

pH as a control on interactions of methanogens and iron reducers

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Abstract

A growing body of evidence demonstrates that methanogenesis and Fe(III) reduction can occur simultaneously. However, environmental controls on interactions between each are poorly understood. In this study we considered pH as a control on interactions between Fe(III) reduction and methanogenesis in anoxic sediment bioreactors. The reactors consisted of 100mL of synthetic aqueous media, and 1 g of marsh sediment amended with goethite (1mmol). One set of reactors received acidic media (pH 6), and the other alkaline media (pH 7.5). Each set received media containing acetate (0.25 mM) to serve as an electron donor. Control reactors, deficient in acetate, were also included. We maintained a fluid residence time of 35 days by sampling and feeding the reactors every seven days.

For pH 6.0 and pH 7.5 reactors, the measured pH of effluent samples averaged 6.33 and 7.37, respectively. The extent of Fe(III) reduction and methanogenesis varied considerably between each set of reactors. More Fe(III) was reduced in the pH 6 reactors (646.39 μ moles on avg.) than the pH 7.5 reactors (31.32 μ moles on avg.). Conversely, more methane formed in pH 7.5 reactors (127.5 μ moles on avg.) than the pH 6 reactors (78.9 μ moles on avg.). Alkalinity concentrations during the middle and end of the experiment averaged 9.6 meq/L and 5.2 meq/L in pH 6 and pH7.5 reactors, respectively. Although much less Fe(III) reduction occurred in pH 7.5 reactors, the relative abundance of Fe(III) reducers in them decreased little from levels observed in the pH 6 reactors. Sequences classified within *Geobacter*, a genus of bacteria known primarily as dissimilatory metal reducers, accounted for 22% and 13.45% of the sequences in the pH 6 and pH 7.5 reactors and only 0.8% of the sequences in the marsh sediment inoculum. In contrast, sequences classified within orders of methanogens were low in abundance, making up only 0.47% and 1.04% of the sequences in the pH 6 and pH 7.5 reactors, respectively.

Mass balance calculations demonstrate that the amount of electron donor consumed by each group varied considerably between the sets of reactors. Expressed as a quantity of acetate, the reactions consumed about 160 μ M of electron donor each in pH 6 reactors. In contrast, methanogenesis consumed over 30 times more electron donor than Fe(III) reduction in the pH 7.5 reactors. Thus, the results of our experiment indicate that the decrease in electron donor consumption by Fe(III) reduction at basic pH was nearly matched by the increase in electron donor consumption by methanogens. Results of geochemical modeling calculations indicate that more energy was available for Fe(III) reduction in the pH 6.0 reactors than the pH 7.5 reactors, matching variation in Fe(III) reduction rates, and that the density of sorbed ferrous iron was higher in pH 6 reactors than pH 7.5 reactors. Thus, the calculation results are consistent with bioenergetics, but not variation in ferrous iron sorption, as a potential mechanism driving variation in the balance between each reaction with pH.

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Chapter 1 - Introduction

Shifts in the balance between carbon storage and carbon emission from soils as a result of climate change represent an important, but not well-understood biogeochemical feedback mechanism on global climate change (Smith and Fang, 2010). Anoxic soils (e.g., soils in wetlands and peatlands) are a particularly potent carbon reservoir because they contain high carbon densities, and serve as a major source of atmospheric methane, a greenhouse gas with 20-fold greater radiative forcing than carbon dioxide. In order to forecast how biogeochemical processes in these soils will respond to climate change we need a clear understanding of environmental controls on interactions of microorganisms within them.

In this study we considered pH as a control on the balance between microbial Fe(III) reduction, and methanogenesis in anoxic soil. During Fe(III) reduction microbes use Fe(III), most commonly found as oxides and oxyhydroxides, as a terminal electron acceptor for cellular respiration to produce Fe(II) (Weber et al., 2006). During methanogenesis microbes make methane using hydrogen and CO₂, methyl-containing C-1 compounds, or the methyl carbon of acetate as their substrates (Whitman et al., 2006)

Both Fe(III) reduction and methanogenesis are known to co-exist in anoxic soils. Competition between the two is common, and most often results in lower amounts of methanogens. (Roden and Wetzel, 1996, 2003; Lovley and Phillips, 1987; Reiche et al., 2008). Evidence suggests that Fe(III) reducers can successfully outcompete methanogens for electron donor in anaerobic environments, resulting in suppression in rates of methane production when there are abundant amounts of microbially reducible Fe(III) oxides (Lovley and Klug 1982, Lovley and Phillips 1987). Interaction between these two groups of microorganisms has the

potential to significantly impact carbon budgets in anoxic systems. Fe(III) reduction has the potential to increase alkalinity of the system through the production of HCO_3^- and OH^- . The increase in alkalinity can cause carbonate mineral precipitation by increasing saturation with respect to carbonate phases (Coleman et al., 1993). Roden and Lovley (1993) showed the precipitation of FeCO_3 during growth of Fe(III) reducing bacteria in a bicarbonate-buffered medium. This process helps to trap carbon in the soil. Methanogenesis, on the other hand, does not generate much alkalinity, and releases carbon through the formation of CH_4 (Kirk et al., 2013; Conrad, 1996).

Previous studies have used thermodynamic calculations to show that pH has the potential to influence interactions between Fe(III) reduction and sulfate reduction (Postma and Jakobsen, 1996; Bethke et al., 2011; Kirk et al., 2013; Kirk et al., 2015a). As shown in Figure 1, each

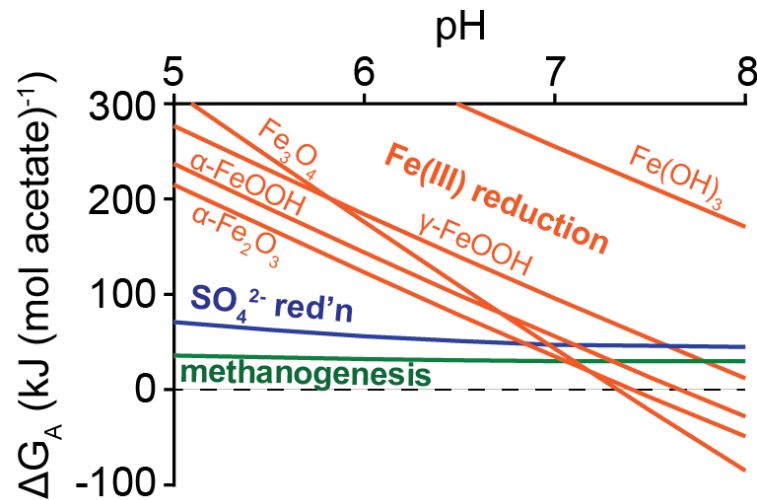


Figure 1: Variation with pH in energy available to Fe(III) reduction, sulfate reduction, and methanogenesis. ΔG_A refers to the amount of energy available to drive reactions forward.

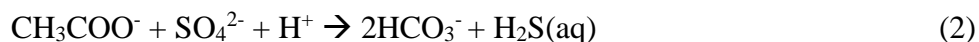
metabolic reaction varies unequally with pH, therefore, changes in pH can alter which reaction is energetically favorable (Postma and Jakobsen, 1996; Bethke et al., 2011). At an acidic pH Fe(III) reduction has a much higher free energy yield than at alkaline pH. This allows Fe(III) reducers to have a more energetically favorable reaction than sulfate reduction at an acidic pH (Bethke et al.

2011; Kirk et al., 2013). We hypothesize that pH may similarly affect interactions between Fe(III) reducers and methanogens. At acidic pH, Fe(III) reduction is thermodynamically more favorable than methanogenesis (Fig. 1). At alkaline pH, however, the opposite may be true.

These changes in free energy yield occur in response to differences in the number of protons consumed by each reaction. Reduction of ferric iron in oxides and oxyhydroxides consumes several protons as shown in the following example reaction which uses goethite as the source of ferric iron and acetate as the electron donor:



As a result of this consumption of protons, the energy yield of the reaction increases rapidly as pH decreases. In contrast, there are much fewer protons consumed during sulfate reduction and methanogenesis, so the energy yield of those reactions vary weakly with pH.



Reaction free energy yields can affect competition between microorganisms. Groups of microbes can conserve energy if their reactions are more energetically favorable. These groups have physiological advantages, including fast reactions and more biomass yield, over those using less favorable reactions (Lovley and Goodwin, 1988; Jin and Bethke, 2007; Roden and Jin, 2011; Jin, 2012). This increase in free energy yield that we see with decreasing pH may allow microbes

capable of Fe(III) reduction to better compete with methanogens in acidic environments than in alkaline environments (Bethke et al., 2011; Kirk et al., 2013).

This study tests the hypothesis that pH can influence interactions between Fe(III) reduction and methanogenesis in a similar way to the relationship between sulfate reduction and Fe(III) reduction. We tested this hypothesis using two sets of bioreactor experiments: one receiving acidic aqueous media (pH 6), and one receiving media with basic pH (pH 7.5). Hereafter, we will refer to these as the pH 6.0 and pH 7.5 sets of bioreactors. Each set of bioreactors was divided in half. One half was fed with acetate during the weekly sampling, and the other was not. We will refer to these as our acetate-fed and control reactors.

Chapter 2 - Materials and Methods

Sample Collection

We collected soil for the experiment from the Big Blue river floodplain at the north end of Tuttle Creek reservoir near Olsburg Kansas (Fig 2). The samples were collected on January 30, 2016. During that time the soil was submerged in about 0.25 m of water. The soil was collected in sterile (autoclaved at 121°C for 30 minutes) mason jars. The jars were rinsed multiple times with water from the marsh before sample collection. The jars were kept sealed in an incubator at 20°C until the bioreactors were assembled. While collecting soil, water samples were also collected in order to analyze the water chemistry of the area. The water samples were filtered using 0.45µm filters and analyzed for major ions, as described below.

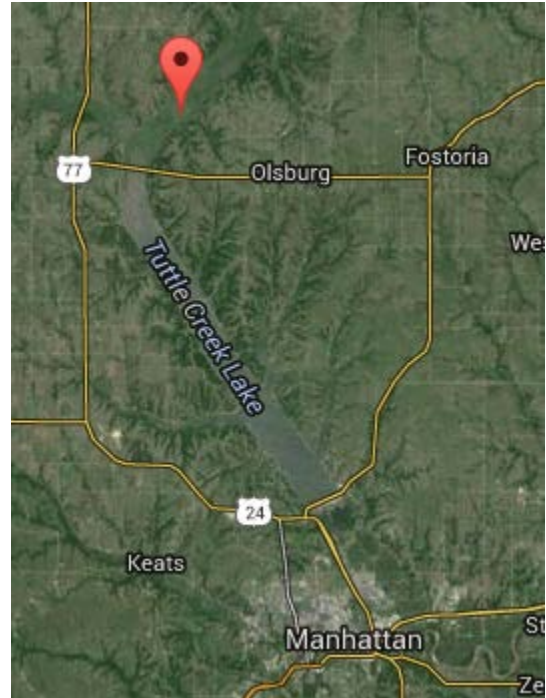


Figure 2: Map showing the location of the sampling site near Olsburg Kansas. (Latitude 039°27'38.988"N Longitude 096°41'25.3428"W)

Medium

We defined a synthetic aqueous media for the experiment based on the composition of the water samples collected from the field site. The media concentrations differ slightly from the water at the sampling site in that we added ammonium and phosphate as important macronutrients (Table 1). Media for the live bioreactors also includes acetate, which can serve as an electron donor for microbes in the reactors. Control media only differed from live media in

that there was no acetate added to control reactors.

To set the pH, and purge oxygen from the media, we sparged the bottles with an oxygen-free gas mixture of nitrogen and carbon dioxide. To achieve a pH of 7.5 we sparged with a gas mixture consisting of 1.1% CO₂ in N₂ for 1 hour. To achieve a pH of 6, we sparged the media with 35% CO₂ in N₂ for 1 hour. After sparging, we sealed the media bottles, autoclaved them for 30 mins at 121°C, and stored them at room temperature (~22°C).

Table 1: Composition of aqueous media.

