–5.44 (m, 2 H), 2.13 (t, J = 6 Hz, 2 H), 2.02–1.88 (m, 4 H), 1.87–1.74 (m, 4 H); ¹³C NMR δ 166.5 (2 C), 133.1 (2 C), 130.8 (2 C), 129.8 (4 C), 128.6 (4 C), 70.9 (2 C), 36.3, 29.1 (2 C), 24.1 (2 C). MS (ESI, MeOH): *m*/*z* = 361.4 ([M + Na]⁺), 249.1, 217.3, 123.1, 102.2. The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane:*i*-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 16.1 min (*R*,*R* enantiomer, major), t_R = 18.8 min (*S*,*S* enantiomer, minor).The racemic *trans*-dibenzoate **40A** were similarly prepared for HPLC/chiral column study from the corresponding racemic *trans*-diols.

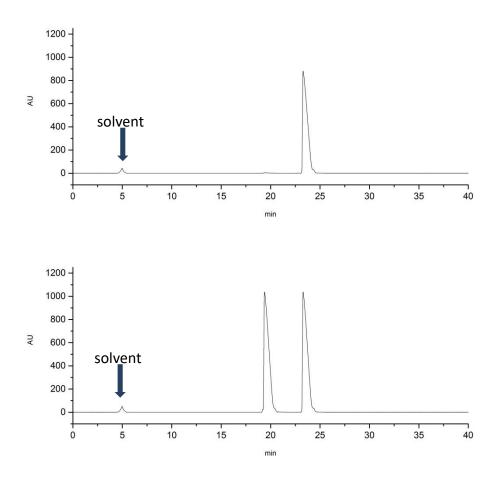


(S)-3-Oxocycloheptyl benzoate (42A). (BH7-107)

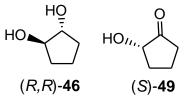


From 10 mg (0.078 mmol) of (*S*)-**42**, 22 mg (0.16 mmol) of benzoyl chloride and 0.1 mL of pyridine in 0.5 mL of dichloromethane, 18 mg (98% yield) of (*S*)-**42A** (99% ee) was obtained. ¹H NMR δ 8.00 (d J = 7.6 Hz, 2 H), 7.56 (t, J = 7.6 Hz, 1 H), 7.43 (t, J = 7.6 Hz, 2 H), 5.54–5.50 (m, 1 H), 2.86 (dd, J = 12, 4.4 Hz, 1 H), 2.81 (dd, J = 12, 8 Hz, 1 H), 2.47–2.40 (m, 2 H), 2.32–2.25 (m, 2 H), 2.15–2.05 (m, 3 H), 1.96–1.86 (m, 1 H); ¹³C NMR δ 208.2, 165.7, 133.7, 129.8 (2 C), 129.0, 128.6 (2 C), 72.4, 46.8, 40.8, 29.1, 21.8, 21.0. MS (ESI, MeOH): m/z = 255.1 ([M + Na]⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak

AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 19$ min (*R* enantiomer, minor), $t_R = 23$ min (*S* enantiomer, major).



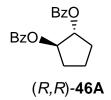
(R,R)- 1,2-Cyclopentanediol (46) & (S)-2-hydroxycyclopentanone (49) (BH7-102)



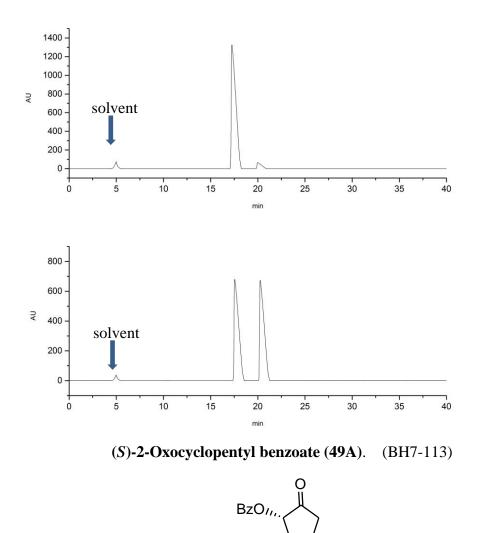
From 30 mg (0.29 mmol) of (\pm)-46, 1.0 mL of Pd/Au (3:1)-1 (0.438 µmol of Pd/0.146 µmol of Au-0.017 µmol of 1) in H₂O, 14 mg (94% yield; based on reacted (*S*,*S*)-46) of (*S*)-49 (99% ee) and 15 mg (50% recovery) of (*R*,*R*)-46. The % ee of (*S*)-49 was determined by HPLC/chiral of

benzoate derivative (*S*)-**49A**. Compound (*S*)-**49**: $[\alpha]_D^{22} = +40.6$ (c 1.0, CHCl₃); Lit.^[61] -37.4 (c 1.0, CHCl₃) for (*R*)-configuration (>99% ee). ¹H NMR δ 4.10 (dd, J = 12, 5 Hz, 1 H, CHO), 3.65–3.55 (bs, 1 H, OH), 2.50–2.34 (m, 2 H), 2.27–2.10 (m, 2 H), 1.86–1.60 (m, 2 H); ¹³C NMR δ 211.9, 75.1, 40.5, 37.6, 26.3. MS (ESI, MeOH): m/z = 123.0 (M+Na⁺). (*R*,*R*)-**46**: The NMR spectra were identical to those of the authentic 1,2-*trans*-cyclopentanediol. $[\alpha]_D^{22} = -19.9$ (c 1.1, CHCl₃); Lit.^[62] -21.3 (c 1.1, CHCl₃) for (*R*,*R*)-configuration (>99% ee).

(*R*,*R*)-1,2-(Diphenylcarbonyloxy)cyclopentane (46A). (reaction was taken by Dr. Man Zhang. HPLC was taken by me)

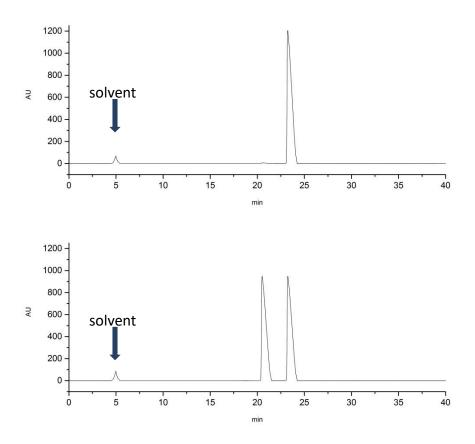


From 14 mg (0.14 mmol) of (*R*,*R*)-**46**, 58 mg (0.41 mmol) of benzoyl chloride in 0.1 mL of pyridine and 2 mL of dichloromethane, gave 29 mg (67% yield) of (*R*,*R*)-**46A**. ¹H NMR δ 8.04–8.00 (m, 4 H), 7.58–7.53 (m, 2 H), 7.46–7.38 (m, 4 H), 5.50 (t, J = 4 Hz, 2 H, CHO), 2.36–2.29 (m, 2 H), 2.00–1.80 (m, 4 H); ¹³C NMR δ 166.2, 133.3, 130.3, 129.9 (4 C), 128.6 (4 C), 79.7, 30.8, 21.9. MS (ESI, MeOH): *m*/*z* = 333.0 (M+Na⁺), 265.3, 189.1, 105.2. The % ee (88%) of (*R*,*R*)-**46A** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 17.3 min (*R*,*R* enantiomer, major), t_R = 20.0 min (*S*,*S* enantiomer, minor). The racemic *trans*-dibenzoate **46A** were similarly prepared for HPLC/chiral column study from the corresponding racemic *trans*-diols.

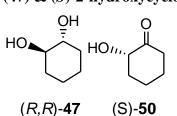


(S)-**49A**

From 10 mg (0.1 mmol) of (*S*)-**49** and 28 mg (0.2 mmol) of benzoyl chloride in 0.1 mL of pyridine and 0.5 mL of dichloromethane, 16.8 mg (82% yield) of (*S*)-**49A** in 99% ee was obtained. ¹H NMR δ 8.08 (d, J = 7.6 Hz, 2 H), 7.57 (t, J = 7.6 Hz, 1 H), 7.45 (t, J = 7.6 Hz, 2 H), 5.46 (dd, J = 9, 3 Hz, 1 H), 2.75–2.65 (m, 1 H), 2.55–2.45 (m, 1 H), 2.17–2.08 (m, 1), 1.95–1.85 (m, 3 H); ¹³C NMR δ 204.6, 169.2, 133.4, 130.1 (2 C), 129.9, 128.6 (2 C), 77.2, 39.9, 31.2, 21.4. MS (ESI, MeOH): m/z = 227.2 (M+Na⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 20.5 min (*R* enantiomer, minor), t_R = 23.3 min (*S* enantiomer, major).

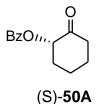


(*R*,*R*)-1,2-Cyclohexanediol (47) & (*S*)-2-hydroxycyclohexanone (50) (BH7-061)

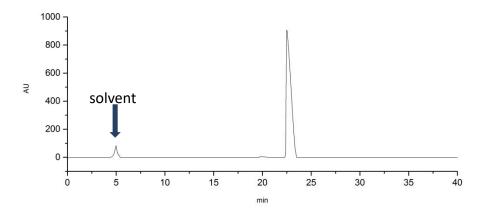


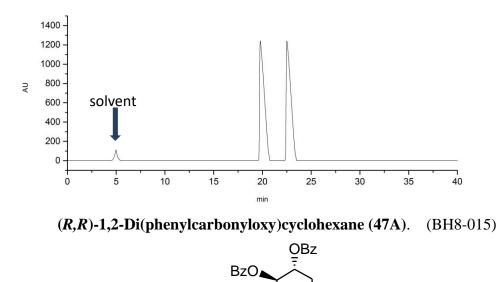
From 161 mg (1.39 mmol) of (±)-47, 4.8 mL of Pd/Au (3:1)-1 (2.1 µmol of Pd/0.7 µmol of Au-0.08 µmol of 1) in H₂O, 72 mg (89% yield; based on reacted (*S*,*S*)-47) of (*S*)-50 (99% ee) and 80 mg (50% recovery) of (*R*,*R*)-47. Compound (*S*)-50: $[\alpha]_D^{22} = -20.7$ (c 0.65, CHCl₃); Lit.^[67] +20.6 (c 1.0, CHCl₃) for (*R*)-50 (>99.0% ee). ¹H NMR δ 4.11 (dd, J = 12, 7 Hz, 1 H, CHO), 3.70–3.55 (bs, 1 H, OH), 2.60–2.30 (m, 3 H), 2.15–1.45 (m, 5 H); ¹³C NMR δ 211.7, 75.6, 39.7, 36.9, 27.8, 23.6. MS (ESI, MeOH): *m*/*z* = 115.1 (M+H⁺). (*R*,*R*)-47: $[\alpha]_D^{22} = -34.7$ (c 1.0, H₂O); Lit.^[61] +37.1 (c 1.0, H₂O) for (*S*,*S*)-29 (>99.0% ee).

(S)-2-Oxocyclohexyl benzoate (50A). (BH7-078)



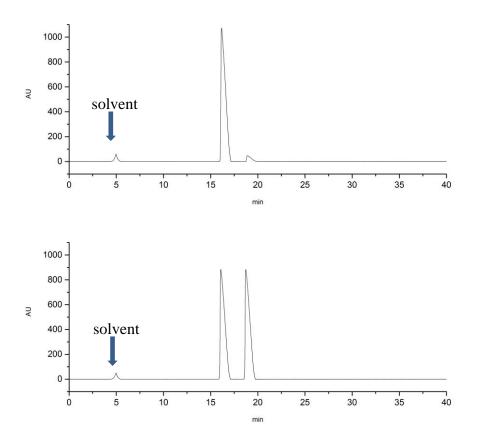
From 50 mg (0.44 mmol) of (*S*)-**50** and 0.122 mg (0.87 mmol) of benzoyl chloride in 0.1 mL of pyridine and 1.5 mL of dichloromethane, 66 mg (69% yield) of (*S*)-**50A** (99% ee). ¹H NMR δ 8.08 (d, J = 8 Hz, 2 H), 7.56 (t, J = 8 Hz, 1 H), 7.45 (t, J = 8 Hz, 2 H), 5.41 (dd, J = 12, 7 Hz, 1 H), 2.60–2.40 (m, 3 H), 2.18–1.64 (m, 5 H); ¹³C NMR δ 204.7, 169.2, 133.3, 130.1 (2 C), 129.9, 128.6 (2 C), 77.2, 40.7, 33.4, 27.4, 24.0. MS (ESI, MeOH): m/z = 241.4 (M+Na⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 20 min (*R* enantiomer, minor), t_R = 22.5 min (*S* enantiomer, major).



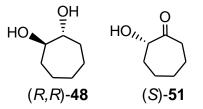


(*R*,*R*)-**47A**

From 20 mg (0.17 mmol) of (*R*,*R*)-**47** and 49 mg (0.34 mmol) of benzoyl chloride in 0.2 mL of pyridine and 0.6 mL of dichloromethane, 51 mg (85% yield) of (*R*,*R*)-**47A** was isolated after silica gel column chromatography. ¹H NMR δ 7.94 (d, J = 7.5 Hz, 4 H), 7.46 (t, J = 7.5 Hz, 2 H), 7.33 (t, J = 7.5 Hz, 4 H), 5.41–5.35 (m, 2 H), 2.27–2.23 (m, 1 H), 2.12–2.06 (m, 1 H), 1.90–1.77 (m, 3 H), 1.63–1.49 (m, 3 H); ¹³C NMR δ 166.2 (2 C), 133.1 (2 C), 129.9 (2 C), 129.1 (4 C), 128.6 (4 C), 74.5 (2 C), 30.4 (2 C), 23.6 (2 C). MS (ESI, MeOH): *m*/*z* = 347.2 (M+Na⁺), 243.2, 123.1, 102.1. The % ee (87%) of (*R*,*R*)-**47A** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 16.2 min (*R*,*R* enantiomer, major), t_R = 18.9 min (*S*,*S* enantiomer, minor). The racemic *trans*-dibenzoate **47A** were similarly prepared for HPLC/chiral column study from the corresponding racemic *trans*-diols.



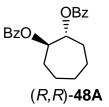
(R,R)-1,2-Cycloheptanediol (48) & (S)-2-hydroxycycloheptanone (51) (BH7-108)



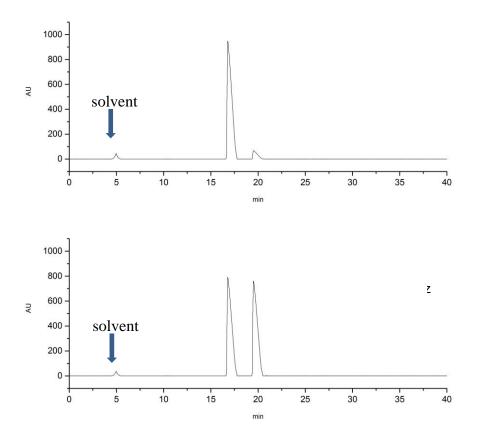
From 30 mg (0.23 mmol) of (±)-cycloheptanediol (**48**), 0.8 mL of Pd/Au (3:1)-**1** (0.35 μmol of Pd/0.12 μmol of Au-0.013 μmol of **1**) in H₂O, 13.5 mg (93% yield based on reacted diols (*S*,*S*)-**48**; 99% ee) of (*S*)-**51** and 16.3 mg (54% recovery) of (*R*,*R*)-**48** (88% ee). The % ee was determined by HPLC/chiral column of benzoate derivative (*S*)-**51A**. Compound (*S*)-**51**: $[\alpha]_D^{22}$ = -143.9 (c 0.8, CHCl₃); Lit.^[61] -86.4 (c 1.0, CHCl₃) for (*R*)-**51** (>99.0% ee). ¹H NMR δ 4.31–4.26 (m, 1 H), 3.83–3.75 (bs, 1 H, OH), 2.71–2.61 (m, 1 H), 2.46 (ddd, J = 17, 11, 4 Hz, 1 H), 2.08–1.55 (m, 7 H), 1.40–1.30 (m, 1 H); ¹³C NMR δ 213.3, 77.20, 39.9, 33.5, 29.4, 26.5, 23.6.

MS (ESI, MeOH): $m/z = 129.2 \text{ (M+H}^+\text{)}$. (*R*,*R*)-48: The NMR spectra were identical to those of the authentic 1,2-*trans*-cycloheptanediol. $[\alpha]_D^{22} = -4.9$ (c 0.7, CHCl₃); Lit.^[61] +10.7 (c 1.0, CHCl₃) for (*S*,*S*)-48 (>99.0% ee).

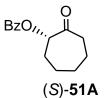
(*R*,*R*)-1,2-Di(phenylcarbonyloxy)cycloheptane (48A). (reaction was taken by Dr. Man Zhang. HPLC was taken by me)



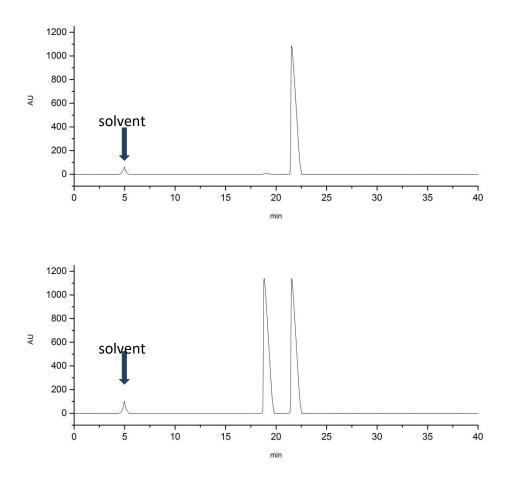
From 10 mg (0.077 mmol) of (*R*,*R*)-**48** and 32 mg (0.23 mmol) of benzoyl chloride in 61 μ L of pyridine and 2 mL of dichloromethane, 23 mg (90% yield) of (*R*,*R*)-**48A** was isolated after silica gel column chromatography. ¹H NMR δ 7.97 (d, J = 7 Hz, 4 H), 7.49 (t, J = 7 Hz, 2 H), 7.38–7.34 (m, 4 H), 5.42–5.40 (m, 2 H, CHO), 2.20–2.00 (m, 2 H), 1.88–1.75 (m, 4 H), 1.75–1.60 (m, 4 H); ¹³C NMR δ 166.3, 133.1, 130.5, 129.8 (4 C), 128.5 (4 C), 77.6, 30.7, 28.5, 23.1. MS (ESI, MeOH): m/z = 361.3 ([M + Na]⁺). The % ee (88%) of (*R*,*R*)-**48A** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min; t_R = 16.8 min (*R*,*R* enantiomer, major), t_R = 19.5 min (*S*,*S* enantiomer, minor). The racemic *trans*-dibenzoate **48A** were similarly prepared for HPLC/chiral column study from the corresponding racemic *trans*-diols.



(S)-2-Oxocycloheptyl benzoate (51A). (BH7-118)

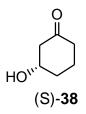


From 10 mg (78 µmol) of (*S*)-**51** and 22 mg (0.16 mmol) of benzoyl chloride in 0.1 mL of pyridine and 0.5 mL of dichloromethane, 17.6 mg (97% yield) of (*S*)-**51A** (99% ee) was obtained. ¹H NMR δ 8.08 (d, J = 8 Hz, 2 H), 7.57 (d, J = 8 Hz, 1 H), 7.45 (t, J = 8 Hz, 2 H), 2.75–2.65 (m, 1 H), 2.55–2.45 (m, 1 H), 2.16–2.10 (m, 1 H), 2.0–1.65 (m, 6 H), 1.51–1.40 (m, 1 H); ¹³C NMR δ 207.4, 165.1, 133.4, 130.1 (2 C), 129.9, 128.6 (2 C), 79.9, 40.8, 30.8, 27.3, 26.1, 22.0. MS (ESI, MeOH): m/z = 255.1 ([M + Na]⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 19 \text{ min } (R \text{ enantiomer, minor}), t_R = 21.5 \text{ min } (S \text{ enantiomer, major}).$

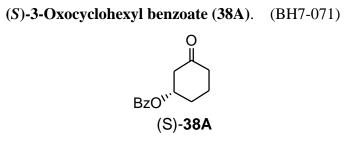


2.6.3 Procedures and Analysis Results for Oxidation of *cis*-Cycloalkanediols.

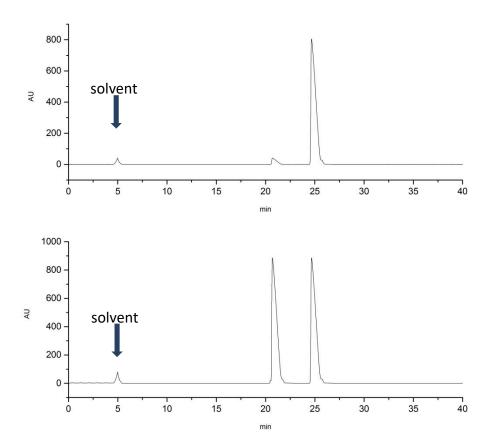
(S)-3-Hydroxycyclohexanone (38) (BH7-053)



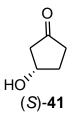
To a solution of 4.8 mL of Pd/Au (3:1)-1 (2.1 µmol of Pd/0.7 µmol of Au-0.08 µmol of 1) in H₂O, were added 16 mL of deionized H₂O, 0.16 g (1.39 mmol; 662 equivalents based on Pd mole atoms) of racemic meso-*cis*-1,3-cyclohexanediol (44) and 68 mg (0.42 mmol) K₂CO₃. The solution was transferred into a high pressure apparatus (Parr Pressure Reactor), stirred at 120°C under 30 psi O₂ for 3 days. The stirring was stopped and reactor was cooled to 25°C. The solution was extracted three times with dichloromethane (20 mL each) and the combined extracts were washed with water and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a mixture of ethyl acetate and hexane (1:1) as an eluent to give 0.141 g (89% yield) of (*S*)-**38** in 91% ee (determined by HPLC/chiral column of the benzoate derivative (*S*)-**38A**). (*S*)-**38**: $[\alpha]_D^{22} = +40.1$ (c 0.8, CHCl₃).; Lit.^[58] for (*S*)-**38**: +37.3 (c 0.80, CHCl₃; 82% ee). ¹H and ¹³C NMR spectra were identical to those of (*S*)-**38** obtained from the oxidation of *trans*-1,3-cyclohexanediol.



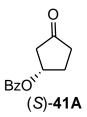
From 0.1 g (0.87 mmol) of (*S*)-**38** and 0.24 g (1.74 mmol) of benzoyl chloride in 0.2 mL of pyridine and 3 mL of dichloromethane, 0.174 g (92% yield) of (*S*)-**38A** (91% ee) was obtained. The % ee of (*S*)-**38a** was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 21 \text{ min}$ (*R* enantiomer, minor), $t_R = 25 \text{ min}$ (*S* enantiomer, major).



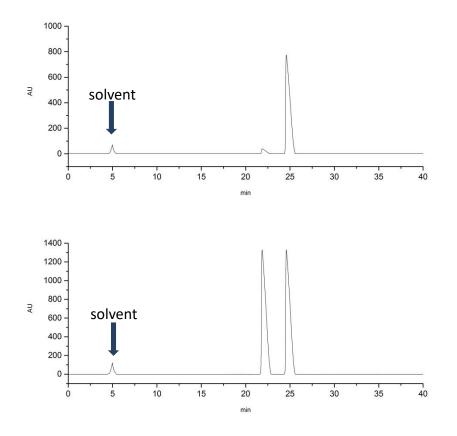
(S)-3-Hydroxycyclopentanone (41) (BH7-069)



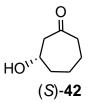
From 30 mg (0.29 mmol) of **43**, 1.0 mL of Pd/Au (3:1)-**1** (0.438 µmol of Pd/0.146 µmol of Au-0.017 µmol of **1**) in H₂O at 120°C and 30 psi of O₂ for 3 days, 28.3 mg (97% yield) of (*S*)-**41** in 90% ee (determined by HPLC/chiral column of the benzoate derivative (*S*)-**41A**) was isolated. (*S*)-**41**: $[\alpha]_D^{22} = -29.9$ (c 0.6, CH₂Cl₂), Lit.^[59] -18.3 (c 0.50, CH₂Cl₂; for S-configuration, 83% ee). ¹H and ¹³C NMR spectra were identical to those of (*S*)-**41** obtained from the oxidation of *trans*-1,3-cyclopentanediol.



From 20 mg (0.20 mmol) of (*S*)-**41** and 56 mg (0.40 mmol) of benzoyl chloride in 0.1 mL of pyridine and 1 mL of dichloromethane, 29 mg (71% yield) of (*S*)-**41A** (90% ee) was obtained. The % ee (90%) of compound (*S*)-**41A** was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 21.8$ min (*R* enantiomer, minor), $t_R = 24.5$ min (*S* enantiomer, major).

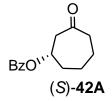


(S)-3-Hydroxycycloheptanone (42). (BH7-080)

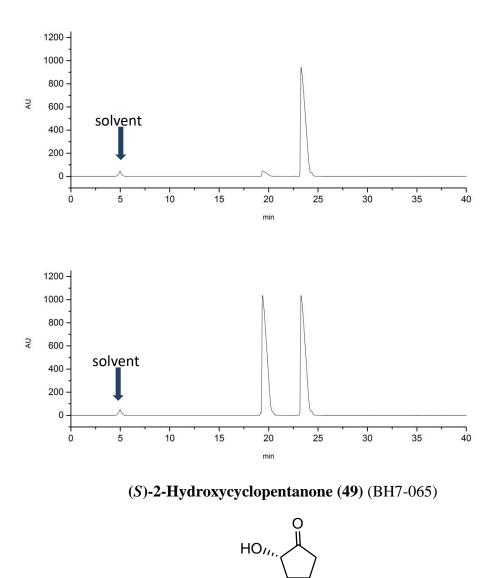


From 30 mg (0.23 mmol) of **45**, 0.79 mL of Pd/Au (3:1)-**1** (0.347 µmol of Pd/0.116 µmol of Au-0.013 µmol of **1**) in H₂O at 120°C and 30 psi of O₂ for 3 days, 28 mg (94.5% yield) of (*S*)-**42** in 92% ee (determined by HPLC/chiral column of the benzoate derivative (*S*)-**42A**) was isolated. [α]_D²² = +15.2 (c 0.8, CHCl₃). ¹H and ¹³C NMR spectra were identical to those of (*S*)-**42** obtained from the oxidation of *trans*-1,3-cycloheptanediol.

(S)-3-Oxocycloheptyl benzoate (42A). (BH7-106)

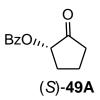


From 25 mg (0.20 mmol) of (*S*)-**42** and 55 g (0.39 mmol) of benzoyl chloride in 0.1 mL of pyridine and 1 mL of dichloromethane, 44 mg (97% yield) of (*S*)-**42A** (92% ee) was obtained. ¹H and ¹³C NMR spectra were identical to those of (*S*)-**42A** as described above. The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 19 min$ (*R* enantiomer, minor), $t_R = 23 min$ (*S* enantiomer, major).

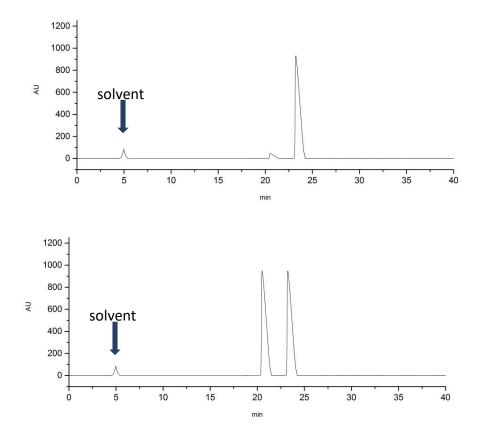


From 60 mg (0.588 mmol) of **52**, 2 mL of Pd/Au (3:1)-**1** (0.89 µmol of Pd/0.296 µmol of Au-0.034 µmol of **1**) in H₂O at 120°C and 30 psi of O₂ for 3 days, 55.4 mg (94.2% yield) of (*S*)-**49** in 91% ee (determined by HPLC/chiral column of the benzoate derivative (*S*)-**49**A). [α]_D²² = +38.4 (c 1.0, CHCl₃); Lit.^[61] -37.4 (c 1.0, CHCl₃) for (*R*)-configuration (>99% ee). ¹H and ¹³C NMR spectra were identical to those of (*S*)-**49** obtained from the oxidation of *trans*-1,2-cyclopentanediol.

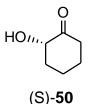
(S)-49



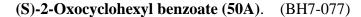
From 40 mg (0.40 mmol) of (*S*)-**49** and 0.112 g (0.80 mmol) of benzoyl chloride in 0.3 mL of pyridine and 1.5 mL of dichloromethane, 80 mg (98% yield) of (*S*)-**49A** (91% ee) was obtained. ¹H and ¹³C NMR spectra were identical to those of (*S*)-**49A** as described above. The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 20.5$ min (*R* enantiomer, minor), $t_R = 23.3$ min (*S* enantiomer, major).

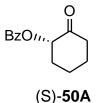


(S)-2-Hydroxycyclohexanone (50) (BH7-058)



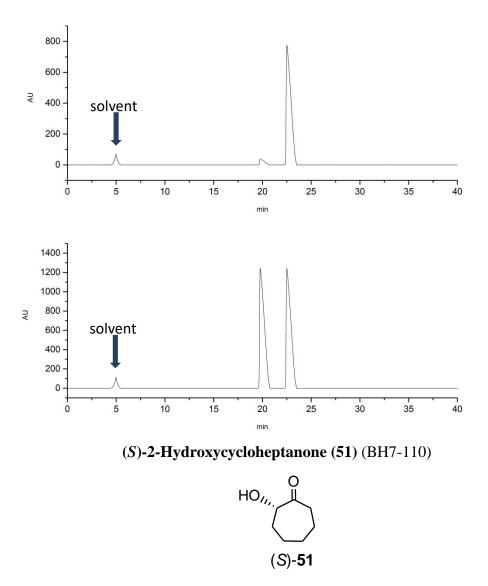
From 0.161 g (1.39 mmol; 662 equivalents based on Pd mole atoms) of **53**, 4.8 mL of Pd/Au (3:1)-**1** (2.1 µmol of Pd/0.7 µmol of Au-0.08 µmol of **1**) in H₂O at 120°C and 30 psi of O₂ for 4 days, 0.142 g (96.5% yield) of (*S*)-**50** in 92% ee [determined by HPLC/chiral column of the benzoate derivative (*S*)-**50A**] was isolated. (*S*)-**50**: $[\alpha]_D^{22} = -18.8$ (c 0.65, CHCl₃); Lit.^[67] +20.6 (c 1.0, CHCl₃) for (*R*)-**50** (>99.0% ee). ¹H and ¹³C NMR spectra were identical to those of (*S*)-**50** obtained from the oxidation of *trans*-1,2-cyclohexanediol.





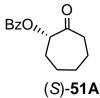
From 0.10 g (0.87 mmol) of (*S*)-**50** and 0.245 g (1.74 mmol) of benzoyl chloride in 0.2 mL of pyridine and 3 mL of dichloromethane, 0.169 g (89% yield) of (*S*)-**50A** (92% ee) was obtained. ¹H and ¹³C NMR spectra were identical to those of (*S*)-**50A** as described above.

The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 20 \text{ min}$ (*R* enantiomer, minor), $t_R = 22.5 \text{ min}$ (*S* enantiomer, major).

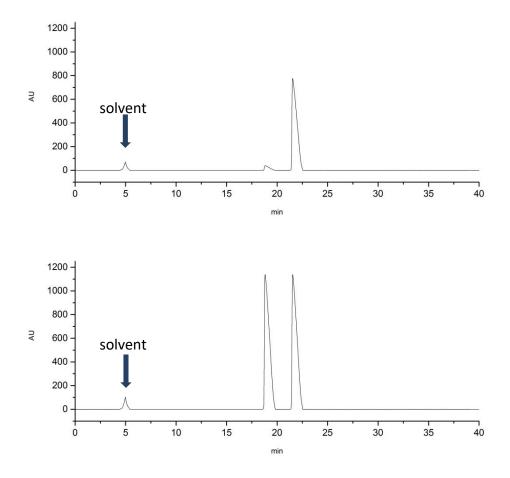


From 30 mg (0.23 mmol) of **54**, 0.79 mL of Pd/Au (3:1)-**1** (0.347 µmol of Pd/0.116 µmol of Au-0.013 µmol of **1**) in H₂O at 120^oC and 30 psi of O₂ for 3 days, 27.6 mg (94% yield) of (*S*)-**51** in 91% ee (determined by HPLC/chiral column of the benzoate derivative (*S*)-**51A**). (*S*)-**51**: $[\alpha]_D^{22} = -130.6$ (c 1.0, CHCl₃). Lit.^[61] -86.4 (c 1.0, CHCl₃) for (*R*)-**51** (>99.0% ee). ¹H and ¹³C NMR spectra were identical to those of (*S*)-**51** obtained from the oxidation of *trans*-1,2-cycloheptanediol.

(S)-2-Oxocycloheptyl benzoate (51A). (add notebook #)



From 20 mg (0.156 mmol) of (*S*)-**51** and 44 mg (0.31 mmol) of benzoyl chloride in 0.2 mL of pyridine and 1 mL of dichloromethane, 35 mg (97% yield) of (*S*)-**51A** (91% ee) was obtained. ¹H and ¹³C NMR spectra were identical to those of (*S*)-**51A** as described above. The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/i-PrOH = 90:10, flow rate: 0.5 mL/min; $t_R = 19$ min (*R* enantiomer, minor), $t_R = 21.5$ min (*S* enantiomer, major).



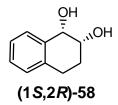
2.6.4 Procedures and Analysis Results for Dihydroxylation of Alkenes.

(**Z**)-cyclooct-4-enone(56) (BH1-071)



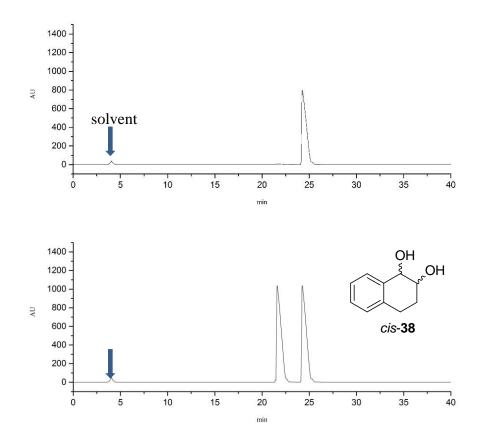
To a 0 °C solution of 23 µmol of Na₂PdCl₄, 7 µmol of HAuCl₄ and 0.8 µmol of PVP in 1.5 mL DMSO, 20 mg (20 µmol) of NaCNBH₃ was added and stired for 30 minutes to give Pd:Au(3:1)-PVP nanoclusters DMSO solution. To it, 5 mL of DMF, 31 mg (312 mmol) of CuCl was added and stirred for 30 minutes under 1 atmospheric of O₂ to give a dark green solution. To it, 0.033 g (0.313 mmol) of cyclooctadiene **55** was added and reacted under 1 atmospheric of O₂ for 7 days, extracted with 3 x 20 mL of diethyl ether, washed combined organic layer with 5 mL water, dried over Na₂SO₄, concentrated under vacuum and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as eluents to give 27 mg (69.6% yield) of **56**. ¹H NMR^[142] δ 5.74–5.63 (m, 2H), 2.52–2.42 (m, 6H), 2.33–2.08 (m,2H), 1.63–1.56 (m, 2H); ¹³C NMR δ 215.0, 130.9, 130.4, 47.4, 40.5, 26.5, 24.1, 22.0; MS (ESI, MeOH): m/z = 125.1 ([M + H]⁺).