Medium	NaHCO ₃ mM	CaCl ₂ mM	MgCl ₂ *6H ₂ O mM	KCl mM	NH ₄ Cl mM	HK ₂ PO ₄ mM	NaCH ₃ COO mM	CO ₂ * %	final pH**
6.0 Live	5	2	0.5	0.25	0.05	0.001	0.25	35	6.0
6.0 Control	5	2	0.5	0.25	0.05	0.001		35	6.0
7.5 Live	5	2	0.5	0.25	0.05	0.001	0.25	1.1	7.6
7.5 Control	5	2	0.5	0.25	0.05	0.001		1.1	7.6

*Proportion of CO₂ in CO₂:N₂ mix.
 ** These are average measured values

Bioreactors

Each set of bioreactor experiments was performed in triplicate. The reactors consisted of 160 mL serum bottles that were filled with 100mL of media, and 1.0 mmol of goethite (Fig 3). We capped the reactors, sparged them with gas, and sterilized them using an autoclave (30 minutes at 121°C).

Next, we placed the reactors in an anaerobic chamber, opened them, and added 1 gram of soil. In addition to the soil, we also added 100 μ M FeCl₂ (final concentration) to act as a reducing agent. We then recapped the reactors with sterile stoppers and seals, and a sterile needle, fitted with a gas-tight syringe valve, was inserted through the stopper. Lastly, the reactors were brought out of the anaerobic chamber to

readjust the pH by sparging with the gas mixtures discussed previously. Once assembled the reactors were kept in an incubator at 20°C.

The reactors were allowed to incubate for seven days. On the seventh day, and every seventh day thereafter, the reactors were sampled and fed. Sampling and feeding consisted of withdrawing 20mL from the serum bottle and replacing the sampled volume with sterile media. Incubation and sampling persisted until electron donor consumption had stabilized for at least one month.

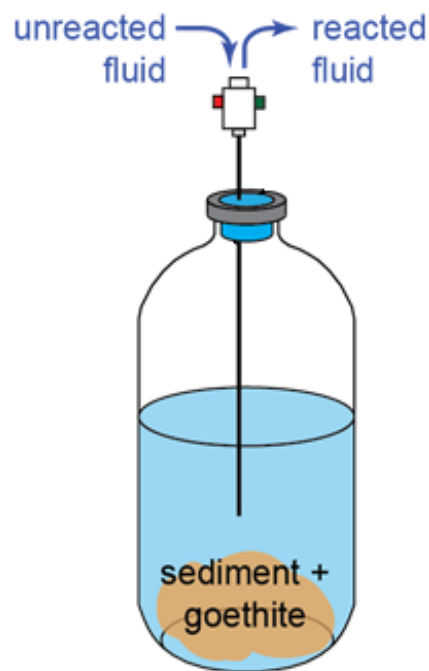


Figure 3: Schematic view of bioreactor. The long needle used for sampling/feeding fluid.

Chemical Analysis

Numerous chemical analyses were performed each week on effluent samples. We also chemically analyzed each batch of fresh media to ensure consistency within our experiment. We used these data to monitor microbial activity during the experiments and provide us with constraints for conducting mass-balance and thermodynamic calculations. Fe(II) concentrations were measured in effluent samples using the ferrozine method (Stookey, 1970) with a Thermo Scientific Genesys 10S UV Vis Spectrophotometer. Total alkalinity was measured using Gran alkalinity titrations with 0.02 N sulfuric acid, and an Oakton PC-300 pH meter. Major anion concentrations (CH_3COO^- , Cl^- , PO_4^{3-} , and SO_4^{2-}) were measured in 0.45 μm filtered samples using a Dionex ICS-1100 ion chromatograph. The system uses carbonate eluent (4.5mM Na_2CO_3 , 1.4mM NaHCO_3) with an AS22 analytical column, and an AERS 500 suppressor. Major cation concentrations (Ca, Na, Mg, NH_4 , K) were measured periodically using a second ICS-1100 ion chromatograph, which uses a sulfuric acid eluent (22mN H_2SO_4 solution) with a CS12 analytical column and a CERS 500 suppressor. For both systems, eluent was made using 18 M Ω DI H_2O . Fe(II) and alkalinity analyses were performed immediately after sampling. Periodically we analyzed the proportion of methane gas in the headspace of the reactors using a GOW MAC series 580 gas chromatograph equipped with a thermal conductivity detector.

Microbial Community Analysis

In order to directly identify which groups of microorganisms were present in the reactors, and help us interpret our geochemical data, we carried out a microbial community analysis of each bioreactor. This analysis was done on samples collected at the end of the experiment for all reactors as well as on sediment collected from the marsh. To collect the microorganisms the

bioreactors were thoroughly mixed, and 3mL of the slurry was removed using sterile 5mL syringes. The slurry was then filtered using a sterile 0.2µm filter. To ensure the filters were sterile, they were first placed in membrane holders, wrapped in aluminum foil, and autoclaved for 30 mins at 121°C. After filtering, the filter was placed in a 2mL centrifuge tube, 0.2 mL of sucrose lysis buffer was added as a preservative, and the filters were frozen at -80°C until DNA extractions were performed.

DNA was extracted from the filters using a MoBio DNA extraction kit. We followed manufacturer's instructions for each step with the exception of using an alternative lysis method to reduce DNA shearing. After extraction, the samples were kept frozen at -80°C. A Nanodrop was used to ensure quality, and measure concentration of the DNA extracted. DNA samples were sent to Argonne National Lab where DNA amplification was carried out using Polymerase Chain Reaction (PCR). PCR amplification was performed using universal bacterial primers 515F (GTGYCAGCMGCCGCGGTAA) and 806R (GGACTACNVGGGTWTCTAAT (Walters, W., 2015). The sequences were then sequenced using the Illumina MiSeq platform. After an initial quality filtering, the lab provided raw sequence data to us in the form of a Fasta file

We analyzed sequence data using QIIME v. 1.9.1. First we split samples according to barcodes, and filtered the sequences to remove low-quality reads (script: split_libraries.py). Next, we generated BIOM formatted OTU (Operational Taxonomic Unit) tables at 97% similarity, and evaluated taxonomy with uclust (script: pick_de_novo_otus.py). This step assigned the most detailed lineage description shared by at least 90% of the sequences within each OTU. Then, singletons were removed, and we created taxonomy tables (scripts: filter_otus_from_table.py, summarize_taxa_through_plots.py). We wanted to compare diversity between samples to examine uniformity of triplicates and influence of pH on community

diversity. In order to do this we first normalized all samples to have the same number of sequences as the sample with the fewest sequences(4834 sequences) (script: `single_rarefaction.py`). From here we computed diversity within samples using alpha diversity (script: `alpha_diversity.py`). Lastly, we computed beta diversity to compare sample communities to one another. Doing this created PCoA plots as well as NMDS plots (script: `beta_diversity_through_plots.py`) (Kirk et al., 2015b).

Chapter 3 - Results

Biogeochemistry

The pH of the effluent samples throughout the experiment from the pH 6.0 and pH 7.5 control reactors averaged 6.33 and 7.37, respectively. The pH of the acetate-fed reactors did not vary substantially from the pH of their corresponding control reactors (Fig 4A).

Initial analysis of the reactor effluent showed substantially higher levels of acetate than was supplied by the media. Average initial acetate concentrations for pH 6.0 and pH 7.5 acetate-fed bioreactors was 2.12mM and 2.10mM, respectively (nearly 8.5 times weekly input value). Average initial acetate concentrations for pH 6.0 and pH 7.5 control bioreactors were 1.74 and 1.57, respectively. Effluent acetate concentrations declined steadily, dropping below input values between incubation day 35 and 42. From incubation day 42 through the end of the experiment, acetate concentrations of effluent samples from all bioreactors remained below detection limit (1mg/L) (Fig 4B).

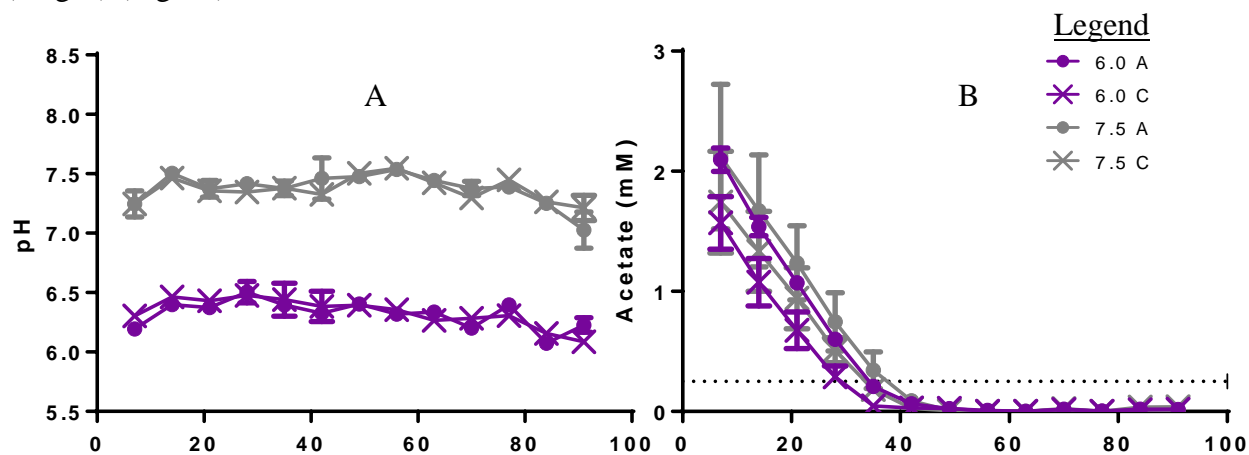


Figure 4: Graphing average pH (A), and acetate values (B) versus time in days. Dotted line illustrates average input of 0.25mM acetate into acetate-fed bioreactors.

Fe(II) concentrations in both acetate-fed and control pH 6.0 bioreactors increased rapidly from day 0, reaching their maximum on day 35 (3,425 μ M) and 28 (3,137 μ M), respectively. Following day 35, Fe(II) concentrations for pH 6.0 bioreactors continued on a downward trend

with the acetate-fed bioreactors maintaining a slightly higher concentration (Fig 6A). Total amounts of Fe(III) reduced in pH 6.0 reactors averaged 6654.7 μM for acetate-fed reactors, and 6273.1 μM for control reactors.

Fe(II) concentrations in both acetate-fed and control pH 7.5 reactors are substantially lower than the pH 6.0 reactors. Maximum concentrations for acetate-fed and control reactors at pH 7.5 reached 207.2 μM and 184.5 μM , respectively. Total Fe(III) reduced during incubation based on mass balance calculations averaged 325.2 and 301.3 μM (Fig 5).

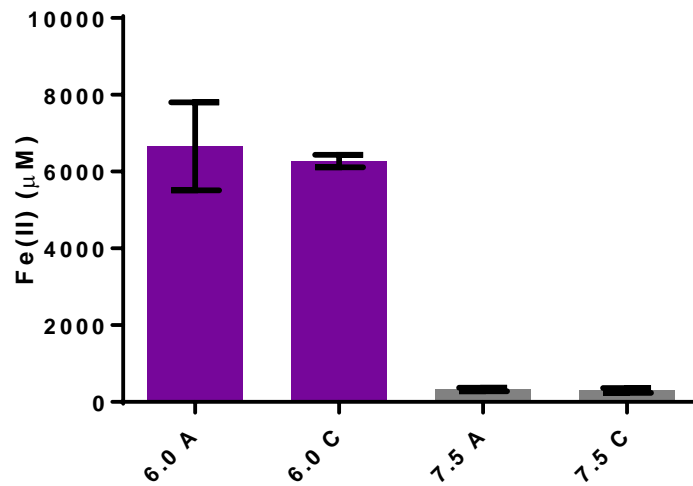


Figure 5: Total amount of Fe(II) in solution at the end of the experiment.

Initially alkalinity data were affected by organic content introduced by the soil. Due to this, reliable alkalinity data were not recorded until day 56. On day 56 average alkalinity values for acetate-fed and control reactors at pH 6.0 were 9.57 and 8.84 meq/L, respectively. The average values for reactors at pH 7.5 were 5.19 and 5.04, respectively (Fig 6B). This higher alkalinity values for pH 6.0 reactors is indicative of Fe(III) reduction as seen in equation (1). On day 91 alkalinity values were substantially lower in the pH 6.0 reactors and relatively unchanged in the pH 7.5 reactors. Average alkalinity values on day 91 for acetate-fed and control reactors at pH 6.0 were 6.68 and 6.64 meq/L, respectively. In pH 7.5 reactors the average values were 5.19

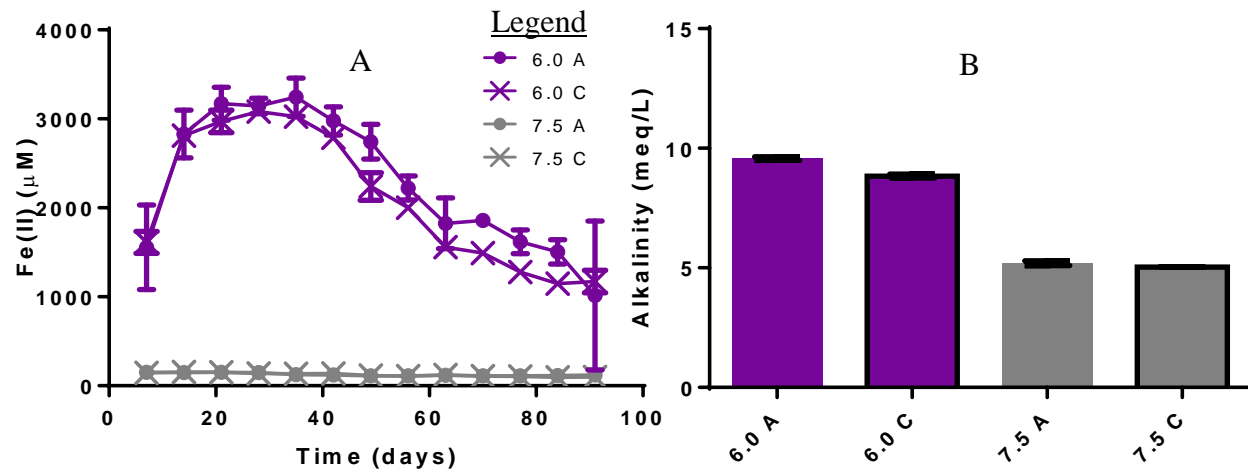


Figure 6: Graph of average Fe(II) concentration of effluent samples (A), and a column chart showing alkalinity values measured on day 56 (B).

and 5.30 meq/L, respectively. This drop in alkalinity between days 56 and 91 in the pH 6.0 bioreactors indicates a decrease in the amount of Fe(III) reduction occurring.

Concentrations of both Ca^{2+} and Mg^{2+} were initially much higher in bioreactors at pH 6.0 versus pH 7.5. On the initial sampling day the average concentration of Ca^{2+} in pH 6.0 and 7.5 bioreactors was 1.3 and 0.65 mM, respectively. The Ca^{2+} concentration was not tested again until day 56. On day 56 the Ca^{2+} concentrations were much closer averaging 0.65 and 0.57 mM, respectively. Although the concentrations converged by day 56, the concentrations remained above the weekly input value of 0.294 mM throughout the entire experiment (Fig 7A).

As stated previously, Mg^{2+} concentrations were initially much higher for pH 6.0 bioreactors versus reactors at pH 7.5. The initial Mg^{2+} concentrations for bioreactors at pH 6.0 and 7.5 were 0.835 and 0.555 mM, respectively. These values would also eventually converge. On day 56 the average concentrations were 0.53 and 0.455 mM, respectively. Unlike Ca^{2+} concentrations which were consistently above the weekly input value, the Mg^{2+} concentrations remained close to the weekly Mg^{2+} input of 0.483 mM after day 56. (Fig 7B).

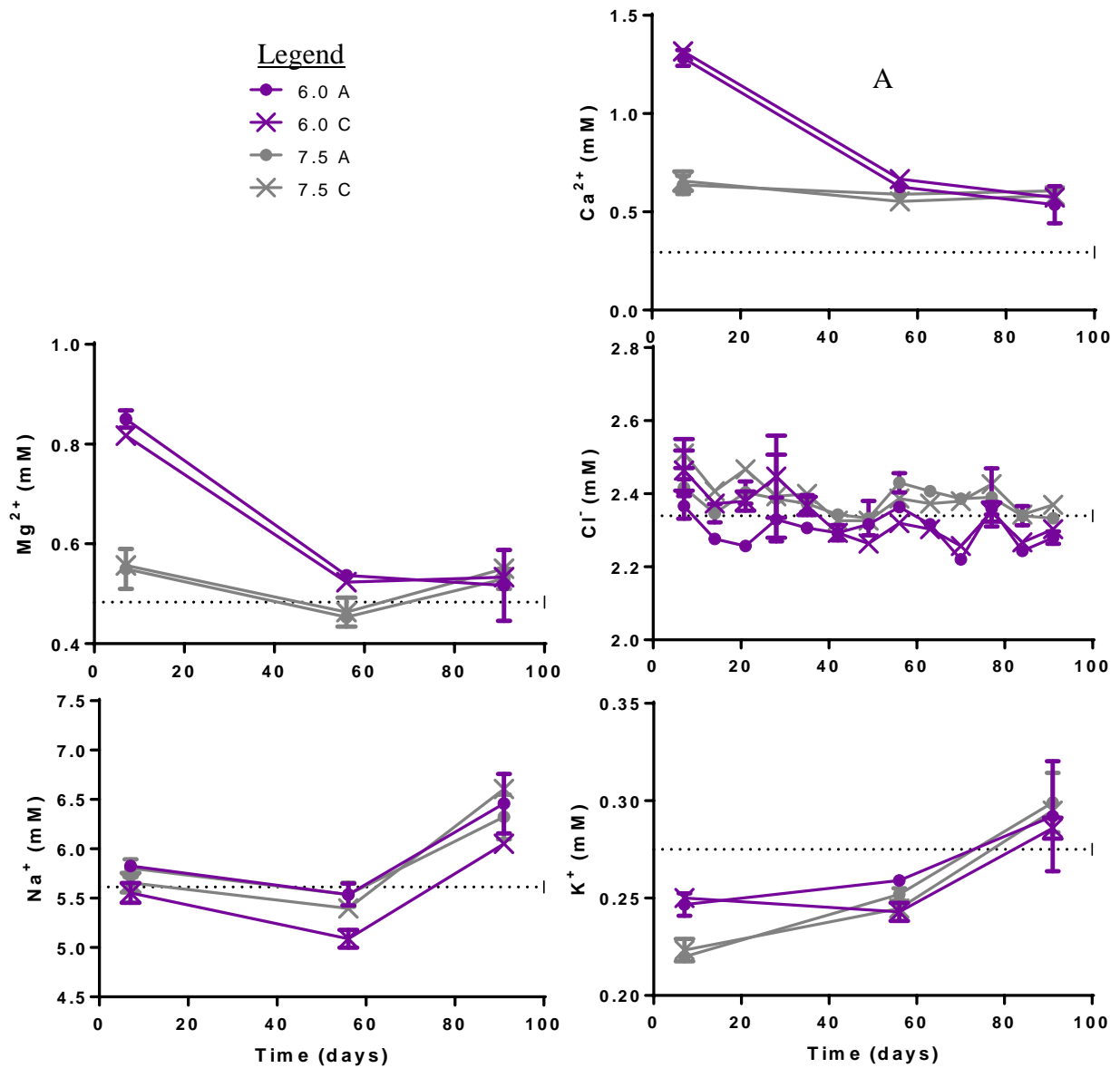


Figure 7: Average concentrations of calcium (A), magnesium (B), chloride (C), sodium (D), and Potassium (E) in effluent samples versus time in days. Average input levels were 0.294mM (A), 0.483mM (B), 2.34mM (C), 5.61mM (D), and 0.275mM (E).

Chloride concentrations throughout the experiment remained consistent. They deviated slightly from the average input of 2.34mM, but no significant trends were noticed (Fig 7C).

B

C

Sodium concentrations of effluent samples were initially near input value of 5.61mM. They dropped slightly below input during day 56 sampling, and then increased by an average of 1.0 mM on the final sampling day (Fig 7D). Potassium concentrations were below input of 0.275mM during the first two samplings. Levels increased slightly above input during the final day (Fig 7E).

Reactor headspace was sampled 3 times throughout the experiment. Methane was detected during every sampling in all 12 bioreactors. Headspace sampling took place before reactor media sampling during the 3rd, 8th, and 12th week of the experiments. Average methane amounts within all pH 6.0 reactors for all three samplings was 27.3, 79, and 78.9 μ moles, respectively. For pH 7.5 reactors, the average amount of methane was 38.5, 119.5, and 127.5 μ moles, respectively (Fig 8B). Total methane production within pH 6.0 reactors averaged 102.4 μ moles for acetate-fed and 55.5 μ moles for control reactors. Averages for pH 7.5 reactors were 163.9 and 91.2 μ moles, respectively (Fig 8A). In order to calculate methane in terms of μ moles we used the ideal gas law ($PV=nRT$). Where (P) is the partial pressure of methane in kPa, (V) is the average volume of headspace within the reactors (0.0594 L), (n) is the number of

moles, (R) is the ideal gas constant (8.314 L kPa/Kmol), and (T) is the temperature in Kelvin (295.5K).

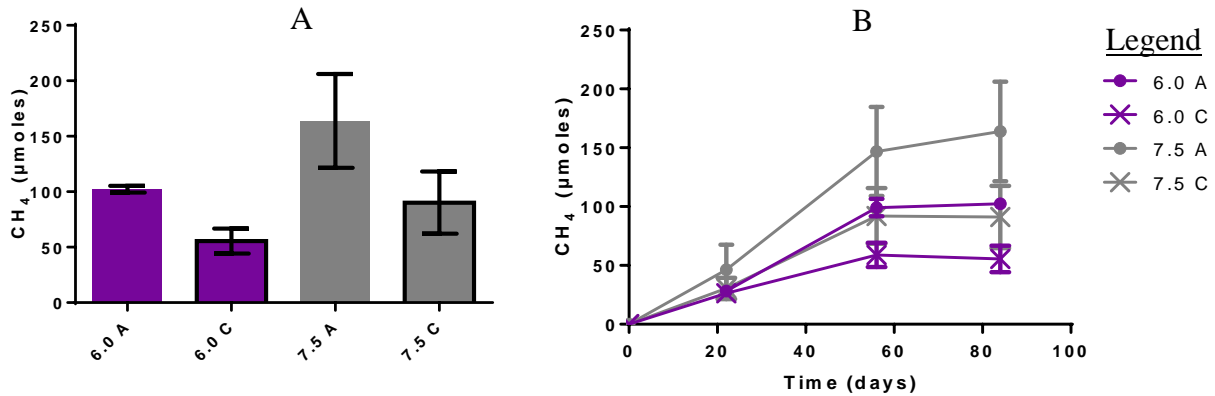


Figure 8: Column chart showing average total methane production (A), and a graph showing methane present in the headspace at each of the 3 sampling days (B).

Microbial Community Composition

Proteobacteria accounted for over 55% of our prokaryotic community (36.4% within marsh sediment. Within this Phylum 27.6% were within the genus *Geobacter* a genus of bacteria known primarily as dissimilatory metal reducers. *Geobacter* had the highest relative abundance within all of our samples (15.29%). Within our prokaryotic community archaea only accounted for 1.34% of sequences. Of the archaea present 51% were within orders of methanogens (0.69% relative abundance). Methanogens found in our samples were from the classes *Methanobacteria*, *Methanomicrobia*, and *Thermoplasmata*.

Relative abundance of *Geobacter* and methanogens within marsh sediment was 0.81% and 0.35%, respectively. Within pH 6.0 reactors, relative abundance of *Geobacter* and Methanogens averaged ~22% and 0.47%, respectively. In the pH 7.5 reactors, *Geobacter* averaged 13.5% and Methanogens averaged around 1% (Figs 9A,B).