(15,2*R*)-1,2,3,4-Tetrahydronaphthalene-1,2-diol (58) (BH7-051)

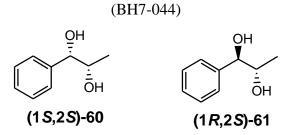


To an aqueous solution of 6.6 mL of Pd/Au (3:1)-1 (2.9 μ mol of Pd/0.96 μ mol of Au-0.10 μ mol of 1) in a Parr pressure reactor, were added 14 mL of deionized H₂O and 0.10 g (0.77 mmol) of 1,2-dihydronaphthalene (57). The apparatus was charged at 30 psi of O₂ and stirred at 25°C for

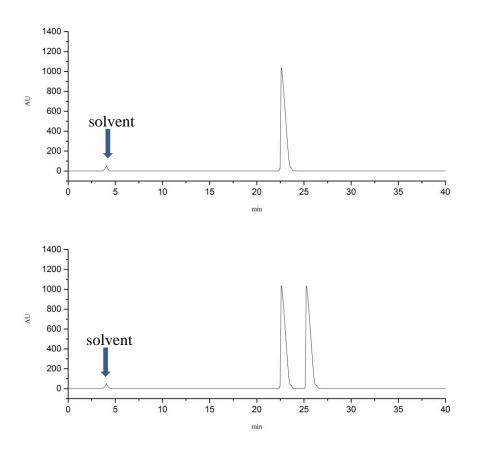
3 days. The solution was then maintained under normal atmosphere, extracted with ethyl acetate three times (20 mL each), and the combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 89 mg (86% yield) of (1*S*,2*R*)-**58** in 99% ee. The optical purity was determined using HPLC-chiral column. $[\alpha]_D^{22} = +35.0$ (c 0.74, CHCl₃). Lit.^[67] +35 (c 0.74, CHCl₃; >98% ee). ¹H NMR δ 7.44–7.41 (m, 1 H), 7.26–7.20 (m, 2 H), 7.13–7.11 (m, 1 H), 4.68 (d, J = 3.6 Hz, 1 H), 4.02–3.98 (m, 1 H), 3.00–2.92 (m, 1 H), 2.82–2.78 (m, 1 H), 2.53–2.45 (bs, 2 H, OH), 2.07–2.00 (m, 1 H), 2.00–1.87 (m, 1 H); ¹³C NMR δ 136.6, 136.4, 130.1, 128.8, 128.4, 126.7, 70.2, 69.8, 27.1, 26.5. MS (ESI, MeOH): *m/z* = 165.1 ([M + H]⁺). The % ee of this compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; t_R = 21.6 min (1*S*,2*R* enantiomer, minor), t_R = 24.2 min (1*R*,2*S* enantiomer, major).



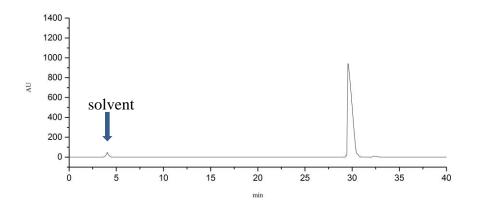
(1S,2S)-1-Phenylpropane-1,2-diol (60) and (1R,2S)-1-phenylpropane-1,2-diol (61)

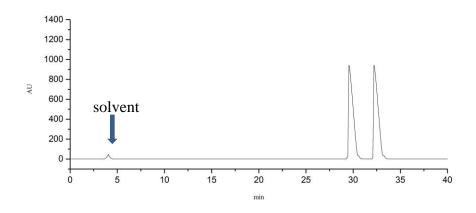


From Pd/Au (3:1)-1 (7.9 µmol of Pd/2.65 µmol of Au-0.29 µmol of 1) and 0.25 g (2.1 mmol) of *trans*-β-methylstyrene (**59**) in 30 mL of deionized water, after stirring at 25°C under 30 psi of O₂. in a Parr pressure reactor for 3 days, 0.281 g (87% yield) of (15,25)-60 (99% ee) and 19 mg (6% yield) of (1R,2S)-61 (97% ee) along with 13 mg of 59 (5% recovery) were obtained after column chromatography. The optical purity was determined using HPLC/chiral column as described above. (1*S*,2*S*)-**60**: $[\alpha]_D^{22} = +51.7$ (c 1.9, CHCl₃). Lit.^[68] +54.3 (c 1.9, CHCl₃). ¹H NMR δ 7.45–7.24 (m, 5 H), 4.37 (d, J = 8 Hz, 1 H), 3.86 (pent, J = 6.5 Hz, 1 H), 2.30–2.15 (bs, 1 H, OH), 1.75–1.60 (bs, 1 H, OH), 1.07 (d, J = 6.5 Hz, 3 H, Me); 13 C NMR δ 142.3, 129.9 (2 C), 128.6, 126.7 (2 C), 79.9, 72.4, 18.1. MS (ESI, MeOH): m/z = 175.1 ([M + Na]⁺). Compound (1R,2S)-61: $[\alpha]_D^{22} = -37.3$ (c 2.52, CHCl₃). Lit.^[68] -29.9 (c 2.52, CHCl₃). ¹H NMR δ 7.36– 7.27 (m, 5 H), 4.68 (d, J = 4.4 Hz, 1 H), 4.03–4.00 (m, 1 H), 2.15–1.80 (bs, 1 H, OH), 1.75–1.50 (bs, 1 H, OH), 1.05 (d, J = 6 Hz, 3 H, Me); 13 C NMR δ 140.4, 128.2 (2 C), 127.9, 126.7 (2 C), 77.5, 71.4, 17.5. MS (ESI, MeOH): m/z = 174.9 ([M + Na]⁺). The % ee (99%) of (1*S*,2*S*)-60 was determined by HPLC using chiral column, Chiralpak AD(-H) column, n-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; $t_R = 22.6 \text{ min } (1S, 2S \text{ enantiomer, major}), t_R = 25.8 \text{ min } (1R, 2R)$ enantiomer, minor).

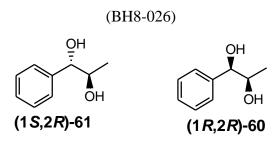


The % ee (97%) of (1*R*,2*S*)-**61** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; $t_R = 29.6$ min (1*R*,2*S* enantiomer, major), $t_R = 32.2$ min (1*S*,2*R* enantiomer, minor).

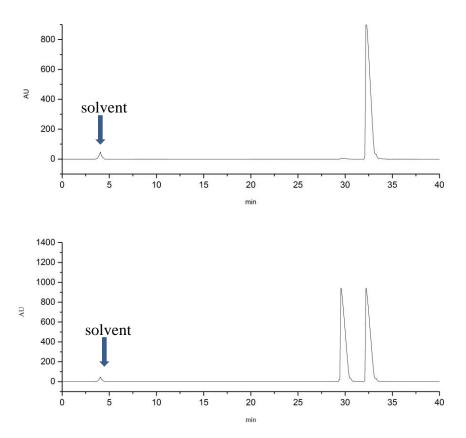




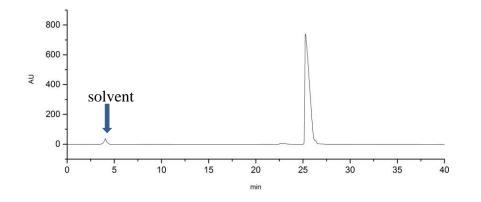
(1S,2R)-1-Phenylpropane-1,2-diol (61) and (1R,2R)-1-phenylpropane-1,2-diol (60)

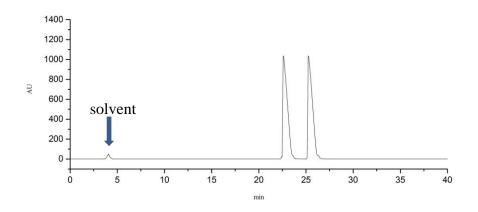


From Pd/Au (3:1)-1 (7.94 μmol of Pd/2.65 μmol of Au-0.29 μmol of 1) and 0.25 g (2.1 mmol) of *cis*-β-methylstyrene (**62**) in 30 mL of deionized water, after stirring at 25°C under 30 psi of O₂ in a Parr pressure reactor for 3 days, 0.291 g (90% yield) of (1*S*,2*R*)-**61** (98% ee) and 26 mg (8% yield) of (1*R*,2*R*)-**60** (98% ee) were obtained after column chromatography. (1*S*,2*R*)-**61**: $[\alpha]_D^{22} = +37.1$ (c 2.52, CHCl₃). Lit.^[68] +36.1 (c 2.52, CHCl₃). ¹H and ¹³C NMR spectra were identical to those of compound (1*R*,2*S*)-**61**. The % ee (98%) of (1*S*,2*R*)-**61A** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; t_R = 29.6 min (1*R*,2*R* enantiomer, minor), t_R = 32.2 min (1*S*,2*R* enantiomer, major).



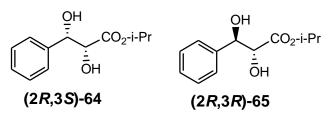
(1R,2R)-60: $[\alpha]_D^{22} = -51.8$ (c 1.9, CHCl₃). Lit.^[68] -51.3 (c 3.5, CHCl₃). ¹H and ¹³C NMR spectra were identical to those of (1S,2S)-60. The % ee (98%) of (1R,2R)-60 was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; t_R = 22.6 min (1S,2S enantiomer, minor), t_R = 25.8 min (1R,2R enantiomer, major).



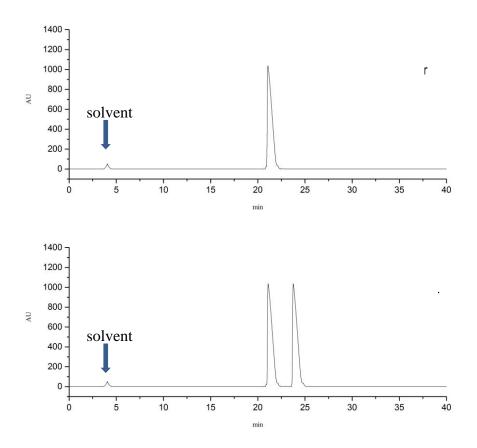


(2R,3S)-Isopropyl 2,3-dihydroxy-3-phenylpropanoate (64) and (2R,3R)-isopropyl

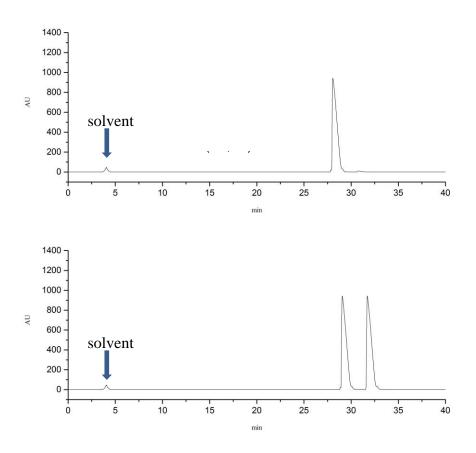
2,3-dihydroxy-3-phenylpropanoate (65) (BH7-116)



From Pd/Au (3:1)-**1** (1.97 μmol of Pd/0.66 μmol of Au-0.073 μmol of **1**) and 0.10 g (0.52 mmol) of *trans*-isoproyl 3-phenylpropenoate (**63**) in 20 mL of deionized water, after stirring at 50°C under 30 psi of O₂ in a Parr pressure reactor for 3 days, 96 mg (82% yield) of (2*R*,3*S*)-**64** (99% ee) and 3.7 mg (3% yield) of (2*R*,3*R*)-**65** (97% ee) were obtained after column chromatography. The optical purity was determined using HPLC/chiral column as described above. (2*R*,3*S*)-**64**: $[\alpha]_D^{22} = +9.7$ (c 0.15, CHCl₃). Lit.^[81] +10.6 (c 1.02, CHCl₃; >99%). ¹H NMR δ 7.40–7.36 (m, 3 H), 7.34–7.32 (m, 2 H), 5.16 (hept, J = 6 Hz, 1 H), 5.01 (d, J = 6 Hz, 1H), 4.36 (d, J = 6 Hz, 1 H), 2.70–2.48 (bs, 2 H, OH), 1.31 (d, J = 6 Hz, 3 H); ¹³C NMR δ 166.8, 144.6, 130.3, 129.1 (2 C), 128.2 (2 C), 86.1, 74.2, 67.9, 22.2. MS (ESI, MeOH): m/z = 225.0 ([M + H]⁺). The % ee (99%) of (2*R*,3*S*)-**64** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; t_R = 21.1 min (2*R*,3*S* enantiomer, major), t_R = 25.8 min (2*S*,3*R* enantiomer, minor).

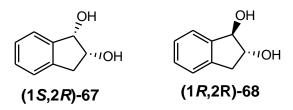


(2R,3R)-**65**: $[\alpha]_D^{22} = +7.3$ (c 0.15, CHCl₃). ¹H NMR δ 7.40–7.35 (m, 3 H), 7.34–7.31 (m, 2 H), 5.14 (hept, J = 6 Hz, 1 H), 5.06 (d, J = 6.8 Hz, 1H), 4.38 (d, J = 5.2 Hz, 1 H), 2.25–2.00 (bs, 2 H, OH), 1.31 (d, J = 6 Hz, 3 H); ¹³C NMR δ 169.4, 145.4, 131.3, 130.0 (2 C), 129.2 (2 C), 87.4, 75.1, 69.1, 23.1. MS (ESI, MeOH): m/z = 225.0 ([M + H]⁺, 100%). The % ee (97%) of (2*R*,3*R*)-**65** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; t_R = 28.1 min (2*R*,3*R* enantiomer, major), t_R = 32.0 min (2*S*,3*S* enantiomer, minor).



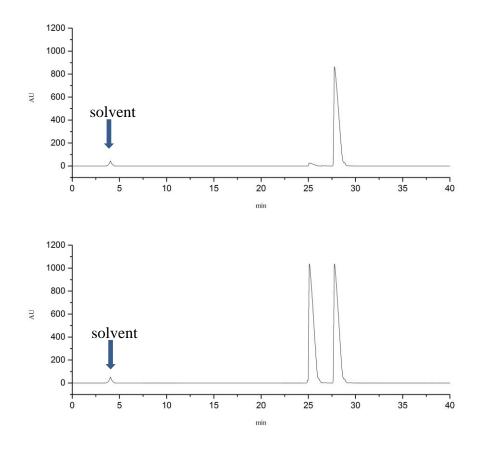
(1S,2R)-2,3-Dihydro-1H-indene-1,2-diol (67) and (1R,2R)-2,3-dihydro-1H-indene-1,2-diol

(68). (BH7-084)

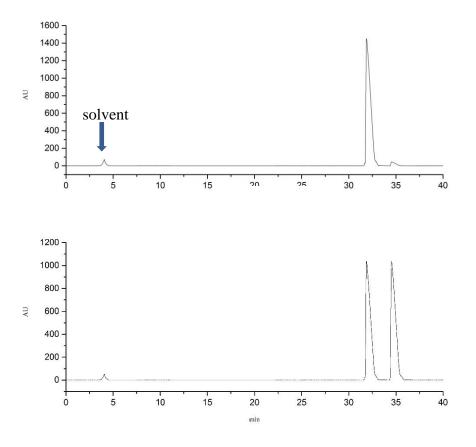


From Pd/Au (3:1)-**1** (19.3 µmol of Pd/6.7 µmol of Au-0.74 µmol of **1**) and 0.30 g (2.59 mmol) of indene (**66**) in 20 mL of deionized water, after stirring at 50°C under 30 psi of O₂ in a Parr pressure reactor for 2 days, 0.261 g (67% yield) of (1*S*,2*R*)-**67** (93% ee) and 44 mg (11% yield) of (1*R*,2*R*)-**68** (94% ee) were obtained after column chromatography. The optical purity was determined using HPLC/chiral column. Compound (1*S*,2*R*)-**67**: $[\alpha]_D^{22} = -46.9$ (c 1.14, CHCl₃). Lit.^[67] -48.0 (c 1.14, CHCl₃). ¹H NMR δ 7.45–7.42 (m, 1 H), 7.31–7.20 (m, 3 H),

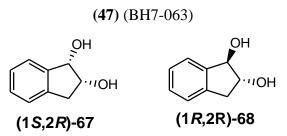
5.03–5.02 (m, 1 H, CH-O), 4.54–4.52 (m, 1 H, CH-O), 3.14 (dd, J = 16, 6 Hz, 1 H), 2.96 (dd, J = 16, 4 Hz, 1 H), 2.50–2.25 (bs, 2 H, OH); ¹³C NMR δ 142.2, 140.2, 129.0, 127.4, 125.6, 125.3, 76.2, 73.7, 38.9. MS (ESI, MeOH): m/z = 173.1 ([M + Na]⁺). Compound (1*R*,2*R*)-67: $[\alpha]_D^{22}$ = -28.8 (c 0.675, EtOH). Lit.^[82] +30.5 (c 0.675, EtOH). ¹H NMR δ 7.40–7.36 (m, 1 H), 7.28–7.18 (m, 3 H), 5.02–5.01 (m, 1 H), 4.40–4.38 (m, 1 H), 3.28 (dd, J = 16, 7.6 Hz, 1 H), 2.83 (dd, J = 16, 8 Hz, 1 H), 2.10–2.09 (m, 1 H, OH), 2.05–2.02 (m, 1 H, OH); ¹³C NMR δ 141.8, 141.3, 130.1, 128.9, 128.3, 126.6, 73.0, 72.4, 37.5 MS (ESI, MeOH): m/z = 151.1 ([M + H]⁺). The % ee (93%) of (1*S*,2*R*)-67 compound was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; t_R = 25.0 min (1*R*,2*S* enantiomer, minor), t_R = 27.8 min (1*S*,2*R* enantiomer, major).



The % ee (94%) of compound (1*R*,2*R*)-**68** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; $t_R = 31.5$ min (1*R*,2*R* enantiomer, major), $t_R = 35$ min (1*S*,2*S* enantiomer, minor).

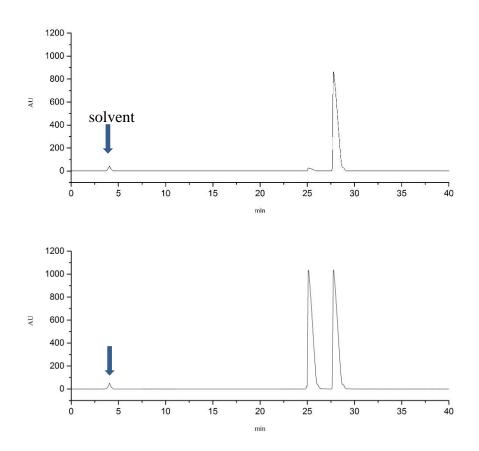


(1S,2R)-2,3-Dihydro-1H-indene-1,2-diol (46) and (1R,2R)-2,3-dihydro-1H-indene-1,2-diol

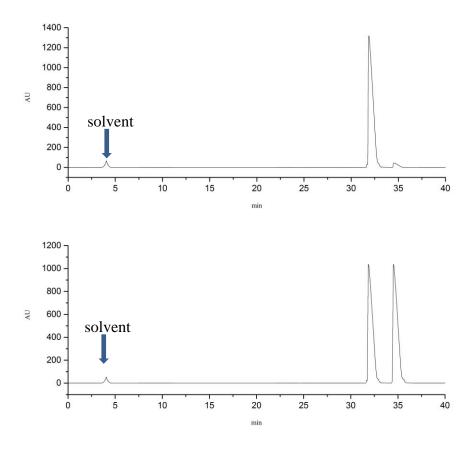


To an aqueous solution of 15 mL of Pd/Au (3:1)-1 (6.5 μ mol of Pd/2.2 μ mol of Au-0.24 μ mol of 1) in a Parr pressure reactor, were added 5 mL of deionized H₂O, 77 mg (0.56 mmol) of K₂CO₃, and 0.20 g (1.75 mmol) of indene (**66**). The apparatus was charged at 30 psi of O₂ and stirred at

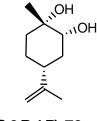
70°C for 5 hours. The solution was then maintained under normal atmosphere, extracted with ethyl acetate three times (20 mL each), and the combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 0.117 g (59% yield based on recovered **66**) of (1*R*,2*R*)-**68** in 91% ee and 10 mg (5% yield based on recovered **66**) of (1*S*,2*R*)-**67** in 89% ee, along with 47 mg (23.5% recovery) of **66**. (1*R*,2*R*)-**68**: $[\alpha]_D^{22} = -27.7$ (c 1.14, CHCl₃). Lit.^[82] +30.5 (c 0.675, EtOH). (1*S*,2*R*)-**67**: $[\alpha]_D^{22} = -44.9$ (c 1.14, CHCl₃). Lit.^[81] -48.0 (c 1.14, CHCl₃). Lit.^[81] -48.0 (c 1.14, CHCl₃). ¹H NMR and ¹³C NMR spectra of **67** and **68** were identical to those described above. No hydroxyindanones were detected. The % ee (89%) of (1*S*,2*R*)-**67** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; t_R = 25.0 min (1*R*,2*S* enantiomer, minor), t_R = 27.8 min (1*S*,2*R* enantiomer, major).



The % ee (91%) of (1*R*,2*R*)-**68** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 85:15, flow rate: 0.5 mL/min; $t_R = 31.5$ min (1*R*,2*R* enantiomer, major), $t_R = 34.6$ min (1*S*,2*S* enantiomer, minor).



(1S,2R,4R)-1-Methyl-4-(prop-1-en-2-yl)cyclohexane-1,2-diol (70) (BH8-030)



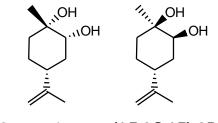
(1S,2R,4R)-70

From Pd/Au (3:1)-1 (2.86 μ mol of Pd/0.99 μ mol of Au-0.11 μ mol of 1) and 52 mg (0.39 mmol) of (*R*)-(+)-limonene (**69**) in 20 mL of deionized water, after stirring at 25°C under 30 psi of O₂ in a Parr pressure reactor for 3 days, 61 mg (92% yield) of (1*S*,2*R*,4*R*)-**70** was obtained as a single

stereoisomer after column chromatography. No other stereoisomers were detected. $[\alpha]_D^{22} = +22.6 \text{ (c } 0.5, \text{ CHCl}_3).$ ¹H NMR δ 4.74 (s, 2 H), 3.92 (s, 1 H), 2.32–2.23 (m, 1 H), 2.18–2.09 (m, 1 H), 2.03–1.94 (m, 1 H), 1.88–1.64 (m, 4 H), 1.73 (s, 3 H), 1.63 (s, 3 H), 1.64–1.58 (bs, 2 H, OH); ¹³C NMR δ 149.3, 109.3, 74.4, 71.7, 37.5, 35.6, 33.9, 29.4, 27.0, 21.1. MS (ESI, MeOH): $m/z = 171.0 ([M + H]^+)$. HRMS-ESI: $m/z [M + Na]^+$ calcd for C₁₀H₁₈NO₂Na⁺: 193.1199; found: 193.1197.

(1S,2R,4R)-1-Methyl-4-(prop-1-en-2-yl)cyclohexane-1,2-diol (70) and

(1*R*,2*S*,4*R*)-1-Methyl-4-(prop-1-en-2-yl)cyclohexane-1,2-diol (85) (BH8-038)

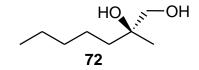


(1*S*,2*R*,4*R*)-70 (1*R*,2*S*,4*R*)-85

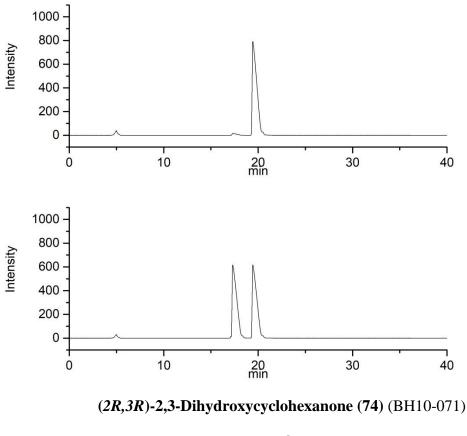
A solution of 0.20 g (1.47 mmol) of (*R*)-(+)-limonene (**69**), 7.5 mg (29 µmol) of OsO₄, and 0.21 g (1.76 mmol) of *N*-methylmorpholine *N*-oxide (NMO) in 2 mL of *t*-BuOH, 7 mL of acetone and 2.5 mL of water was stirred 0°C for 30 min and 25°C for 14 h. The reaction mixture was filtered through Celite and concentrated on a rotary evaporator to remove acetone and t-BuOH, diluted with water (20 mL), and extracted three times with dichloromethane (20 mL each). The combined extract was washed with brine, dried (anhydrous Na₂SO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ether as eluent to give 27 mg (11% yield) of (1*S*,2*R*,4*R*)-**70** and 0.118 g (47% yield) of (1*R*,2*S*,4*R*)-**85**^[83] along with 46 mg (23% recovery) of (*R*)-limonene. (1*S*,2*R*,4*R*)-**70**: $[\alpha]_D^{22} = +22.6$ (c 0.5, CHCl₃). ¹H and ¹³C NMR spectra were identical to those of compound **70** obtained from the catalytic asymmetric dihydroxylation reaction described above. (1*R*,2*S*,4*R*)-**85**: $[\alpha]_D^{22} = +41.8$ (c 1.0, MeOH); Lit.^[81]

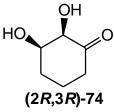
OH), 1.93–1.78 (m, 3 H), 1.74–1.67 (m, 1 H), 1.71 (s, 3 H, Me), 1.55–1.50 (m, 1 H), 1.42 (s, 1 H, OH), 1.34–1.25 (m, 2 H), 1.25 (s, 3 H, Me); ¹³C NMR δ 148.9, 108.7, 73.5, 71.2, 37.2, 33.6, 33.3, 26.1, 25.7, 20.6.

(S)-2-Methylheptane-1,2-diol (72) (BH10-143)



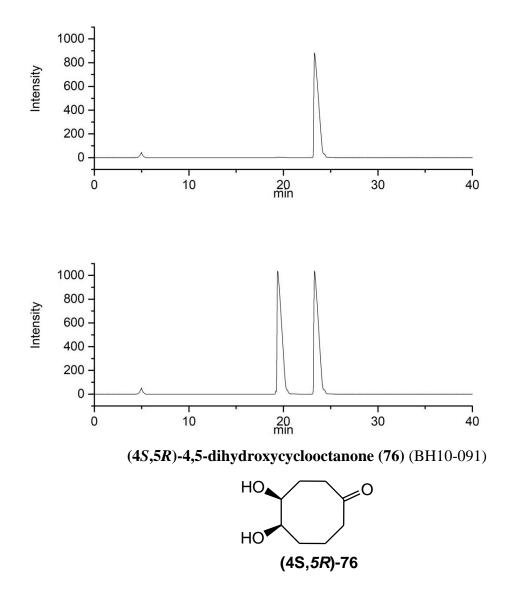
To an aqueous solution of 2 mL of Pd/Au (3:1)-2 (10 µmol of Pd, 3.3 µmol of Au, and 0.16 µmol of **26**) in a Parr pressure reactor, were added 18 mL of deionized H₂O and 30 mg (0.27 mmol) of 2-methylhept-1-ene (**71**). The apparatus was maintained under 30 psi of O₂, stirred at 50 °C for 3 days, vented to normal atmosphere, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 28.6 mg (73.4% yield) of (**S**)-**72** in 97% ee. $[\alpha]_D^{22} = -2.28$ (c 1.2, CHCl₃), Lit.^[80] +2.3 (c 1.21, CHCl₃; 97% ee, *R*-configuration); ¹H NMR δ 3.46 (dd, *J* = 11.2, 3.6 Hz, 1H), 1.48-1.45 (m, 2H), 1.35-1.30 (m, 6H), 1.16 (s, 3H), 0.89 (t, *J* = 7.2 Hz, 3H); ¹³C NMR δ 73.2, 69.8, 38.7, 32.5, 23.5, 23.0, 22.3, 14.2; MS (ESI, MeOH): m/z = 147.3 ([M + H]⁺). The % *ee* (97%) of (*S*)-**72** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 85:15, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 17.8 min (*R* enantiomer, minor), t_R = 19.9 min (*S* enantiomer, major).



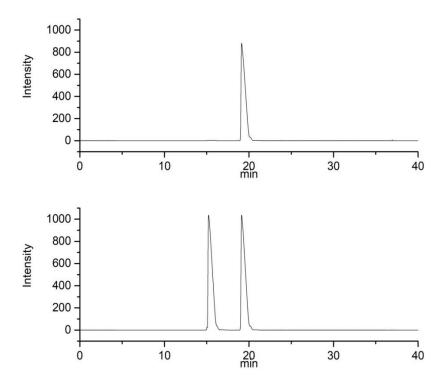


To an aqueous solution of 2 mL of Pd/Au (3:1)-**26** (0.84 µmol of Pd, 0.27 µmol of Au, and 0.03 µmol of **26**) in a Parr pressure reactor were added 18 mL of deionized H₂O and 21 mg (0.21 mmol) of cyclohex-2-enone (**73**). The apparatus was maintained under 30 psi of O₂, stirred at 50 °C for 3 days, vented to normal atmosphere, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 24 mg (86% yield) of (2*R*,3*R*)-**74** in 99% ee. $[\alpha]_D^{22} = -13.8$ (c 0.6, CHCl₃); Lit. ^[89]+1.48 (neat, 1 = 2 dm; for 2*S*,3*S* configuration). ¹H NMR δ 4.41(d, *J* =

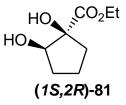
3.2, 1H), 4.21 (dd, J = 6, 2.8 Hz, 1H), 3.92 (b, 1H), 2.59–2.52 (m, 2H), 2.36 (td, J = 14.8, 1.6 Hz, 1H), 2.17–2.04 (m, 2H), 1.95–1.82 (m, 2H); ¹³C NMR δ 210.1, 82.6, 72.3, 38.0, 31.0, 18.9; MS (ESI, MeOH): m/z = 267.1 ([M + Na]⁺). The % *ee* (99%) of (2*R*,3*R*)-**74** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 85:15, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 19.6 min (2*S*,3*S* enantiomer, minor), t_R = 23.8 min (2*R*,3*R* enantiomer, major)



To an aqueous solution of 2 mL of Pd/Au (3:1)-**26** (0.75 µmol of Pd, 0.25 µmol of Au, and 0.014 µmol of **26**) in a Parr pressure reactor, were added 18 mL of deionized H₂O and 25 mg (0.2 mmol) of (Z)-cyclooct-4-enone (**56**). The apparatus was maintained under 30 psi of O₂, stirred at 50 °C for 3 days, vented to normal atmosphere, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 26.8 mg (85% yield) of (4*S*,5*R*)-**76** in 99% ee. $[\alpha]_D^{22} = -16.4$ (c 0.5, CHCl₃); ¹H NMR δ 3.78 (d, *J* = 7.8, 1H), 3.39–3.36 (m, 1H), 2.51–2.29 (m, 4H), 2.02–1.97 (b, 1H), 1.85–1.52 (m, 5H), 1.40–1.25 (m, 2H), 1.35–1.25 (m, 2H); ¹³C NMR δ 211.6, 79.3, 78.3, 42.2, 33.5, 32.2, 26.5, 17.5; MS (ESI, MeOH): m/z = 181.1 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₈H₁₅O₃⁺: 159.1021, found: 159.1023. The % *ee* (99%) of (*4S*,5*R*)-**21** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 85:15, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 15.7 min (*4R*,5*S* enantiomer, minor), t_R = 19.4 min (*4S*,5*R* enantiomer, major).

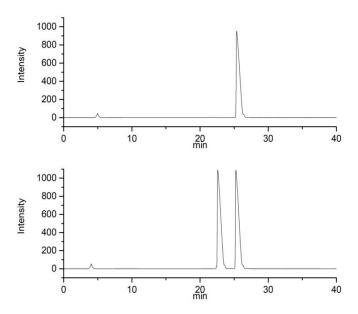


(1S,2R)-Ethyl 1,2-dihydroxycyclopentanecarboxylate (81). (BH10-062)

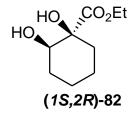


To an aqueous solution of 2 mL of Pd/Au (3:1)-**26** (0.84 µmol of Pd/0.27 µmol of Au-0.0295 µmol of **26**) in a Parr pressure reactor were added 18 mL of deionized H₂O and 0.03 g (0.214 mmol) of ethyl cyclopent-1-enecarboxylate (**77**). The apparatus was maintained under 30 psi of O₂, stirred at 50 °C for 3 days, vented to normal atmosphere, and extracted three times with ethyl acetate (20 mL each). The combined extracts were washed with water, and brine, dried (MgSO₄), and concentrated to dryness. The crude product was column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as eluents to give 28.3 mg (77% yield) of (1*S*,2*R*)-**81** in 99% ee. $[\alpha]_D = -36.8 (1.0, CHCl_3); {}^{1}H NMR \delta 4.26 (q,$ *J*= 7.2 Hz, 2H), 4.04 (t,*J*= 8.8 Hz, 1H), 3.12 (b, 1H), 2.64 (b, 1H), 2.20 - 2.16 (m, 1H), 2.14 - 2.03 (m, 1H), 1.93-1.80 (m, 2H), 1.73-1.58 (m, 2H), 1.30 (t,*J* $= 6.8 Hz, 3H); {}^{1}C NMR$

δ 176.4, 79.1, 75.9, 62.0, 33.4, 27.5, 25.6, 14.0; MS (ESI, MeOH): m/z = 197.3 ([M + Na]⁺) HRMS-ESI: m/z [M + H]⁺ calcd for C₈H₁₅O₄⁺: 175.0970, found: 175.0962. The % *ee* (99%) of (*1S*,2*R*)-**81** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 85:15, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 22.6 min (*1R*,2*S* enantiomer, minor), t_R = 25.3 min (*1S*,2*R* enantiomer, major).

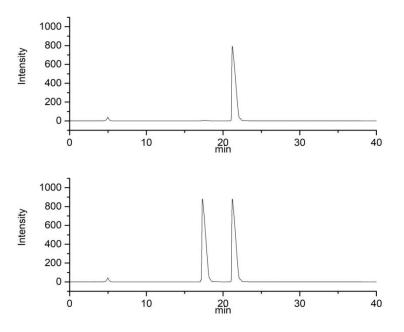


(1S,2R)-Ethyl 1,2-dihydroxycyclohexanecarboxylate (82). (BH9-069)

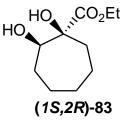


To an aqueous solution of 2 mL of Pd/Au (3:1)-**26** (2.44 μ mol of Pd, 0.81 μ mol of Au, and 0.09 μ mol of **26**) in a Parr pressure reactor were added 18 mL of deionized H₂O and 0.1 g (0.65 mmol) of ethyl cyclohex-1-enecarboxylate (**78**). The apparatus was maintained under 30 psi of O₂, stirred at 50 °C for 3 days, vented to normal atmosphere, and extracted with ethyl acetate three times (20 mL each). The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and

ethyl acetate as eluents to give 102 mg (83% yield) of (1*S*,2*R*)-**82** in 99% ee. $[\alpha]_{D}^{22} = -29.6$ (c 1.0, CHCl₃); ¹H NMR δ 4.29 (q, J = 6.8 Hz, 2H), 3.92 (t, J = 9.6 Hz, 1H), 3.48 (b, 1H), 2.22 ^{13}C 1.30 3H); (d, 1H), 1.93-1.42 8H), (t, J6.8 Hz, NMR (m, = δ 177.9, 79.1, 73.8, 60.8, 33.2, 28.5, 22.4, 20.3, 14.4; MS (ESI, MeOH): m/z = 211.2 ([M + Na]⁺). HRMS-ESI: m/z $[M + H]^+$ calcd for C₉H₁₇O₄⁺: 189.1127, found: 189.1129. The % *ee* (99%) of (1S,2R)-82 was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 85:15, flow rate 0.5 mL/min, detected at 220 nm wavelength; $t_R = 17.6$ min (1R,2S enantiomer, minor), $t_R = 22.3 \text{ min} (1S,2R \text{ enantiomer, major})$.

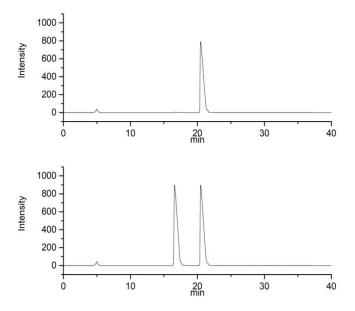


(1S,2R)-Ethyl 1,2-dihydroxycycloheptanecarboxylate (83). (BH9-025)

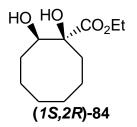


To an aqueous solution of 2 mL of Pd/Au (3:1)-26 (1.12 μ mol of Pd, 0.37 μ mol of Au, and 0.041 μ mol of 26) in a Parr pressure reactor, were added 18 mL of deionized H₂O and 50 mg (0.30

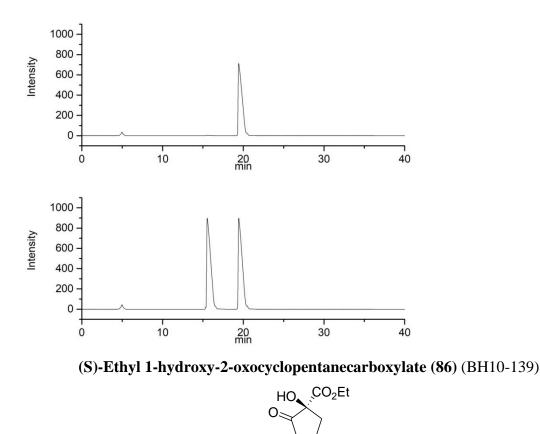
mmol) of ethyl cyclohept-1-enecarboxylate (**79**). The apparatus was maintained at 30 psi of O₂, stirred at 50 °C for 3 days, vented to normal atmosphere, and extracted with ethyl acetate three times (20 mL each). The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 51 mg (85% yield) of (1*S*,2*R*)-**83** in 99% ee. $[\alpha]_D^{22} = -71.6$ (c 1.0, CHCl₃); ¹H NMR δ 4.27 (q, *J* = 5.2 Hz, 1 H), 3.72 (t, *J* = 4.8 Hz, 1 H), 3.48 (b, 1 H), 2.24 (d, *J* = 9.6 Hz, 1H), 1.85 – 1.39 (m, 9 H,), 1.30 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 177.2, 78.8, 76.4, 62.3, 35.2, 32.2, 27.6, 23.2, 20.8, 14.4. MS (ESI, MeOH): m/z = 225.3 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₀H₁₈O₄⁺: 203.1283 found: 203.1279; The % *ee* (99%) of (*1S*,2*R*)-**83** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 85:15, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 16.2 min (*1R*,2*S* enantiomer, minor), t_R = 20.6 min (*1S*,2*R* enantiomer, major).



(1S,2R)-Ethyl 1,2-dihydroxycyclooctanecarboxylate (84). (BH10-083)



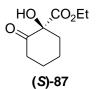
To an aqueous solution of 2 mL of Pd/Au (3:1)-**26** (1.53 µmol of Pd, 0.51 µmol of Au, and 0.056 µmol of **26**) in a Parr pressure reactor, were added 18 mL of deionized H₂O and 74 mg (0.41 mmol) of ethyl cyclooct-1-enecarboxylate (**80**). The apparatus was maintained under 30 psi of O₂, stirred at 50 °C for 3 days, vented to normal atmosphere, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 72.3 mg (81% yield) of (1*S*,2*R*)-**84** in 99% ee. $[\alpha]_D^{22} = -16.2$ (c 1.0, CHCl₃); ¹H NMR δ 4.29 (q, *J* = 6.8 Hz, 2H), 3.12 (t, J = 9.6 Hz, 1H), 3.51 (b, 1H), 2.25 (d, 1H), 2.08 (m, 1H), 1.93–1.42 (m, 12H), 1.30 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 176.9, 79.4, 73.8, 62.2, 33.5, 31.2, 27.4, 26.5, 24.7, 21.2, 14.3; MS (ESI, MeOH): m/z = 239.4 ([M + Na]⁺); HRMS-ESI: m/z [M + H]⁺ calcd for C₁₁H₂₁O₄⁺: 217.1440, found: 217.1429; The % *ee* (99%) of (*1S*,2*R*)-**84** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 85:15, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 15.3 min (*1R*,2*S* enantiomer, minor), t_R = 20.0 min (*1S*,2*R* enantiomer, major).



(S)-86

To a solution of 17.4 mg (0.1 mmol) (*IS*,*2R*)-ethyl 1,2-dihydroxycyclopentanecarboxylate in 1 mL DMSO, 56 mg (0.2 mmol) of IBX was added and stirred at room temperature for 10 hours. The resulting solution was diluted with 2 mL water, extracted with 3 x 10 mL ethyl acetate, washed with brine, dried over Na₂SO₄, concentrated under vacuum and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 12.1 mg (70% yield) of the titled compound. $[\alpha]_D$ -14.6 (0.2, CHCl₃); Lit.^[84] $[\alpha]_D$ +12 (1.0, CHCl₃ for R configuration); ¹H NMR δ 4.19–4.14 (q, *J* = 7.2 Hz, 2H), 3.84 (b, 1H), 2.44–2.34 (m, 3H), 2.09–1.98 (m, 3H), 1.24 (t, J = 6.8 Hz, 3H); ¹³C NMR δ 213.5, 171.5, 79.9, 62.4, 35.8, 34.8, 18.8, 13.8. MS (ESI, MeOH): 173.1([M + H]⁺).

Ethyl 1-hydroxy-2-oxocyclohexanecarboxylate (87) (BH10-137)



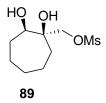
To a solution of 18.8 mg (0.1 mmol) (1S,2R)-ethyl 1,2-dihydroxycyclopentanecarboxylate in 1 mL DMSO, 56 mg (0.2 mmol) of IBX was added and stired at room temperature for 10 hours. The resulting solution was diluted with 2 mL water, extracted with 3 x 10 mL ethyl acetate, washed with brine, dried over Na₂SO₄, concentrated under vacuum and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 11.6 mg (61% yield) of the titled compound. $[\alpha]_D$ -126.8 (0.1, CHCl₃); Lit.^[84] $[\alpha]_D$ +134 (1.0, CHCl₃ for R configuration); ¹H NMR δ 4.33 (s, 1H), 4.21 (q, J = 7.2 Hz, 2H), 2.68–2.49 (m, 3H), 2.06–1.97(m, 1H), 1.86–1.76 (m, 2H), 1.73–1.59 (m, 2H), 1.27 (t, J = 6.8 Hz, 3H); ¹³C NMR δ 207.4, 170.1, 80.7, 62.1, 39.0, 37.7, 27.1, 22.0, 14.1. MS (ESI, MeOH): 187.1([M + H]⁺).

(1R,2R)-1-(Hydroxymethyl)cycloheptane-1,2-diol (88) (BH9-044)



To a cold (0 °C) solution of 18 mg (0.09 mmol) of (1*S*,2*R*)-**83** in 1 mL of THF under argon was added 5.1 mg (0.13 mmol) of LiAlH₄. The mixture was warmed to room temperature, stirred for 2 hours, and added 2 g of Na₂SO₄•10 H₂O and one drop of CH₃COOH. The resulting solution was filtered through Celite, washed with 20 mL of methylene chloride, and concentrated under vacuum to give 14.1 mg of compound **88**. This material was used in the subsequent step without further purification. $[\alpha]_D^{22} = -22.3$ (c 0.2, CHCl₃); ¹H NMR δ 3.96 (t, J = 9.6 Hz, 1H), 3.58 (m, 2H), 2.88 (b, 1H), 2.62 (b, 1H), 1.91–1.46 (m, 10H); ¹³C NMR δ 81.4, 71.9, 65.8, 32.6, 29.3, 28.5, 25.9, 22.8; MS (ESI, MeOH): m/z = 161.2 ([M + H]⁺).

[(1*R*,2*R*)-1,2-Dihydroxycycloheptyl]methyl methanesulfonate (89) (BH9-045)



To a solution of 12 mg (0.075 mmol) of (1*R*,2*R*)-**88** in 4 mL of distilled methylene chloride under argon were added 7.5 mg (0.075 mmol) of Et₃N and 8.5 mg (0.075 mmol) of methanesulfonyl chloride (MsCl). It was stirred for 5 hours, diluted with 5 mL of H₂O, and extracted with 10 mL of CH₂Cl₂ twice. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a mixture of diethyl ether and hexane in a ratio of 6:1 as an eluent to give 11 mg (60% yield) of (*1R*,2*R*)-**89**. $[\alpha]_D^{22} =$ -80.7 (c 0.1, CHCl₃); ¹H NMR δ 4.06 (t, J = 9.6 Hz, 1H), 3.92 (m, 2H), 2.91 (s, 3H), 2.45 (b, 1H), 2.03 (d, 1H), 1.86–1.46 (m, 10H); ¹³C NMR δ 78.9, 76.4, 69.2, 37.9, 35.5, 31.4, 27.6, 22.9, 20.6; MS (ESI, MeOH): m/z = 261.1 ([M + Na]⁺).

(1S,2R)-1-Methylcycloheptane-1,2-diol (90) (BH9-051)



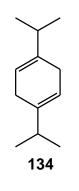
To a cold (0 °C) solution of 9.8 mg (0.041 mmol) of (*1R*,2*R*)-**89** in 1 mL of distilled THF under argon was added 1.56 mg (0.041 mmol) of LiAlH₄. The mixture was warmed to room temperature and stirred for 6 hours, diluted with 1 mL of water and 0.5 mL of HCl, and extracted twice with 10 mL each of ethyl acetate. The combined extracts were washed with brine, dried (MgSO₄), and concentrated to give (*1S*,2*R*)-**90**. This material was used in the subsequent step without further purification. ¹H NMR δ 3.91 (t, *J* = 9.2 Hz, 1H), 3.39 (b, 1H), 2.23 (d, 1H), 1.88–1.42 (m, 10H), 1.30 (s, 3H); ¹³C NMR δ 80.6, 73.6, 32.5, 28.5, 27.0, 24.7, 22.6, 20.5; MS (ESI, MeOH): m/z = 183.4 ([M + K]⁺).

(S)-2-Hydroxy-2-methylcycloheptanone (91) (BH9-053)



To a solution of 14 mg (0.048 mmol) of 2-iodoxybenzoic acid (IBX) in 1 mL of DMSO under argon was added 6.3 mg (0.044 mmol) of (*1S*,2*R*)-**90**. The solution was stirred for 8 hours, dilute with 20 mL of CH₂Cl₂, and washed three times with 5 mL each of H₂O. The combined organic layer was dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an eluent to give 5.2 mg (83% yield) of (*S*)-**91**. $[\alpha]_D^{22} = -71.2$ (c = 0.24, CHCl₃); Lit.^[85] -71 (c 0.24, CHCl₃; *S*-configuration); ¹H NMR δ 3.42 (b, 1 H), 2.73 – 2.67 (m, 1 H), 2.55–2.48 (m,1 H), 2.16–2.10 (m, 1 H), 1.99 – 1.60 (m, 6 H,), 1.23 (s, 3 H); ¹³C NMR δ 216.3, 89.2, 36.9, 33.5, 31.4, 27.2, 25.9, 21.3. MS (ESI, MeOH): m/z = 165.3 ([M + Na]⁺).

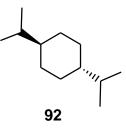
2.6.5 Procedures and Analysis Results for Oxidation of Cycloalkanes.



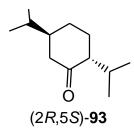
1,4-Diisopropyl-1,4-cyclohexadiene (134) (BH4-110)

To a cold (-78°C) solution of 40 mL of liquid NH₃ under argon was added 0.86 g (0.12 mol) of lithium wire, and the resulting blue solution was stirred at -35°C for 1 h. To it, a solution of 4.0 g (24.7 mmol) of 1,4-diisopropylbenzene in 5.4 g of *t*-BuOH was added, and the solution was stirred for 6 h at -35°C under argon. The reaction solution was diluted with aqueous NH₄Cl (10 mL), extracted twice with diethyl ether, and the combined extract was washed with water, brine, and dried (MgSO₄). The solvents were removed by distillation under normal pressure, and residue was column chromatographed on silica gel using hexane as eluent to give 2.81 g (70% yield) of 1,4-diisopropyl-1,4-cyclohexadiene (**134**) along with 0.96 g (24% recovery) of 1,4-diisopropylbenzene. ¹H NMR δ 5.46 (s, 2 H, =CH), 2.64 (s, 4 H, CH₂), 2.19 (hept, J = 7 Hz, 2 H), 1.01 (d, J = 7 Hz, 12 H); ¹³C NMR δ 140.9, 116.4, 34.7, 27.8, 21.5. MS (ESI, MeOH): m/z = 165.0 ([M + H]⁺).

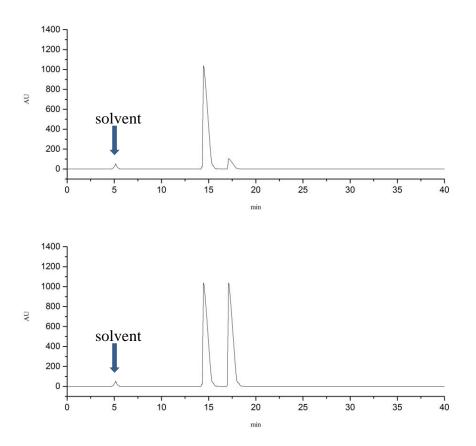




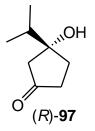
A mixture of 2.0 g (12.2 mmol) of 1,4-diisopropyl-1,4-cyclohexadiene and 0.14 g of 10% Pd/C in 30 mL of ethyl acetate in a bottle was shaken under 30 psi of H₂ on a Parr hydrogenator for 16 h and then maintained under normal pressure. The mixture was filtered through Celite, and the filtrate was concentrated by distillation under normal pressure and column chromatographed on silica gel using pentane as solvent to give 1.3 g (65% yield) of pure **92** along with 0.50 g of **92** and **134** (2:1). Compound **92**: ¹H NMR δ 1.73–1.68 (m, 2 H), 1.59–1.05 (m, 7 H), 0.95–0.90 (m, 3 H), 0.84 (d, J = 7.2 Hz, 12 H); ¹³C NMR δ 42.6, 30.2, 26.7, 20.1. MS (ESI, MeOH): *m/z* = 207.0 ([M + K]⁺).



To a solution of 2 mL of Cu/Au (3:1)-1 (4.46 µmol of Cu/1.49 µmol of Au-0.16 µmol of 1) were added 3 mL of acetonitrile, 0.10 g (0.60 mmol) of **92** and 3 mL of 30% H₂O₂, and the resulting solution was stirred at 50°C for 7 days. The solution was cooled to 25°C and extracted three times with diethyl ether (15 mL each). The combined extracts were washed with water and then brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as eluents to give 94 mg (87% yield) of (2*R*,5*S*)-**93** in 81% ee along with 7 mg (7% recovery) of **92**. The optical purity was determined using HPLC/chiral column. Compound (2*R*,5*S*)-**93**: $[\alpha]_D^{22} = +12.1$ (c 1.5, CHCl₃). Lit.^[101] +14 (c 1.51, CHCl₃). ¹H NMR δ 2.38 (ddd, J = 12, 3.4, 2.2 Hz, 1 H), 2.13–2.05 (m, 1 H), 1.83–1.52 (m, 3 H), 1.43–0.85 (m, 5 H), 1.01 (d, J = 7 Hz, 6 H), 0.84 (d, J = 7 Hz, 6 H); ¹³C NMR δ 213.0, 56.2, 46.5, 32.6, 28.8, 27.9, 25.9, 21.2, 19.6 (2 C), 18.6 (2 C). MS (ESI, MeOH): *m/z* = 205.1 ([M + Na]⁺; 100%), 139.0. The % ee (81%) of (1*R*,5*S*)-**93** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, detected at 220 nm wavelength; t_R = 14.5 min (2*R*,5*S* enantiomer, major), t_R = 17.2 min (2*S*,5*R* enantiomer, minor).

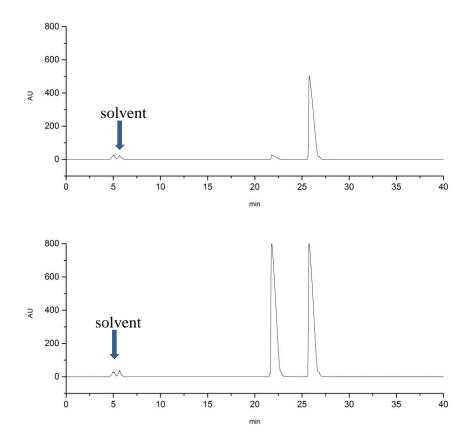


(*R*)-3-Hydroxy-3-isopropylcyclopentanone (97). (BH7-091)

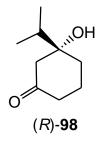


From 90 mg (0.70 mmol) of 1-isopropylcyclopentanol (**94**), 2.4 mL of Cu/Au (3:1)-**1** (5.27 μmol of Cu/1.75 μmol of Au-0.19 μmol of **1**), 2 mL of H₂O₂, and 1 mL of CH₃CN stirring at 50°C for 7 days, 91 mg (91% yield) of (*R*)-**97** in 92% ee. $[\alpha]_D^{22} = +13.7$ (c 0.20, CHCl₃). ¹H NMR δ 2.41–2.30 (m, 2 H, C2 Hs), 2.30–2.17 (m, 1 H, C5 H), 2.07–1.80 (m, 4 H, C5 H, C4 Hs, & C<u>H</u>Me₂), 1.72–1.60 (m, 2 H, C4 H & OH), 1.02 (d, J = 7 Hz, 6 H). The proton NMR assignments were derived from 2D COSY spectrum. ¹³C NMR δ 205.9, 73.3, 53.6, 42.4, 32.5, 22.0, 16.7. MS (ESI, MeOH): m/z = 143.0 ([M + H]⁺), HRMS-Hexane Atmospheric

Pressure Chemical Ionization (HAPCI): $m/z [M]^+$ calcd for C₈H₁₄O₂: 142.0994; found: 142.0049. The % ee (92%) of (*R*)-**97** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, detected at 220 nm wavelength; t_R = 23.5 min (*S* enantiomer, minor), t_R = 27.5 min (*R* enantiomer, major).

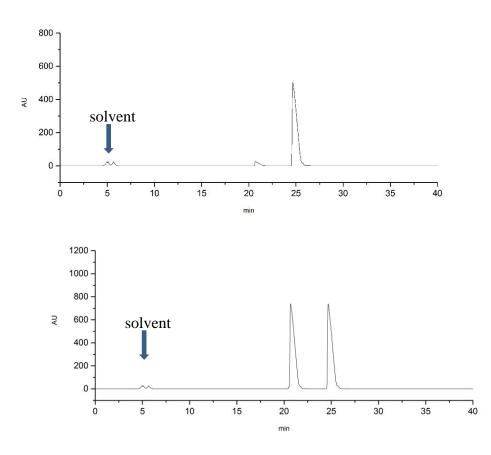


(*R*)-3-Hydroxy-3-isopropylcyclohexanone (98) (BH7-098)



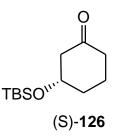
From 0.10 g (0.70 mmol) of 1-isopropylcyclohexanol (95), 2.4 mL of Cu/Au (3:1)-1 (5.27 μ mol of Cu/1.75 μ mol of Au-0.19 μ mol of 1), 2 mL of H₂O₂, and 1 mL of CH₃CN stirring at 50°C for

7 days, 0.107 g (98% yield) of (*R*)-**98** in 91.4% ee. $[\alpha]_D^{22} = +6.1$ (c 0.20, CHCl₃). ¹H NMR δ 2.45–2.40 (m, 2 H), 2.15–2.10 (m, 2 H), 1.96–1.88 (m, 2 H), 1.77–1.60 (m, 4 H), 0.88 (d, J = 6.8 Hz, 6 H). The proton NMR assignments were derived from 2D COSY spectrum. ¹³C NMR δ 205.1, 73.2, 54.2, 42.4, 34.3, 29.3, 23.7, 17.5. MS (ESI, MeOH): m/z = 157.3 ([M + H]⁺). HRMS-Hexane Atmospheric Pressure Chemical Ionization (HAPCI): m/z [M+H]⁺ calcd for C₉H₁₇O₂: 157.1229; found: 157.1221. The % ee (91.4%) of (*R*)-**56** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, detected at 220 nm wavelength; t_R = 20.8 min (*S* enantiomer, minor), t_R = 24.7 min (*R* enantiomer, major).



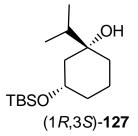
Alternate synthetic route for (R)-98 to determine the absolutely configuration

(S)-3-(t-Butyldimethylsilyloxy)cyclohexanone (126) (BH8-034)



To a solution of 40 mg (0.35 mmol) of (*S*)-3-hydroxycyclohexanone (**38**), 4 mg (0.036 mmol) of 4-dimethylaminopyridine, and 0.10 g (1.0 mmol) of trimethylamine in 3 mL of dichloromethane at 0°C under argon, was added 77 mg (0.51 mmol) of *t*-butyldimethylsilyl chloride. The solution was stirred for 4 h, diluted with 10 mL of water, and extracted three times with ethyl acetate (10 mL each). The combined extract was washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel to give 28 mg (35% yield) of (*S*)-3-(*t*-butyldimethylsilyloxy)cyclohexanone (**126**) and 17 mg (49% yield) of 2-cyclohexenone along with 4 mg (10% recovery) of starting 3-hydroxyketone **38**. ¹H NMR δ 4.17–4.12 (m, 1 H), 2.62 (dd, J = 14, 4 Hz, 1 H), 2.36 (dd, J = 14, 7.5 Hz, 1 H), 2.28 (t, J = 6.5 Hz, 2 H), 2.15–2.00 (m, 2 H), 1.82–1.65 (m, 2 H), 1.00 (s, 9 H), 0.38 (s, 6 H).

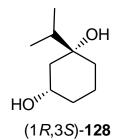
(1R,3S)-3-(t-Butyldimethylsilyloxy)-1-isopropylcyclohexanone (127) (BH10-040)



To a mixture of 1.7 mg (13 μ mol) of anhydrous ZnCl₂ in 2 mL of dry THF under argon at 25°C, was added a solution of 0.18 mmol of isopropylmagnesium bromide in 1 mL of THF, and the solution was stirred for 1 h and then cooled to 0°C. To it, was added a solution of 28 mg (0.12 mmol) of (*S*)-**126** in 0.5 mL of THF via cannula under argon and the resulting solution was stirred at 0°C for 4 h. The solution was diluted with aqueous NH₄Cl and water (10 mL),

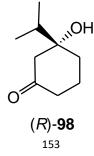
extracted three times with ethyl acetate (20 mL each), and the combined extract was washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluent to give 23 mg (70% yield) of (1*R*,3*S*)-3-(*t*-butyldimethylsilyloxy)-1-isopropylcyclohexanone (**127**). No other stereoisomer was found. ¹H NMR δ 4.37 (m, 1 H), 1.96–1.50 (m, 8 H), 1.32–1.24 (m, 2 H), 0.97 (s, 9 H), 0.91 (d, J = 7 Hz, 6 H), 0.36 (s, 6 H); ¹³C NMR δ 73.3, 66.2, 37.5, 35.6, 33.1, 26.2, 23.9 (3 C), 20.9, 18.6, 16.9 (2 C), -1.15 (2 C).

(1R,3S)-1-Isopropyl-1,3-cyclohexanediol (128) (BH8-046)

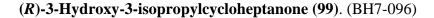


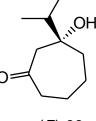
A solution of 14 mg (52 μ mol) of the above (1*R*,3*S*)-**127** and 0.10 mL (0.10 mmol) of *n*-Bu₄NF (1 M in THF) in 0.5 mL of THF was stirred under argon at 0°C for 1 h and then at 25°C for 1.5 h. The solution was diluted with water (10 mL), and extracted three times with diethyl ether (10 mL each). The combined extract was washed with brine, dried (MgSO₄), and concentrated under vacuum to give 8.7 mg of (1*R*,3*S*)-1-isopropyl-1,3-cyclohexanediol, which was used immediately in the following reaction without further purification.

(R)-3-Hydroxy-3-isopropylcyclohexanone (98) (BH8-046)



To a solution of **128** in 0.5 mL of DMSO was added 16 mg (57 µmol) of IBX under argon at 25°C. The resulting mixture was stirred for 14 h, filtered to remove white solid, and rinsed with 20 mL of diethyl ether. The ether filtrate was washed three times with water (10 mL each), dried (MgSO₄), concentrated, and column chromatographed on silica gel using a mixture of hexane and ethyl acetate (1:2) as eluent to give 7.6 mg (95% yield) of (*R*)-3-hydroxy-3-isopropylcyclohexanone (**98**). $[\alpha]_D^{22} = +6.4$ (c 0.20, CHCl₃). ¹H and ¹³C NMR spectra were identical those of (*R*)-**98** synthesized from the Pd/Au-**1** catalytic oxidation reaction.

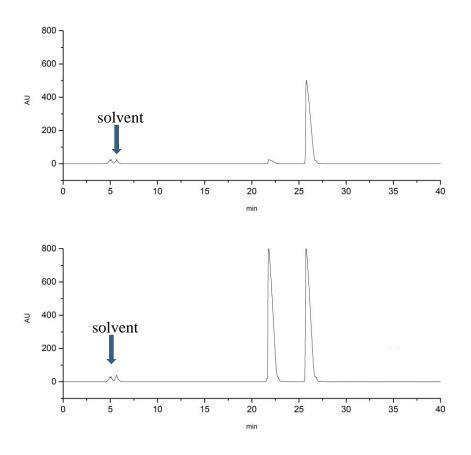




(R)-99

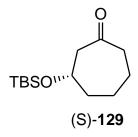
From 42 mg (0.27 mmol) of 1-isopropylcycloheptanol (**96**), 0.92 mL of Cu/Au (3:1)-**1** (2.0 µmol of Cu/0.65 µmol of Au-0.074 µmol of **1**), 1 mL of H₂O₂, and 1 mL of CH₃CN stirring at 50°C for 7 days, 41 mg (89% yield) of (*R*)-**99** in 93% ee. $[\alpha]_D^{22} = +23.8$ (c 0.20, CHCl₃). ¹H NMR δ 2.35 (d, J = 2 Hz, 2 H, C2 Hs), 2.16–2.18 (m, 2 H, C7 Hs), 1.97–1.93 (m, 2 H, C6 Hs), 1.90–1.83 (m, 1 H, C<u>H</u>Me₂), 1.75–1.70 (m, 1 H, C4 H), 1.67–1.58 (m, 3 H, C5 Hs & C4 H), 1.30–1.20 (bs, 1 H, OH), 0.86 (d, J = 8 Hz, 6 H). The proton NMR assignments were derived from 2D COSY spectrum. ¹³C NMR δ 206.1, 74.4, 52.8, 41.6, 37.4, 33.9, 27.0, 21.0, 17.1 (2 C). MS (ESI, MeOH): m/z = 171.1 ([M + H]⁺). HRMS-Hexane Atmospheric Pressure Chemical Ionization (HAPCI): m/z [M+H]⁺ calcd for C₁₀H₁₉O₂: 171.1385; found: 171.1381. The % ee (93%) of (*R*)-**99** was determined by HPLC using chiral column, Chiralpak AD(-H) column,

n-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, detected at 220 nm wavelength; $t_R = 21.7$ min (*S* enantiomer, minor), $t_R = 25.8$ min (*R* enantiomer, major).



Alternate synthetic route for (R)-99 to determine the absolutely configuration

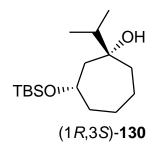
(S)-3-(t-Butyldimethylsilyloxy)cycloheptanone (129) (BH8-044)



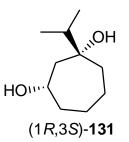
To a solution of 15 mg (0.12 mmol) of (*S*)-3-hydroxycycloheptanone (**42**), 1.5 mg (0.012 mmol) of 4-dimethylaminopyridine, and 35 mg (0.35 mmol) of trimethylamine in 1 mL of dichloromethane at 0°C under argon, was added 25 mg (0.17 mmol) of *t*-butyldimethylsilyl chloride. The solution was stirred for 8 h, diluted with 10 mL of water, and extracted three $_{155}$

times with ethyl acetate (10 mL each). The combined extract was washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel to give 19 mg (76% yield) of (*S*)-3-(*t*-butyldimethylsilyloxy)cycloheptanone and 3.2 mg (25% yield) of 2-cycloheptenone. The silyloxycycloheptanone was used in the following step without purification. (*S*)-3-(*t*-Butyldimethylsilyloxy)cycloheptanone: ¹H NMR δ 4.12–4.10 (m, 1 H), 2.85–2.75 (m, 2 H), 2.55–2.40 (m, 2 H), 1.95–1.55 (m, 6 H), 0.98 (s, 9 H), 0.37 (s, 6 H).

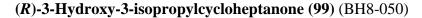
(1R,3S)-3-(t-Butyldimethylsilyloxy)-1-isopropylcycloheptanone (130) (BH8-048)

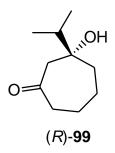


To a mixture of 1.1 mg (8 µmol) of anhydrous ZnCl₂ in 0.5 mL of dry THF under argon at 25°C, was added a solution of 0.12 mmol of isopropylmagnesium bromide in 0.2 mL of THF, and the solution was stirred for 1 h and then cooled to 0°C. To it, was added a solution of 19 mg (0.078 mmol) of (*S*)-3-(*t*-butyldimethylsilyloxy)cycloheptanone in 0.2 mL of THF via cannula under argon and the resulting solution was stirred at 0°C for 3 h. The solution was diluted with aqueous NH₄Cl and water (10 mL), extracted three times with ethyl acetate (10 mL each), and the combined extract was washed with brine, dried (anh. Na₂SO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluent to give 14 mg (62% yield) of (1*R*,3*S*)-3-(*t*-butyldimethylsilyloxy)-1-isopropylcycloheptanone (**130**). No other stereoisomer was found. ¹H NMR δ 4.42–4.35 (m, 1 H), 2.02–1.50 (m, 9 H), 1.35–1.20 (m, 3 H), 0.97 (s, 9 H), 0.91 (d, J = 7 Hz, 6 H), 0.36 (s, 6 H); ¹³C NMR δ 75.6, 70.9, 35.0, 33.1, 30.6, 29.6, 25.8 (3 C), 25.5, 24.3, 20.0, 17.2 (2 C), -1.6 (2 C).

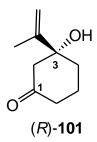


A solution of 7 mg (24 μ mol) of the above (1*R*,3*S*)-**130** and 48 μ L (48 μ mol) of *n*-Bu₄NF (1 M in THF) in 0.5 mL of THF was stirred under argon at 25°C for 2 h. The solution was diluted with water (10 mL), and extracted three times with diethyl ether (10 mL each). The combined extract was washed with brine, dried (MgSO₄), and concentrated under vacuum to give 4.2 mg of (1*R*,3*S*)-1-isopropyl-1,3-cycloheptanediol, which was used immediately in the following reaction without further purification.

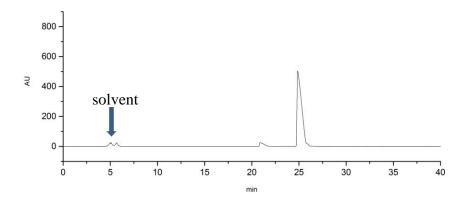


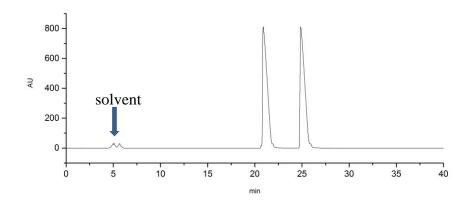


To a solution of **131** in 0.5 mL of DMSO was added 8 mg (26 µmol) of IBX under argon at 25°C. The resulting mixture was stirred for 14 h, filtered to remove white solid, and rinsed with 10 mL of diethyl ether. The ether filtrate was washed three times with water (10 mL each), dried (anh. Na₂SO₄), concentrated, and column chromatographed on silica gel using a mixture of hexane and 4.0ethyl acetate (1:2)eluent give (96%) vield) as to mg of (*R*)-3-hydroxy-3-isopropylcycloheptanone (99). $[\alpha]_D^{22} = +25.9$ (c 0.20, CHCl₃). ¹H and ¹³C NMR spectra were identical those of 99 synthesized by the oxidation reaction from 1-isopropyl-1-cycloheptanol with Pd/Au-1 catalyst.

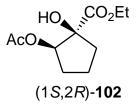


From 70 mg (0.50 mmol) of 1-isopropenylcyclohexanol (**100**), 1.7 mL of Cu/Au (3:1)-**1** (3.75 μ mol of Cu/1.25 μ mol of Au-0.13 μ mol of **1**), 1.5 mL of H₂O₂, and 1.5 mL of CH₃CN stirring at 50°C for 8 days, 76 mg (98% yield) of (*R*)-**101** in 93% ee. [α]_D²² = +17.3 (c 1.0, CHCl₃). ¹H NMR δ 5.07 (d, J = 0.8 Hz, 1 H, =CH), 4.95 (d, J = 0.8 Hz, 1 H, =CH), 2.35–2.32 (m, 2 H, C2 Hs), 2.16–2.15 (m, 2 H, C6 Hs), 2.00–1.95 (m, 2 H, C4 Hs), 1.85 (s, 3 H), 1.85–1.7 (m, 2 H, C5 Hs), 1.7–1.55 (bs, 1 H, OH). The proton NMR assignments were derived from 2D COSY spectrum. ¹³C NMR δ 209.5, 152.2, 109.2, 76.1, 52.9, 40.8, 38.8, 23.0, 17.8. MS (ESI, MeOH): m/z = 177.1 ([M + Na]⁺). HRMS-hexane Atmospheric Pressure Chemical Ionization (HAPCI): m/z [M+H]⁺ calcd for C₉H₁₅O₂: 155.1072; found: 155.1088. The % ee (93%) of (*R*)-**101** was determined by HPLC using chiral column, Chiralpak AD(-H) column, *n*-hexane/*i*-PrOH = 90:10, flow rate: 0.5 mL/min, detected at 220 nm wavelength; t_R = 20.8 min (*S* enantiomer, minor), t_R = 24.9 min (*R* enantiomer, major).