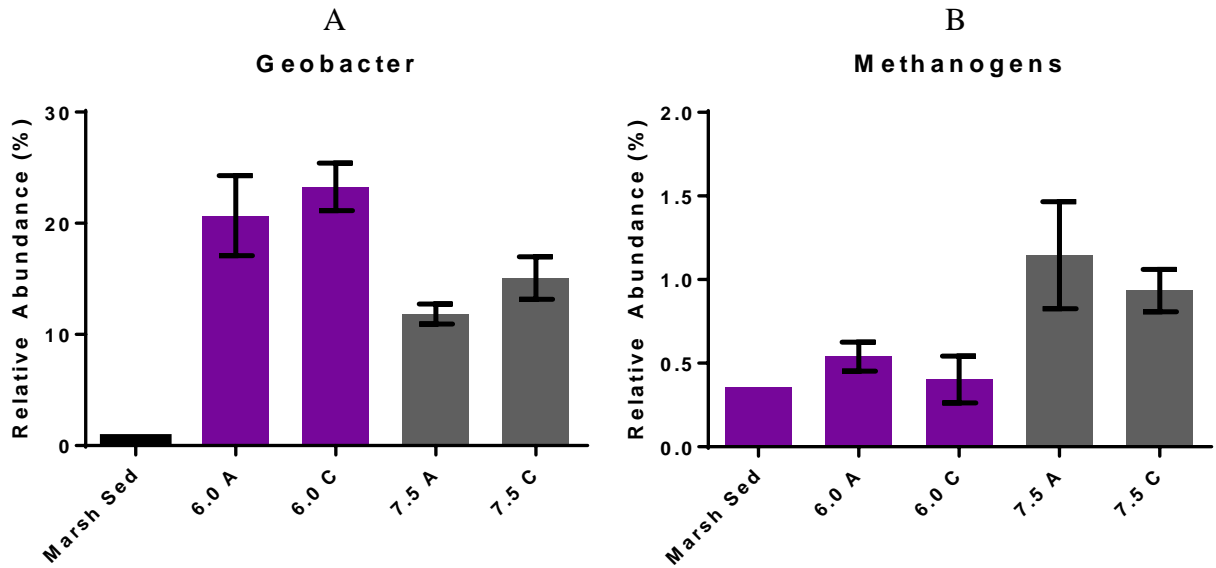


Figure 9: Column charts showing relative abundance of *Geobacter* (A) and methanogens (B) within marsh sediment and all groups of bioreactors.

Chapter 4 - Discussion

Microbial Reactions

The observed changes in aqueous chemistry and headspace composition are consistent with the growth of Fe(III)- reducers and methanogens. Early in the experiment there was a noticeable decrease in effluent acetate concentrations, an increase in Fe(II) and alkalinity, as well as noticeable methane production. Results indicate growth within populations of both methanogens and Fe(III) reducers. Stabilization of effluent acetate concentrations below detection (0.01mM) following day 42 indicates that the microbial community had grown enough to consume nearly all of the influent acetate each week.

Mass-balance calculations based on aqueous chemistry demonstrate that the extent to which Fe(III) reduction occurred differed considerably between each set of reactors. In bioreactors at pH 6.0, the acetate consumed for Fe(III) reduction in acetate-fed and control reactors averaged 83.18 and 78.41 μ moles, respectively. The amount of acetate consumed in pH 7.5 bioreactors for Fe(III) reduction in acetate-fed and control reactors averaged 4.06 and 3.77 μ moles, respectively (Fig 10).

On average, methanogenesis required more acetate than Fe(III) reduction. The amount of acetate consumed by methanogens in the acetate-fed and control reactors at pH 6.0 was 102.4 and 55.50 μ moles, respectively. Within the pH7.5 reactors, methanogens consumed 163.9 and 91.2 μ moles, respectively (Fig 10).

Contrary to the pH 6.0 reactors, the difference in acetate consumption for methanogenesis and Fe(III) reduction in pH 7.5 reactors was substantial. Within acetate-fed reactors at pH 7.5, methanogens consumed 163.9 μ moles of acetate and Fe(III) reducers only 5.06 μ moles. This

resulted in minimal Fe(II) production, and an increased amount of methane production in pH 7.5 reactors.

We understand that acetate was not the only organic molecule being metabolized by these microbes. Degradation of organic matter within the soil most likely provided other forms of electron donor, but for simplification we are comparing them based upon 8 electron transfer. In addition we understand that acetate could have been consumed by other electron acceptors such as NO_3^- , SO_4^{2-} , or Mn(IV). Given that SO_4^{2-} and NO_3^- levels were below detection we feel their impact is minimal. Mn(IV) on the other hand was not quantified, so this provides possible uncertainty.

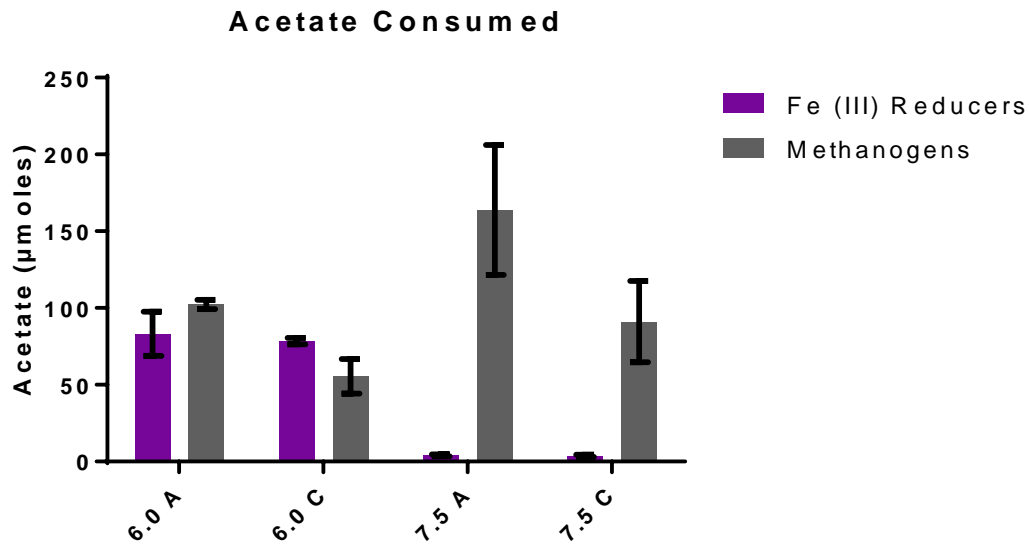


Figure 10: Amount of acetate consumed throughout the experiment by Fe(III) reducers and methanogens in all sets of bioreactors.

Although methanogens had the more energetically favorable reaction within the pH 7.5 reactors, it is evident that Fe(III) reducers were still active. The total amount of Fe(III) reduced in the pH 7.5 reactors averaged only 313.25 μ M (~6500 μ M in pH 6.0 reactors). Evidence of Fe(III) reducer activity is shown in the growth of microorganisms within the *Geobacter* genus (Fig 11). Average relative abundance of *Geobacter* within marsh sediment was only 0.8%, but within the pH 6.0 and pH 7.5 bioreactors they averaged 22% and 13.45%, respectively. So even though very little Fe(III) was reduced within pH 7.5 reactors, the population of Fe(III) reducers increased more than 16 times. Fe(III) reducers may possess a kinetic advantage over methanogens that allows them to consume electron donor more quickly (Bethke et al., 2008). The extent to which they can exploit this advantage may depend in part upon thermodynamics

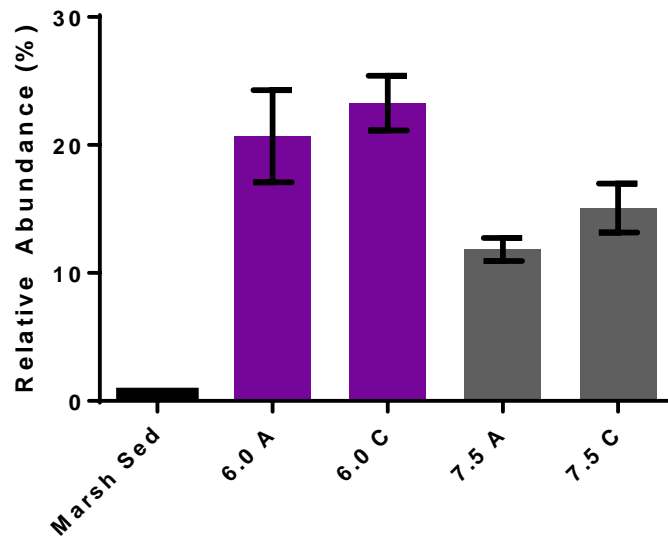


Figure 11: Relative abundance of *Geobacter* within marsh sediment and bioreactors.

(Bethke et al., 2011). It seems that even at alkaline pH, where methanogenesis is more thermodynamically favorable, Fe(III) reducers are still able to exploit this advantage, but not to reduce Fe(III). A possible explanation for this phenomena is Direct Interspecies Electron Transfer (DIET) (Lovley, 2017). While still not extensively studied, DIET and cable bacteria

have demonstrated that microbial cells can exchange electrons over distances of μm to cm lengths through electrical connections (Lovley, 2017). DIET is potentially an effective form of syntrophy in methanogenic communities (Rotaru et al., 2014a). Electrically connected microbial communities (e-communities) are currently primarily restricted to anaerobic environments in which bacteria and archaea electrically connect to either produce (Morita et al., 2011; Kato et al., 2012; Rotaru et al., 2014b) or consume (McGlynn et al., 2015; Wegener et al., 2015) methane. Under alkaline conditions *Geobacter* may be consuming acetate, but instead of utilizing it to reduce Fe(III) they may be transferring the electrons to methanogens (Fig 12).

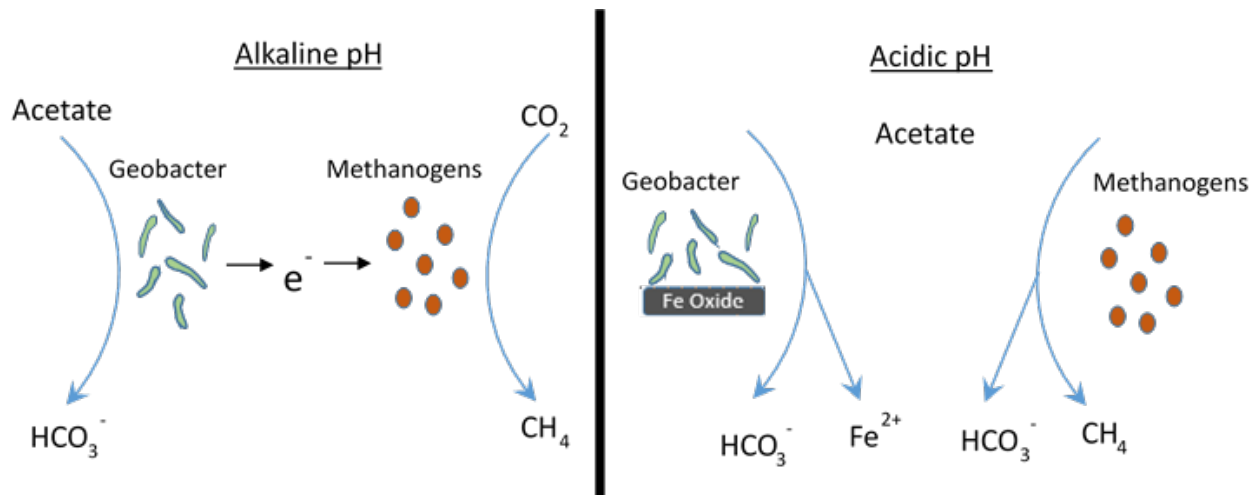


Figure 12: Diagram illustrating Direct Interspecies Electron Transfer (DIET) between *Geobacter* and methanogens. At alkaline pH *Geobacter* utilizes its kinetic advantage to transfer electrons to methanogens. At acidic pH *Geobacter* reduces Fe(III) because that reaction is more favorable.

Potential Mechanisms

Potential mechanisms that could cause changes in the interactions between these two microbial reactions are bioenergetics, and variations in the ability and extent of sorption of Fe(II) onto goethite. In the following sections we consider each of these possibilities in more detail.

Bioenergetics

Reduction of ferric iron oxides and oxyhydroxides consumes several protons as shown in the following reaction which exemplifies the reaction that is occurring within our bioreactors.



As stated previously, due to this consumption of protons, the energy yield of the reaction increases rapidly as pH decreases (Fig 1). For this reason, Fe(III) reduction is more energetically favorable in acidic environments. The consumption of protons by Fe(III) reduction also leads to an increase in pH. The peak of Fe(III) reduction (~35 days) correlates with the highest pH levels recorded within the pH6.0 reactors.

Contrary to Fe(III) reduction, methanogenesis consumes few protons and thus varies little with changes in pH (Equation 3). Due to this, methanogenesis remains energetically favorable along a wide range of pH. So, although Fe(III) reduction has more energy available at low pH, methanogenesis can still occur. This is why we see methane production within our pH 6.0 reactors.



We carried out thermodynamic calculations to determine amounts of energy available to drive the reactions forward. Using Geochemist's Workbench we derived activity values for all gaseous and aqueous species, as well as log K values for each reaction. Using the log K values we calculated standard state Gibbs Free Energy (ΔG°_T) (Equation 4).

$$\Delta G^{\circ}_T = -2.303RT \log(K) \quad (4)$$

In this equation R represents the gas constant (8.314 J/Kmol) and T is temperature. Using the activity values we then calculated the reaction quotient (Q), and applied these values to the reaction free energy equation to find the available free energy.

$$\Delta G_r = \Delta G^{\circ}_T + RT \ln(Q) \quad (5)$$

Microbes can only run their metabolism when the energy available outside the cell exceeds the internal store (Jin and Bethke, 2009). Usable free energy (ΔG_U) is the difference between the energy in the environment and within the cell. ΔG_U is calculated using the following equation (Jin and Bethke, 2002, 2003, 2005, 2007).

$$\Delta G_U = \Delta G_A - m\Delta G_P \quad (6)$$

Our calculations reveal that there is generally more usable energy available to Fe(III) reducers at acidic pH, and more usable energy available to methanogens at alkaline pH (Fig 13).

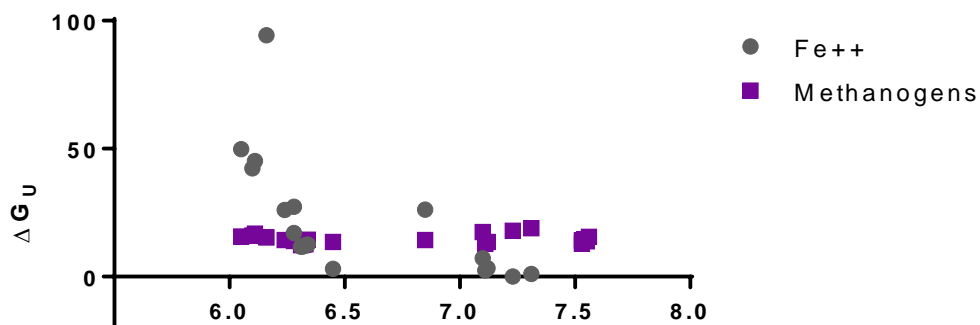


Figure 13: Graph showing shift in usable free energy available to methanogens and Fe(III) reducers with changing pH.

As the amount of usable energy decreases, the reaction rates start to slow down. As pH increases the amount of usable energy available to Fe(III) reducers begin to decrease. Fe(III) reducers are able to run their reactions at high rates at acidic pH, but at alkaline pH they are limited by thermodynamics. Methanogens are not affected in the same way by thermodynamics. Therefore, under alkaline conditions, where Fe(III) reducers are lacking in available energy, methanogens are able to thrive.

Ferrous Iron Sorption

Another potential mechanism for controls on the microbial reactions within our experiments is Fe(II) sorption onto goethite. When Fe(II) sorbs to a ferric surface, in this case goethite, valence electrons are taken up and conducted toward high potential sites (Williams and Scherer, 2004; Larese-Casanova and Scherer, 2007). The electrons lower the effective redox potential of the surface, degrading its ability to act as an electron acceptor (Handler and other, 2009; Rosso and others, 2010). Under acidic conditions Fe(II) generally doesn't sorb as well. So as pH increases, Fe(II) sorption should increase (Dixit and Hering, 2006), and the rate of Fe(III)

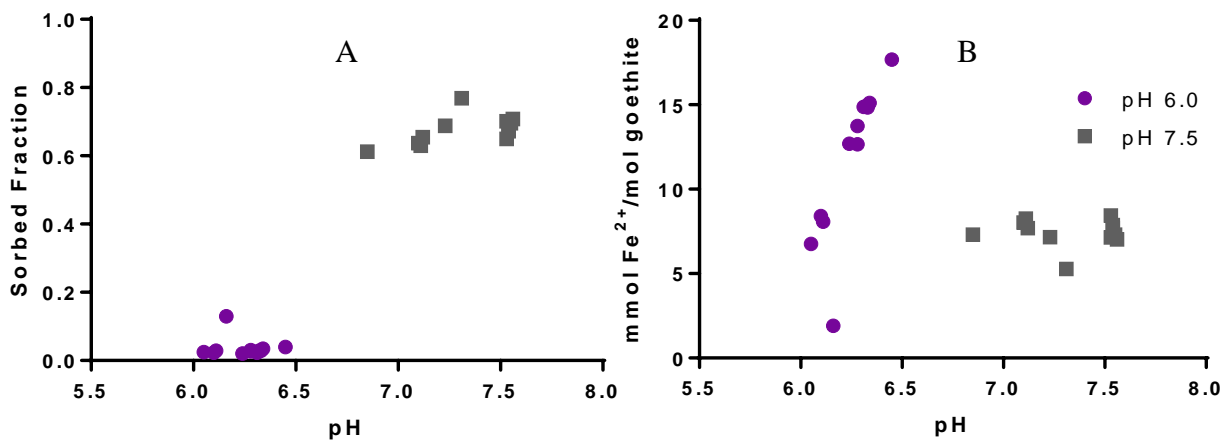


Figure 14: Graphs showing fraction of Fe(II) that has sorbed onto the goethite surface (A), and the density of Fe(II) sorbed on goethite within the bioreactors (B).

reduction should decrease. We created a sorption model using Geochemist's Workbench and from this we calculated sorbed fractions of Fe(II) as well as the density of sorbed Fe(II) on goethite (Fig 14). Sorbed fraction refers to the fraction of the total Fe(II) in solution that has sorbed onto goethite. Sorbed density is referring to the total amount of Fe(II) that sorbed. Through these calculations it is evident that there is a higher fraction of Fe(II) sorbing at basic pH, but due to the increased Fe(II) present in solution there is a much higher density of sorbed Fe(II) at acidic pH. This is inconsistent with sorption being a potential mechanism driving shifts in interactions with pH.

Implications

Small ponds, lakes, and marshes, like the one we sampled for this study, have a large impact on our greenhouse gas budgets. A recent article discusses the contribution of small inland lakes and ponds on CO₂ and CH₄ emissions. They found that inland water actively transforms carbon, and plays an important role in natural and anthropogenic greenhouse gas budgets, including CO₂ and CH₄. According to the study, ponds that fell within the smallest size range have a disproportionately large contribution to carbon flux relative to their size. In fact, they estimate that although small ponds only make up 8.6% of the global surface area of lakes and ponds, they comprise 15.1% of all CO₂ emissions and 40.6% of all diffusive CH₄ emissions from lentic freshwaters (Holgerson and Raymond, 2016).

In order to understand how these systems will respond to changing climate, we need to understand the environmental controls on microbial reaction within them. As temperature increases, so do rates of organic matter degradation (Davidson, 2006). This increased rate of degradation could increase the levels of CO₂ in anoxic soil and water, and in turn lower the pH.

This change in pH has the potential to increase the ability of Fe(III) reducers to compete with methanogens. This could work to lower the amount of methane being released from these systems. Moreover, because Fe(III) reduction consumes a lot of protons, the reaction is very effective at generating carbonate alkalinity (Kirk et al., 2013), which would help these systems store additional carbon. Methanogenesis generates little alkalinity and releases carbon in the form of CH₄. Carbon can also be stored through precipitation of carbon bearing minerals from solution. After calculating mineral saturations using Geochemist's Workbench we found that siderite, an iron carbonate mineral (FeCO₃), was super saturated in nearly 60% of samples with a higher percentage of them being in the pH 6.0 reactors. Thus, shifts in interactions between these two groups with pH would have significant implications for carbon budgets in anoxic systems.

Chapter 5 - Conclusions

The balance between methanogenesis and Fe(III) reduction in our experiments shifted considerably with pH. The reactions consumed nearly equivalent amounts of electron donors in reactors with acidic pH, but methanogenesis consumed 40X more at alkaline pH. Despite this shift, the abundance of Fe(III) reducers and methanogens appears to have changed relatively little. We hypothesize that Fe(III) reducer's abundance remained high despite the low extent of Fe(III) reduction because they increasingly interacted with methanogens via interspecies electron transfer at alkaline pH. Thermodynamic calculations suggest that thermodynamic controls on microbial reactions plays an important role in the shift between Fe(III) reduction and methanogenesis. Fe(II) sorption onto goethite, however, appears to have had less of an impact on the balance of each reaction based on surface complexation model calculations. Shifts in the balance between methanogenesis and Fe(III) reduction can have significant implications on carbon budgets within anoxic systems. As Fe(III) reduction increases these systems can become important carbon storage reservoirs by increasing carbonate alkalinity, decreasing methane production, and favoring siderite precipitation.

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Appendix A - Water Chemistry

6.0A Water Chemistry														
date	time days	mass (g)			pH			Media pH	Fe(II) (µM)			Alk (meq/L)		
		6A-1	6A-2	6A-3	6A-1	6A-2	6A-3		6A-1	6A-2	6A-3	6A-1	6A-2	6A-3
4/26/2016	7	212.32	212	212.09	6.21	6.22	6.15	6.04	1072.2	2021.4	1576.9			
6/2/2016	14	211.91	212.98	212.41	6.37	6.44	6.39	5.92	2563.7	3101.6	2821.7			
6/9/2016	21	210.92	212.63	212.08	6.4	6.37	6.35	5.9	3061.4	3383.9	3068.3			
6/16/2016	28	210.87	212.75	212.05	6.6	6.49	6.42	5.9	3130.0	3068.3	3239.8			
6/23/2016	35	212.25	212.84	212.68	6.4	6.35	6.44	6.05	3006.5	3425.0	3301.5			
6/30/2016	42	212.08	212.98	212.65	6.37	6.31	6.3	5.88	2800.7	3020.3	3109.5			
7/7/2016	49	211.46	212.56	212.37	6.37	6.42	6.42	6.05	2533.2	2917.4	2780.2			
7/14/2016	56	211.71	212.36	212.39	6.32	6.31	6.33	6.07	2238.2	2347.9	2080.4	9.48	9.61	9.62
7/21/2016	63	211.81	212.66	212.11	6.3	6.35	6.36	5.99	1613.9	1716.8	2149.0			
7/28/2016	70	211.49	212.66	211.93	6.17	6.25	6.18	6.09	1943.2	1806.0	1826.6			
8/4/2016	77	210.8	213.12	212.25	6.42	6.39	6.38	6.05	1469.8	1661.9	1723.6			
8/11/2016	84	211.81	212.95	211.88	6.03	6.07	6.12	5.97	1408.1	1449.2	1661.9			
8/18/2016	91	N/A	N/A	N/A	6.16	6.28	6.24	N/A	70.3	1312.0	1661.9	5.11	7.45	7.49