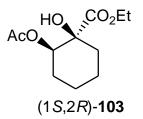


(15,2R)-Ethyl 2-acetoxy-1-hydroxycyclopentanecarboxylate (102) (BH10-068)



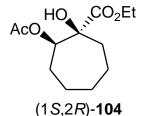
A solution of 20 mg (0.12 mmol) of (1*S*,2*R*)-**81**, 12 mg (0.11 mmol) of acetic anhydride and 9.1 mg (0.11 mmol) of pyridine in 1 mL of CH₂Cl₂ was stirred under argon for 12 hours, acidified with 1*N* HCl to pH 5, and extracted twice with 10 mL each of CH₂Cl₂. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an eluent to give 24 mg (96% yield) of (**1S**,2*R*)-**102**. $[\alpha]^{22}_{D}$ = +125.6 (0.2, CHCl₃); ¹H NMR δ 5.09 (dd, *J* = 11.2, 2.4 Hz, 1H), 4.22 (q *J* = 7.2 Hz, 2H), 3.50 (bs, 1H), 1.98 (s, 3H), 1.86–1.55 (m, 6H), 1.27 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 175.4, 170.2, 78.5, 78.0, 62.2, 28.7, 23.9, 21.0, 17.9, 14.3; MS (ESI, MeOH): m/z = 239.1 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₀H₁₇O₅⁺: 217.1076, found: 217.1080.

(1S,2R)-Ethyl 2-acetoxy-1-hydroxycyclohexanecarboxylate (103) (BH10-030)



To a solution of 50 mg (0.27 mmol) of (1*S*,2*R*)-**82** in 2 mL of CH₂Cl₂ under argon were added 27 mg (0.27 mmol) of acetic anhydride and 21 mg (0.27 mmol) of pyridine. The solution was stirred for 6 hours, acidified with 1*N* HCl to pH 5, and extracted twice with 10 mL each of CH₂Cl₂. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an eluent to give 26 mg (42% yield) of (1*S*,2*R*)-**103** and 19 mg (38% recovery) of (1*S*,2*R*)-**82** . $[\alpha]^{22}{}_{D}$ = +28.4 (0.2, CHCl₃); ¹H NMR δ 5.08 (dd, *J* = 11.2, 2.4 Hz, 1H), 4.19 (q *J* = 7.2 Hz, 2H), 2.85 (b, 1H), 1.98 (s, 3H), 1.78–1.55 (m, 8H), 1.25 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 176.0, 170.3, 79.1, 78.3, 62.2, 36.2, 29.4, 28.7, 20.9, 19.8, 14.2; MS (ESI, MeOH): m/z = 231.4 ([M + H]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₁H₁₉O₅⁺: 231.1232, found: 231.1235.

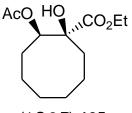
(1S,2R)-Ethyl 2-acetoxy-1-hydroxycycloheptanecarboxylate (104) (BH9-034)



To a solution of 48 mg (0.24 mmol) of (1S,2R)-83 in 2 mL of CH₂Cl₂, 24 mg (0.24 mmol) of acetic anhydride and 18 mg (0.24 mmol) of pyridine were added. The reaction was stirred under argon for 3 hours, acidified with 1*N* HCl to pH 5, and extracted with 10 mL of CH₂Cl₂ twice. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ether as an eluent

to give 26 mg (45% yield) of (1*S*,2*R*)-**104** and 24 mg (50% recovery) of (1*S*,2*R*)-**83**. $[\alpha]^{22}{}_{D}$ = +48.6 (0.5, CHCl₃); ¹H NMR δ 5.07 (dd, *J* = 11.2, 2.4 Hz, 1H), 4.20 (q *J* = 7.2 Hz, 2H), 3.25 (b, 1H), 1.98 (s, 3H), 1.94–1.55 (m, 10H), 1.25 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 175.9, 169.9, 78.6, 78.0, 62.3, 35.6, 28.6, 27.3, 23.3, 21.4, 21.2, 14.3; MS (ESI, MeOH): m/z = 267.1 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₂H₂₁O₅⁺: 245.1389, found: 245.1392.

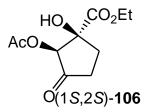
(1S,2R)-Ethyl 2-acetoxy-1-hydroxycyclooctanecarboxylate (105) (BH10-159)



(1S,2R)-105

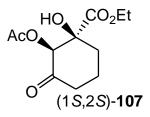
To a solution of 20 mg (0.093 mmol) of (1*S*,2*R*)-**84** in 1 mL of CH₂Cl₂, 9.48 mg (0.093 mmol) of acetic anhydride and 7.4 mg (0.093 mmol) of pyridine were added. The reaction was stirred under argon for 5 hours, acidified with 1*N* HCl to pH 5, and extracted three times with 10 mL of CH₂Cl₂ each. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ether as an eluent to give 17.6 mg (74% yield) of (1*S*,2*R*)-**105**. $[\alpha]^{22}{}_{D}$ =+16.2 (0.1, CHCl₃); ¹H NMR δ 5.03 (d, *J* = 10.8 Hz, 1H), 4.20 (q *J* = 7.2 Hz, 2H), 2.41 (b, 1H), 1.98 (s, 3H), 1.90–1.55 (m, 12H), 1.25 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 175.4, 170.9, 78.6, 77.8, 62.3, 30.4, 28.9, 28.6, 26.1, 23.8, 21.0, 18.4, 14.1; MS (ESI, MeOH): m/z = 281.1 ([M + Na]⁺).]; HRMS-ESI: m/z [M + H]⁺ calcd for C₁₃H₂₃O₅⁺: 259.1546, found: 259.1550.

(15,25)-Ethyl 2-acetoxy-1-hydroxy-3-oxocyclopentanecarboxylate (106) (BH10-082)



To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (0.69 µmol of Cu, 0.23 µmol of Au, and 0.025 µmol of **26**), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 20 mg (0.092 mmol) of (*1S*,2*R*)-**102**. The resulting solution was stired at 50°C for 3 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 17.7 mg (83% yield) of (*1S*,2*S*)-**106**. $[\alpha]^{22}_{D}$ =+134.2 (0.2, CHCl₃); ¹H NMR δ 5.21 (s, 1H), 4.22 (q, *J* = 7.2 Hz, 2H), 2.52–2.48 (m, 2H), 1.96 (s, 3H), 1.95–1.86 (m, 2H) 1.24 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 212.0, 177.1, 173.1, 100.2, 76.4, 62.2, 27.6, 20.5, 17.6, 14.2; MS (ESI, MeOH): m/z = 231.1 ([M + H]⁺); HRMS-ESI: m/z [M + H]⁺ calcd for C₁₀H₁₅O₆⁺: 231.0869, found: 231.0870.

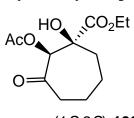
(1S,2S)-Ethyl 2-acetoxy-1-hydroxy-3-oxocyclohexanecarboxylate (107) (BH10-037)



To an aqueous solution of 3 mL of Cu/Au (3:1)-26 (0.82 µmol of Cu, 0.27 µmol of Au, and 0.030 µmol of 26), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 25 mg (0.109 mmol) of (*1S*,2*R*)-103. The resulting solution was stired at 50°C for 3 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 22 mg (83% yield) of (*1S*,2*S*)-107. $[\alpha]^{22}{}_{\rm D}$ =+20.2 (0.1, CHCl₃); ¹H NMR δ 5.10 (s, 1H), 4.22 (q, *J* = 7.2 Hz, 2H), 3.81 (b, 1H),

2.52–2.48 (m, 2H), 1.96 (s, 3H), 1.90–1.72 (m, 4H) 1.24 (t, J = 6.8 Hz, 3H); ¹³C NMR δ 212.0, 177.2, 173.0, 100.0, 76.3, 62.4, 32.1, 27.9, 23.5, 20.7, 14.2; MS (ESI, MeOH): m/z = 267.1 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₁H₁₇O₆⁺: 245.1025, found: 245.1031.

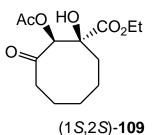
(15,25)-Ethyl 2-acetoxy-1-hydroxy-3-oxocycloheptanecarboxylate (108). (BH9-040)



(1*S*,2*S*)-**108**

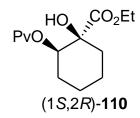
To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (0.80 µmol of Cu, 0.26 µmol of Au, and 0.029 µmol of **26**), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 26 mg (0.107 mmol) of (*1S*,2*R*)-**104**. The resulting solution was stirred at 50°C for 3 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 21 mg (76% yield) of (*1S*,2*S*)-**108**. $[\alpha]^{22}_{D}$ =+18.6 (0.1, CHCl₃); ¹H NMR δ 5.22 (s, 1H), 4.20 (q, *J* = 7.2 Hz, 2H), 3.48 (b, 1H), 2.52–2.48 (m, 2H), 1.96 (s, 3H), 1.78–1.54 (m, 6H) 1.24 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 211.2, 177.5, 172.4, 100.0, 76.5, 61.4, 32.1, 30.6, 25.6, 24.5, 20.7, 14.2; MS (ESI, MeOH): m/z = 281.1 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₂H₁₉O₆⁺: 259.1182, found: 259.1176.

(1S,2S)-Ethyl 2-acetoxy-1-hydroxy-3-oxocyclooctanecarboxylate (109) (BH11-015)



To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (0.43 µmol of Cu, 0.15 µmol of Au, and 0.016 µmol of **26**), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 15 mg (0.058 mmol) of (*IS*,*2R*)-**105**. The resulting solution was stirred at 50°C for 4 days, cooled to 25°C, and extracted three times with 20 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 13.2 mg (83% yield) of (*IS*,*2S*)-**109** and recover 0.8 mg (5.3% yield) of (*IS*,*2R*)-**105**. $[\alpha]^{22}_{D} = +43.8$ (0.1, CHCl₃); ¹H NMR δ 5.12 (s, 1H), 4.18 (q, *J* = 7.2 Hz, 2H), 2.51–2.47 (m, 2H), 1.96 (s, 3H), 1.90–1.48 (m, 8H) 1.24 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 211.8, 177.5, 173.7, 100.2, 76.4, 62.2, 34.4, 32.2, 26.8, 25.3, 23.7, 21.3, 14.2; MS (ESI, MeOH): m/z = 295.1 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₃H₂₁O₆⁺: 273.1138, found: 273.1133.

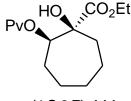
(1S,2R)-Ethyl 1-hydroxy-2-(pivaloyloxy)cyclohexanecarboxylate (110) (BH10-140)



To a solution of 50 mg (0.27 mmol) of (1*S*,2*R*)-**82** in 2 mL of CH_2Cl_2 under argon were added 32 mg (0.27 mmol) of pivaloyl chloride and 21.3 mg (0.27 mmol) of pyridine. The solution was stirred for 10 hours, acidified with 1*N* HCl to pH 5, and extracted twice with 20 mL each of CH_2Cl_2 . The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an

eluent to give 57 mg (80% yield) of (1*S*,2*R*)-**110**. $[\alpha]^{22}{}_{D}$ =+21.2 (0.4, CHCl₃); ¹H NMR δ 5.02 (d, *J* = 10.6 Hz, 1H), 4.26 (q, *J* = 6.8 Hz, 2H), 2.01–1.54 (m, 8H), 1.31 (t, *J* = 6.8 Hz, 3H), 1.26 (s, 9H); ¹³C NMR δ 177.0, 172.6, 79.5, 79.1, 62.2, 38.9, 31.8, 28.5, 27.5, 22.8, 19.5, 14.4; MS (ESI, MeOH): m/z = 273.1 ([M + H]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₄H₂₅O₅⁺: 273.1702, found: 273.1700.

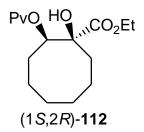
(1S,2R)-Ethyl 1-hydroxy-2-(pivaloyloxy)cycloheptanecarboxylate (111) (BH9-057)



(1S,2R)-111

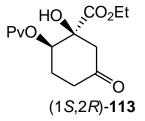
To a solution of 80 mg (0.396 mmol) of (1*S*,2*R*)-**83** in 3 mL of CH₂Cl₂ under argon were added 48 mg (0.396 mmol) of pivaloyl chloride and 31.3 mg (0.396 mmol) of pyridine. The solution was stirred for 15 hours, acidified with 1*N* HCl to pH 5, and extracted twice with 20 mL each of CH₂Cl₂. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an eluent to give 39 mg (34% yield) of (1*S*,2*R*)-**111** and 42 mg (52% recovery) of (1*S*,2*R*)-**83**. $[\alpha]^{22}{}_{\rm D}$ = +14.6 (0.5, CHCl₃); ¹H NMR δ 5.04 (d, *J* = 10.6 Hz, 1H), 4.26 (q, *J* = 6.8 Hz, 2H), 3.49 (bs, 1H), 1.92–1.42 (m, 10H), 1.31 (t, *J* = 6.8 Hz, 3H), 1.26 (s, 9H); ¹³C NMR δ 176.9, 172.6, 79.4, 79.3, 62.2, 38.7, 32.1, 28.9, 27.3, 26.4, 24.7, 21.1, 14.2; MS (ESI, MeOH): m/z = 287.1 ([M + H]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₅H₂₇O₅⁺: 287.1859, found: 287.1864.

(1S,2R)-Ethyl 1-hydroxy-2-(pivaloyloxy)cyclooctanecarboxylate (112) (BH10-141)



To a solution of 40 mg (0.185 mmol) of (1*S*,2*R*)-**84** in 2 mL of CH₂Cl₂ under argon were added 22 mg (0.185 mmol) of pivaloyl chloride and 14.6 mg (0.185 mmol) of pyridine. The solution was stirred for 10 hours, acidified with 1*N* HCl to pH 5, and extracted twice with 20 mL each of CH₂Cl₂. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an eluent to give 51 mgheld) of (1*S*,2*R*)-**112**. $[\alpha]^{22}_{D} = +67.3$ (0.2, CHCl₃); ¹H NMR δ 5.04 (d, *J* = 10.6 Hz, 1H), 4.23 (q, *J* = 6.8 Hz, 2H), 2.78 (bs, 1H), 1.86–1.44 (m, 12H), 1.31 (t, *J* = 6.8 Hz, 3H), 1.26 (s, 9H); ¹³C NMR δ 176.5, 172.2, 79.5, 79.0, 62.2, 39.1, 31.8, 30.0, 29.3, 28.5, 25.5, 22.9, 17.1, 14.4; MS (ESI, MeOH): m/z = 315.7 ([M + H]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₆H₂₉O₅⁺: 315.2015, found: 315.2017.

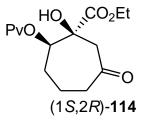
(1S,2R)-Ethyl 1-hydroxy-5-oxo-2-(pivaloyloxy)cyclohexanecarboxylate (113) (BH11-023)



To an aqueous solution of 6 mL of Cu/Au (3:1)-26 (2.05 μ mol of Cu, 0.69 μ mol of Au, and 0.075 μ mol of 26), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 15 mg (0.055 mmol) of (*1S*,2*R*)-110. The resulting solution was stirred at 50°C for 6 days, cooled to 25°C, and extracted three times with 20 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 13.1 mg (83%)

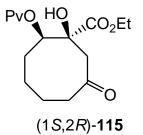
yield) of (1S,2S)-**113** and recover 1.1 mg (7.2% yield) of (1S,2R)-**110**. $[\alpha]^{22}_{D} = +61.3$ (0.5, CHCl₃); ¹H NMR δ 4.25 (q, J = 6.8 Hz, 2H, CH₂O), 3.62 – 3.60 (m, 1H, CHOPv), 2.71 (dd, J = 7.2, 1.6 Hz, 1H, CHC=O), 2.59 (dd, J = 7.2, 1.6 Hz, 1H, CHC=O), 2.40 – 2.32 (m, 1H), 2.28–2.22 (m, 1H), 2.04 – 2.00 (m, 2H), 1.51 (s, 9H), 1.31 (t, J = 6.8 Hz, 3H); The proton NMR assignments were derived from 2D COSY spectrum; ¹³C NMR δ 211.4, 176.9, 174.3, 80.6, 79.3, 62.2, 47.5, 38.8, 33.2, 26.4, 21.4, 14.4; MS (ESI, MeOH): m/z = 309.4 ([M + H]⁺); HRMS-ESI: m/z [M + H]⁺ calcd for C₁₄H₂₃O₆⁺: 287.1495, found: 287.1501.

(1S,2R)-Ethyl 1-hydroxy-6-oxo-2-(pivaloyloxy)cycloheptanecarboxylate (114) (BH9-059)



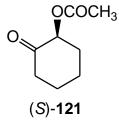
To an aqueous solution of 6 mL of Cu/Au (3:1)-26 (2.50 µmol of Cu, 0.83 µmol of Au, and 0.09 µmol of 26), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 20 mg (0.066 mmol) of (*1S*,2*R*)-111. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 18.3 mg (87.5% yield) of (*1S*,2*S*)-113 and recover 1.3 mg (6.5% yield) of (*1S*,2*R*)-111. $[\alpha]^{22}_{D}$ = +16.4 (0.5, CHCl₃); ¹H NMR δ 4.25 (q, *J* = 6.8 Hz, 2H, CH₂O), 3.82 – 3.83 (m, 1H, CHOPv), 2.81 (d, *J* = 7.2 Hz, 1H, CHC=O), 2.60 (d, *J* = 7.2, 1.6 Hz, 1H, CHC=O), 2.37 – 2.35 (m, 2H), 1.96 – 1.62 (m, 4H), 1.51 (s, 9H), 1.31 (t, *J* = 6.8 Hz, 3H); The proton NMR assignments were derived from 2D COSY spectrum; ¹³C NMR δ 210.4, 176.9, 173.1, 80.2, 79.3, 62.1, 46.5, 43.5, 38.7, 27.4, 26.5 21.2, 14.0; MS (ESI, MeOH): m/z = 323.4 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₅H₂₅O₆⁺: 301.1651, found: 301.1651.

(1S,2R)-Ethyl 1-hydroxy-7-oxo-2-(pivaloyloxy)cyclooctanecarboxylate (115) (BH11-025)

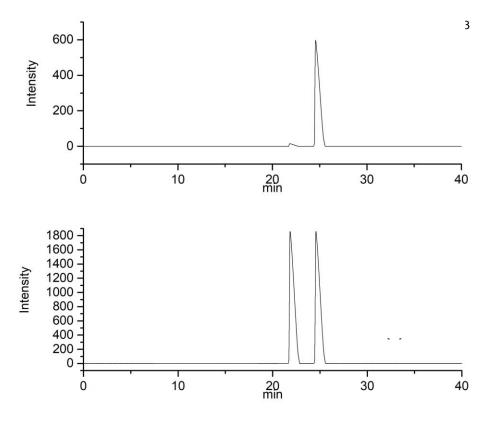


To an aqueous solution of 6 mL of Cu/Au (3:1)-26 (1.9 µmol of Cu, 0.63 µmol of Au, and 0.069 µmol of 26), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 15 mg (0.05 mmol) of (*1S*,2*R*)-112. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 14.1 mg (90% yield) of (*1S*,2*S*)-115. $[\alpha]^{22}_{D} = +21.7$ (0.7, CHCl₃); ¹H NMR δ 4.40 (bs, 1H, OH), 4.24 (q, *J* = 6.8 Hz, 2H, CH₂O), 3.76 – 3.74 (m, 1H), 2.73 (dd, *J* = 7.2, 1.6 Hz, 1H), 2.61 (dd, *J* = 7.2, 1.2 Hz, 1H), 2.46–2.23 (m, 3H), 2.10–1.96 (m, 2H), 1.76-1.72 (m, 3H), 1.51 (s, 9H), 1.31 (t, *J* = 6.8 Hz, 3H); The proton NMR assignments were derived from 2D COSY spectrum; ¹³C NMR δ 210.2, 176.9, 172.9, 80.6, 79.1, 62.1, 41.6, 40.6, 38.7, 33.5, 27.4, 24.7, 21.2, 14.3; MS (ESI, MeOH): m/z = 337.7 ([M + Na]⁺); HRMS-ESI: m/z [M + H]⁺ calcd for C₁₆H₂₇O₆⁺: 315.1808, found: 315.1807.

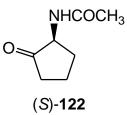




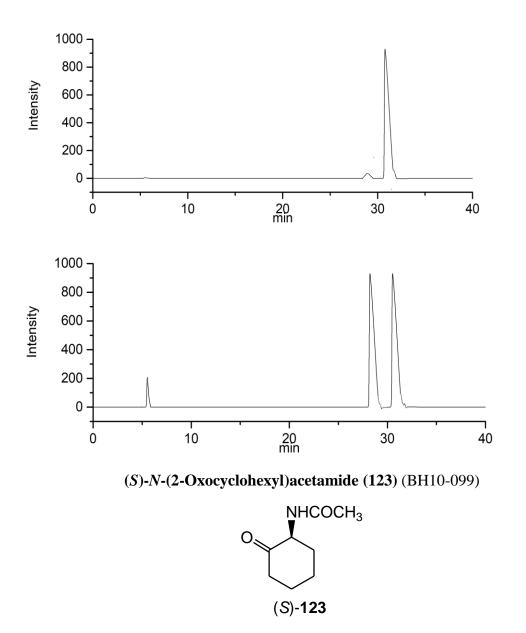
To an aqueous solution of 5 mL of Cu/Au (3:1)-**26** (7.0 µmol of Cu, 2.3 µmol of Au, and 0.132 µmol of **26**), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 30 mg (0.188 mmol) of **116**. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 30.7 mg (94% yield) of (*S*)-**121**. $[\alpha]_D^{22} =$ -80.4 (c 1.2, MeOH); Lit.^[103] $[\alpha]_D^{22} =$ +75.8 (c 1.2, MeOH for 88% *ee* R configuration); ¹H NMR : δ 5.17 (dd, *J* = 11.2, 6.0 Hz, 1H), 2.35–2.54 (m, 3H), 2.16 (s, 3H), 1.78–1.89 (m, 3H), 1.66-1.76 (m, 2H); ¹³C NMR : δ = 204.2, 170.3, 76.3, 40.1, 32.7, 26.9, 23.2, 20.1; MS (ESI, MeOH): m/z = 179.5 ([M + Na]⁺); The % *ee* (94%) of (*S*)-**121** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 95:5, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 21.9 min (*R* enantiomer, minor), t_R = 25.0 min (*S* enantiomer, major).



(S)-N-(2-Oxocyclopentyl)acetamide (122) (BH12-095)

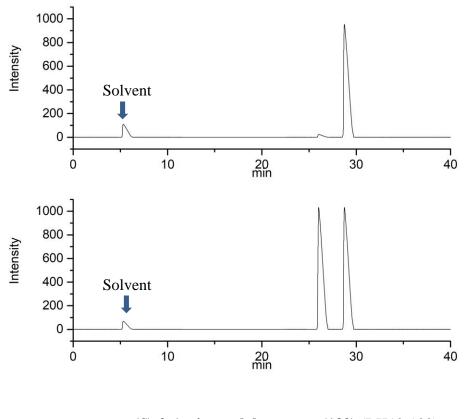


To an aqueous solution of 6 mL of Cu/Au (3:1)-**26** (6.7 µmol of Cu, 2.2 µmol of Au, and 0.132 µmol of **26**), were added 0.5 mL of 30% H₂O₂, 2 mL of CH₃CN and 22.5 mg (0.178 mmol) of **117**. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 10 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 17.5 mg (76% yield) of (*S*)-**122** and recover 3.8 mg (16.9% yield) of **117**. $[\alpha]_D^{22} = -41.6$ (c 0.2, CHCl₃); ¹H NMR δ 4.62–4.51 (m, 1H), 4.11 (bs, 1H, NH), 2.68–2.48 (m, 2H), 2.10 (s, 3H), 2.04–1.75 (m, 4H); ¹³C NMR δ 210.2, 170.5, 60.5, 32.3, 23.3, 14.2, 12.9; HRMS-ESI: m/z [M + H]+ calcd for C₇H₁₂NO₂⁺: 142.0868, found: 142.0861. The % ee (92%) of (*S*)-**122** was determined by HPCL using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/i-PrOH = 95:5, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 28.6 min (R enantiomer, minor), t_R = 31.0 min (S enantiomer, major).

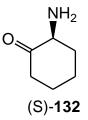


To an aqueous solution of 6 mL of Cu/Au (3:1)-**26** (13.4 µmol of Cu, 4.5 µmol of Au, and 0.263 µmol of **26**), were added 1 mL of 30% H₂O₂, 4 mL of CH₃CN and 50 mg (0.357 mmol) of **118**. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 39.2 mg (72% yield) of (*S*)-**123** and recover 8.6 mg (17.2% yield) of **118**. $[\alpha]_D^{22} = -62.4$ (c 0.5, CHCl₃); ¹H NMR^[104] δ 5.90 (bs, 1H,

NH), 4.51-4.44 (m, 1H), 2.70–2.35 (m, 3H), 2.19-2.11 (m, 1H), 2.02 (s, 3H), 1.95- 1.79 (m, 2H), 1.75-1.66 (m, 1H), 1.35-1.26 (m, 1H); ¹³C NMR δ 209.9, 171.4, 60.0, 43.0, 37.4, 30.0, 25.4, 25.0; The % *ee* (93%) of (*S*)-**123** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 95:5, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 26.1 min (*R* enantiomer, minor), t_R = 29.5 min (*S* enantiomer, major).



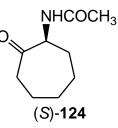
(S)-2-Aminocyclohexanone (132) (BH10-133)



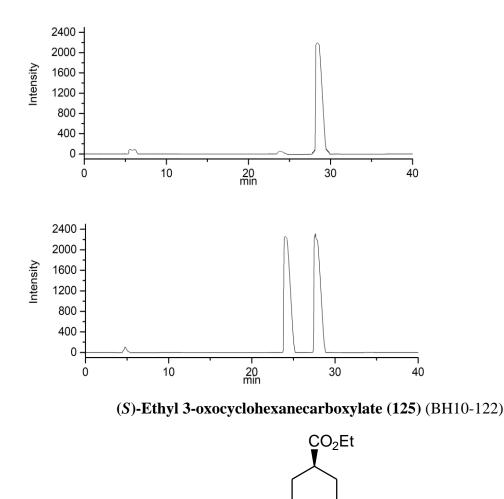
A solution of 15.5 mg (0.1 mmol) of **123** in 1 mL of hydrazine was heated to 70 °C under argon for 12 hours until no starting material remaining. The resulting solution was purified by

column chromatography on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 11.4 mg (99% yield) of (*S*)-**132**. $[\alpha]_D^{22} = -71.3$ (c 0.5, CHCl₃) Lit.^[103] $[\alpha]_D^{22} = +68.1$ (c 0.5, CHCl₃ for *R* configuration); ¹H NMR : δ 3.35–3.27 (m, 1H), 2.40–2.12 (m, 3H), 1.96–1.50 (m, 5H); ¹³C NMR : δ 209.3, 57.7, 38.7, 27.6, 26.7, 20.8; MS (ESI, MeOH): m/z = 114 ([M + H]⁺).





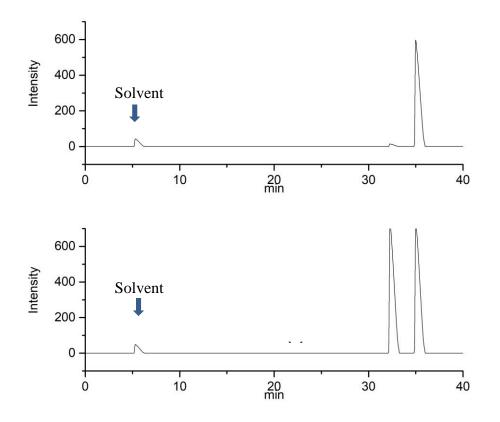
To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (6.7 µmol of Cu, 2.2 µmol of Au, and 0.132 µmol of **26**), were added 0.5 mL of 30% H₂O₂, 2 mL of CH₃CN and 27.5 mg (0.178 mmol) of **119**. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 10 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 25.6 mg (84% yield) of (*S*)-**124** and recover 2 mg (7.3% yield) of **119**. $[\alpha]_D^{22} = -127.2$ (c 0.2, CHCl₃); ¹H NMR^[104] : δ 6.25 (br, 1H), 4.69-4.59 (m, 1H), 2.71–2.41 (m, 2H), 2.14-2.05 (m, 1H), 2.02 (s, 3H), 1.95-1.67 (m, 5H), 1.50-1.41 (m, 1H), 1.35-1.25 (m, 1H); ¹³C NMR δ 210.4, 169.5, 59.0, 41.6, 32.9, 29.1, 27.6, 23.4, 23.2. The % ee (93%) of (*S*)-**124** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/i-PrOH = 95:5, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 24.2 min (R enantiomer, minor), t_R = 28.0 min (S enantiomer, major).



(S)-125

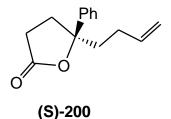
To an aqueous solution of 8 mL of Cu/Au (3:1)-**26** (20.5 µmol of Cu, 6.8 µmol of Au, and 0.383 µmol of **26**), were added 1 mL of 30% H₂O₂, 3 mL of CH₃CN and 80 mg (0.548 mmol) of **120**. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 76.9 mg (88% yield) of (*S*)-**125**. $[\alpha]_D^{22} = +2.62$ (c 1.61, MeOH) Lit.^[105] $[\alpha]_D^{22} = +2.7$ (c 1.6, MeOH, 96% *ee*) The % *ee* (93%) of (*S*)-**46** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 95:5, flow

rate 0.5 mL/min, detected at 220 nm wavelength; $t_R = 32.4$ min (*R* enantiomer, minor), $t_R = 35.2$ min (*S* enantiomer, major); ¹H NMR : δ 4.15 (q, J = 7.2 Hz, 2H), 3.79–3.77 (m, 1H), 2.60–2.58 (m, 2H), 2.78–2.25 (m, 2H), 2.14–2.05 (m, 2H), 1.89–1.78 (m, 2H), 1.25 (t, J = 6.8 Hz, 3H); ¹³C NMR δ 209.3, 173.7, 61.1, 43.2, 43.1, 40.9, 27.6, 24.3, 14.3; MS (ESI, MeOH): m/z = 193.2 ([M + Na]⁺).

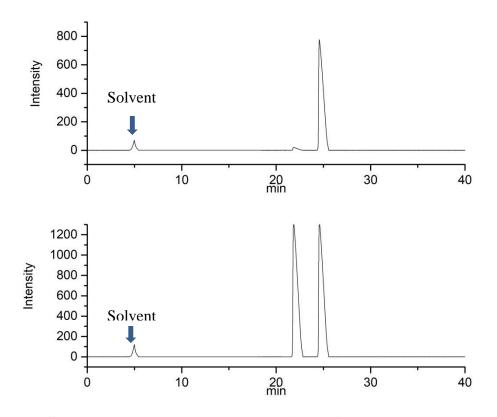


2.6.6 Procedures and Analysis Results for Oxidative Ring Closing Reaction of Dienes.

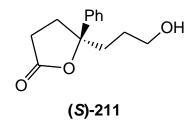
(S)-5-(3-Butenyl)-5-phenyl-dihydrofuran-2(3H)-one (200) (BH10-021)



To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (3.7 µmol of Cu, 1.2 µmol of Au, and 0.13 µmol of **26**), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 25 mg (0.116 mmol) of 5-phenylnona-1,8-dien-5-ol (**195**). The resulting solution was stirred at 50°C for 3 days, cooled to 25°C, and extracted three times with 10 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 35.3 mg (71% yield) of (**S**)-**200** in 96% ee. $[\alpha]_D^{22} = -14.6$ (0.3, CHCl₃); ¹H NMR δ 7.39–7.22 (m, 5H), 5.84–5.74 (m, 1H), 4.95 (d, *J* = 16.4 Hz, 1H), 4.92 (d, *J* = 9.6 Hz, 1H), 2.53 (m, 2H), 2.14–1.79 (m, 6H); ¹³C NMR δ 175.2, 142.3, 138.9, 127.2, 126.2, 125.1, 115.4, 92.2, 40.1, 34.4, 32.2, 28.1; MS (ESI, MeOH): m/z = 217.1 ([M + H]⁺); HRMS-ESI: m/z [M + H]⁺ calcd for C₁₄H₁₇O₂⁺: 217.1229, found: 217.1227. The % *ee* (96%) of (*S*)-**200** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 95:5, flow rate 0.5 mL/min, detected at 254 nm wavelength; t_R = 22.2 min (*R* enantiomer, minor), t_R = 25.0 min (*S* enantiomer, major).



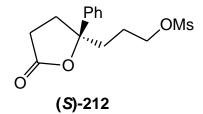
(S)-5-(3-Hydroxypropyl)-5-phenyl-dihydrofuran-2(3H)-one (211) (BH11-056)



To 6 mL of methylene chloride, ozone was bubbled into it under -78 °C for 3 minutes and gave a blue solution. To it, 19 mg of **200** (0.0879 mmol) was added and stir for 5 minutes under -78 °C until no starting material remaining. After the ozone evaporates, 6.7 mg (0.176 mmol) of NaBH₄ was added, the mixture was warmed up to room temperature and stirred for 1 hour under Ar. The reaction was quenched by 2 mL of sat. NH₄Cl solution and extracted three times with 20 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 13 mg (67% yield) of **211**. $[\alpha]_D^{22} = -143.1$

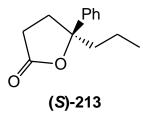
(0.05, CHCl₃); ¹H NMR δ 7.32–7.20 (m, 5H), 3.79 (t, J = 8.8 Hz, 1H), 2.25–2.19 (m, 2H), 2.10–1.52 (m, 7H); ¹³C NMR δ 175.1, 142.3, 127.2, 126.2, 125.1, 92.2, 34.3, 33.0, 28.6, 28.0; MS (ESI, MeOH): m/z = 243.4 ([M + Na]⁺).

(S)-3-(5-Oxo-2-phenyl-tetrahydrofuran-2-yl)propyl methanesulfonate (212) (BH11-062)



To a mixture of **211** (13 mg, 0.059 mmol) and dry triethylamine (6.6 mg, 0.065 mmol) in 1 mL of dry dichloromethane under argon, methanesulfonyl chloride (15.4 mg, 0.135 mmol) was added dropwise at 0°C. The reaction mixture was stirred for 8 hours at room temperature, quenched with 0.5 mL of water and extracted with methylene chloride (3 x 10 mL). The combined organic layer was washed with brine, dried over anhydrous sodium sulfate and concentrated under vacuum to give 17 mg (96.5% yield) **212**. The residue was used in the subsequent step without further purification. ¹H NMR δ 7.32–7.20 (m, 5H), 4.13–4.02 (m, 2H), 2.95 (s, 3H), 2.25–2.19 (m, 2H), 2.07–1.52 (m, 6H); ¹³C NMR δ 175.4, 142.3, 127.1, 125.7, 124.6, 91.9, 71.7, 35.7, 32.9, 27.66, 26.8, 21.6; MS (ESI, MeOH): m/z = 299.1 ([M + H]⁺).