date	time days	CH3COO-						Adjusted Cl-					
		6A-1		6A-2		6A-3		6A-1		6A-2		6A-3	
		mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM
4/26/2016	7	125.41	2.12	128.67	2.18	117.35	1.99	82.77	2.33	85.11	2.40	84.04	2.37
6/2/2016	14	90.10	1.53	95.46	1.62	87.08	1.47	80.51	2.27	80.98	2.28	80.71	2.28
6/9/2016	21	60.47	1.02	66.57	1.13	62.47	1.06	79.82	2.25	79.86	2.25	80.50	2.27
6/16/2016	28	32.01	0.54	36.55	0.62	37.79	0.64	80.53	2.27	82.58	2.33	84.77	2.39
6/23/2016	35	9.58	0.16	12.32	0.21	15.15	0.26	81.63	2.30	81.97	2.31	82.04	2.31
6/30/2016	42	2.34	0.04	3.63	0.06	5.57	0.09	81.75	2.31	80.58	2.27	81.46	2.30
7/7/2016	49	1.22	0.02	1.32	0.02	1.55	0.03	81.79	2.31	82.19	2.32	82.33	2.32
7/14/2016	56	0.56	0.01	0.54	0.01	0.53	0.01	83.68	2.36	83.46	2.35	84.36	2.38
7/21/2016	63	0.00	0.00	0.00	0.00	0.00	0.00	81.72	2.31	81.95	2.31	82.67	2.33
7/28/2016	70	0.91	0.02	1.07	0.02	0.89	0.02	78.22	2.21	78.78	2.22	79.02	2.23
8/4/2016	77	0.00	0.00	0.00	0.00	0.32	0.01	83.43	2.35	84.49	2.38	83.27	2.35
8/11/2016	84	1.30	0.02	1.04	0.02	1.37	0.02	78.96	2.23	79.81	2.25	79.74	2.25
8/18/2016	91	0.85	0.01	1.05	0.02	1.02	0.02	80.14	2.26	81.09	2.29	81.18	2.29

date	time days	Na+						K+											
		6A-1		6A-2		6A-3		6A-1		6A-2		6A-3							
		mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM						
4/26/2016	7	133.32	5.80	134.26	5.84	134.38	5.84	9.85	0.25	9.56	0.24	9.59	0.25						
6/2/2016	14																		
6/9/2016	21																		
6/16/2016	28																		
6/23/2016	35																		
6/30/2016	42																		
7/7/2016	49																		
7/14/2016	56	128.42	5.59	124.38	5.41	129.08	5.61	10.15	0.26	10.10	0.26	10.14	0.26						
7/21/2016	63																		
7/28/2016	70																		
8/4/2016	77																		
8/11/2016	84																		
8/18/2016	91	143.17	6.23	156.26	6.80	145.94	6.35	10.30	0.26	12.51	0.32	11.45	0.29						
Mg++																			
date	time days	6A-1			6A-2			6A-3			6A-1			6A-2			6A-3		
		mg/L	mM		mg/L	mM		mg/L	mM		mg/L	mM		mg/L	mM		mg/L	mM	
4/26/2016	7	20.84	0.86		20.93	0.86		20.13	0.83		51.89	1.29		53.10	1.32		49.72	1.24	
6/2/2016	14																		
6/9/2016	21																		
6/16/2016	28																		
6/23/2016	35																		
6/30/2016	42																		
7/7/2016	49																		
7/14/2016	56	13.08	0.54		12.87	0.53		13.24	0.54		25.41	0.63		25.01	0.62		25.13	0.63	
7/21/2016	63																		
7/28/2016	70																		
8/4/2016	77																		
8/11/2016	84																		
8/18/2016	91	10.80	0.44		14.01	0.58		12.98	0.53		17.10	0.43		24.29	0.61		22.80	0.57	

6.0C Water Chemistry														
date	time days	mass (g)			pH			Media pH	Fe(II) (µM)			Alk (meq/L)		
		6C-1	6C-2	6C-3	6C-1	6C-2	6C-3		6C	6C-1	6C-2	6C-3	6C-1	6C-2
4/26/2016	7	212.53	212.42	213.13	6.33	6.3	6.28	6.19	1634.5	1478.1	1722.3			
6/2/2016	14	212.32	212.37	212.74	6.5	6.43	6.46	5.99	2851.9	2876.6	2700.9			
6/9/2016	21	212.03	211.99	211.94	6.4	6.43	6.46	5.91	3102.6	2848.8	2965.4			
6/16/2016	28	212.63	212.74	211.83	6.41	6.5	6.52	5.99	3095.7	3136.9	3013.4			
6/23/2016	35	212.53	212.54	211.65	6.37	6.35	6.6	6.14	2944.8	3095.7	3034.0			
6/30/2016	42	212.77	212.48	211.77	6.32	6.53	6.3	6.87	2807.6	2711.6	2841.9			
7/7/2016	49	212.35	212.26	214.03	6.37	6.4	6.41	6.08	2293.1	2361.7	2066.7			
7/14/2016	56	212.45	212	214.04	6.34	6.28	6.45	5.88	1970.6	2025.5	2004.9	8.85	8.91	8.75
7/21/2016	63	212.13	212.05	214.34	6.31	6.26	6.23		1545.3	1655.0	1476.7			
7/28/2016	70	212.76	212.19	212.36	6.27	6.27	6.31	5.89	1456.1	1511.0	1504.1			
8/4/2016	77	212.81	212.13	212.83	6.34	6.32	6.26	6.01	1305.2	1298.3	1229.7			
8/11/2016	84	213.04	212.41	213.1	6.21	6.16	6.1	5.89	1154.2	1174.8	1113.1			
8/18/2016	91	N/A	N/A	N/A	6.11	6.1	6.05	N/A	1092.5	1318.9	1106.2	6.52	6.82	6.58

CH3COO-																
date	time days	6C-1			6C-2			6C-3			Adjusted Cl-					
		mg/L	mM		mg/L	mM		mg/L	mM		mg/L	mM	mm	mg/L	mM	
4/26/2016	7	81.73	1.38		89.64	1.52		107.01	1.81		85.39	2.41	87.10	2.46	89.33	2.52
6/2/2016	14	53.40	0.90		61.54	1.04		76.33	1.29		83.23	2.35	84.44	2.38	84.80	2.39
6/9/2016	21	31.86	0.54		38.42	0.65		49.44	0.84		83.28	2.35	84.65	2.39	84.91	2.40
6/16/2016	28	13.22	0.22		15.60	0.26		23.12	0.39		83.42	2.35	85.72	2.42	91.04	2.57
6/23/2016	35	2.32	0.04		3.55	0.06		2.47	0.04		82.83	2.34	84.15	2.37	84.72	2.39
6/30/2016	42	1.05	0.02		1.57	0.03		2.35	0.04		81.01	2.29	81.77	2.31	80.86	2.28
7/7/2016	49	1.48	0.03		1.58	0.03		1.06	0.02		79.26	2.24	80.36	2.27	80.89	2.28
7/14/2016	56	0.00	0.00		0.00	0.00		0.00	0.00		82.14	2.32	81.94	2.31	82.56	2.33
7/21/2016	63	0.00	0.00		0.00	0.00		0.00	0.00		81.17	2.29	82.20	2.32	81.45	2.30
7/28/2016	70	0.65	0.01		0.92	0.02		0.96	0.02		79.34	2.24	80.04	2.26	80.31	2.27
8/4/2016	77	0.00	0.00		0.00	0.00		0.00	0.00		84.16	2.37	83.75	2.36	82.42	2.32
8/11/2016	84	0.82	0.01		0.84	0.01		1.04	0.02		80.80	2.28	80.20	2.26	80.09	2.26
8/18/2016	91	1.01	0.02		0.98	0.02		0.95	0.02		81.65	2.30	82.13	2.32	81.06	2.29

date	time days	Na+						K+								
		6C-1 mg/L	6C-1 mM	6C-2 mg/L	6C-2 mM	6C-3 mg/L	6C-3 mM	6C-1 mg/L	6C-2 mM	6C-3 mg/L	6C-3 mM					
4/26/2016	7	125.31	5.45	130.00	5.65	127.86	5.56	9.69	0.25	9.90	0.25	9.74	0.25			
6/2/2016	14															
6/9/2016	21															
6/16/2016	28															
6/23/2016	35															
6/30/2016	42															
7/7/2016	49															
7/14/2016	56	118.78	5.17	117.44	5.11	114.74	4.99	9.67	0.25	9.51	0.24	9.31	0.24			
7/21/2016	63															
7/28/2016	70															
8/4/2016	77															
8/11/2016	84															
8/18/2016	91	140.16	6.10	137.29	5.97	139.97	6.09	10.97	0.28	11.19	0.29	11.39	0.29			
Mg++																
date	time days	6C-1			6C-2			6C-3			Ca++					
		mg/L	mM	0.81	mg/L	mM	0.83	mg/L	mM	0.81	mg/L	mM	1.36	mg/L	mM	1.31
4/26/2016	7	19.75	0.81	20.27	0.83	19.77	0.81	54.35	1.36	52.48	1.31	51.41	1.28			
6/2/2016	14															
6/9/2016	21															
6/16/2016	28															
6/23/2016	35															
6/30/2016	42															
7/7/2016	49															
7/14/2016	56	12.85	0.53	12.84	0.53	12.44	0.51	27.22	0.68	26.78	0.67	26.10	0.65			
7/21/2016	63															
7/28/2016	70															
8/4/2016	77															
8/11/2016	84															
8/18/2016	91	12.91	0.53	13.00	0.53	13.12	0.54	22.38	0.56	22.92	0.57	23.54	0.59			

7.5A Water Chemistry																	
date	time days	mass (g)			pH			Media pH	Fe(II) (µM)			Alk (meq/L)					
		7.5A-1	7.5A-2	7.5A-3	7.5A-1	7.5A-2	7.5A-3		7.5A-1	7.5A-2	7.5A-3	7.5A-1	7.5A-2	7.5A-3			
4/26/2016	7	212.01	212.41	212.04	7.37	7.21	7.16	8.05	116.9	128.9	207.2						
6/2/2016	14	211.06	212.17	212.06	7.55	7.5	7.46	7.61	124.2	148.9	176.3						
6/9/2016	21	211.12	212.35	211.57	7.44	7.38	7.3	7.53	125.2	139.2	194.6						
6/16/2016	28	211.32	212.1	211.59	7.42	7.46	7.37	7.6	157.7	120.3	164.4						
6/23/2016	35	211.96	213.01	212.43	7.31	7.43	7.39	7.6	110.9	117.2	150.4						
6/30/2016	42	211.86	212.95	212.23	7.36	7.66	7.36	7.5	110.9	115.6	142.2						
7/7/2016	49	211.07	212.75	211.16	7.45	7.51	7.47	7.54	107.4	104.6	118.9						
7/14/2016	56	211.19	212.44	211.29	7.53	7.55	7.53	7.59	101.9	104.9	128.2	5.28		5.22			
7/21/2016	63	211.07	212.29	211.37	7.44	7.45	7.44	7.53	109.1	111.2	130.9						
7/28/2016	70	211.36	212.53	212.55	7.37	7.44	7.32	7.57	107.4	105.4	119.7						
8/4/2016	77	210.99	212.94	212.38	7.35	7.41	7.41	7.44	111.8	103.2	115.0						
8/11/2016	84	212.64	213.01	212.34	7.21	7.3	7.24	7.51	119.2	105.4	115.9						
8/18/2016	91	N/A	N/A	N/A	6.85	7.12	7.11	N/A	117.5	115.9	127.9	4.99		4.97	5.24		
CH3COO-																	
date	time days	7.5A-1			7.5A-2			7.5A-3			Adjusted Cl-						
		mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM
4/26/2016	7	99.71	1.69	110.22	1.87	165.77	2.81		84.67	2.39	86.32	2.43		86.26	2.43		
6/2/2016	14	78.73	1.33	87.31	1.48	129.99	2.20		82.20	2.32	84.02	2.37		83.17	2.35		
6/9/2016	21	60.19	1.02	64.68	1.10	93.87	1.59		84.06	2.37	86.08	2.43		85.41	2.41		
6/16/2016	28	33.19	0.56	38.98	0.66	60.04	1.02		83.85	2.37	84.97	2.40		84.71	2.39		
6/23/2016	35	12.29	0.21	18.31	0.31	30.31	0.51		83.70	2.36	84.95	2.40		83.83	2.36		
6/30/2016	42	2.73	0.05	5.02	0.09	7.76	0.13		82.92	2.34	83.39	2.35		82.81	2.34		
7/7/2016	49	0.65	0.01	0.48	0.01	0.83	0.01		83.13	2.35	80.95	2.28		84.14	2.37		
7/14/2016	56	0.00	0.00	0.42	0.01	0.44	0.01		85.18	2.40	86.40	2.44		86.94	2.45		
7/21/2016	63	0.00	0.00	0.00	0.00	0.00	0.00		84.69	2.39	85.66	2.42		85.43	2.41		
7/28/2016	70	1.02	0.02	0.88	0.01	0.77	0.01		83.90	2.37	85.08	2.40		84.58	2.39		
8/4/2016	77	0.00	0.00	0.00	0.00	0.00	0.00		81.68	2.30	87.01	2.45		85.92	2.42		
8/11/2016	84	0.87	0.01	0.83	0.01	0.70	0.01		82.09	2.32	82.48	2.33		84.08	2.37		
8/18/2016	91	0.70	0.01	0.56	0.01	0.64	0.01		82.29	2.32	82.83	2.34		82.81	2.34		

date	time days	Na+			K+								
		7.5A-1 mg/L	mM	7.5A-2 mg/L	mM	7.5A-3 mg/L	mM						
4/26/2016	7	130.98	5.70	134.88	5.87	134.15	5.84	8.47	0.22	8.64	0.22	8.72	0.22
6/2/2016	14												
6/9/2016	21												
6/16/2016	28												
6/23/2016	35												
6/30/2016	42												
7/7/2016	49												
7/14/2016	56	128.27	5.58	124.08	5.40	129.40	5.63	9.97	0.25	9.72	0.25	9.86	0.25
7/21/2016	63												
7/28/2016	70												
8/4/2016	77												
8/11/2016	84												
8/18/2016	91	144.85	6.30	140.44	6.11	150.79	6.56	11.71	0.30	11.09	0.28	12.28	0.31

date	time days	Mg++			Ca++								
		7.5A-1 mg/L	mM	7.5A-2 mg/L	mM	7.5A-3 mg/L	mM						
4/26/2016	7	12.40	0.51	13.42	0.55	14.37	0.59	24.65	0.61	24.32	0.61	27.66	0.69
6/2/2016	14												
6/9/2016	21												
6/16/2016	28												
6/23/2016	35												
6/30/2016	42												
7/7/2016	49												
7/14/2016	56	11.13	0.46	10.71	0.44	11.16	0.46	23.26	0.58	23.09	0.58	24.61	0.61
7/21/2016	63												
7/28/2016	70												
8/4/2016	77												
8/11/2016	84												
8/18/2016	91	12.99	0.53	12.46	0.51	13.31	0.55	24.03	0.60	23.47	0.59	25.44	0.63

7.5C Water Chemistry														
date	time days	mass (g)			pH			Media pH	Fe(II) (µM)			Alk (meq/L)		
		7.5C-1	7.5C-2	7.5C-3	7.5C-1	7.5C-2	7.5C-3		7.5C	7.5C-1	7.5C-2	7.5C-3	7.5C-1	7.5C-2
4/26/2016	7	211.9	211.99	212.16	7.34	7.22	7.18	7.56	120.8	152.9	161.4			
6/2/2016	14	210.86	211.68	211.41	7.54	7.42	7.43	7.52	126.9	151.6	184.5			
6/9/2016	21	210.38	211.44	211.39	7.41	7.33	7.33	7.53	131.8	159.8	165.8			
6/16/2016	28	210.55	212.18	211.72	7.28	7.39	7.37	7.41	124.1	144.7	149.6			
6/23/2016	35	211.38	212.07	212.99	7.35	7.36	7.43	7.49	118.6	137.3	147.4			
6/30/2016	42	211.25	212.07	212.97	7.38	7.29	7.31	7.54	115.6	157.0	138.6			
7/7/2016	49	210.41	211.74	212.57	7.53	7.5	7.48	7.6	102.4	121.4	120.3			
7/14/2016	56	211.37	212.1	212.46	7.56	7.54	7.54	7.57	98.9	113.1	117.2	5.04	5.04	5.03
7/21/2016	63	211.59	212.01	212.72	7.44	7.42	7.41	7.53	112.1	129.1	127.8			
7/28/2016	70	211.29	212.05	212.67	7.34	7.28	7.27	7.49	95.0	114.2	113.4			
8/4/2016	77	211.31	212.31	212.98	7.45	7.44	7.46	7.46	95.8	110.4	108.7			
8/11/2016	84	211.84	212.19	212.09	7.34	7.19	7.26	7.42	73.9	104.6	104.6			
8/18/2016	91	N/A	N/A	N/A	7.31	7.23	7.1	N/A	68.4	102.2	123.0	5.23	5.14	5.52
CH3COO-														
date	time days	7.5C-1			7.5C-2			7.5C-3			Adjusted Cl-			
		mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L	mM	mg/L
4/26/2016	7	75.75	1.28	108.64	1.84	124.51	2.11		90.27	2.55	87.62	2.47	89.08	2.51
6/2/2016	14	57.28	0.97	83.07	1.41	95.37	1.62		85.46	2.41	85.03	2.40	85.42	2.41
6/9/2016	21	39.00	0.66	60.33	1.02	67.98	1.15		87.43	2.47	87.11	2.46	87.52	2.47
6/16/2016	28	22.78	0.39	32.84	0.56	34.54	0.58		83.66	2.36	81.48	2.30	89.41	2.52
6/23/2016	35	7.00	0.12	12.43	0.21	13.07	0.22		84.99	2.40	84.75	2.39	85.47	2.41
6/30/2016	42	1.29	0.02	1.58	0.03	1.89	0.03		82.61	2.33	82.37	2.32	82.58	2.33
7/7/2016	49	0.37	0.01	0.00	0.00	0.66	0.01		82.85	2.34	81.72	2.31	82.68	2.33
7/14/2016	56	0.00	0.00	0.00	0.00	0.00	0.00		84.12	2.37	84.83	2.39	84.94	2.40
7/21/2016	63	0.00	0.00	0.00	0.00	0.00	0.00		83.89	2.37	84.26	2.38	84.00	2.37
7/28/2016	70	1.32	0.02	1.56	0.03	1.32	0.02		83.70	2.36	84.23	2.38	85.02	2.40
8/4/2016	77	0.00	0.00	0.00	0.00	0.00	0.00		86.39	2.44	86.12	2.43	85.51	2.41
8/11/2016	84	1.68	0.03	2.19	0.04	2.64	0.04		82.55	2.33	83.41	2.35	83.29	2.35
8/18/2016	91	2.26	0.04	2.31	0.04	2.37	0.04		84.02	2.37	83.66	2.36	84.31	2.38

date	time days	Na+			K+								
		7.5C-1 mg/L mM	7.5C-2 mg/L mM	7.5C-3 mg/L mM	7.5C-1 mg/L mM	7.5C-2 mg/L mM	7.5C-3 mg/L mM						
4/26/2016	7	128.57	5.59	132.67	5.77	128.89	5.61	8.68	0.22	9.07	0.23	8.78	0.22
6/2/2016	14												
6/9/2016	21												
6/16/2016	28												
6/23/2016	35												
6/30/2016	42												
7/7/2016	49												
7/14/2016	56	124.73	5.43	123.83	5.39	123.54	5.37	9.64	0.25	9.55	0.24	9.52	0.24
7/21/2016	63												
7/28/2016	70												
8/4/2016	77												
8/11/2016	84												
8/18/2016	91	152.67	6.64	150.91	6.56	152.13	6.62	11.71	0.30	11.38	0.29	11.50	0.29
Mg++													
date	time days	Mg++			Ca++								
		7.5C-1 mg/L mM	7.5C-2 mg/L mM	7.5C-3 mg/L mM	7.5C-1 mg/L mM	7.5C-2 mg/L mM	7.5C-3 mg/L mM						
4/26/2016	7	13.17	0.54	13.97	0.57	13.70	0.56	24.05	0.60	27.72	0.69	27.45	0.68
6/2/2016	14												
6/9/2016	21												
6/16/2016	28												
6/23/2016	35												
6/30/2016	42												
7/7/2016	49												
7/14/2016	56	11.67	0.48	10.34	0.43	11.65	0.48	22.86	0.57	20.59	0.51	23.21	0.58
7/21/2016	63												
7/28/2016	70												
8/4/2016	77												
8/11/2016	84												
8/18/2016	91	13.32	0.55	13.35	0.55	13.41	0.55	23.29	0.58	23.43	0.58	23.75	0.59

Appendix B - Gas Chemistry

Gas Chemistry

6/10/2016	6.0 A-1	6.0 A-2	6.0 A-3	6.0 C-1	6.0 C-2	6.0 C-3	7.5 A-1	7.5 A-2	7.5 A-3	7.5 C-1	7.5 C-2	7.5 C-3
O2 (%)	0.00	0.01	0.01	0.01	0.01	0.05	0.02	0.01	0.01	0.00	0.01	0.01
N2 (%)	85.43	94.18	94.84	84.09	94.06	96.19	128.51	128.45	127.21	134.14	134.19	133.56
CH4 (%)	1.32	1.14	1.01	0.86	1.16	1.20	1.18	1.65	2.86	0.93	1.15	1.65
Guage Pressure (kPa)	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
Total Pressure (kPa)	101.33	101.33	101.33	101.33	101.33	101.33	101.33	101.33	101.33	101.33	101.33	101.33
µmoles CH4	54.1225	47.0433	41.4459	35.5192	47.8665	49.4716	48.525	68.0338	117.547	38.359	47.1668	68.075
partial pressure (kPa)	2.22	1.93	1.70	1.46	1.96	2.03	1.99	2.79	4.82	1.57	1.94	2.79
Partial Pressure (atm)	0.01315	0.01143	0.01007	0.00863	0.01163	0.01202	0.01179	0.01653	0.02856	0.00932	0.01146	0.01654
7/14/2016	6.0 A-1	6.0 A-2	6.0 A-3	6.0 C-1	6.0 C-2	6.0 C-3	7.5 A-1	7.5 A-2	7.5 A-3	7.5 C-1	7.5 C-2	7.5 C-3
O2 (%)	0.009	0.005	0.007	0.008	0	0.02	0.007	0.005	0.005	0.006	0.005	0
N2 (%)	95.259	93.405	93.006	94.067	93.065	93.955	129.615	130.306	127.96	133.704	135.133	121.318
CH4 (%)	3.655	3.856	3.464	1.858	2.281	2.627	4.257	5.088	7.196	2.135	3.486	3.93
Guage Pressure (kPa)	13.75	11.25	7	7	5	7.75	10	10.5	7	25.5	17.5	15
Total Pressure (kPa)	115.075	112.575	108.325	108.325	106.325	109.075	111.325	111.825	108.325	126.825	118.825	116.325
µmoles CH4	102.507	105.795	91.452	49.0525	59.1081	69.8348	115.5	138.667	189.979	65.9917	100.954	111.417
Partial Pressure (kPa)	4.20599	4.34089	3.75238	2.01268	2.42527	2.8654	4.73911	5.68966	7.79507	2.70771	4.14224	4.57157
Partial Pressure (atm)	0.04151	0.04284	0.03703	0.01986	0.02394	0.02828	0.04677	0.05615	0.07693	0.02672	0.04088	0.04512
µmolar of methane	58.1139	59.9778	51.8463	27.809	33.5098	39.591	65.4799	78.6136	107.704	37.4123	57.233	63.1651
8/11/2016	6.0 A-1	6.0 A-2	6.0 A-3	6.0 C-1	6.0 C-2	6.0 C-3	7.5 A-1	7.5 A-2	7.5 A-3	7.5 C-1	7.5 C-2	7.5 C-3
O2 (%)	0.007	0.015	0.023	0.006	0.006	0	0.007	0.005	0.033	0.01	0.004	0.005
N2 (%)	93.522	100.198	93.864	93.788	91.32	92.67	128.004	129.126	128.143	131.384	131.953	130.629
CH4 (%)	3.596	3.408	3.977	1.678	2.088	2.519	4.847	5.207	7.884	2.024	3.358	4.103
Guage Pressure (kPa)	17	19	8	8	8	8.5	9.75	18	9.5	27.5	15.75	15
Total Pressure (kPa)	118.325	120.325	109.325	109.325	109.325	109.825	111.075	119.325	110.825	128.825	117.075	116.325
µMoles CH4	103.701	99.9406	105.965	44.7093	55.6335	67.4242	131.213	151.428	212.947	63.5473	95.8145	116.322
Partial Pressure (kPa)	4.25497	4.10068	4.34786	1.83447	2.28271	2.76649	5.38381	6.21325	8.73744	2.60742	3.93138	4.77281
Partial Pressure (atm)	0.04199	0.04047	0.04291	0.0181	0.02253	0.0273	0.05313	0.06132	0.08623	0.02573	0.0388	0.0471
µmolar	58.7906	56.6587	60.074	25.3468	31.54	38.2244	74.3876	85.8481	120.725	36.0265	54.3196	65.9456