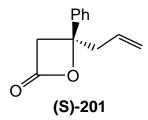
(S)-5-Phenyl-5-propyl-dihydrofuran-2(3H)-one (213) (BH11-063)



To a mixture of **212** (17 mg, 0.057 mmol) and *t*-BuOH (21 mg, 0.285 mmol) in 1 mL 1,2-dimethoxyethane, NaBH₄ (6.4 mg, 0.171 mmol) was added at 0° C. The reaction solution

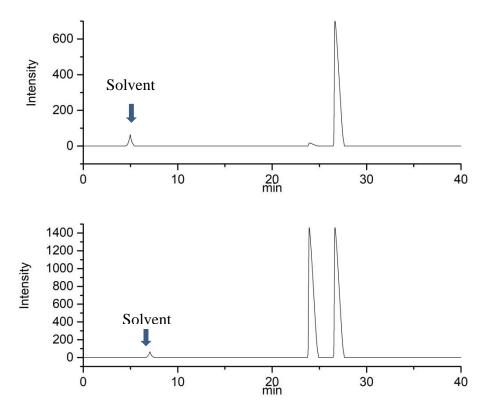
was stirred for 30 minutes at room temperature, heated to reflux overnight, quenched with 15 mL of water and extracted with ethyl acetate (3 x 20 mL). The combined organic layer was washed with brine, dried over anhydrous sodium sulfate and concentrated under vacuum and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 8.4 mg (72% yield) **213**. $[\alpha]_D^{22} = -42.1$ (c 0.02, MeOH), Lit.^[118] -33.0 (c 0.01, MeOH for (S)-**58C** with 74%ee); ¹H NMR δ 7.32–7.20 (m, 5H), 2.55–2.33 (m, 4H), 1.91–1.81 (m, 2H), 1.34–1.25 (m, 1H) 1.08–1.01 (m, 1H), 0.79 (t, J = 7.2 Hz); ¹³C NMR δ 177.3, 143.5, 128.7, 127.7, 125.0, 90.0, 45.1, 35.5, 29.1, 17.6, 14.3; MS (ESI, MeOH): m/z = 205.3 ([M + H]⁺).

(S)-4-Allyl-4-phenyloxetan-2-one (201) (BH10-053)

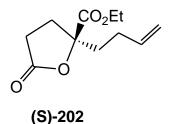


To an aqueous solution of 3 mL of Cu/Au (3:1)-26 (3.9 µmol of Cu, 1.3 µmol of Au, and 0.073 µmol of 26), were added 1 mL of 30% H₂O₂, 8 mL of CH₃CN 15 mL of water and 25 mg (0.133 mmol) of 4-phenylhepta-1,6-dien-4-ol (196). The resulting solution was heated to 60°C for 6 days, cooled to 25°C, and extracted three times with 10 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried ($MgSO_4$), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 10.9 mg (44% yield) of (S)-201 in 92% ee and 7.3 mg (29% recovery) of 196. $[\alpha]_D^{22} =$ -16.8 (0.2, CHCl₃); ¹H NMR δ 7.43–7.22 (m, 5H), 5.66–5.56 (m, 1H), 5.06 (d, J = 12 Hz, 1H), 4.98 (d, J = 4.8 Hz, 1H), 3.70 (d, J = 12.8 Hz, 1H), 3.48 (d, J = 12.8 Hz, 1H), 2.70 (dd, J = 14, ^{13}C 14, 1H); 6.4 Hz. 1H), 2.52 (dd, J= 6.4 Hz, NMR δ 174.9, 142.9, 138.9, 127.3, 126.3, 125.2, 115.5, 92.8, 43.7, 37.6; MS (ESI, MeOH): m/z = 189.1 ($[M + H]^+$); HRMS-ESI: m/z $[M + H]^+$ calcd for $C_{12}H_{13}O_2^+$: 189.0916, found: 189.0918.

The % *ee* (92%) of (*S*)-**201** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 95:5, flow rate 0.5 mL/min, detected at 254 nm wavelength; $t_R = 24.3 \text{ min}$ (*R* enantiomer, minor), $t_R = 27.0 \text{ min}$ (*S* enantiomer, major).

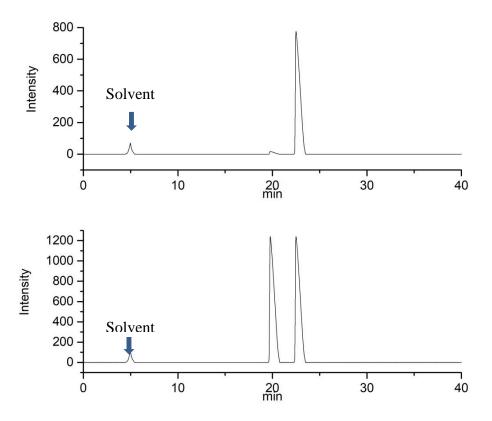


(S)-Ethyl 2-(but-3-enyl)-5-oxo-tetrahydrofuran-2-carboxylate (202). (BH11-051)

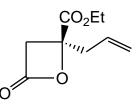


To an aqueous solution of 5 mL of Cu/Au (3:1)-26 (2.9 μ mol of Cu, 1.0 μ mol of Au, and 0.11 μ mol of 26), were added 1 mL of 30% H₂O₂, 5 mL of CH₃CN and 20 mg (0.095 mmol) of ethyl 2-(but-3-enyl)-2-hydroxyhex-5-enoate (197). The resulting solution was heated to 50 °C for 3 days, and extracted three times with 10 mL each of ethyl acetate. The combined extracts were

washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 16.7 mg (84% yield) of (**S**)-**202** in 93% ee and recovered 0.6 mg (3% yield). $[\alpha]_D^{22} = +35.7$ (0.4, CHCl₃); ¹H NMR δ 5.85 – 5.75 (m, 1H), 5.03 (d, J = 17.2 Hz, 1H), 4.98 (dd, J = 12, 6 Hz, 1H), 4.21 (q, J = 6.8 Hz, 2H), 2.59 (m, J = 9.2, 6.4 Hz, 2H), 2.25–1.63 (m, 6H), 1.28 (t, J = 7.2 Hz, 3H); ¹³C NMR δ 177.4, 174.9, 138.9, 115.4, 97.6, 61.5, 40.1, 34.4, 30.2, 23.8, 14.9; MS (ESI, MeOH): m/z = 234.0 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₁H₁₇O₄⁺: 213.1127, found: 213.1127. The % *ee* (93%) of (*S*)-**202** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 90:10, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 20.0 min (*R* enantiomer, minor), t_R = 23.1 min (*S* enantiomer, major).

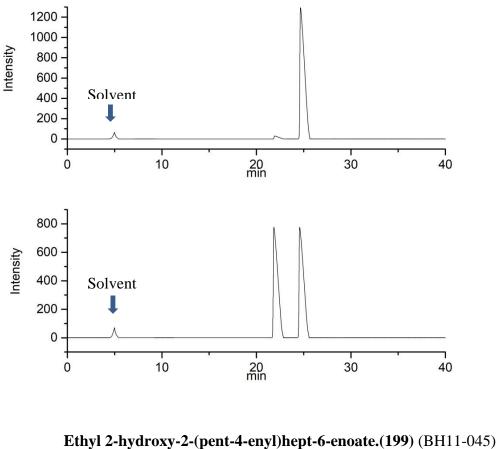


(S)-Ethyl 2-allyl-4-oxooxetane-2-carboxylate (203) (BH11-013)



(S)-203

To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (3.4 µmol of Cu, 1.4 µmol of Au, and 0.13 µmol of **26**), were added 2 mL of 30% H₂O₂, 3 mL of CH₃CN and 20 mg (0.115 mmol) of ethyl 2-allyl-2-hydroxypent-4-enoate (**198**). The resulting solution was stirred to 50°C for 7 days, and extracted three times with 15 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 6.2 mg (33% yield) of (**S**)-**203** in 93% ee and 1 mg (5% recovery) of **198**. $[\alpha]_D^{22} = +28.9$ (0.3, CHCl₃); ¹H NMR δ 5.83 – 5.73 (m, 1H), 5.11 (d, *J* = 14.4 Hz, 1H), 5.10 (d, *J* = 11.2 Hz, 1H), 4.21 (q, *J* = 6.8 Hz, 2H), 3.70 (d, *J* = 12.8 Hz, 1H), 3.49 (d, *J* = 12.8 Hz, 1H), 2.52–2.40 (m, 2H), 1.28 (t, *J* = 7.2 Hz, 3H); ¹³C NMR δ 176.9, 173.3, 137.7, 115.3, 86.6, 61.4, 40.2, 25.8, 14.8; MS (ESI, MeOH): m/z = 207.2 ([M + Na]⁺); HRMS-ESI: m/z [M + H]⁺ calcd for C₉H₁₃O₄⁺: 185.0814, found: 185.0811. The % *ee* (93%) of (*S*)-**203** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 90:10, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 22.3 min (*R* enantiomer, minor), t_R = 25.1 min (*S* enantiomer, major).



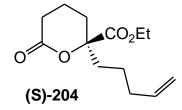
Ethyl 2-hydroxy-2-(pent-4-enyl)hept-6-enoate.(199) (BH11-045) HO_COOEt

199

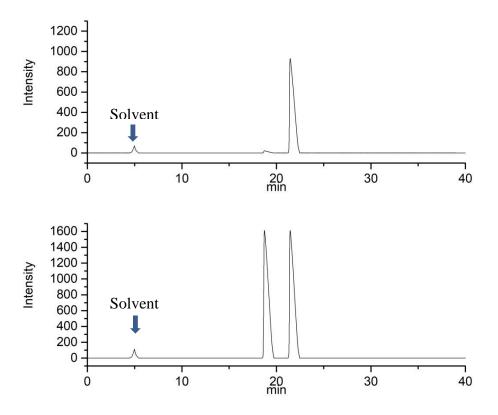
To a suspension of 70.88 mg (2.95 mmol) Mg in 5 mL anhydrous THF, 100 mg (0.671 mmol) of 5-bromopent-1-ene was added and heated with heat gun to initiate the reaction. To it, 300 mg (2.031 mmol) of 5-bromopent-1-ene was added and heated to 50 °C under argon until no Mg remaining. 111.1 mg (1.61 mmol) of diethyl oxalate in 5 mL of anhydrous THF was dropwise cannulated into the Grignard reagent under -78 °C, stirred at room temperature for 3 hours, quenched with sat. NH₄Cl solution, extracted three times of 30 mL each of methylene chloride and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an eluent to give 112 mg (35% yield) of the titled compound. ¹H NMR δ 5.74 (m, 2H), 4.96 (dd, *J* = 17.2 1.6 Hz, 2H), 4.92 (d, *J* = 9.6 Hz, 2H), 4.14 (q, *J* = 6.8 Hz, 2H), 2.04 – 1.98 (m, 4H),

1.75 - 1.35 (m, 8H), 1.24 (t, J = 7.2 Hz, 3H); 13 C NMR δ 177.5, 138.4, 114.8, 79.6, 61.0, 43.0, 35.8, 24.2, 14.5; MS (ESI, MeOH): m/z = 267.1 ([M + Na]+).

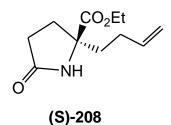
(S)-ethyl 6-oxo-2-(pent-4-enyl)-tetrahydro-2H-pyran-2-carboxylate (204). (BH11-049)



To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (4.8 µmol of Cu, 1.6 µmol of Au, and 0.17 µmol of **26**), were added 2 mL of 30% H₂O₂, 4 mL of CH₃CN and 40 mg (0.0845 mmol) of ethyl 2-hydroxy-2-(pent-4-enyl)hept-6-enoate (**199**). The resulting solution was heated to 50 °C for 6 days, and extracted three times with 25 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 33 mg (83% yield) of (**S**)-**204** in 93% ee and recovered 2 mg (6% yield) of **199**. $[\alpha]_D^{22} =$ +17.5 (0.3, CHCl₃); ¹H NMR δ 5.84–5.74 (m, 1H), 4.95 (d, *J* = 16.4 Hz, 1H), 4.92 (d, *J* = 9.6 Hz, 1H), 4.21 (q, *J* = 6.8 Hz, 2H), 2.50 (t, 8.0 Hz, 2H), 2.18–1.40 (m, 10H), 1.25 (t, *J* = 7.2 Hz, 3H); ¹³C NMR δ 177.7, 174.7, 138.9, 115.4, 89.0, 61.4, 37.5, 34.4, 32.9, 29.3, 24.9, 23.8, 14.8; MS (ESI, MeOH): m/z = 235.0 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₃H₂₁O₄⁺: 241.1440, found: 241.1440. The % *ee* (93%) of (*S*)-**204** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 90:10, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 18.6 min (*R* enantiomer, minor), t_R = 22.0 min (*S* enantiomer, major).

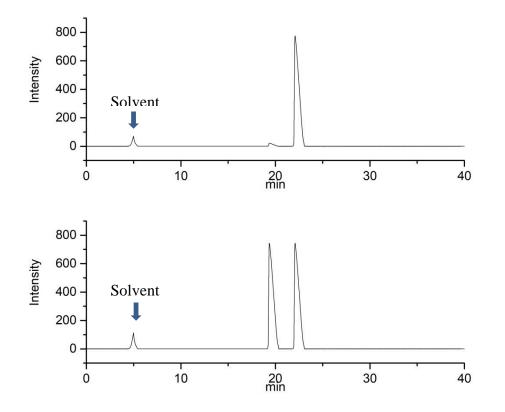


(S)-Ethyl 2-(but-3-enyl)-5-oxopyrrolidine-2-carboxylate (208). (BH10-051)

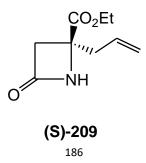


To an aqueous solution of 15 mL of Cu/Au (3:1)-**26** (21.3 µmol of Cu, 7.1 µmol of Au, and 0.7 µmol of **26**), were added 5 mL of 30% H₂O₂, 5 mL of CH₃CN and 150 mg (0.711 mmol) of ethyl 2-amino-2-(but-3-enyl)hex-5-enoate (**205**). The resulting solution was stirred to 50°C for 3 days, cooled to room temperature, and extracted three times with 40 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 126 mg (84% yield) of (**S**)-**208** in 95% ee. $[\alpha]_D^{22} = +18.6$ (0.2, CHCl₃); ¹H NMR δ 8.18 (bs, NH), 5.83–5.71 (m, 1H), 5.10 (d, *J* = 17.2 Hz, 1H), 4.95 (d, *J* =

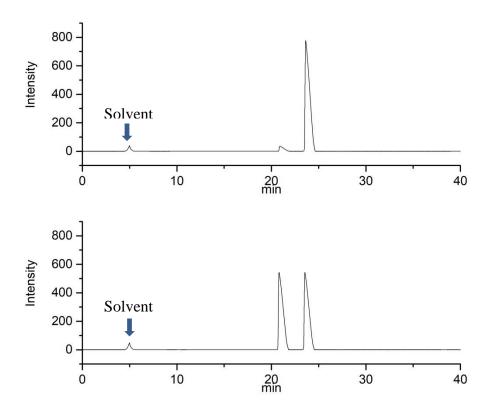
9.2 Hz, 1H), 4.24 (q, J = 6.8 Hz, 2H), 2.62 (t, J = 8.4 Hz, 1H), 2.21–1.64 (m, 6H), 1.28 (t, J = 7.2 Hz, 3H); ¹³C NMR δ 176.9, 173.5, 138.1, 115.3, 61.6, 27.0, 40.1, 36.7, 33.4, 23.2, 14.8; MS (ESI, MeOH): m/z = 234.6 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₁H₁₈NO₃⁺: 212.1287, found: 212.1288. The % *ee* (95%) of (*S*)-**208** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 90:10, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 19.6 min (*R* enantiomer, minor), t_R = 22.2 min (*S* enantiomer, major).



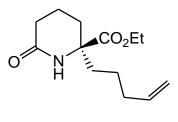
(S)-Ethyl 2-allyl-4-oxoazetidine-2-carboxylate (209). (BH11-060)



To an aqueous solution of 2 mL of Cu/Au (3:1)-**26** (3.2 µmol of Cu, 1.0 µmol of Au, and 0.06 µmol of **26**), were added 1 mL of 30% H₂O₂, 2 mL of CH₃CN and 20 mg (0.1093 mmol) of ethyl 2-allyl-2-aminopent-4-enoate (**206**). The resulting solution was heated to 50 °C for 7 days, and extracted three times with 20 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 12.8 mg (64% yield) of (**S**)-**209** in 92% ee. $[\alpha]_D^{22} = +33.7$ (0.2, CHCl₃); ¹H NMR δ 7.28 (bs, 1H), 5.80–5.70 (m, 1H), 5.16 (d, *J* = 19.2 Hz, 1H), 5.15 (d, *J* = 10.8 Hz, 1H), 4.21 (q, *J* = 6.8 Hz, 2H), 3.21–3.12 (m, 2H), 2.62–2.45 (m, 1H), 2.31–2.28 (m, 1H), 1.28 (t, *J* = 7.2 Hz, 3H); ¹³C NMR δ 177.0, 173.3, 137.7, 115.3, 61.8, 57.0, 38.4, 24.3, 14.0; MS (ESI, MeOH): m/z = 206.2 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₉H₁₄NO₃⁺: 184.0974, found: 184.0963. The % *ee* (92%) of (*S*)-**209** was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/*i*-PrOH = 90:10, flow rate 0.5 mL/min, detected at 220 nm wavelength; t_R = 20.9 min (*R* enantiomer, minor), t_R = 24.1 min (*S* enantiomer, major).



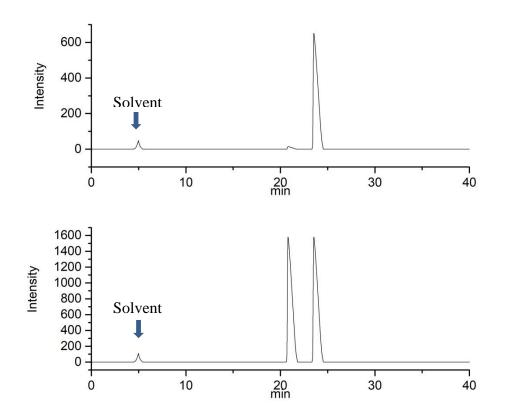
(S)-Ethyl 6-oxo-2-(pent-4-enyl)piperidine-2-carboxylate (210) (BH11-037)



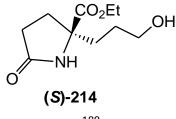
(S)-210

To an aqueous solution of 2 mL of Cu/Au (3:1)-**26** (2.5 µmol of Cu, 0.8 µmol of Au, and 0.046 µmol of **26**), were added 1 mL of 30% H₂O₂, 5 mL of CH₃CN and 20 mg (0.0845 mmol) ethyl 2-amino-2-(pent-4-enyl)hept-6-enoate (**207**). The resulting solution was stirred at 50°C for 5 days, cooled to room temperature, and extracted three times with 15 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 18.6 mg (93% yield) of (*S*)-**210** in 94% ee. $[\alpha]_D^{22} = +48.7 (0.5, 0.5)$

CHCl₃); ¹H NMR δ 5.79 – 5.69 (m, 1H), 4.96 (d, J = 17.2 Hz, 1H), 4.92 (d, J = 10.0 Hz, 1H), 4.19 (q, J = 7.2 Hz, 2H), 2.32 (td, J = 10.2, 2.4 Hz, 2H) 1.77–1.37 (m, 10H), 1.24 (t, J = 7.2 Hz, 3H); ¹³C NMR δ 177.9, 172.4, 138.9, 116.4, 61.4, 56.5, 39.0, 34.4, 31.1, 27.0, 23.5, 22.8, 14.8; MS (ESI, MeOH): m/z = 262.2 ($[M + Na]^+$). HRMS-ESI: m/z $[M + H]^+$ calcd for $C_{13}H_{22}NO_3^+$: 240.1660, found: 240.1651. The % ee (94%) of (S)-210 was determined by HPLC using chiral column, Chiralpak AD(-H) column [Chiralpak AD(-H) column, size: 0.46 cm x 25 cm, from Daicel Chemical Industries], n-hexane/i-PrOH = 90:10, flow rate 0.5 mL/min, detected at 220 nm wavelength; $t_R = 21.2 \text{ min} (R \text{ enantiomer, minor}), t_R = 24.5 \text{ min} (S \text{ enantiomer, major}).$

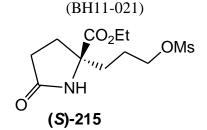


(S)-Ethyl 2-(3-hydroxypropyl)-5-oxopyrrolidine-2-carboxylate (214) (BH11-019)



To a solution of 12 mL of methylene chloride under argon at -78°C, ozone was bubbled into it for 3 minutes while the argon was turned off, to give a blue solution. To it, 35 mg of **208** (0.167 mmol) was added and stirred for 15 minutes at -78 °C (until no starting material remaining). The remaining ozone was evaporated by introducing argon into the system for 10 minutes. To it, 12.6 mg (0.33 mmol) of NaBH₄ and 0.5 mL of methanol were added, and the solution was warmed up to room temperature and stirred for 1 hour. The reaction was quenched by 2 mL of saturated aqueous NH₄Cl solution and extracted three times with 40 mL each of methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 26.2 mg (75% yield) of **214**. $[\alpha]_D^{22} = +31.7$ (0.05, CHCl₃); ¹H NMR δ 4.29 (q, *J* = 7.2 Hz, 2H), 3.53–3.61 (m, 2H), 2.55–2.45 (m, 1H), 2.25–1.62 (m, 8H), 1.32 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 177.0, 173.4, 65.8, 62.6, 61.6, 35.0, 29.6, 27.2, 24.1, 14.8; MS (ESI, MeOH): m/z = 216.3 ([M + H]⁺).

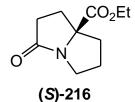
(S)-Ethyl 2-(3-(methylsulfonyloxy)propyl)-5-oxopyrrolidine-2-carboxylate (215)



To a cold (0°C) solution of **214** (26 mg, 0.123 mmol) and triethylamine (13.6 mg, 0.135 mmol) in 2 mL of dichloromethane under argon was added methanesulfonyl chloride (15.4 mg, 0.135 mmol) dropwise. The reaction solution was stirred for 8 hours at room temperature, diluted with 0.5 mL of water and extracted with methylene chloride (3 x 10 mL). The combined organic layer was washed with brine, dried over anhydrous sodium sulfate, and concentrated on a rotary evaporator and then under vacuum to give 29 mg (80.4% yield) **215**. The residue was

used in the subsequent step without further purification. $[\alpha]_D^{22} = +131.6 (0.1, \text{CHCl}_3)$; ¹H NMR δ 4.22 (q, J = 7.2 Hz, 2H), 3.96–3.85 (m, 2H), 2.95 (s, 3H), 2.53–2.45 (m, 1H), 2.22–1.64 (m, 8H), 1.32 (t, J = 6.8 Hz, 3H); ¹³C NMR δ 177.0, 173.4, 70.0, 65.9, 61.6, 37.9, 35.0, 29.9, 26.1, 20.5, 14.9; MS (ESI, MeOH): m/z = 316.3 ([M + Na]⁺).

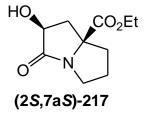
(S)-Ethyl 3-oxo-hexahydro-1H-pyrrolizine-7a-carboxylate (216) (BH11-027)



To a cold (0°C) solution of 29 mg (0.099 mmol) of **215** in 2 mL distilled THF under argon was added NaH (2.5 mg, 0.104 mmol), and the solution was stirred at room temperature for 3 hour, quenched with 0.5 mL of water, and extracted with methylene chloride (3 x 10 mL). The combined organic layer was washed with brine, dried over anhydrous sodium sulfate, concentrated under vacuum, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 16 mg (78% yield) of **216.** $[\alpha]_D^{22} = -41.6$ (0.05, CHCl₃); ¹H NMR δ 4.20 (q, *J* = 7.2 Hz, 2H), 3.61–3.48 (m, 2H), 2.55 (dd, *J* = 4.8, 1.2 Hz, 1H), 2.38 (d, *J* = 4.4 Hz, 1H), 2.25–1.80 (m, 6H), 1.30 (t, *J* = 6.8 Hz, 3H); ¹³C NMR δ 175.4, 173.4, 67.7, 61.6, 43.2, 42.4, 32.2, 27.2, 20.0, 14.8; MS (ESI, MeOH): m/z = 220.1 ([M + Na]⁺).

(2S,7aS)-Ethyl 2-hydroxy-3-oxo-hexahydro-1H-pyrrolizine-7a-carboxylate (217)

(BH11-053)

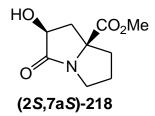


To a cold (-78°C) solution of 0.11 mL (1.05 mmol) of diisopropylamine in 5 mL distilled THF 191

under argon was added 1 mL of *n*-BuLi (1M in hexanes). The resulting solution was warmed up to room temperature and stirred for 1 hour to give a LDA solution. To a cold (-78°C) solution of **216** (19 mg, 0.096 mmol) in 1 mL of distilled THF under argon, 0.6 mL of LDA solution was added dropwise and stirred at room temperature for 30 minutes. To 62.5 mg (0.144 mmol) of MoOPH at -30°C under argon was added the above anion solution via cannula, and the solution was stirred for 40 minutes, warmed to room temperature, diluted with 2 mL of saturated aqueous Na_2SO_3 solution, and extracted with ethyl acetate (3 x 10 mL). The combined organic layer was washed with brine, dried over anhydrous sodium sulfate, concentrated to dryness, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 11.7 mg (57% yield) of (2S,7aS)-217 and 1.6 mg (7.8% yied) of (2*R*,7**a***S*)-217. $[\alpha]_D^{22} = -8.9$ (c 0.88, CHCl₃), ¹H NMR δ 5.91 (b, 1H), 4.41 (d, J =6.8, 1H), 4.20 (q, J = 7.2 Hz, 2H), 3.64 (d, J = 10.0 Hz, 1H), 3.28 (dd, J = 10.4, 2.0 Hz, 1H), 2.54 (d, J = 12.8 Hz, 1H), 2.32 (d, J = 12.8 Hz, 1H), 2.26 (d, J = 11.2 Hz, 1H), 2.14-1.98 (m, 2H),1.62 (m, 1H), 1.24 (t, J = 6.8 Hz, 3H); ¹³C NMR δ 174.4, 173.7, 74.1, 72.2, 61.6, 41.0, 39.9, 36.0, 25.5, 14.9; $m/z = 236.1 ([M + Na]^+)$.

(2S,7aS)-Methyl 2-hydroxy-3-oxo-hexahydro-1H-pyrrolizine-7a-carboxylate (218)

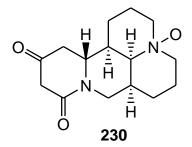
(BH11-067)

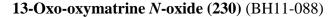


To a solution of (11.5 mg,0.054 mmol) **217** (11.5 mg, 0.054 mmol) in 2 mL of distilled MeOH under argon was added 1 mg of MeONa, and the solution was stirred at room temperature for 8 hours, concentrated on a rotary evaporator, and diluted with 20 mL of methylene chloride. The solution was washed with 5 mL of water, and 5 mL of brine, dried over anhydrous sodium

sulfate, concentrated under vacuum, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 4.2 mg (37% yield) of **218** and 4.8 mg (41% recovery) of **217**. Compound **218**: $[\alpha]_D^{22} = -10.2$ (c 0.88, CHCl₃), Lit.^[120] -10.2 (c 0.88, CHCl₃); ¹H NMR δ 4.95 (bs, 1H), 4.37 (d, *J* = 6.8, 1H), 3.76 (s, 3H), 3.64 (dt, *J* = 10.4, 1.2 Hz, 1H), 3.28 (dd, *J* = 10.4, 2.0 Hz, 1H), 2.54 (d, *J* = 12.8 Hz, 1H), 2.35 (d, *J* = 12.8 Hz, 1H), 2.23 (d, *J* = 11.2 Hz, 1H), 2.14-2.00 (m, 2H), 1.63-1.61 (m, 1H); ¹³C NMR δ 174.4, 173.8, 74.1, 72.2, 52.8, 41.2, 39.8, 36.0, 25.5; m/z = 222.1 ([M + Na]⁺).

2.6.7 Procedures and Analysis Results for Late-stage C-H Oxidation of Complex Molecules.

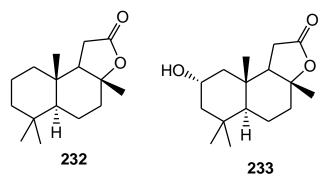




To an aqueous solution of 6 mL of Cu/Au (3:1)-**26** (41.6 µmol of Cu, 13.9 µmol of Au, and 0.76 µmol of **26**), were added 3 mL of 30% H₂O₂, 4 mL of CH₃CN and 49 mg (0.185 mmol) of oximatrine *N*-oxide (**229**). The resulting solution was stirred at 80°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 31 mg (60% yield) of **230** and recover 16.3 mg (33.3% yield) of **229**. $[\alpha]_D^{22} = + 14.6$ (c 0.2, MeOH); ¹H NMR δ 5.15 – 5.09 (m, 1H), 4.41 (dd, *J* = 12, 5.2 Hz, 1H), 4.20 (t, *J* = 12.4 Hz, 1H), 3.50–3.25 (m, 2H), 3.22–3.02 (m, 5H), 2.82–2.60 (m, 2H), 2.48–2.37(m, 1H), 2.32 (t, *J* = 4.8 Hz, 1H),

2.10–2.04 (m, 1H), 1.89–1.48 (m, 7H); ¹³C NMR^[132] δ 210.1, 169.1, 69.1, 68.3, 66.6, 52.3, 45.2, 43.3, 42.0, 41.5, 34.5, 26.0, 24.4, 17.3, 17.0; MS (ESI, MeOH): m/z = 301.4 ([M + Na]⁺). HRMS-ESI: m/z [M + H]⁺ calcd for C₁₅H₂₃N₂O₃⁺: 279.1709, found: 279.1714. The ¹³C NMR data is in agreement with that reported^[132] and the structure was further verified by NMR 2D COSY experiment.

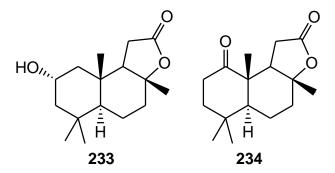
Sclareolide (232) & (2S)-2-Hydroxysclareolide (233). (BH11-086)



To an aqueous solution of 8 mL of Cu/Au (3:1)-**26** (15.9 µmol of Cu, 5.3 µmol of Au, and 0.30 µmol of **26**), were added 2 mL of 30% H₂O₂, 4 mL of CH₃CN and 100 mg (0.423 mmol) of ambroxide **231**. The resulting solution was stirred at 80 °C for 5 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 78.3 mg (73.9% yield) of **232**, 6.7 mg (5.9% yield) of **233** and recover 3.2 mg (3.2% yield) of **231**. **232**: ¹H NMR δ 3.39 (dd, *J* = 14.2, 1.2 Hz, 1H), 2.16 (dd, J = 16.4, 6.8 Hz, 1 H), 2.06 (dt, J = 12, 3.2 Hz, 1 H), 1.95 (dd, J = 14.4, 6.4 Hz, 1 H), 1.89–1.83 (m, 1 H), 1.71–1.62 (m, 2 H), 1.47–1.33 (m, 2 H), 1.32 (s, 3 H), 1.22–1.13 (m, 1 H), 1.058 (dd, J = 13, 3 Hz, 1 H), 0.89 (s, 3 H), 0.86 (s, 3 H), 0.82 (s, 3 H); ¹³C NMR δ 86.6, 59.3, 56.8, 42.4, 39.7, 38.9, 36.3, 33.4, 33.3, 28.9, 21.8, 21.1, 20.8, 18.3, 15.3; MS (ESI, MeOH): m/z = 251.2 (M+H⁺). **233**: $[\alpha]_D^{22} = +78$ (c 0.1, MeOH). Litt.^[136] -100 (c 0.006, MeOH). ¹H NMR δ 4.03–3.94 (m, 1 H), 2.42 (t, J = 14.4 Hz, 1 H), 2.27 (dd, J = 16.4, 6.4 Hz, 1 H), 2.10 (dt, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 12, 3.2 Hz, 1 H), 2.01 (dd, J = 14.4, 6.4 Hz, 1 H), 1.94–1.80 (m, 14), 2.01 (dd, J = 14.4, 6.

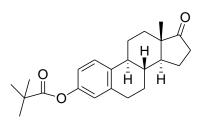
3 H), 1.74–1.66 (m, 1 H), 1.40–1.35 (m, 1 H), 1.33 (s, 3 H), 1.28–1.25 (m, 1 H), 1.17 (t, J = 11.6 Hz, 1 H), 1.08 (dd, J = 13, 3 Hz, 1 H), 0.96 (s, 3 H), 0.95 (s, 3 H), 0.89 (s, 3 H); ¹³ C NMR δ 176.0, 86.0, 64.4, 58.9, 56.3, 51.6, 48.4, 38.5, 37.5, 34.7, 33.2, 28.7, 21.9, 21.7, 20.1, 16.2; MS (ESI, MeOH): m/z = 289.4 ([M + Na]⁺).

(2S)-2-Hydroxysclareolide (233) & 1-Oxosclareolide (234) (BH8-098)

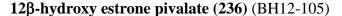


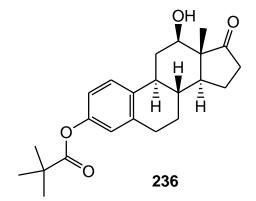
To a solution of 12 mL of Cu/Au (3:1)-1 (24 µmol of Cu/8.0 µmol of Au-0.88 µmol of 1) in H₂O (as described above) were added 12 mL of acetonitrile, 100 mg (0.40 mmol) of (+)-sclareolide (**232**) and 1.5 mL of 30% H₂O₂, and the resulting solution was stirred at 60°C for 6 days. The solution was cooled to 25°C, diluted with water, and extracted three times with diethyl ether (20 mL each). The combined extracts were washed with water and then brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate, followed by dichloromethane and methanol as eluents to give 40 mg (37.5% yield) of **233** and 3.3 mg (3% yield) of **234** along with 46 mg (46% recovery) of **232** and 10 mg of unidenficable polymers. The NMR data of **233** is the same as the one synthesis from **231**. **234**: ¹H NMR δ 2.96 (dd, J = 17, 7 Hz, 1 H), 2.68 (ddd, J = 16, 8.8, 6.4 Hz, 1 H), 2.53 (dd, J = 16.8, 14.4 Hz, 1 H), 2.29 (ddd, J = 15.6, 8.8, 5.2 Hz, 1 H), 2.15 (dd, J = 14.4, 6.4 Hz, 1 H), 2.10– 2.05 (m, 1 H), 1.94–1.80 (m, 2 H), 1.70–1.60 (m, 2 H), 1.35 (s, 3 H), 1.19 (s, 3 H), 1.05 (s, 3 H), 1.04–0.96 (m, 1 H), 1.02 (s, 3 H), 0.91–0.86 (m, 1 H). MS (ESI, MeOH): m/z = 287.3 ([M + Na]⁺).

Estrone pivalate (235) (BH12-062)



To a solution of 100 mg (0.37 mmol) of estrone in 3 mL of CH₂Cl₂ under argon were added 44 mg (0.37 mmol) of pivaloyl chloride and 29.2 mg (0.37 mmol) of pyridine. The solution was stirred for 10 hours, acidified with 1N HCl to pH 5, and extracted three times with 20 mL each of CH₂Cl₂. The combined extracts were washed with brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and diethyl ether as an eluent to give 117 mg (89% yield) of the titled compound. $[\alpha]_D^{22} = +97$ (c 0.1, CHCl₃); ¹H NMR^[143] δ 7.28 (d, J = 8.4 Hz, 1H), 6.84 (d, J = 8.4 Hz, 1H), 6.79 (s, 1H), 2.94 – 2.88 (m, 2H), 2.53 – 2.30 (m, 3H), 2.16 – 1.89 (m, 4H), 1.67 – 1.38 (m, 6H), 1.35 (s, 9H), 0.91 (s, 3H); ¹³C NMR δ 177.0, 172.6, 79.5, 79.1, 62.2, 38.9, 31.8, 28.5, 27.5, 22.8, 19.5, 14.4.

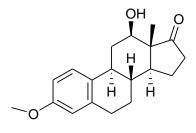




To an aqueous solution of 5 mL of Cu/Au (3:1)-**26** (7.5 μ mol of Cu, 2.5 μ mol of Au, and 0.28 μ mol of **26**), were added 3 mL of 30% H₂O₂, 2 mL of CH₃CN and 70.8 mg (0.2 mmol) of **235**. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine,

dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 42.6 mg (58.2% yield) of **236** and recover 10.9 mg (15.4% yield) of **235**. $[\alpha]_D^{22} = -5.6$ (c 0.1, CHCl₃); ¹H NMR δ 7.25 (dd, J = 9.6, 2.0 Hz, 1H), 6.71 (dd, J = 8.6, 2.8 Hz, 1H), 6.63 (d, J = 2.7 Hz, 1H), 3.98 (dd, J = 10.8, 3.6 Hz, 1H), 2.94–2.87 (m, 2H), 2.59–2.46 (m, 2H), 2.41–2.30 (m, 1H), 2.20–1.98 (m, 3H), 1.78 – 1.72 (m, 1H), 1.63–1.35 (m, 5H), 1.36 (s, 9H), 0.98 (s, 3H); ¹³C NMR δ 222.0, 174.1, 149.3, 136.0, 131.2, 126.0, 117.9, 116.9, 72.9, 52.1, 48.5, 42.8, 38.6, 36.6, 35.8, 32.6, 29.6, 27.0, 21.1, 9.4. HRMS-ESI: m/z [M + H]⁺ calcd for C₂₃H₃₁O₄⁺: 371.2222, found: 371.2209.

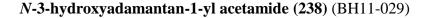
12β-Hydroxy-3-methoxy-estra-1.3.5(10)-triene-17-one (241). (BH12-107)

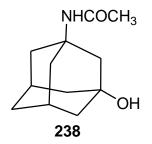


241

To a solution of 18.5 mg (0.05 mmol) of **236** in 1 mL MeOH, add 3.6 mg (0.15 mmol) of LiOH. The resulting solution was stirred at room temperature for 6 hours until no starting material remaining, concentrated under vacuum and dissolve with 10 mL methylene chloride, adjusted the pH to 2 with HCl and extracted 2 times with each 10 mL methylene chloride. The combined organic solution was concentrated under vacuum. To the residue, 2 mL of MeOH, 6.9 mg (0.05mmol) of K_2CO_3 , 7.1 mg (0.05 mmol) of CH_3I was added and heated to reflux for 6 hours. After cooling down, the solution was concentrated under vacuum, dissolve with 10 mL methylene chloride. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to

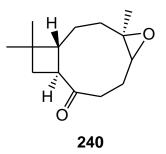
give 10.1 mg (67.6% yield) of **241.** $[\alpha]_D^{22} = -40.2$ (c 0.1, CHCl₃); ¹H NMR^[138] δ 7.16 (dd, J = 8.8, 2.0 Hz, 1H), 6.71 (dd, J = 8.6, 2.8 Hz, 1H), 6.63 (d, J = 2.7 Hz, 1H), 3.98 (dd, J = 10.8, 3.6 Hz, 1H), 3.78 (s, 3H), 2.96–2.87 (m, 2H), 2.59–2.46 (m, 2H), 2.41–2.30 (m, 1H), 2.20–1.98 (m, 3H), 1.78 - 1.72 (m, 1H), 1.63–1.35 (m, 5H), 0.98 (s, 3H); ¹³C NMR δ 222.8, 157.9, 137.7, 131.3, 126.2, 114.2, 111.7, 72.8, 55.5, 52.3, 48.5, 42.4, 37.4, 36.0, 33.3, 29.7, 26.4, 21.6, 8.5.





To an aqueous solution of 4 mL of Cu/Au (3:1)-**26** (9.7 µmol of Cu, 3.2 µmol of Au, and 0.18 µmol of **26**), were added 2 mL of 30% H₂O₂, 4 mL of CH₃CN and 50 mg (0.259 mmol) of **237**. The resulting solution was stirred at 50°C for 7 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 32.1 mg (74% yield) of **238** and recover 6.4 mg (12.8% yield) of **237**. ¹H NMR^[140] δ 5.25–5.17 (bs, 1H), 2.29–2.22 (m, 2H), 2.03–1.95 (m, 2H), 1.95–1.88 (m, 7H), 1.72–1.67 (m, 4H), 1.58–1.53 (m, 2H); ¹³C NMR δ 169.8, 69.6, 54.8, 49.5, 44.5, 40.8, 35.3, 31.0, 25.0.

Kobusone (240) (BH10-080)



To an aqueous solution of 3 mL of Cu/Au (3:1)-**26** (8.5 µmol of Cu, 2.9 µmol of Au, and 0.31 µmol of **26**), were added 2 mL of 30% H₂O₂, 2 mL of CH₃CN and 50 mg (0.23 mmol) of **239**. The resulting solution was stirred at 80°C for 4 days, cooled to 25°C, and extracted three times with 20 mL each of ethyl acetate. The combined extracts were washed with water, and brine, dried (MgSO₄), concentrated, and column chromatographed on silica gel using a gradient mixture of hexane and ethyl acetate as eluents to give 34.0 mg (67.4% yield) of **240** and recover 8.7 mg (17.4% yield) of **239**. $[\alpha]_D^{22} = -131.6$ (c 0.2, CHCl₃); ¹H NMR ^[141] δ 3.01–3.09 (m, 1H), 2.67 (dd, *J* = 4.0, 8.0 Hz, 1H), 2.58-2.49 (m, 2H), 2.42-2.33 (m, 1H), 2.13 (dt, *J* = 2.4, 8.0 Hz, 1H), 2.07-2.03 (m, 1H), 1.94-1.90 (m, 1H), 1.66-1.60 (m, 2H), 1.54-1.46 (m, 1H), 1.46-1.38 (m, 1H), 1.29 (s, 3H), 1.02 (s, 6H), 0.93 (td, *J* = 10.8, 3.2 Hz, 1H), ¹³C NMR δ 214.6, 61.9, 59.2, 52.8, 51.6, 39.2, 37.9, 35.5, 34.7, 29.6, 26.7, 25.0, 22.4, 16.4; MS (ESI, MeOH): m/z = 245.3 ([M + Na]⁺).

Chapter 3 - Synthesis of

6-(Dimethylamino)-2-phenylisoindolin-1-one derivative.

3.1 Background and Significance

Bioluminescence is the production and emission of light by a living organism. It is a form of chemiluminescence. Bioluminescence occurs widely in marine vertebrates and invertebrates, as well as in some fungi, microorganisms including some bioluminescent bacteria and terrestrial [144] invertebrates such as fireflies. Bioluminescence is widely used in high throughput screening (HTS) as a light based detection method to assay biological activities of molecules, proteins or genes against a defined set of enzymes or receptor targets.^[145] In a general sense, the principal chemical reaction in bioluminescence involves some light-emitting molecule and an enzyme, generally called the luciferin and the luciferase, respectively. Luciferase is a generic term for the class of oxidative enzymes that produce bioluminescence, and is usually distinguished from a photo protein. The luciferase requires other cofactors such as calcium or magnesium ions, and sometimes also the energy-carrying molecule adenosine triphosphate (ATP). ^[144] Hence firefly luciferase is used as a sensor of the ATP content in cells as a measure of cell viability, and in biochemical assays to measure ATP-dependent enzyme reactions such as kinases.^[146] Besides that, firefly luciferase has been commonly used as a reporter in cells expressing a luciferase gene or its enzymatic activity under the control of a promoter of interest to assess its transcriptional activity.^[147, 148] However, it has been found that some molecules can directly inhibit firefly reporter enzyme in cells, independently from promoter-specific transcriptional activity.^[149] In contrast, some inhibitors have found to form stable firefly enzyme-inhibitor complexes which are more resistant to degradation than the free firefly enzyme.^[150, 151] This leads to a reporter enzyme accumulation and signal activation in

cells, independent of effects on transcription/translation.^[150, 151] These phenomena give rise to false results in cell-based HTS assays.^[150] Therefore, to avoid these misleading activities, study of firefly luciferase inhibitors and their mechanisms of inhibition are highly relevant.

Crystal structure of firefly luciferase has shown two distinct domains, a large N-terminal domain (residues 1–436) and a small C-terminal domain (residues 440–550), separated by a wide cleft.^[152] It is suggested that, seven residues (Gly200, Lys206, Glu344, Asp422, Arg437, Gly446 and Glu455) play a crucial role in the binding of ATP and in adenylate formation.^[152] Hence, the active site of luciferase is proposed to locate on the surface of both domains facing each other across a large cleft.^[152] Further, it is believed that upon substrate binding, the two domains move together to form the active center.^[152] Site-directed mutation studies have shown that Lys529 is crucial for effective substrate orientation and for transition state stabilization which lead to efficient adenylate formation.^[153] (**Figure 15**)

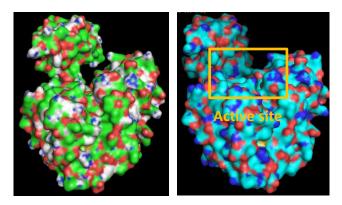


Figure 15. Surface view of (a) firefly luciferase without its substrate (Protein Data Bank accession number 1LCI)^[152] (b) firefly luciferase in complex with bromoform (Protein Data Bank accession number 1BA3).^[154]

Natural substrate of luciferase enzyme is the D-isomer of firefly luciferin (D-LH₂) which is chemically defined as (S)-2-(6'-hydroxy-2'-benzothiazolyl)-2-thiazoline-4-carboxylic acid (**Figure 16**).^[155] The bioluminescent chemical reaction catalyzed by luciferase is a two-step

process.^[146, 155] In the first step of the bioluminescent reaction, D-luciferyl-adenylate (D-LH₂-AMP) intermediate is formed by the reaction of D-luciferin (D-LH₂) and ATP in the presence of $Mg^{2+,[146, 155]}$ Oxidation of luciferyl-adenylate (D-LH₂-AMP) intermediate with molecular oxygen in the following step produces AMP, oxyluciferin and light.^[146, 155] (Scheme **20**, eq 1–2)

It is reported that the primary structure of firefly luciferase has a high sequence similarity to a long-chain acyl-CoA synthetase.^[156] Therefore, besides the light emitting reactions, firefly luciferase catalyzes fatty acyl-CoA synthesis as well.^[157] First, long-chain fatty acids are adenylated in the presence of ATP and Mg²⁺, and the subsequent step is the thioesterification with CoA.^[157, 158] (Scheme 20, eq 3–4) It is suggested that luciferase catalyzes the adenylation of fatty acids via carboxylic acid moiety in a mechanism similar to the adenylation step of the bioluminescence reaction of luciferase.^[159] As reported, long chain fatty acids (C12-C20) can be a competitive inhibitors of firefly luciferase against luciferin in micromolar concentration level.^[158, 160]

luciferase + D-luciferin + ATP-Mg²⁺
$$\implies$$
 luciferase.D-luciferyl-AMP + PPi-Mg²⁺ (1)

luciferase.D-luciferyl-AMP +
$$O_2$$
 \longrightarrow luciferase + AMP + CO_2 + oxyluciferin + hv (2)

luciferase + fatty acid + ATP-Mg²⁺
$$\implies$$
 luciferase.fatty acyl-AMP + PPi-Mg²⁺ (3)

$$luciferase.fatty acyl-AMP + CoA \implies fatty acyl-CoA + AMP + luciferase$$
(4)

$$luciferase.dehydroluciferyl-AMP + CoA \longrightarrow luciferase + AMP + luciferyl-CoA$$
(5)

Scheme 20. Luciferase Catalyzed Reactions.

Common inhibitors that inhibit firefly luciferase activity are found to be substrates related compounds, intermediates or products of the luciferase catalyzed reaction and fatty acids.^[155] Substrates related compounds such as L-luciferin, dehydroluciferin, intermediates such as dehydroluciferyl-adenylate (L-AMP), and products such as pyrophosphate (PPi) and oxyluciferin

were found to inhibit bioluminescent light emission in a competitive or noncompetitive manner.^[161-165] However, coenzyme A (CoA) was found to prevent this inhibition by reacting with dehydroluciferyl-adenylate (L-AMP).^[166] CoA reaction with L-AMP results free luciferase and luciferyl-CoA (L-CoA) which is found to be a weaker inhibitor in the bioluminescence reaction.^[160, 166] (Scheme 20, eq 5)

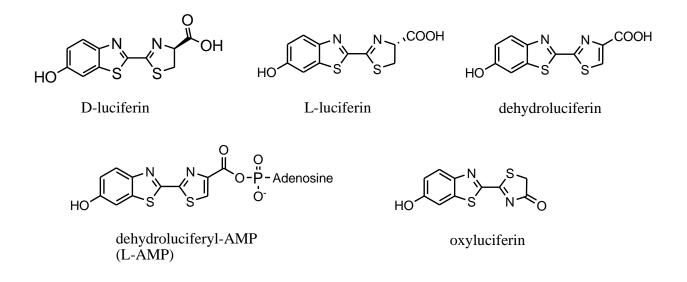
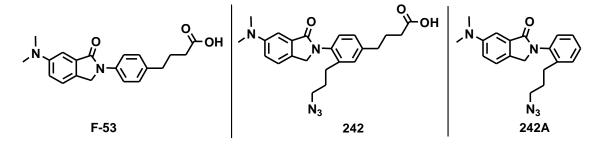


Figure 16. Structure of D-luciferin and Firefly Luciferase Inhibitors.

Nakagomi and coworkers have reported an aromatic carboxylic acid, F-53 which inhibits the enzymatic activity of firefly luciferase by covalently binding to a regulatory lysine residue (Lys529) *via* an amide bond formation.^[167] Based on experimental results, they proposed that the carboxylic acid of F-53 is first activated to its CoA-thioester derivative by luciferase via its acyl-CoA synthetase activity.^[167] This F-53-CoA derivative is proposed to inactivate luciferase *via* acylation of Lys-529 by an unknown cellular acetyltransferase.^[167] With knowledge of above mentioned results, our research efforts focused on synthesizing two analogous probes of F-53 possessing ortho 3-azidopropyl substituent on the phenyl ring with (**242**) and without (**243**) butyric acid group (**Scheme 21**). We hypothesized that compound **242** acts on luciferase by

competing for the binding site of the luciferin substrate which is involved in the light emission. Therefore, these two probe molecules will be used to study the mechanism of luciferase inhibition further for understanding the effects of carboxylic acid-containing drugs.



Scheme 21. Structure of F-53, 242 and 242A.

3.2 Synthetic Route and Discussions

Before the synthesis of compound 242, similar compounds F-53 and 242A were successfully made by Dr. Ai Ito in Research Foundation ITSUU Laboratory and Dr. Medha Gunaratna in Dr. Hua's Lab, respectively. With the general idea of the multi-step synthetic routes for F-53 and 242A, the synthesis of 242 was designed. However, due to the side chain on the phenyl ring, the synthesis of 242 becomes more complicated. Base on the retrosynthetic analysis, 242 can be disconnected into three parts, the isoindolinone, the aniline derivative and δ -side chain. (Figure 17) The final synthetic route to obtain 242 was shown in Scheme 22.

For the first part, O-toluic acid, **243** in conc. sulfuric acid was treated with a solution of potassium nitrate in conc. sulfuric acid following a reported procedure.^[168] (**Scheme 22**) In this reaction, potassium nitrate reacts with sulfuric acid to form nitric acid and potassium bisulfate. Sulfuric acid being a stronger acid than nitric acid facilitates generation of NO^{2+} in situ. Importantly, to avoid the formation of di-nitrated product, potassium nitrate in conc. sulfuric

acid should be added dropwise to keep the nitronium ion concentration low and the reaction should be carried out at 0 °C to slow down the reaction. During workup, since mixing concentrated sulfuric acid with water gives off much heat, reaction mixture should be added to ice rather than water. The resulting product was a mixture of 2-methyl-5-nitrobenzoic acid, **244** (85%) and 2-methyl-3-nitrobenzoic acid (15%). The compound can be purified by recrystallization through 2:1 ether: dichloromethane.

Since both products are very polar and they both have very similar Rf values, the mixture was directly treated with thionyl chloride in methanol.^[169] Reaction mixture was refluxed to give a mixture of corresponding methyl esters **245** and its isomer in a quantitative yield. Resultant compound, **245** was able to purify using hexane *via* recrystallization. Benzylic bromination of compound **245** using *N*-bromosuccinimide and a radical initiator, benzoyl peroxide afforded compound **246**,^[168] as a key precursor for **242**.

The para-side chain synthesis was start from 3-butyn-1-ol (**247**). Because of the active hydroxyl group, TBS protection of alcohol was needed. With the presence of imidazole as a base in methylene chloride, TBSCl was added into **247** to yield **248**.^[170]

2-Amino-5-iodobenzoric acid (249), as a cheap amino acid, was used as the substrate for the aniline derivative part of 242. Firstly, the carboxylic acid was reduced by LiAlH₄ in THF to give corresponding alcohol 250. The coupling of 250 with 246 in EtOH at reflux temperature for 24 h resulted its condensation product 251.^[167] Low temperature and less reaction times resulted uncyclized product. From the appearance of uncyclized by-product, the amino group attack Br with a S_N^2 reaction was much faster than the amidation. The isolated uncyclized product was cyclized again in reflux ethanol to give 251. At the same time, the alcohol on ortho position was also attacked the Br to give ether type product in ~10% yield.

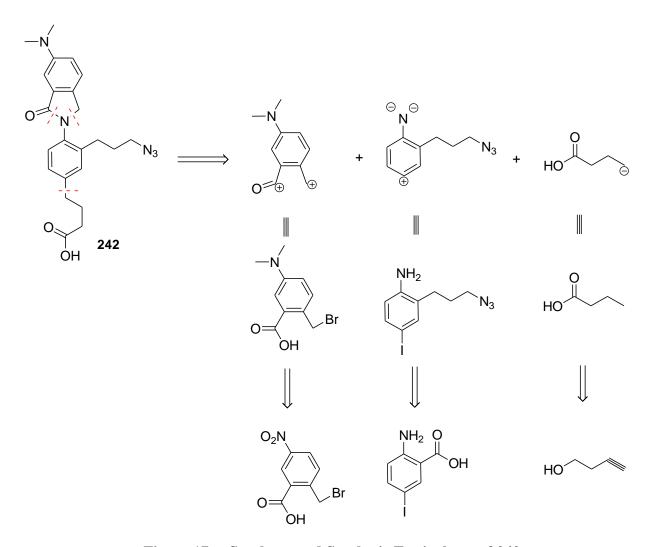
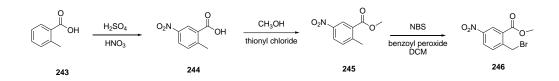


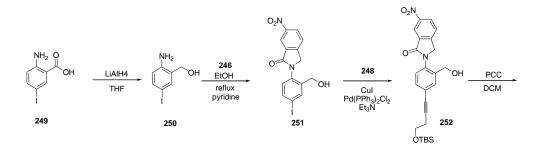
Figure 17. Synthons and Synthetic Equivalents of 242.

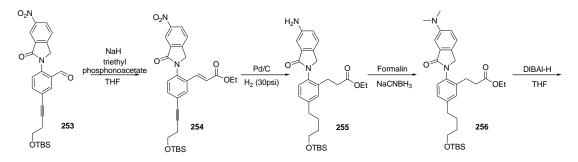
The connection between **251** and **248** was constructed by Sonogashira Reaction, coupling the benzyl iodide and C-C triple bond. The Suzuki Coupling was tried with both pinacol-borane and catechol-borane, the alkenylboron was not easy to make and yield of the reaction was not consistence (32-68% yield). Therefore, a milder route, Sonogashira Coupling was carried out with the catalysis of Pd(PPh₃)₂Cl₂ and cuprous iodide as a co-catalyst in trimethylamine as solvent. The reaction was needed to be done in a dry box and gave only desired organic product **252** in a quantitative yield.

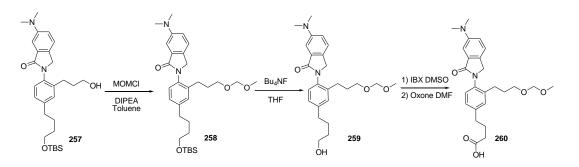




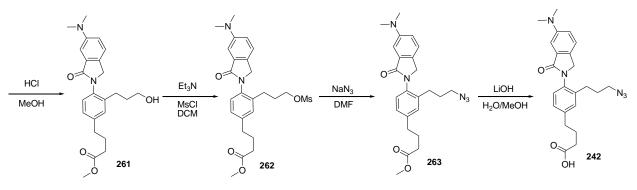








(To be continued)



Scheme 22. The Synthetic Route for Compound 242.

Compound **252** was oxidized by PCC at room temperature in dry methylene chloride and gave quantitative yield of corresponding aldehyde **253**. The work up process was the key to the reaction since large amount of sticky PCC may cover the product and make a lower yield. After the filter through Celite, the sonication of Celite in methylene chloride was necessary, even though partial PCC was dissolve back into the solution. The adding of molecular sieve into the reaction was also helpful to avoid the sticky PCC. Compound **253** was finally purified by chromatography column.

With the optimized Horner-Wadsworth-Emmons reaction, **253** was reacted with stabilized phosphorus ylide (phosphonate carbanion) and leaded to **254** with *E*-selectivity. Sodium Hydride was used to make the phosphorus ylide with triethylphosphonoacetate.^[171] The product **254** was hydrogenated with the catalysis of Pd/C to reduce C-C double bond, and C-C triple bond and NO₂ group at the same time. Reductive methylation of aryl amino group of compound **255** was achieved using formaldehyde and sodium cyanoborohydride by reductive amination to obtain *N*, *N*-dimethyl product **256**, followed with the reduction of ester group to alcohol with DIBAI-H with 95% yield.

Compound **257** was protected with MOMCl with DIPEA as an amine and remove TBS protecting group with TBAF in THF for the preparation of functionalization on para-side chain.

The terminal alcohol was oxidized to carboxylic acid (**260**) with two steps, (1) oxidation to aldehyde with IBX/DMSO and (2) oxidation to carboxylic acid with oxone.^[172] To avoid the mesylation of the carboxylic acid, the methyl ester was made with HCl/MeOH. At the same time, MOM was removed in the same reaction to give **261**. The hydroxyl group of compound **261** was mesylated using MsCl/Et₃N and followed by the azidation with sodium azide to give compound **263** in a quantitative yield. Finally, the methyl ester was hydrolysis with LiOH and yield the final product **242**.

3.3 Future Work

Both probe molecules **242** and **243** were sent to Dr. Madoka Nakagomi's lab for biology studies. They will be used for identifying target protein(s) in the firefly luciferase inhibition through click chemistry. Probe molecule **242** consists of three elements which are important in the activity-based protein profiling studies, (1) a reactive carboxylic group that can covalently reacts with a residue in the active site of the target enzyme (2) a spacer for selectivity and (3) an azide group that can undergo Cu(I)-catalyzed [3+2] dipolar cycloaddition with an alkyne bearing a fluorescent detection tag.^[173-175] Probe molecule **243**, which does not have a butyric acid group will be used as the negative control.

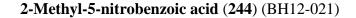
3.4 Synthetic Experimental Procedures

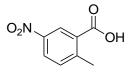
3.4.1 General

¹H NMR spectra (400 MHz) and ¹³C NMR spectra (100 MHz) were measured from a solution in CDCl₃ unless otherwise mentioned. The chemical shift data for each signal on ¹H NMR are given in units of δ relative to TMS ($\delta = 0$) or CHCl₃ ($\delta = 7.26$). For ¹³C NMR spectra, the chemical shifts are recorded relative to CDCl₃ ($\delta = 77.0$). Low-resolution mass spectra were

taken from an API 2000-triple quadrupole ESI-MS/MS mass spectrometer. High-resolution mass spectra were obtained using a Waters LCT Premier time of flight mass spectrometer. IR spectra were measured directly in solid form from Agilent Cary 630 FTIR. Chemicals were purchased from Fisher Scientific Co., Aldrich Chemical Co., Chem-Impex International, and VWR International.

3.4.2 Representative Synthesis

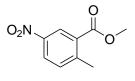




244

To a solution of **243** (15 g, 110.17 mmol) in conc. sulfuric acid (90 mL), a solution of potassium nitrate (15 g, 143.22 mmol) in conc. sulfuric acid (90 mL) was added dropwise over 1 h at 0 °C. The reaction mixture was added to a 100 g of ice chips and stirred vigorously until all ice melted. The resulting precipitate was re-dissolved in diethyl ether. The ether layer was washed with water and dried with anhydrous sodium sulfate. The solution was concentrated under vacuum to obtain 18.62 g (93.2% yield) of a mixture of compound **244** (85%) and 2-methyl-3-nitrobenzoic acid (15%) as a white solid. The compound can be purified by recrystallization through 2:1 ether: dichloromethane. ¹H NMR^[168] δ ppm 8.95 (s, 1H), 8.30 (d, J = 8.6 Hz, 1H), 7.50 (m, 1H), 2.80 (s, 3H); ¹³C NMR δ ppm 166.7, 146.8, 145.2, 132.5, 131.2, 126.1, 125.2, 20.9.

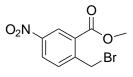
2-Methyl-5-nitrobenzoic acid methyl ester (245). (BH12-022)



245

To a mixture of **244** (85%) and 2-methyl-3-nitrobenzoic acid (15%) (9.05 g, 50 mmol) in methanol (120 mL), thionyl chloride (7.26 mL, 100 mmol) was added dropwise at 0 °C and it was allowed to reflux for 4 h at 70 °C. The solution was concentrated under vacuum, then re-dissolved in ethyl acetate (150 mL). A solution of saturated sodium bicarbonate (70 mL) was added to adjust the pH to 8. The product was extracted from the aqueous layer using ethyl acetate twice. The combined organic layer was washed with brine (70 mL) and dried over anhydrous sodium sulfate. The solvent was evaporated under vacuum and recrystallized with hexane to obtain 8.3 g (99% yield based on **244**) of compound **245** as a white solid; mp. 66~67 °C; ¹H NMR^[176] δ ppm 8.78 (d, *J* = 2.73 Hz, 1H), 8.25 (dd, *J* = 2.54, 8.40 Hz, 1H), 7.44 (d, *J* = 8.20 Hz, 1H), 3.96 (s, 3H), 2.73 (s, 3H); ¹³C NMR δ ppm 165.8, 147.8, 145.9, 132.7, 130.4, 126.0, 125.6, 52.3, 21.8.

Methyl 2-(bromomethyl)-5-nitrobenzoate (246) (BH12-023)

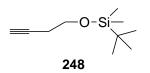


246

A mixture of **245** (12.6 g, 40 mmol) and *N*-bromosuccinimide (18.4 g, 44 mmol) in dichloroethane (200 mL), in the presence of a catalytic amount of benzoyl peroxide (780 mg) was refluxed for 15 h under argon. The reaction mixture was neutralized with saturated sodium bicarbonate and extracted with dichloromethane. The organic layer was dried over anhydrous sodium sulfate. The solvent was evaporated under reduced pressure and the residue was purified by silica gel column chromatography (hexanes: ethyl acetate = 9:1) to obtain 10.90 g

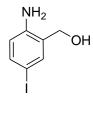
(62.1% yield) of compound **246** as a white solid; mp. 77~78 °C; ¹H NMR^[177] δ 8.82 (d, J = 1.95 Hz, 1H), 8.34 (dd, J = 8.40, 1.95 Hz, 1H), 7.69 (d, J = 8.59 Hz, 1H), 5.01 (s, 2H), 4.02 (s, 3H); ¹³C NMR δ ppm 165.0, 147.4, 146.1, 133.0, 130.3, 126.9, 126.4, 53.0, 29.3.

3-(tert-Butyldimethylsiloxy)-1-butyne (248) (BH12-026)



To a solution of 247 (2.8 g, 40 mmol) in 200 mL anhydrous methylene chloride, imidazole (5.8 g, 80 mmol) and TBSCl (9.68 g, 60 mmol) were added and stirred under argon at room temperature overnight. The reaction solution was washed with water (2 x 50 mL) and brine (50 mL), dried over anhydrous Na₂SO₄ and concentrate under rotary evaporator and vacuum. The residue was purified by silica gel column chromatography (hexanes: ethyl acetate = 5:1) to obtain 6.9 g (96.5%) yield) of compound **248** as a colorless oil. ¹H NMR^[178] δ 3.74 (t, J = 7.2 Hz, 2H), 2.39 (m, 2H), 1.96 (s, 1H), 0.10 (s, 9H), 0.07 (s, 6H); ¹³C NMR δ ppm 82.0, 70.1, 61.8, 26.1, 24.3, 18.6, -5.4.

(2-Amino-5-iodophenyl)methanol (250) (BH12-013)

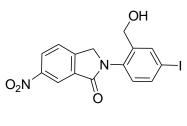


250

To a cold (0 °C) solution of 3 g (11.4 mmol) of 2-amino-5-iodobenzoic acid (249) in 100 mL of dry THF, 0.44 g (11.4 mmol) of LiAlH₄ was added in portion over 20 minutes and the reaction mixture was stirred at room temperature for 3 hours to give a yellow solution. Progress of the reaction is monitored by TLC until no starting material remains. To it, H₂O (3 mL) and 10% 212

aqueous NaOH solution (3 mL) were added to the solution, filtered to remove Al(OH)₃ and washed with of ethyl acetate (3 x 50 mL). The filtrate was concentrated and the residue was purified by silica gel column chromatography using a gradient mixture of hexane and diethyl ether as an eluent to give 1.96 g (70% yield) of the titled compound as a light yellow solid; mp. 181~183 °C; ¹H NMR^[179] δ 7.40–7.35 (m, 2H), 6.48 (d, *J* = 8 Hz, 1H), 4.61 (d, *J* = 2.8 Hz, 2H), 4.21 (broad, 1H); ¹³C NMR δ ppm 147.2, 138.0, 136.4, 128.6, 117.5, 76.6, 62.0.

2-(2-(Hydroxymethyl)-4-iodophenyl)-6-nitroisoindolin-1-one (251) (BH12-024)

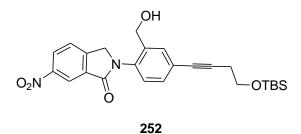




To a solution of **250** (4 g, 16 mmol) and **246** (4.37 g, 16 mmol) in absolute ethanol (200 mL) under argon, dry pyridine (1.64 g, 20.8 mmol) was added. The reaction mixture was refluxed at 80 °C for 30 h. After cooling to room temperature, the reaction was neutralized with saturated sodium bicarbonate. The reaction mixture was extracted with ethyl acetate. The combined organic layer was washed with saturated sodium chloride (100 mL) and dried over anhydrous sodium sulfate. The solvent was evaporated under vacuum and the residue was purified by silica gel column chromatography (hexanes:ethyl acetate = 1:1) to obtain 1.92 g (52% yield) of compound **251** as a yellow oil. ¹H NMR δ 8.80 (s, 1H), 8.49 (dd, *J* = 2.15, 8.40 Hz, 1H), 7.68 (d, *J* = 8.59 Hz, 1H), 7.32–7.42 (m, 3H), 4.62 (m, 4H); ¹³C NMR δ ppm 166.7, 151.8, 148.6, 139.2, 134.2, 130.9, 129.3, 128.1, 128.0, 126.4, 124.9, 119.8, 88.6, 61.2, 53.1. HRMS-ESI: m/z [M + H]⁺ calcd for C₁₅H₁₂IN₂O₄: 410.9842 found: 410.9839.

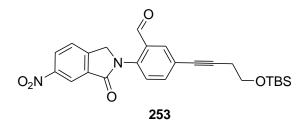
2-(4-(4-(tert-Butyldimethylsilyloxy)but-1-ynyl)-2-(hydroxymethyl)phenyl)-6-nitroisoindolin

-1-one (252) (BH12-029)



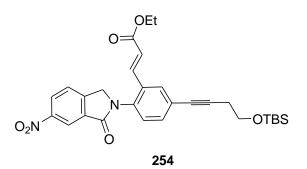
To a yellow solution of 251 (1.9 g, 2.5 mmol) in 200 mL triethylamine, CuI (17.6 mg, 0.09 mmol) and bis(triphenylphosphine)palladium chloride (64.9 mg, 0.09 mmol) were added under nitrogen in a dry box. The reaction mixture was stirred under argon for 20 minutes and (but-3-ynyloxy)(tert-butyl)dimethylsilane (248) (937 mg, 5.097 mmol) was added into it to give a green solution. The solution was stirred under nitrogen at room temperature overnight, concentrated under vacuum, and re-dissolve in 100 mL of ethyl acetate and 50 mL of H₂O. The mixture was extracted with ethyl acetate (2 x 100 mL) and the combined organic layer was washed with saturated sodium chloride (50 mL) and dried over anhydrous sodium sulfate. The solvent was evaporated under vacuum and the residue was purified by silica gel column chromatography (hexanes: ethyl acetate = 1:1) to obtain 1.87 g (86.6% yield) of compound 252 as a yellow oil. ¹H NMR δ 8.80 (s, 1H), 8.50 (dd, J = 2.15, 8.40 Hz, 1H), 7.87 (d, J = 4 Hz, 1H) 7.60 (d, J = 8 Hz, 1H), 7.31 (d, J = 4.4 Hz, 1H), 7.14 (s, 1H), 4.62 (m, 4H), 3.79 (t, J = 7.2Hz, 2H), 2.59 (t, J = 4.8 Hz, 2H), 0.90 (s, 9H), 0.09 (s, 6H) ; ¹³C NMR δ ppm 166.4, 153.2, 140.3, 137.2, 135.6, 133.1, 129.8, 128.1, 127.3, 125.4, 123.0, 120.8, 118.1, 116.0, 82.4, 63.6, 61.2, 54.3, 27.6, 24.0, 19.0 (3C), -5.1(2C). HRMS-ESI: $m/z [M + H]^+$ calcd for $C_{25}H_{31}N_2O_5Si$: 467.2002 found: 467.2013.

5-(4-(tert-Butyldimethylsilyloxy)but-1-ynyl)-2-(6-nitro-1-oxoisoindolin-2-yl)benzaldehyde (253). (BH12-031)



To a solution of **252** (1.9 g, 4.07 mmol) in 50 mL anhydrous methylene chloride under argon, activated 4 Å molecular sieve (2 g) and pyridinium chlorochromate (1.05 g, 4.88 mmol) were added and stirred at room temperature for 3 hours to give a dark green solution. The solution was filtered through Celite and washed with 100 mL of methylene chloride. The filtrate was concentrated under an evaporator and vacuum and purified by silica gel column chromatography (hexanes:ethyl acetate = 1:1) to obtain 1.82 g (96% yield) of compound **253** as a yellow oil. ¹H NMR δ 10.36 (s, 1H), 8.80 (s, 1H), 8.49 (dd, *J* = 2.15, 8.40 Hz, 1H), 8.38 (d, *J* = 4.8 Hz, 1H), 7.70 (d, *J* = 8.0 Hz, 1H), 7.62 (s, 1H), 7.40 (d, *J* = 4.0 Hz 1H), 4.62 (s, 2H), 3.79 (t, *J* = 7.2 Hz, 2H), 2.60 (t, *J* = 4.8 Hz, 2H), 0.90 (s, 9H), 0.08 (s, 6H); ¹³C NMR δ ppm 195.0, 162.1, 153.1, 141.9, 139.6, 139.2, 132.1, 131.4, 128.6, 120.3, 118.6, 117.6, 114.2, 102.6, 88.2, 81.2, 61.8, 60.4, 26.0, 24.0, 18.8 (3C), -5.1 (2C). HRMS-ESI: m/z [M + H]⁺ calcd for C₂₅H₂₉N₂O₅Si: 465.1846 found: 465.1862.

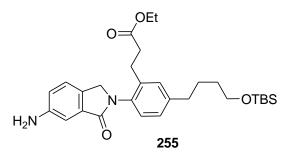
Ethyl-3-(5-(4-(tert-butyldimethylsilyloxy)but-1-ynyl)-2-(6-nitro-1-oxoisoindolin-2-yl)phenyl)acrylate (254) (BH12-033)



To a cold (0°C) solution of triethyl phosphonoacetate (878 mg, 3.92 mmol) in 30 mL anhydrous THF under argon, NaH (98 mg, 3.92 mmol) was added in portion and the solution was stirred at 215

room temperature for 2 hours to give a colorless solution. To it, a solution of **253** (1.82 g, 3.92 mmol) in 30 mL THF was added via cannula and stirred under argon at room temperature for another 2 hours. The reaction was quenched by 2 mL of H₂O and extracted three times with 50 mL each of methylene chloride. The combined organic layer was washed with brine (20 mL), dried over anhydrous Na₂SO₄, concentrated under vacuum, and purified by silica gel column chromatography (hexanes:ethyl acetate = 1:2) to obtain 2.0 g (96% yield) of compound **254** as a yellow oil. ¹H NMR δ 8.80 (s, 1H), 8.49 (dd, *J* = 2.15, 8.40 Hz, 1H), 7.70 (m, 2H), 7.51 (s, 1H), 7.33 (d, *J* = 8.0 Hz 1H), 6.67 (s, 1H), 6.35 (d, *J* = 8 Hz, 1H), 4.62 (s, 2H), 4.23 (q, *J* = 6.8, 2H), 3.78 (t, *J* = 7.2 Hz, 2H), 2.58 (t, *J* = 4.8 Hz, 2H), 1.30 (t, *J* = 7.2 Hz, 3H), 0.90 (s, 9H), 0.07 (s, 6H); ¹³C NMR δ ppm 167.5, 163.2, 152.4, 138.9, 137.1, 134.2, 133.2, 130.8, 129.6, 125.8, 124.3, 122.6, 121.4, 116.8, 114.8, 111.0, 88.2, 81.6, 61.8, 60.6, 34.8, 27.2, 25.0, 23.9, 19.0 (3C), -5.0 (2C). HRMS-ESI: m/z [M + H]⁺ calcd for C₂₉H₃₅N₂O₆Si: 535.2264 found: 535.2259.

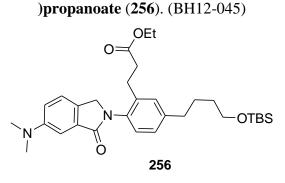
Ethyl-3-(2-(6-amino-1-oxoisoindolin-2-yl)-5-(4-(tert-butyldimethylsilyloxy)butyl)phenyl)pro panoate (255). (BH12-035)



To a solution of **254** (2.0 g, 3.76 mmol) in 30 mL of ethyl acetate, 50 mg of 10% palladium/carbon was added and shake under 30 psi hydrogen gas overnight. The reaction mixture was filtered through Celite and concentrated under vacuum to give 1.91 g (99% yield) of compound **255** as a yellow oil. This molecule was used in the subsequent step without purification. ¹H NMR δ 7.30–6.90 (m, 6H), 4.62 (s, 2H), 4.20 (q, *J* = 6.8, 2H), 3.64 (t, *J* = 7.2 Hz, 2H), 2.89 (m, 2H), 2.62 (m, 4H), 1.70–1.55 (m, 4H), 1.30 (t, *J* = 7.2 Hz, 3H), 0.90 (s,

9H), 0.07 (s, 6H); ¹³C NMR δ ppm 168.0, 165.0, 147.6, 138.4, 136.1, 131.4, 130.6, 128.4, 128.0, 127.2, 124.0, 119.6, 116.2, 109.8, 63.0, 61.2, 34.0, 31.2, 29.9, 28.6, 27.4, 26.6, 25.4, 24.8, 18.0 (3C), -5.4 (2C). HRMS-ESI: m/z [M + H]⁺ calcd for C₂₉H₄₃N₂O₄Si: 511.2992 found: 511.3001.

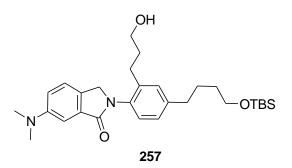
Ethyl



To a solution of **255** (1.91 g, 3.76 mmol) in MeOH, formalin (37% W/W, 6.11 g, 37.6 mmol) and sodium cyanoborohydride (1.42 g, 22.5 mmol) were added, and the solution was stirred for 24 h at room temperature. The reaction solution was neutralized with saturated sodium bicarbonate and extracted with ethyl acetate three times. The combined organic layer was washed with saturated sodium chloride (70 mL) and dried over anhydrous sodium sulfate. The solvent was evaporated under reduced pressure and the residue was purified by silica gel column chromatography (hexanes:ethyl acetate = 1:1) to obtain 1.92 g (94.9% yield) of compound **256** as a yellow oil. ¹H NMR δ 7.35–6.90 (m, 6H), 4.58 (s, 2H), 4.20 (q, *J* = 6.8, 2H), 3.61 (t, *J* = 7.2 Hz, 2H), 2.99 (s, 6H), 2.82 (m, 2H), 2.58 (m, 4H), 1.70–1.55 (m, 4H), 1.30 (t, *J* = 7.2 Hz, 3H), 0.86 (s, 9H), 0.04 (s, 6H); ¹³C NMR δ ppm 168.2, 165.0, 148.2, 140.1, 138.2, 132.3, 131.9, 129.8, 129.6, 126.7, 120.9, 115.6, 113.0, 106.4, 62.8, 60.1, 41.8(2C), 34.6, 32.0, 29.6, 27.3, 26.9, 26.1, 25.8, 24.9, 18.0 (3C), -5.2 (2C). HRMS-ESI: m/z [M + H]⁺ calcd for C₃₁H₄₇N₂O₄Si: 539.3305 found: 539.3305.

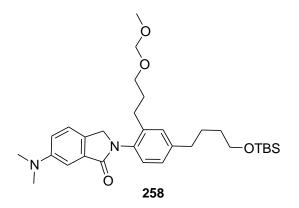
2-(4-(4-(tert-Butyldimethylsilyloxy)butyl)-2-(3-hydroxypropyl)phenyl)-6-(dimethylamino)is

oindolin-1-one (257) (BH12-047)



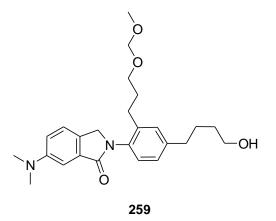
To a cold (-78 °C) solution of **256** (1.92 g, 3.5 mmol) in 40 mL anhydrous THF under argon, 9 mL (9 mmol) of diisobutylaluminum hydride (1M in toluene) solution was added dropwise. After adding, the reaction was stirred at -30 °C for 2 hours until no starting material remained, quenched with 2 mL of H₂O, filtered and extracted the filtrate three times with ethyl acetate (60 mL each). The combined organic layer was washed with brine, dried over anhydrous sodium sulfate and concentrate under vacuum. The residue was purified by silica gel column chromatography (hexanes:ethyl acetate = 1:2) to obtain 1.68 g (95% yield) of compound **257** as a yellow oil. ¹H NMR δ 7.35–6.90 (m, 6H), 4.60 (s, 2H), 3.61 (m, 4H), 2.99 (s, 6H), 2.69 (t, *J* = 6.4 Hz, 2H), 2.55 (t, *J* = 8.4 Hz, 2H), 1.86 (m, 2H), 1.70–1.40 (m, 4H), 0.88 (s, 9H), 0.03 (s, 6H); ¹³C NMR δ ppm 168.8, 149.8, 139.1, 136.5, 131.6, 129.0, 128.7, 128.2, 127.9, 126.5, 122.2, 116.0, 105.1, 63.8, 60.1, 53.8, 50.4, 39.8 (2C), 37.5, 35.2, 28.4, 27.3, 26.2, 18.4 (3C), -5.1 (2C). HRMS-ESI: m/z [M + H]⁺ calcd for C₂₉H₄₅N₂O₃Si: 497.3199 found: 497.3221.

2-(4-(4-(tert-Butyldimethylsilyloxy)butyl)-2-(3-(methoxymethoxy)propyl)phenyl)-6-(dimeth ylamino)isoindolin-1-one (258) (BH12-051)



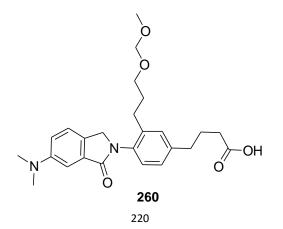
To a solution of **257** (1.68 g, 3.38 mmol) in 100 mL anhydrous methylene chloride under argon, *N*,*N*-diisopropylethylamine (1.09 g, 8.45 mmol) and chloromethyl methyl ether (0.544 g, 6.76 mmol) were added. The reaction was reflux for 12 hours, diluted with 100 mL methylene chloride, washed with 10% citric acid (50 mL), saturated NaHCO₃ solution (50 mL), and brine (50 mL), dried over anhydrous sodium sulfate and concentrated under vacuum. The residue was purified by silica gel column chromatography (hexanes:ethyl acetate = 1:1) to obtain 1.70 g (93% yield) of compound **258** as a yellow oil. ¹H NMR δ 7.35–6.90 (m, 6H), 5.22 (s, 2H), 4.57 (s, 2H), 3.97 (t, *J* = 6.4 Hz, 2H), 3.61 (m, 5H), 2.99 (s, 6H), 2.59 (m, 4H), 1.90–1.55 (m, 6H), 0.87 (s, 9H), 0.02 (s, 6H); ¹³C NMR δ ppm 168.7, 149.7, 139.1, 136.5, 131.9, 129.1, 128.8, 128.3, 127.9, 126.5, 122.2, 116.1, 105.1, 99.9, 63.8, 62.1, 60.6, 53.8, 50.4, 39.8 (2C), 35.1, 32.2, 28.4, 27.3, 26.2, 18.5 (3C), -4.9 (2C). HRMS-ESI: m/z [M + H]⁺ calcd for C₃₁H₄₉N₂O₄Si: 541.3462 found: 541.3459.

6-(Dimethylamino)-2-(4-(4-hydroxybutyl)-2-(3-(methoxymethoxy)propyl)phenyl)isoindolin-1-one (259). (BH12-055)



To a solution of **258** (1.70 g, 3.18 mmol) in 20 mL THF under argon, 3.18 mL of 1M tetra-*n*-butylammonium fluoride THF solution was added. The reaction was stirred under room temperature for 4 hours until no starting material remained, washed with water (2 x 20 mL) and brine (20 mL), dried over anhydrous sodium sulfate and concentrated under vacuum to obtain 1.34 g (99% yield) of compound **259** as a yellow oil. This material was used in the subsequent step without further purification. ¹H NMR δ 7.35–6.90 (m, 6H), 5.23 (s, 2H), 4.60 (s, 2H), 3.97 (t, *J* = 6.4 Hz, 2H), 3.61 (m, 5H), 2.99 (s, 6H), 2.58 (m, 4H), 1.88–1.56 (m, 6H); ¹³C NMR δ ppm 169.9, 151.5, 140.1, 137.5, 133.2, 129.9, 129.5, 128.7, 128.2, 127.5, 123.2, 117.1, 106.3, 99.9, 63.8, 62.1, 53.8, 50.2, 41.0 (2C), 36.4, 35.1, 31.6, 28.8, 25.1. HRMS-ESI: m/z [M + H]⁺ calcd for C₂₅H₃₅N₂O₄: 427.2597 found: 427.2612.

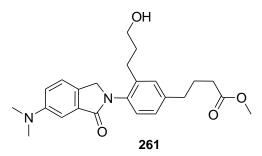
4-(4-(6-(Dimethylamino)-1-oxoisoindolin-2-yl)-3-(3-(methoxymethoxy)propyl)phenyl)butan oic acid (260). (BH12-058)



To a solution of **259** (1.065 g, 2.5 mmol) in 20 mL DMSO under argon, IBX (1.05 g, 3.75 mmol) was added and stirred at room temperature overnight. To it, 200 mL of ethyl acetate was added and washed with water (3 x 50 mL). The organic layer was concentrated and the residue was re-dissolved with 30 mL of DMF. To it, oxone (0.57 g, 3.75 mmol) was added and stirred for 6 hours. The solution was basified with 100 mL of saturated NaHCO₃, washed with 50 mL of methylene chloride, acidified 10% HCl until pH = 4, and extracted with ethyl acetate (4 x 100 mL). The combined organic layer was washed with brine (50 mL), dried over anhydrous sodium sulfate, and concentrated under vacuum to obtain 0.711 g (64.6% yield) of compound **260** as a pale yellow solid. This material was used in the subsequent step without further purification. mp. 191~192 °C; ¹H NMR δ 7.35–6.90 (m, 6H), 5.21 (s, 2H), 4.60 (s, 2H), 3.97 (t, *J* = 6.4 Hz, 2H), 3.61 (s, 3H), 2.99 (s, 6H), 2.58 (m, 4H), 2.39 (t, *J* = 6.4 Hz, 2H), 1.88 (m, 2H) 1.54 (m, 2H); ¹³C NMR δ ppm 176.3, 169.6, 151.3, 140.2, 137.6, 132.6, 129.9, 129.6, 129.1, 128.6, 127.5, 123.1, 117.1, 106.7, 99.8, 62.0, 53.8, 50.2, 40.8 (2C), 37.8, 33.2, 31.2, 27.0, 25.1. HRMS-ESI: m/z [M + H]⁺ calcd for C₂₅H₃₂N₂O₅: 440.2311 found: 440.2317.

Methyl-4-(4-(6-(dimethylamino)-1-oxoisoindolin-2-yl)-3-(3-hydroxypropyl)phenyl)butanoat

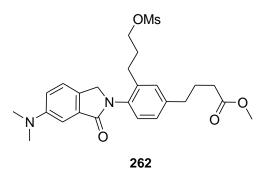
e (261). (BH12-064)



To a solution of **260** (0.711 g, 1.61 mmol) in 20 mL of MeOH, 2 mL of conc. HCl is added and stirred for 24 hours. To it, 50 mL of saturated NaHCO₃ solution was added, and concentrated under vacuum to remove MeOH. The residue was extracted with ethyl acetate (4 x 50 mL) and the combined organic layer was washed with brine (20 mL), dried over anhydrous sodium sulfate,

and concentrated under vacuum. The residue was purified by silica gel column chromatography (hexanes:ethyl acetate = 1:1) to obtain 0.584 g (88% yield) of compound **261** as a yellow oil. ¹H NMR δ 7.35–6.90 (m, 6H), 4.57 (s, 2H), 3.71 (s, 3H), 3.61 (t, *J* = 6 Hz, 2H), 2.99 (s, 6H), 2.58 (m, 4H), 2.39 (t, *J* = 6.4 Hz, 2H), 1.88 (m, 2H) 1.54 (m, 2H); ¹³C NMR δ ppm 174.8, 169.8, 151.1, 140.1, 137.6, 132.5, 129.9, 129.6, 129.2, 128.4, 127.5, 123.1, 117.1, 106.6, 61.4, 53.8, 51.2, 40.8 (2C), 37.8, 32.2, 31.8, 27.1, 25.1. HRMS-ESI: m/z [M + H]⁺ calcd for C₂₄H₃₁N₂O₄: 411.2284 found: 411.2279.

Methyl-4-(4-(6-(dimethylamino)-1-oxoisoindolin-2-yl)-3-(3-(methylsulfonyloxy)propyl)phen yl)butanoate (262) (BH12-078)

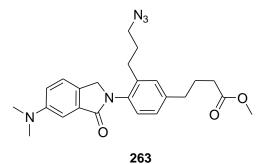


To a mixture of **261** (0.6 g, 1.467 mmol) and dry triethylamine (306 µL, 2.19 mmol) in 20 mL of dry dichloromethane under argon, methanesulfonyl chloride (0.251 g, 2.19 mmol) was added dropwise at 0°C. The reaction mixture was stirred for 8 hours at room temperature, quenched with 5 mL of water, and extracted with methylene chloride (3 x 50 mL). The combined organic layer was washed with brine (20 mL), dried over anhydrous sodium sulfate, and concentrated under vacuum. The residue was purified by silica gel column chromatography (hexanes:ethyl acetate = 1:1) to obtain 0.713 g (99% yield) of compound **262** as a yellow oil. ¹H NMR δ 7.35–6.90 (m, 6H), 4.61 (s, 2H), 4.31 (t, *J* = 6.4 Hz, 2 H), 3.72 (s, 3H), 3.07 (s, 3H), 2.99 (s, 6H), 2.58 (m, 4H), 2.40 (t, *J* = 6.4 Hz, 2H), 1.88 (m, 2H) 1.60 (m, 2H); ¹³C NMR δ ppm 174.8, 169.8, 151.2, 140.1, 137.6, 132.4, 130.2, 129.6, 128.6, 128.2, 127.5, 123.0, 117.1, 106.4, 68.2,

53.8, 51.2, 41.2 (2C), 37.6, 32.2, 31.8, 31.4, 27.0, 25.3. HRMS-ESI: $m/z [M + H]^+$ calcd for $C_{25}H_{33}N_2O_6S$: 489.2059 found: 489.2023.

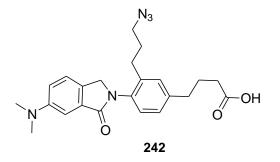
Methyl-4-(3-(3-azido propyl)-4-(6-(dimethylamino)-1-oxoisoindolin-2-yl) phenyl) but an oate the second se

(263) (BH12-082)



To a solution of **262** (0.713 g, 1.46 mmol) in 30 mL of dry DMF under argon, NaN₃ (379 mg, 5.83 mmol) was added. The reaction mixture was stirred at room temperature for 18 hours. To it, NaHCO₃ solution (10 mL) was added and extracted with methylene chloride (3 x 50 mL). The combined organic layer was washed with brine (20 mL), dried over anhydrous sodium sulfate, and concentrated under vacuum. The residue was purified by silica gel column chromatography (hexanes:ethyl acetate = 1:1) to obtain 0.617 g (97.2% yield) of compound **263** as a yellow oil. ¹H NMR δ 7.35–6.90 (m, 6H), 4.61 (s, 2H), 3.70 (s, 3H), 3.25 (t, *J* = 6.0 Hz, 2H), 3.01 (s, 6H), 2.58 (m, 4H), 2.38 (t, *J* = 6.4 Hz, 2H), 1.88 (m, 2H) 1.59 (m, 2H); ¹³C NMR δ ppm 176.2, 169.9, 151.6, 140.8, 138.1, 134.6, 132.8, 131.1, 130.6, 130.2, 129.8, 126.0, 115.2, 105.0, 53.8, 53.4, 50.9, 40.6 (2C), 35.0, 32.4, 31.8, 29.3, 28.7. HRMS-ESI: m/z [M + H]⁺ calcd for C₂₄H₃₀N₅O₃: 436.2349 found: 436.2361.

4-(3-(3-Azidopropyl)-4-(6-(dimethylamino)-1-oxoisoindolin-2-yl)phenyl)butanoic acid (242). (BH12-090)



To a solution of **263** (0.617 g, 1.42 mmol) in 30 mL of MeOH, 2 mL of 10% aqueous LiOH solution was added and stirred at room temperature for 6 hours. The resulting solution was concentrated under vacuum to remove MeOH and washed with 50 mL of methylene chloride. The aqueous layer was acidified with 10% aqueous HCl until pH = 4, concentrated under vacuum and purified by a short column chromatography to obtain 0.532 g (89% yield) of compound **242** as a light yellow solid; mp. $183\sim186$ °C; ¹H NMR δ 7.35–6.95 (m, 6H), 4.62 (s, 2H), 3.24 (t, *J* = 6.0 Hz, 2H), 3.02 (s, 6H), 2.58 (m, 4H), 2.39 (t, *J* = 8.0 Hz, 2H), 1.90 (m, 2H) 1.59 (m, 2H); ¹³C NMR δ ppm 178.2, 169.1, 150.6, 139.2, 137.6, 133.4, 129.8, 129.2, 128.7, 128.3, 127.6, 124.3, 117.5, 106.4, 53.6, 50.9, 40.8 (2C), 35.0, 32.4, 31.8, 29.5, 28.9; HRMS-ESI: m/z [M + H]⁺ calcd for C₂₃H₂₈N₅O₃: 422.2192 found: 422.2187.

Reference

- Hao, B., M.J. Gunaratna, M. Zhang, S. Weerasekara, S.N. Seiwald, V.T. Nguyen, A. Meier, and D.H. Hua, *Chiral-Substituted Poly-N-vinylpyrrolidinones and Bimetallic Nanoclusters in Catalytic Asymmetric Oxidation Reactions*. Journal of the American Chemical Society, 2016. 138(51): p. 16839-16848.
- 2. Knolker, H.J., H. Goesmann, H. Hermann, D. Herzberg, and G. Rohde, *Transition metal* complexes in organic synthesis, part 49. Development of novel chiral catalysts for the asymmetric catalytic complexation of prochiral cyclohexa-1,3-dienes by the tricarbonyliron fragment Mechanism of the asymmetric catalysis and involvement of a dinuclear iron cluster. Synlett, 1999(4): p. 421-425.
- 3. Hajos, Z.G. and D.R. Parrish, *Asymmetric synthesis of bicyclic intermediates of natural product chemistry*. The Journal of Organic Chemistry, 1974. **39**(12): p. 1615-1621.
- 4. Schmid, A., J.S. Dordick, B. Hauer, A. Kiener, M. Wubbolts, and B. Witholt, *Industrial biocatalysis today and tomorrow*. Nature, 2001. **409**: p. 258.
- 5. List, B., *Introduction: Organocatalysis.* Chemical Reviews, 2007. **107**(12): p. 5413-5415.
- 6. Swiegers, G.F., (Ed.), Mechanical Catalysis: Methods of Enzimatic, Homogeneous and Heterogeneous Catalysis, Wiley&Sons, New Jersey, 2008.
- Schmid, G., M. Baumle, M. Geerkens, I. Heim, C. Osemann, and T. Sawitowski, *Current and future applications of nanoclusters*. Chemical Society Reviews, 1999. 28(3): p. 179-185.
- 8. Schmid, G., *CLUSTERS AND COLLOIDS BRIDGES BETWEEN MOLECULAR AND CONDENSED MATERIAL*. Endeavour, 1990. **14**(4): p. 172-178.
- 9. Demirer, G.S., A.C. Okur, and S. Kizilel, *Synthesis and design of biologically inspired biocompatible iron oxide nanoparticles for biomedical applications*. Journal of Materials Chemistry B, 2015. **3**(40): p. 7831-7849.
- Hou, W., N.A. Dehm, and R.W.J. Scott, Alcohol oxidations in aqueous solutions using Au, Pd, and bimetallic AuPd nanoparticle catalysts. Journal of Catalysis, 2008. 253(1): p. 22-27.
- Kanaoka, S., N. Yagi, Y. Fukuyama, S. Aoshima, H. Tsunoyama, T. Tsukuda, and H. Sakurai, *Thermosensitive Gold Nanoclusters Stabilized by Well-Defined Vinyl Ether Star Polymers: Reusable and Durable Catalysts for Aerobic Alcohol Oxidation*. Journal of the American Chemical Society, 2007. **129**(40): p. 12060-12061.
- 12. Sophiphun, O., J. Wittayakun, R.N. Dhital, S. Haesuwannakij, A. Murugadoss, and H. Sakurai, *Gold/Palladium Bimetallic Alloy Nanoclusters Stabilized by Chitosan as Highly Efficient and Selective Catalysts for Homocoupling of Arylboronic Acid.* Australian Journal of Chemistry, 2012. **65**(9): p. 1238-1243.

- 13. Teraguchi, M., M. Ohtake, H. Inoue, A. Yoshida, T. Aoki, T. Kaneko, and K. Yamanaka, *Preparation of chiral polystyrene monoliths by utilizing W/O emulsion polymerization and their optical resolution ability.* Journal of Polymer Science Part A: Polymer Chemistry, 2005. **43**(11): p. 2348-2357.
- Hohberger, C., K. Beckerle, and J. Okuda, *Towards chiral polystyrene based materials:* controlled polymerization of p-(2,2 ' -diphenylethyl)styrene. Polymer Chemistry, 2010. 1(4): p. 534.
- 15. Zhang, H., T. Watanabe, M. Okumura, M. Haruta, and N. Toshima, *Catalytically highly active top gold atom on palladium nanocluster*. Nature Materials, 2011. **11**: p. 49.
- 16. Miyamura, H. and S. Kobayashi, *Tandem Oxidative Processes Catalyzed by Polymer-Incarcerated Multimetallic Nanoclusters with Molecular Oxygen.* Accounts of Chemical Research, 2014. **47**(4): p. 1054-1066.
- 17. Rodriguez, J., R.A. Campbell, and D.W. Goodman, *The nature of metal-metal bonding at bimetallic interfaces*. Surface Science, 1994. **307-309**: p. 377-383.
- 18. Zhang, H., T. Watanabe, M. Okumura, M. Haruta, and N. Toshima, *Catalytically highly active top gold atom on palladium nanocluster*. Nat Mater, 2011. **11**(1): p. 49-52.
- Shi, J., X. Hu, J. Zhang, W. Tang, H. Li, X. Shen, and N. Saito, One-step facile synthesis of Pd nanoclusters supported on carbon and their electrochemical property. Progress in Natural Science: Materials International, 2014. 24(6): p. 593-598.
- 20. Qian, H. and R. Jin, *Ambient Synthesis of Au144(SR)60 Nanoclusters in Methanol*. Chemistry of Materials, 2011. **23**(8): p. 2209-2217.
- 21. Leadbeater, N.E. and M. Marco, *Preparation of Polymer-Supported Ligands and Metal Complexes for Use in Catalysis*. Chemical Reviews, 2002. **102**(10): p. 3217-3274.
- 22. Cohen, N.B., B. L.; Laurenzano, A. J., *2,3-O-Isopropylidene-D-erythronolactone*. Org. Synth., 1984. **63**.
- Smrcina, M., P. Majer, E. Majerová, T.A. Guerassina, and M.A. Eissenstat, *Facile* stereoselective synthesis of γ-substituted γ-amino acids from the corresponding α-amino acids. Tetrahedron, 1997. 53(38): p. 12867-12874.
- 24. Kerr, M.S., J. Read de Alaniz, and T. Rovis, *An Efficient Synthesis of Achiral and Chiral* 1,2,4-Triazolium Salts: Bench Stable Precursors for N-Heterocyclic Carbenes. The Journal of Organic Chemistry, 2005. **70**(14): p. 5725-5728.
- Ward, B.D., H. Risler, K. Weitershaus, S. Bellemin-Laponnaz, H. Wadepohl, and L.H. Gade, 2-Aminopyrrolines: New Chiral Amidinate Ligands with a Rigid Well-Defined Molecular Structure and Their Coordination to TiIV. Inorganic Chemistry, 2006. 45(19): p. 7777-7787.
- 26. van Leeuwen, S.H., P.J.L.M. Quaedflieg, Q.B. Broxterman, Y. Milhajlovic, and R.M.J. Liskamp, *The synthesis of amides and dipeptides from unprotected amino acids by a simultaneous protection–activation strategy using boron trifluoride diethyl etherate.* Tetrahedron Letters, 2005. **46**(4): p. 653-656.

- J.S. McClanahan, 27. Digenis, G.A., P.-L. Chen, *Synthesis* ofand [2,3,4,5-14C]-1-vinyl-2-pyrrolidinone. Labelled Journal of Compounds and Radiopharmaceuticals, 1993. 33(1): p. 11-17.
- 28. Zhai, L., T. Shi, and H. Wang, *Preparation of polyvinylpyrrodione microspheres by dispersion polymerization*. Frontiers of Chemistry in China, 2009. **4**(1): p. 83.
- 29. Flebbe, T., R. Hentschke, E. Hädicke, and C. Schade, *Modeling of polyvinylpyrrolidone and polyvinylimidazole in aqueous solution*. Macromolecular Theory and Simulations, 1998. **7**(6): p. 567-577.
- 30. Hanessian, S., *Sugar lactams. III. Synthesis of five-,six-, and seven-membered analogs.* The Journal of Organic Chemistry, 1969. **34**(3): p. 675-681.
- Dirk Elend, M.F., J.; Pryde, D. , Synthesis of 5-O-benzyl-2-C- &-fluoromethyl-1,2,3-tri-O-acetyl-D-ribofuranose. ARKIVOC, 2006. 11: p. 114-127.
- 32. Ferrando, R., J. Jellinek, and R.L. Johnston, *Nanoalloys: From Theory to Applications of Alloy Clusters and Nanoparticles*. Chemical Reviews, 2008. **108**(3): p. 845-910.
- 33. Yao, H. and R. Kobayashi, *Chiral monolayer-protected Au–Pd bimetallic nanoclusters: Effect of palladium doping on their chiroptical responses.* Journal of Colloid and Interface Science, 2014. **419**: p. 1-8.
- 34. Lu, J. and P.H. Toy, Organic Polymer Supports for Synthesis and for Reagent and Catalyst Immobilization. Chemical Reviews, 2009. **109**(2): p. 815-838.
- Deng, D., Y. Jin, Y. Cheng, T. Qi, and F. Xiao, Copper Nanoparticles: Aqueous Phase Synthesis and Conductive Films Fabrication at Low Sintering Temperature. ACS Applied Materials & Interfaces, 2013. 5(9): p. 3839-3846.
- Murugadoss, A., K. Okumura, and H. Sakurai, *Bimetallic AuPd Nanocluster Catalysts* with Controlled Atomic Gold Distribution for Oxidative Dehydrogenation of Tetralin. The Journal of Physical Chemistry C, 2012. **116**(51): p. 26776-26783.
- Weir, M.G., M.R. Knecht, A.I. Frenkel, and R.M. Crooks, *Structural Analysis of PdAu Dendrimer-Encapsulated Bimetallic Nanoparticles*. Langmuir, 2010. 26(2): p. 1137-1146.
- 38. Poole, C.P.J.O., F. J., *Introduction to Nanotechnology*. 2003: John Wiley & Sons: New Jersey.
- 39. Zielińska, S., E. Ortyl, R. Barille, and S. Kucharski, *Preparation and characteristics of new chiral photochromic copolymers*. Optical Materials, 2009. **32**(1): p. 198-206.
- 40. Xu, Q., S. Kumar, S. Jin, H. Qian, M. Zhu, and R. Jin, *Chiral 38-gold-atom nanoclusters: synthesis and chiroptical properties.* Small, 2014. **10**(5): p. 1008-14.
- 41. Ackermann, J., M. Matthes, and C. Tamm, *Approaches to the Synthesis of Cytochalasans Part9. A versatile concept leading to all structural types of cytochalasans.* Helvetica Chimica Acta, 1990. **73**(1): p. 122-132.

- 42. Vanhessche, K., E. Van der Eycken, M. Vandewalle, and H. Röper, *L-Ribulose: A novel chiral pool compound*. Tetrahedron Letters, 1990. **31**(16): p. 2337-2340.
- 43. Ankamwar, B., C. Damle, A. Ahmad, and M. Sastry, *Biosynthesis of Gold and Silver Nanoparticles Using Emblica Officinalis Fruit Extract, Their Phase Transfer and Transmetallation in an Organic Solution.* Vol. 5. 2005. 1665-71.
- 44. Nishimura, S., Y. Yakita, M. Katayama, K. Higashimine, and K. Ebitani, *The role of negatively charged Au states in aerobic oxidation of alcohols over hydrotalcite supported AuPd nanoclusters*. Catalysis Science & Technology, 2013. **3**(2): p. 351-359.
- 45. Zhang, S., Y. Shao, H.-g. Liao, J. Liu, I.A. Aksay, G. Yin, and Y. Lin, *Graphene Decorated with PtAu Alloy Nanoparticles: Facile Synthesis and Promising Application for Formic Acid Oxidation*. Chemistry of Materials, 2011. **23**(5): p. 1079-1081.
- Wang, F., Y. Jiang, X. Wen, J. Xia, G. Sha, and R. Amal, Confined Au-Pd Ensembles in Mesoporous TiO2 Spheres for the Photocatalytic Oxidation of Acetaldehyde. ChemCatChem, 2013. 5(12): p. 3557-3561.
- 47. Dhital, R.N., C. Kamonsatikul, E. Somsook, K. Bobuatong, M. Ehara, S. Karanjit, and H. Sakurai, Low-Temperature Carbon–Chlorine Bond Activation by Bimetallic Gold/Palladium Alloy Nanoclusters: An Application to Ullmann Coupling. Journal of the American Chemical Society, 2012. 134(50): p. 20250-20253.
- 48. Dhital, R.N. and H. Sakurai, Anomalous Efficacy of Bimetallic Au/Pd Nanoclusters in C-Cl Bond Activation and Formal Metathesis-type C–B Bond Activation at Room Temperature. Chemistry Letters, 2012. **41**(6): p. 630-632.
- 49. Yoo, W.-J., H. Miyamura, and S. Kobayashi, *Polymer-Incarcerated Gold–Palladium Nanoclusters with Boron on Carbon: A Mild and Efficient Catalyst for the Sequential Aerobic Oxidation–Michael Addition of 1,3-Dicarbonyl Compounds to Allylic Alcohols.* Journal of the American Chemical Society, 2011. **133**(9): p. 3095-3103.
- Lee, C.K., B.S. Koo, Y.S. Lee, H.K. Cho, and K.J. Lee, Oxidation and bromodehydroxymethylation of benzylic alcohols using NaBrO3/NaHSO3 reagent. Bulletin of the Korean Chemical Society, 2002. 23(11): p. 1667-1670.
- 51. Menger, F.M. and C. Lee, *Synthetically useful oxidations at solid sodium permanganate surfaces.* Tetrahedron Letters, 1981. **22**(18): p. 1655-1656.
- 52. Lee, D.G. and U.A. Spitzer, *Aqueous dichromate oxidation of primary alcohols*. The Journal of Organic Chemistry, 1970. **35**(10): p. 3589-3590.
- 53. Meng, S.-S., Y. Liang, K.-S. Cao, L. Zou, X.-B. Lin, H. Yang, K.N. Houk, and W.-H. Zheng, *Chiral Phosphoric Acid Catalyzed Highly Enantioselective Desymmetrization of 2-Substituted and 2,2-Disubstituted 1,3-Diols via Oxidative Cleavage of Benzylidene Acetals.* Journal of the American Chemical Society, 2014. **136**(35): p. 12249-12252.
- 54. Moritani, J., Y. Hasegawa, Y. Kayaki, and T. Ikariya, Aerobic oxidative desymmetrization of meso-diols with bifunctional amidoiridium catalysts bearing chiral *N*-sulfonyldiamine ligands. Tetrahedron Letters, 2014. **55**(6): p. 1188-1191.

- 55. Mandal, S.K., D.R. Jensen, J.S. Pugsley, and M.S. Sigman, *Scope of Enantioselective Palladium(II)-Catalyzed Aerobic Alcohol Oxidations with (-)-Sparteine*. The Journal of Organic Chemistry, 2003. **68**(11): p. 4600-4603.
- 56. Jakka, K. and C.-G. Zhao, *Highly Enantioselective CH Oxidation of vic-Diols with Shi's Oxazolidinone Dioxiranes.* Organic Letters, 2006. **8**(14): p. 3013-3015.
- 57. Lehtonen, A., R. Kivekäs, and R. Sillanpää, *The separation of cis- and trans-1,3-cyclohexanediol isomers by copper complexation. Crystal structures of cis-1,3-cyclohexanediol and copper(II) chloride cis-1,3-cyclohexanediol complex.* Polyhedron, 2002. **21**(11): p. 1133-1138.
- 58. Kobayashi, S., P. Xu, T. Endo, M. Ueno, and T. Kitanosono, *Chiral Copper(II)-Catalyzed Enantioselective Boron Conjugate Additions to α,β-Unsaturated Carbonyl Compounds in Water*. Angewandte Chemie International Edition, 2012. **51**(51): p. 12763-12766.
- 59. Chen, B.-S. and U. Hanefeld, *Enantioselective preparation of (R) and (S)-3-hydroxycyclopentanone by kinetic resolution*. Journal of Molecular Catalysis B: Enzymatic, 2013. **85-86**: p. 239-242.
- 60. Lifchits, O., M. Mahlau, C.M. Reisinger, A. Lee, C. Farès, I. Polyak, G. Gopakumar, W. Thiel, and B. List, *The Cinchona Primary Amine-Catalyzed Asymmetric Epoxidation and Hydroperoxidation of α,β-Unsaturated Carbonyl Compounds with Hydrogen Peroxide*. Journal of the American Chemical Society, 2013. **135**(17): p. 6677-6693.
- Zhang, J., T. Xu, and Z. Li, Enantioselective Biooxidation of Racemic trans-Cyclic Vicinal Diols: One-Pot Synthesis of Both Enantiopure (S,S)-Cyclic Vicinal Diols and (R)-α-Hydroxy Ketones. Advanced Synthesis & Catalysis, 2013. 355(16): p. 3147-3153.
- 62. Liu, Y., R.-R. Li, and X.-B. Lu, *Microstructure Analysis of Poly(cyclopentene carbonate)s at the Diad Level*. Macromolecules, 2015. **48**(19): p. 6941-6947.
- 63. Singh, G. and J. Aube, *Synthesis of cyclic 1,3-diols as scaffolds for spatially directed libraries.* Organic & Biomolecular Chemistry, 2016. **14**(18): p. 4299-4303.
- 64. Stahl, S.S., J.L. Thorman, R.C. Nelson, and M.A. Kozee, *Oxygenation of Nitrogen-Coordinated Palladium(0): Synthetic, Structural, and Mechanistic Studies and Implications for Aerobic Oxidation Catalysis.* Journal of the American Chemical Society, 2001. **123**(29): p. 7188-7189.
- Stahl, S.S., Palladium Oxidase Catalysis: Selective Oxidation of Organic Chemicals by Direct Dioxygen-Coupled Turnover. Angewandte Chemie International Edition, 2004.
 43(26): p. 3400-3420.
- 66. Talsi, E.P. and K.P. Bryliakov, *Chemo- and stereoselective CH oxidations and epoxidations/cis-dihydroxylations with H2O2, catalyzed by non-heme iron and manganese complexes.* Coordination Chemistry Reviews, 2012. **256**(13): p. 1418-1434.
- 67. Boyd, D.R., N.D. Sharma, M.V. Berberian, M. Cleij, C. Hardacre, V. Ljubez, G. McConville, P.J. Stevenson, L.A. Kulakov, and C.C.R. Allen, *Arene cis-Diol*

Dehydrogenase-Catalysed Regio- and Stereoselective Oxidation of Arene-, Cycloalkaneand Cycloalkene-cis-diols to Yield Catechols and Chiral α -Ketols. Advanced Synthesis & Catalysis, 2015. **357**(8): p. 1881-1894.

- 68. Kihumbu, D., T. Stillger, W. Hummel, and A. Liese, *Enzymatic synthesis of all stereoisomers of 1-phenylpropane-1,2-diol.* Tetrahedron: Asymmetry, 2002. **13**(10): p. 1069-1072.
- 69. Pr évost, C., Sur un complexe iodo-argento-benzo que et son application à l'oxydation des combinaisons éthyléniques en α-glycols. Comptes Rendus, 1933. **196**: p. 1129.
- VanRheenen, V., R.C. Kelly, and D.Y. Cha, An improved catalytic OsO4 oxidation of olefins to cis-1,2-glycols using tertiary amine oxides as the oxidant. Tetrahedron Letters, 1976. 17(23): p. 1973-1976.
- 71. Woodward, R.B. and F.V. Brutcher, *cis-Hydroxylation of a Synthetic Steroid Intermediate with Iodine, Silver Acetate and Wet Acetic Acid.* Journal of the American Chemical Society, 1958. **80**(1): p. 209-211.
- 72. Spivey, A.C., R. Hanson, N. Scorah, and S.J. Thorpe, *Sharpless Asymmetric Dihydroxylation: Effect of Alkene Structure on Rates and Selectivity.* Journal of Chemical Education, 1999. **76**(5): p. 655.
- 73. Bataille, C.J.R. and T.J. Donohoe, *Osmium-free direct syn-dihydroxylation of alkenes*. Chemical Society Reviews, 2011. **40**(1): p. 114-128.
- 74. Stoltz, B.M., *Palladium Catalyzed Aerobic Dehydrogenation: From Alcohols to Indoles and Asymmetric Catalysis.* Chemistry Letters, 2004. **33**(4): p. 362-367.
- 75. Neisius, N.M. and B. Plietker, *Diastereoselective Ru-Catalyzed Cross-Metathesis–Dihydroxylation Sequence. An Efficient Approach toward Enantiomerically Enriched syn-Diols.* The Journal of Organic Chemistry, 2008. **73**(8): p. 3218-3227.
- 76. de Boer, J.W., W.R. Browne, S.R. Harutyunyan, L. Bini, T.D. Tiemersma-Wegman, P.L. Alsters, R. Hage, and B.L. Feringa, *Manganese catalysed asymmetric cis-dihydroxylation with H2O2*. Chemical Communications, 2008(32): p. 3747-3749.
- 77. Zang, C., Y. Liu, Z.-J. Xu, C.-W. Tse, X. Guan, J. Wei, J.-S. Huang, and C.-M. Che, *Highly Enantioselective Iron-Catalyzed cis-Dihydroxylation of Alkenes with Hydrogen Peroxide Oxidant via an FeIII-OOH Reactive Intermediate*. Angewandte Chemie International Edition, 2016. 55(35): p. 10253-10257.
- 78. Becker, H., S.B. King, M. Taniguchi, K.P.M. Vanhessche, and K.B. Sharpless, New Ligands and Improved Enantioselectivities for the Asymmetric Dihydroxylation of Olefins. The Journal of Organic Chemistry, 1995. 60(13): p. 3940-3941.
- 79. Choudary, B.M., K. Jyothi, S. Madhi, and M. Lakshmi Kantam, *Catalytic Asymmetric Dihydroxylation of Aliphatic Olefins with Reusable Resin-Osmium Tetroxide*. Advanced Synthesis & Catalysis, 2003. **345**(11): p. 1190-1192.

- Zeng, X.-P. and J. Zhou, Me2(CH2Cl)SiCN: Bifunctional Cyanating Reagent for the Synthesis of Tertiary Alcohols with a Chloromethyl Ketone Moiety via Ketone Cyanosilylation. Journal of the American Chemical Society, 2016. 138(28): p. 8730-8733.
- 81. Jin, Y.Y., Z.; Zhang, S.; Jiang, R.; Sun, X, Application of Recyclable Ligand [QN(OH)2]2PYDZ to Asymmetric Dihy-droxylation and Asymmetric Aminohydroxylation of Olefins. Chinese J. Org. Chem., 2007. 2007(27): p. 602.
- 82. Sengupta, S. and S. Mondal, *A highly diastereoselective chiral pool based synthesis of cisandtrans- indan-1, 2-diols.* Tetrahedron Letters, 1999. **40**(17): p. 3469-3470.
- 83. Kumar, S., M. Javagal, P. Srinivas, and B. Kempaiah, *Eco-friendly kinetic separation of trans-limonene and carvomenthene oxides*. Vol. 126. 2014. 875-880.
- 84. Buisson, D., X. Baucherel, E. Levoirier, and S. Juge, *Baker's yeast reduction of* α -alkyl- α -hydroxy- β -keto esters1. Tetrahedron Letters, 2000. **41**(9): p. 1389-1392.
- 85. Fujisawa, T., M. Watanabe, and T. Sato, *Diastereoface-differentiating addition of organometallics toward chiral .ALPHA.-ketoenamines.* Chemistry Letters, 1984: p. 2055-2058.
- Ahrgren, L. and L. Sutin, *Sharpless Asymmetric Dihydroxylation on an Industrial Scale*. Organic Process Research & Development, 1997. 1(6): p. 425-427.
- 87. Fuji, K., K. Tanaka, and H. Miyamoto, *Asymmetric cis-dihydroxylation of olefins by utilizing chiral bispiperazine*. Tetrahedron Letters, 1992. **33**(28): p. 4021-4024.
- 88. Malla Reddy, S., M. Srinivasulu, Y. Venkat Reddy, M. Narasimhulu, and Y. Venkateswarlu, *Catalytic asymmetric dihydroxylation of olefins using polysulfone-based novel microencapsulated osmium tetroxide*. Tetrahedron Letters, 2006. **47**(30): p. 5285-5288.
- Vadivel, S.K., S. Vardarajan, R.I. Duclos, J.T. Wood, J. Guo, and A. Makriyannis, *Conformationally constrained analogues of 2-arachidonoylglycerol.* Bioorganic & Medicinal Chemistry Letters, 2007. 17(21): p. 5959-5963.
- 90. Newhouse, T. and P.S. Baran, *If C H Bonds Could Talk: Selective C H Bond Oxidation*. Angewandte Chemie International Edition, 2011. **50**(15): p. 3362-3374.
- Powers, D.C., D.Y. Xiao, M.A.L. Geibel, and T. Ritter, On the Mechanism of Palladium-Catalyzed Aromatic C-H Oxidation. Journal of the American Chemical Society, 2010. 132(41): p. 14530-14536.
- 92. Moriyama, K., M. Takemura, and H. Togo, Direct and Selective Benzylic Oxidation of Alkylarenes via C-H Abstraction Using Alkali Metal Bromides. Organic Letters, 2012. 14(9): p. 2414-2417.
- 93. Chen, M.S., N. Prabagaran, N.A. Labenz, and M.C. White, *Serial Ligand Catalysis: A Highly Selective Allylic C-H Oxidation*. Journal of the American Chemical Society, 2005. 127(19): p. 6970-6971.

- 94. Gormisky, P.E. and M.C. White, *Catalyst-Controlled Aliphatic C–H Oxidations with a Predictive Model for Site-Selectivity*. Journal of the American Chemical Society, 2013. 135(38): p. 14052-14055.
- 95. Osberger, T.J., D.C. Rogness, J.T. Kohrt, A.F. Stepan, and M.C. White, *Oxidative diversification of amino acids and peptides by small-molecule iron catalysis*. Nature, 2016. **537**(7619): p. 214-219.
- 96. Canta, M., D. Font, L. Gómez, X. Ribas, and M. Costas, *The Iron(II) Complex* [*Fe(CF3SO3)2(mcp)*] as a Convenient, Readily Available Catalyst for the Selective Oxidation of Methylenic Sites in Alkanes. Advanced Synthesis & Catalysis, 2014. **356**(4): p. 818-830.
- 97. Boorman, T.C. and I. Larrosa, *Gold-mediated C-H bond functionalisation*. Chemical Society Reviews, 2011. **40**(4): p. 1910-1925.
- 98. Gutekunst, W.R. and P.S. Baran, *C-H functionalization logic in total synthesis*. Chemical Society Reviews, 2011. **40**(4): p. 1976-1991.
- 99. Lu, H. and X.P. Zhang, *Catalytic C-H functionalization by metalloporphyrins: recent developments and future directions.* Chemical Society Reviews, 2011. **40**(4): p. 1899-1909.
- 100. Collet, F., C. Lescot, and P. Dauban, *Catalytic C-H amination: the stereoselectivity issue*. Chemical Society Reviews, 2011. **40**(4): p. 1926-1936.
- 101. Santos Pisoni, D.d., J. Sobieski da Costa, D. Gamba, C.L. Petzhold, A. C ésar de Amorim Borges, M.A. Ceschi, P. Lunardi, and C.A. Saraiva Gonçalves, *Synthesis and AChE inhibitory activity of new chiral tetrahydroacridine analogues from terpenic cyclanones*. European Journal of Medicinal Chemistry, 2010. **45**(2): p. 526-535.
- 102. Hatano, M., S. Suzuki, and K. Ishihara, *Highly Efficient Alkylation to Ketones and Aldimines with Grignard Reagents Catalyzed by Zinc(II) Chloride*. Journal of the American Chemical Society, 2006. **128**(31): p. 9998-9999.
- Mizar, P. and T. Wirth, *Flexible Stereoselective Functionalizations of Ketones through* Umpolung with Hypervalent Iodine Reagents. Angewandte Chemie International Edition, 2014. 53(23): p. 5993-5997.
- 104. Kim, Y., H.K. Pak, Y.H. Rhee, and J. Park, *Catalytic transformation of esters of 1,2-azido alcohols into [small alpha]-amido ketones.* Chemical Communications, 2016. 52(39): p. 6549-6552.
- 105. Brenna, E., M. Crotti, F.G. Gatti, D. Monti, F. Parmeggiani, A. Pugliese, and F. Tentori, Biocatalytic synthesis of chiral cyclic [gamma]-oxoesters by sequential C-H hydroxylation, alcohol oxidation and alkene reduction. Green Chemistry, 2017. 19(21): p. 5122-5130.
- 106. Conde, A., L. Vilella, D. Balcells, M.M. D áz-Requejo, A. Lledós, and P.J. Pérez, *Introducing Copper as Catalyst for Oxidative Alkane Dehydrogenation*. Journal of the American Chemical Society, 2013. **135**(10): p. 3887-3896.

- 107. Pan, G.-H., R.-J. Song, and J.-H. Li, Radical-mediated synthesis of [gamma]-lactones by copper-catalyzed intermolecular carboesterification of alkenes with [small alpha]-carbonyl alkyl bromides and H2O. Organic Chemistry Frontiers, 2018. 5(2): p. 179-183.
- 108. Caruano, J., G.G. Muccioli, and R. Robiette, *Biologically active gamma-lactams: synthesis and natural sources.* Org Biomol Chem, 2016. **14**(43): p. 10134-10156.
- 109. Shu, C., M.-Q. Liu, S.-S. Wang, L. Li, and L.-W. Ye, Gold-Catalyzed Oxidative Cyclization of Chiral Homopropargyl Amides: Synthesis of Enantioenriched γ-Lactams. The Journal of Organic Chemistry, 2013. 78(7): p. 3292-3299.
- 110. Rhee, J.U. and M.J. Krische, *Highly Enantioselective Reductive Cyclization of Acetylenic Aldehydes via Rhodium Catalyzed Asymmetric Hydrogenation*. Journal of the American Chemical Society, 2006. **128**(33): p. 10674-10675.
- Schaus, S.E., J. Br ånalt, and E.N. Jacobsen, Asymmetric Hetero-Diels-Alder Reactions Catalyzed by Chiral (Salen)Chromium(III) Complexes. The Journal of Organic Chemistry, 1998. 63(2): p. 403-405.
- 112. Wang, H., B. Dong, Y. Wang, J. Li, and Y. Shi, A Palladium-Catalyzed Regioselective Hydroesterification of Alkenylphenols to Lactones with Phenyl Formate as CO Source. Organic Letters, 2014. 16(1): p. 186-189.
- 113. Wilent, J. and K.S. Petersen, *Enantioselective Desymmetrization of Diesters*. The Journal of Organic Chemistry, 2014. **79**(5): p. 2303-2307.
- Sinai, Á., Á. Mészáros, T. Gáti, V. Kudar, A. Palló, and Z. Novák, *Copper-Catalyzed Oxidative Ring Closure and Carboarylation of 2-Ethynylanilides*. Organic Letters, 2013. 15(22): p. 5654-5657.
- 115. Weng, Y., H. Zhou, C. Sun, Y. Xie, and W. Su, Copper-Catalyzed Cyclization for Access to 6H-Chromeno[4,3-b]quinolin-6-ones Employing DMF as the Carbon Source. J Org Chem, 2017. 82(17): p. 9047-9053.
- 116. Aradi, K., P. Bombicz, and Z. Novak, Modular Copper-Catalyzed Synthesis of Chromeno[4,3-b]quinolines with the Utilization of Diaryliodonium Salts. J Org Chem, 2016. 81(3): p. 920-31.
- 117. Hu, T. and C. Li, *Synthesis of Lactams via Copper-Catalyzed Intramolecular Vinylation of Amides*. Organic Letters, 2005. **7**(10): p. 2035-2038.
- 118. Kerrigan, N.J., P.C. Hutchison, T.D. Heightman, and D.J. Procter, Development of a solid-phase 'asymmetric resin-capture-release' process: application of an ephedrine chiral resin in an approach to [gamma]-butyrolactones. Organic & Biomolecular Chemistry, 2004. 2(17): p. 2476-2482.
- 119. Theodore, L.J. and W.L. Nelson, *Stereospecific synthesis of the enantiomers of verapamil and gallopamil.* The Journal of Organic Chemistry, 1987. **52**(7): p. 1309-1315.

- 120. Cordero, F.M., F. Pisaneschi, M. Salvati, S. Valenza, C. Faggi, and A. Brandi, *Application of Mosher's method for absolute configuration assignment and resolution of* 2-hydroxypyrrolizidinones. Chirality, 2005. **17**(3): p. 149-153.
- Vedejs, E., D.A. Engler, and J.E. Telschow, *Transition-metal peroxide reactions*. Synthesis of .alpha.-hydroxycarbonyl compounds from enolates. The Journal of Organic Chemistry, 1978. 43(2): p. 188-196.
- 122. Hartman, G.D. and L.M. Weinstock, *Thiazoles from Ethyl Isocyanoacetate and Thiono Esters: Ethyl Thiazole-4-Carboxylate.* Organic Syntheses, 2003.
- 123. Newhouse, T. and P.S. Baran, *If C–H Bonds Could Talk Selective C–H Bond Oxidation*. Angewandte Chemie (International ed. in English), 2011. **50**(15): p. 3362-3374.
- 124. Stout, E.P., M.Y. Choi, J.E. Castro, and T.F. Molinski, *Potent Fluorinated Agelastatin Analogues for Chronic Lymphocytic Leukemia: Design, Synthesis, and Pharmacokinetic Studies.* Journal of Medicinal Chemistry, 2014. **57**(12): p. 5085-5093.
- 125. He, J., L.G. Hamann, H.M.L. Davies, and R.E.J. Beckwith, *Late-stage C–H functionalization of complex alkaloids and drug molecules via intermolecular rhodium-carbenoid insertion.* Nature Communications, 2015. **6**: p. 5943.
- 126. Guti érrez-Bonet, Á., C. Remeur, J.K. Matsui, and G.A. Molander, *Late-Stage C–H Alkylation of Heterocycles and 1,4-Quinones via Oxidative Homolysis of 1,4-Dihydropyridines*. Journal of the American Chemical Society, 2017. **139**(35): p. 12251-12258.
- 127. Huang, X., T.M. Bergsten, and J.T. Groves, *Manganese-Catalyzed Late-Stage Aliphatic C–H Azidation*. Journal of the American Chemical Society, 2015. **137**(16): p. 5300-5303.
- 128. Loskot, S.A., D.K. Romney, F.H. Arnold, and B.M. Stoltz, *Enantioselective Total Synthesis of Nigelladine A via Late-Stage C–H Oxidation Enabled by an Engineered P450 Enzyme*. Journal of the American Chemical Society, 2017. **139**(30): p. 10196-10199.
- 129. Nanjo, T., E.C. de Lucca, and M.C. White, *Remote, Late-Stage Oxidation of Aliphatic C– H Bonds in Amide-Containing Molecules*. Journal of the American Chemical Society, 2017. **139**(41): p. 14586-14591.
- 130. Stang, E.M. and M. Christina White, *Total synthesis and study of 6-deoxyerythronolide B by late-stage C–H oxidation*. Nature Chemistry, 2009. **1**: p. 547.
- 131. Shen, X.-C., Y.-P. Yang, T.-T. Xiao, J. Peng, and X.-D. Liu, Protective effect of oxymatrine on myocardial fibrosis induced by acute myocardial infarction in rats involved in TGF-β1-Smads signal pathway. Journal of Asian Natural Products Research, 2011. 13(3): p. 215-224.
- 132. GU, X., A. ZHANG, M. NI, and X. MAO, Sophocarpidine and oxymatrine derivatives and preparation method and application thereof, in CN106518873 (A). 2017: China.
- 133. Bai, G.Y., Wang, D.Q., Ye, C.H. et al., 1H and 13C Chemical Shift Assignments and Stereochemistry

of Matrine and Oxymatrine Applied Magnetic Resonance, 2002. 23: p. 113-121.

- 134. Jana, S., M. Ghosh, M. Ambule, and S. Sen Gupta, *Iron Complex Catalyzed Selective C– H Bond Oxidation with Broad Substrate Scope*. Organic Letters, 2017. **19**(4): p. 746-749.
- 135. Chen, M.S. and M.C. White, *Combined Effects on Selectivity in Fe-Catalyzed Methylene Oxidation*. Science, 2010. **327**(5965): p. 566.
- Choudhary, M.I., S.G. Musharraf, A. Sami, and R. Atta ur, *Microbial Transformation of Sesquiterpenes*, (-)-Ambrox® and (+)-Sclareolide. Helvetica Chimica Acta, 2004. 87(10): p. 2685-2694.
- 137. Kuhl, H., *Pharmacology of estrogens and progestogens: influence of different routes of administration*. Climacteric, 2005. **8**(sup1): p. 3-63.
- See, Y.Y., A.T. Herrmann, Y. Aihara, and P.S. Baran, *Scalable C–H Oxidation with Copper: Synthesis of Polyoxypregnanes*. Journal of the American Chemical Society, 2015. 137(43): p. 13776-13779.
- 139. Schleyer, P.v.R., J.E. Williams, and K.R. Blanchard, *Evaluation of strain in hydrocarbons. The strain in adamantane and its origin.* Journal of the American Chemical Society, 1970. **92**(8): p. 2377-2386.
- 140. Fresno, N., R. Pérez-Fern ández, C. Goicoechea, I. Alkorta, A. Fern ández-Carvajal, R. de la Torre-Mart nez, S. Quirce, A. Ferrer-Montiel, M.I. Mart negative for the second sec
- 141. Duhamel, N., D. Martin, R. Larcher, B. Fedrizzi, and D. Barker, *Convenient synthesis of deuterium labelled sesquiterpenes*. Tetrahedron Letters, 2016. **57**(40): p. 4496-4499.
- 142. Arrington, K.J., C.B. Murray, E.C. Smith, H. Marand, and J.B. Matson, *Precision Polyketones by Ring-Opening Metathesis Polymerization: Effects of Regular and Irregular Ketone Spacing*. Macromolecules, 2016. **49**(10): p. 3655-3662.
- 143. Xi, X., T. Chen, J.-S. Zhang, and L.-B. Han, *Efficient and selective hydrogenation of C-O bonds with a simple sodium formate catalyzed by nickel.* Chemical Communications, 2018. 54(12): p. 1521-1524.
- 144. Kricka, L.J. and G.H.G. Thorpe, *Chemiluminescent and bioluminescent methods in Analytical Chemistry. A review.* Analyst, 1983. **108**(1292): p. 1274-1296.
- 145. Thorne, N., D.S. Auld, and J. Inglese, Apparent activity in high-throughput screening: origins of compound-dependent assay interference. Current Opinion in Chemical Biology, 2010. 14(3): p. 315-324.
- 146. Auld, D.S., N.T. Southall, A. Jadhav, R.L. Johnson, D.J. Diller, A. Simeonov, C.P. Austin, and J. Inglese, *Characterization of chemical libraries for luciferase inhibitory activity*. Journal of medicinal chemistry, 2008. 51(8): p. 2372-2386.
- 147. Thorne, N., M. Shen, W.A. Lea, A. Simeonov, S. Lovell, D.S. Auld, and J. Inglese, Firefly luciferase in chemical biology: a compendium of inhibitors, mechanistic

evaluation of chemotypes, and suggested use as a reporter. Chemistry & biology, 2012. **19**(8): p. 1060-1072.

- 148. Fan, F. and K.V. Wood, *Bioluminescent assays for high-throughput screening*. Assay and drug development technologies, 2007. **5**(1): p. 127-136.
- 149. Braeuning, A., *Firefly luciferase inhibition: a widely neglected problem*. Archives of toxicology, 2015. **89**(1): p. 141-142.
- 150. Auld, D.S., N. Thorne, D.-T. Nguyen, and J. Inglese, A specific mechanism for nonspecific activation in reporter-gene assays. 2008, ACS Publications.
- 151. Thompson, J.F., L.S. Hayes, and D.B. Lloyd, *Modulation of firefly luciferase stability* and impact on studies of gene regulation. Gene, 1991. **103**(2): p. 171-177.
- 152. Conti, E., N.P. Franks, and P. Brick, *Crystal structure of firefly luciferase throws light on a superfamily of adenylate-forming enzymes.* Structure, 1996. **4**(3): p. 287-298.
- 153. Branchini, B.R., M.H. Murtiashaw, R.A. Magyar, and S.M. Anderson, *The role of lysine* 529, a conserved residue of the acyl-adenylate-forming enzyme superfamily, in firefly *luciferase*. Biochemistry, 2000. **39**(18): p. 5433-5440.
- 154. Franks, N., A. Jenkins, E. Conti, W. Lieb, and P. Brick, *Structural basis for the inhibition of firefly luciferase by a general anesthetic*. Biophysical journal, 1998. **75**(5): p. 2205-2211.
- 155. Leit ão, J.M. and J.C.E. da Silva, *Firefly luciferase inhibition*. Journal of Photochemistry and Photobiology B: Biology, 2010. **101**(1): p. 1-8.
- 156. Suzuki, H., Y. Kawarabayasi, J. Kondo, T. Abe, K. Nishikawa, S. Kimura, T. Hashimoto, and T. Yamamoto, *Structure and regulation of rat long-chain acyl-CoA synthetase*. Journal of Biological Chemistry, 1990. 265(15): p. 8681-8685.
- 157. Oba, Y., M. Ojika, and S. Inouye, *Firefly luciferase is a bifunctional enzyme: ATP dependent monooxygenase and a long chain fatty acyl CoA synthetase.* FEBS letters, 2003. 540(1-3): p. 251-254.
- 158. Inouye, S., *Firefly luciferase: an adenylate-forming enzyme for multicatalytic functions.* Cellular and molecular life sciences, 2010. **67**(3): p. 387-404.
- 159. Marques, S.M. and J.C. Esteves da Silva, *Firefly bioluminescence: a mechanistic approach of luciferase catalyzed reactions*. IUBMB life, 2009. **61**(1): p. 6-17.
- Matsuki, H., A. Suzuki, H. Kamaya, and I. Ueda, Specific and non-specific binding of long-chain fatty acids to firefly luciferase: cutoff at octanoate. Biochimica et Biophysica Acta (BBA)-General Subjects, 1999. 1426(1): p. 143-150.
- 161. Airth, R., W. Rhodes, and W. McElroy, *The function of coenzyme A in luminescence*. Biochimica et biophysica acta, 1958. **27**: p. 519-532.
- McElroy, W., J. Hastings, J. Coulombre, and V. Sonnenfeld, *The mechanism of action of pyrophosphate in firefly luminescence*. Archives of biochemistry and biophysics, 1953.
 46(2): p. 399-416.

- 163. Nakamura, M., S. Maki, Y. Amano, Y. Ohkita, K. Niwa, T. Hirano, Y. Ohmiya, and H. Niwa, *Firefly luciferase exhibits bimodal action depending on the luciferin chirality*. Biochemical and biophysical research communications, 2005. **331**(2): p. 471-475.
- 164. Lemasters, J.J. and C.R. Hackenbrock, *Kinetics of product inhibition during firefly luciferase luminescence*. Biochemistry, 1977. **16**(3): p. 445-447.
- 165. da Silva, L.P. and J.C.E. da Silva, *Kinetics of inhibition of firefly luciferase by dehydroluciferyl-coenzyme A, dehydroluciferin and L-luciferin.* Photochemical & Photobiological Sciences, 2011. **10**(6): p. 1039-1045.
- 166. Fontes, R., A. Dukhovich, A. Sillero, and M.A.G. Sillero, Synthesis of dehydroluciferin by firefly luciferase: effect of dehydroluciferin, coenzyme A and nucleoside triphosphates on the luminescent reaction. Biochemical and biophysical research communications, 1997. 237(2): p. 445-450.
- 167. Nakagomi, M., N. Fujimaki, A. Ito, T. Toda, H. Fukasawa, K. Shudo, and R. Tomita, A Novel Aromatic Carboxylic Acid Inactivates Luciferase by Acylation of an Enzymatically Active Regulatory Lysine Residue. PloS one, 2013. 8(9): p. e75445.
- 168. Gobbi, S., A. Cavalli, M. Negri, K.E. Schewe, F. Belluti, L. Piazzi, R.W. Hartmann, M. Recanatini, and A. Bisi, *Imidazolylmethylbenzophenones as highly potent aromatase inhibitors*. Journal of medicinal chemistry, 2007. 50(15): p. 3420-3422.
- 169. Ghosh, A.K., J. Takayama, Y. Aubin, K. Ratia, R. Chaudhuri, Y. Baez, K. Sleeman, M. Coughlin, D.B. Nichols, and D.C. Mulhearn, *Structure-Based Design, Synthesis, and Biological Evaluation of a Series of Novel and Reversible Inhibitors for the Severe Acute Respiratory Syndrome- Coronavirus Papain-Like Protease.* Journal of medicinal chemistry, 2009. **52**(16): p. 5228-5240.
- 170. Corey, E.J. and A. Venkateswarlu, *Protection of hydroxyl groups as tert-butyldimethylsilyl derivatives*. Journal of the American Chemical Society, 1972.
 94(17): p. 6190-6191.
- 171. Claridge, T.D.W., S.G. Davies, J.A. Lee, R.L. Nicholson, P.M. Roberts, A.J. Russell,
 A.D. Smith, and S.M. Toms, *Highly (E)-Selective Wadsworth–Emmons Reactions Promoted by Methylmagnesium Bromide*. Organic Letters, 2008. 10(23): p. 5437-5440.
- 172. Travis, B.R., M. Sivakumar, G.O. Hollist, and B. Borhan, *Facile Oxidation of Aldehydes* to Acids and Esters with Oxone. Organic Letters, 2003. **5**(7): p. 1031-1034.
- 173. Speers, A.E. and B.F. Cravatt, *Profiling enzyme activities in vivo using click chemistry methods*. Chemistry & biology, 2004. **11**(4): p. 535-546.
- 174. Kolb, H.C. and K.B. Sharpless, *The growing impact of click chemistry on drug discovery*. Drug discovery today, 2003. **8**(24): p. 1128-1137.
- 175. Li, L. and Z. Zhang, *Development and applications of the copper-catalyzed azide-alkyne cycloaddition (CuAAC) as a bioorthogonal reaction.* Molecules, 2016. **21**(10): p. 1393.
- 176. Ghosh, A.K., J. Takayama, Y. Aubin, K. Ratia, R. Chaudhuri, Y. Baez, K. Sleeman, M. Coughlin, D.B. Nichols, D.C. Mulhearn, B.S. Prabhakar, S.C. Baker, M.E. Johnson, and

A.D. Mesecar, Structure-Based Design, Synthesis, and Biological Evaluation of a Series of Novel and Reversible Inhibitors for the Severe Acute Respiratory Syndrome-Coronavirus Papain-Like Protease. Journal of Medicinal Chemistry, 2009. **52**(16): p. 5228-5240.

- 177. Saint-Louis, C.J., R.N. Shavnore, C.D.C. McClinton, J.A. Wilson, L.L. Magill, B.M. Brown, R.W. Lamb, C.E. Webster, A.K. Schrock, and M.T. Huggins, Synthesis, computational, and spectroscopic analysis of tunable highly fluorescent BN-1,2-azaborine derivatives containing the N-BOH moiety. Organic & Biomolecular Chemistry, 2017. 15(48): p. 10172-10183.
- Vedachalam, S., N. Murugesh, P. Chakraborty, R. Karvembu, and X.-W. Liu, NHC catalyzed enantioselective Coates-Claisen rearrangement: a rapid access to the dihydropyran core for oleuropein based secoiridoids. New Journal of Chemistry, 2018. 42(3): p. 1832-1839.
- 179. Sabbatini, P., P. Wellendorph, S. Høg, M.H.F. Pedersen, H. Bräuner-Osborne, L. Martiny, B. Frølund, and R.P. Clausen, *Design, Synthesis, and in Vitro Pharmacology of New Radiolabeled γ-Hydroxybutyric Acid Analogues Including Photolabile Analogues with Irreversible Binding to the High-Affinity γ-Hydroxybutyric Acid Binding Sites.* Journal of Medicinal Chemistry, 2010. 53(17): p. 6506-6510.

Appendix A. ¹H-NMR, ¹³C NMR and Mass Spectrum