Appendix C - Mass Balance Calculations

6.0A Mass Balance Calculations

date	Time Days	Acetate Consumed Weekly			Rate of Acetate Oxidation			Total Acetate Consumed			Acetate consumption by Fe reducers			
		6A-1 uM	6A-2 uM	6A-3 uM	6A-1 uM/Day	6A-2 uM/Day	6A-3 uM/Day	6A-1 uM	6A-2 uM	6A-3 uM	6.0 A-1 uM/Day	6.0 A-2 uM/Day	6.0 A-3 uM/Day	
4/26/2016	7	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
6/2/2016	14	1747.65	1791.75	1638.57	249.66	255.96	234.08	1747.65	1791.75	1638.57	30.46	26.51	27.86	
6/9/2016	21	1269.70	1342.25	1228.79	181.39	191.75	175.54	3017.35	3134.01	2867.36	18.04	16.12	14.48	
6/16/2016	28	868.83	951.33	895.77	124.12	135.90	127.97	3886.18	4085.34	3763.13	12.16	6.45	14.02	
6/23/2016	35	483.56	545.05	561.73	69.08	77.86	80.25	4369.74	4630.38	4324.86	8.97	17.33	12.67	
6/30/2016	42	179.70	216.86	255.13	25.67	30.98	36.45	4549.44	4847.24	4579.99	7.06	5.00	8.36	
7/7/2016	49	81.75	99.14	125.50	11.68	14.16	17.93	4631.19	4946.39	4705.49	5.22	8.95	5.22	
7/14/2016	56	66.58	67.86	70.96	9.51	9.69	10.14	4697.76	5014.25	4776.45	3.78	0.25	-2.57	
7/21/2016	63	57.64	57.37	57.15	8.23	8.20	8.16	4755.41	5071.63	4833.60	-3.15	-2.89	8.66	
7/28/2016	70	49.98	49.98	49.98	7.14	7.14	7.14	4805.39	5121.61	4883.58	11.64	7.72	1.92	
8/4/2016	77	62.39	64.48	62.03	8.91	9.21	8.86	4867.78	5186.09	4945.62	-1.51	3.88	4.69	
8/11/2016	84	49.98	49.98	54.32	7.14	7.14	7.76	4917.76	5236.07	4999.93	4.15	2.14	5.05	
8/18/2016	91	67.62	64.04	68.56	9.66	9.15	9.79	4985.37	5300.11	5068.49	-18.86	2.73	5.94	

6.0A Mass Balance Calculations													
date	Time Days	Fe Reduced Weekly			Rate of Iron reduction			Total Iron reduced			Total Methane Produced		
		6.0 A-1 uM	6.0 A-2 uM	6.0 A-3 uM	6.0 A-1 uM/Day	6.0 A-2 uM/Day	6.0 A-3 uM/Day	6.0 A-1 uM	6.0 A-2 uM	6.0 A-3 uM	6.0 A-1 μmoles	6.0 A-2 μmoles	6.0 A-3 μmoles
4/26/2016	7	972.218	1921.42	1476.87	138.888	274.489	210.981	972.218	1921.42	1476.87			
6/2/2016	14	1705.96	1484.45	1560.19	243.708	212.065	222.885	2678.18	3405.88	3037.06			
6/9/2016	21	1010.45	902.601	810.945	144.35	128.943	115.849	3688.62	4308.48	3848	53.6920585	46.6692189	41.116276
6/16/2016	28	680.891	361.194	785.17	97.2702	51.5992	112.167	4369.51	4669.67	4633.17			
6/23/2016	35	502.519	970.402	709.705	71.7885	138.629	101.386	4872.03	5640.07	5342.88			
6/30/2016	42	395.496	280.241	468.217	56.4995	40.0344	66.8882	5267.53	5920.31	5811.1			
7/7/2016	49	292.59	501.147	292.59	41.7985	71.5925	41.7985	5560.12	6421.46	6103.69			
7/14/2016	56	211.636	14.0555	-143.73	30.2338	2.00793	-20.534	5771.76	6435.52	5959.95	101.692121	104.95374	90.7246958
7/21/2016	63	-176.66	-161.57	484.682	-25.238	-23.082	69.2403	5595.09	6273.94	6444.63			
7/28/2016	70	652.077	432.543	107.358	93.1539	61.7918	15.3368	6247.17	6706.49	6551.99			
8/4/2016	77	-84.735	217.125	262.404	-12.105	31.0178	37.4862	6162.43	6923.61	6814.39			
8/11/2016	84	232.218	119.706	282.985	33.174	17.1009	40.4264	6394.65	7043.32	7097.38			
8/18/2016	91	-1056.2	152.637	332.38	-150.88	21.8052	47.4829	5338.48	7195.96	7429.76	102.876253	99.145817	105.122097
		Acetate needed			Acetate needed			Acetate Needed			Acetate Needed		
		83.1841			83.1841			102.381389			102.381389		

6.0C Mass Balance Calculations														
date	Time Days	Acetate Consumed Weekly			Rate of Acetate Oxidation			Total Acetate Consumed			Acetate consumption by Fe reducers			
		6C-1 uM	6C-2 uM	6C-3 uM	6C-1 uM/Day	6C-2 uM/Day	6C-3 uM/Day	6C-1 uM	6C-2 uM	6C-3 uM	6.0 C-1 uM/Day	6.0 C-2 uM/Day	6.0 C-3 uM/Day	
4/26/2016	7	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
6/2/2016	14	202.95	172.21	157.05	28.99	24.60	22.44	202.95	172.21	157.05	27.58	30.25	23.63	
6/9/2016	21	183.92	183.25	196.85	26.27	26.18	28.12	386.87	355.46	353.90	14.66	9.78	14.37	
6/16/2016	28	207.74	256.35	278.33	29.68	36.62	39.76	594.61	611.81	632.23	10.96	15.32	11.45	
6/23/2016	35	139.85	151.20	271.50	19.98	21.60	38.79	734.45	763.01	903.73	8.36	10.47	11.13	
6/30/2016	42	13.70	21.45	-6.36	1.96	3.06	-0.91	748.16	784.47	897.36	8.07	4.20	7.41	
7/7/2016	49	-10.87	-5.40	13.87	-1.55	-0.77	1.98	737.29	779.06	911.23	0.84	3.44	-3.69	
7/14/2016	56	20.07	21.37	14.36	2.87	3.05	2.05	757.35	800.44	925.59	2.43	2.43	6.28	
7/21/2016	63	0.00	0.00	0.00	0.00	0.00	0.00	757.35	800.44	925.59	-0.56	0.62	-2.27	
7/28/2016	70	-10.98	-15.54	-16.28	-1.57	-2.22	-2.33	746.37	784.90	909.31	3.93	3.34	5.76	
8/4/2016	77	8.79	12.43	13.02	1.26	1.78	1.86	755.16	797.33	922.34	2.51	1.60	0.47	
8/11/2016	84	-13.93	-14.30	-17.67	-1.99	-2.04	-2.52	741.23	783.02	904.67	1.97	2.43	2.31	
8/18/2016	91	-5.91	-5.22	-1.97	-0.84	-0.75	-0.28	735.32	777.81	902.70	3.02	6.77	3.85	

6.0C Mass Balance Calculations

date	Time Days	Fe Reduced Weekly			Rate of Iron reduction			Total Iron reduced			Total Methane Produced		
		6.0 C-1 uM	6.0 C-2 uM	6.0 C-3 uM	6.0 C-1 uM/Day	6.0 C-2 uM/Day	6.0 C-3 uM/Day	6.0 C-1 uM/Day	6.0 C-2 uM/Day	6.0 C-3 uM/Day	6.0 C-1 μmoles	6.0 C-2 μmoles	6.0 C-3 μmoles
4/26/2016	7	1534.49	1378.08	1622.31	219.213	196.868	231.758	1534.49	1378.08	1622.31			
6/2/2016	14	1544.28	1694.11	1323.1	220.611	242.016	189.014	3078.77	3072.18	2945.4			
6/9/2016	21	821.098	547.504	804.633	117.3	78.2148	114.948	3899.87	3619.69	3750.04	35.2366893	47.4858281	49.0782162
6/16/2016	28	613.659	857.891	641.1	87.6655	122.556	91.5858	4513.53	4477.58	4391.14			
6/23/2016	35	468.217	586.217	623.263	66.8882	83.7453	89.0376	4981.74	5063.8	5014.4			
6/30/2016	42	451.752	234.962	414.706	64.536	33.566	59.2437	5433.5	5298.76	5429.11			
7/7/2016	49	46.9856	192.427	-206.85	6.71223	27.4896	-29.55	5480.48	5491.18	5222.25			
7/14/2016	56	136.171	136.171	351.59	19.4531	19.4531	50.2271	5616.65	5627.36	5573.84	48.6623801	58.6380631	69.2794185
7/21/2016	63	-31.223	34.6368	-127.27	-4.4605	4.94812	-18.181	5585.43	5661.99	5446.57			
7/28/2016	70	219.869	186.939	322.776	31.4098	26.7055	46.1108	5805.3	5848.93	5769.35			
8/4/2016	77	140.288	89.5204	26.4043	20.0411	12.7886	3.77204	5945.59	5938.45	5795.75			
8/11/2016	84	110.102	136.171	129.311	15.7288	19.4531	18.473	6055.69	6074.62	5925.07			
8/18/2016	91	169.102	379.031	215.753	24.1574	54.1473	30.8218	6224.79	6453.65	6140.82	44.3537539	55.1910835	66.8880167
		Acetate needed			78.4136			Acetate Needed			55.477618		

7.5A Mass Balance Calculations																
date	Time Days	Acetate Consumed			Rate of Acetate Oxidation			Total Acetate Consumed			Acetate consumption by Fe reducers					
		7.5-A-1 uM	7.5-A-2 uM	7.5-A-3 uM	7.5-A-1 uM/Day	7.5-A-2 uM/Day	7.5-A-3 uM/Day	7.5-A-1 uM	7.5-A-2 uM	7.5-A-3 uM	7.5-A-1 uM/Day	7.5-A-2 uM/Day	7.5-A-3 uM/Day			
4/26/2016	7	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
6/2/2016	14	1399.69	1541.94	2293.91	199.96	220.28	327.70	1399.69	1541.94	2293.91	0.55	0.82	0.19			
6/9/2016	21	1115.67	1231.86	1809.66	159.38	175.98	258.52	2515.36	2773.80	4103.57	0.46	0.36	0.96			
6/16/2016	28	864.97	925.73	1320.85	123.57	132.25	188.69	3380.33	3699.53	5424.42	1.03	0.16	0.16			
6/23/2016	35	499.45	577.87	862.92	71.35	82.55	123.27	3879.77	4277.40	6287.33	-0.27	0.38	0.34			
6/30/2016	42	216.53	297.96	460.58	30.93	42.57	65.80	4096.31	4575.35	6747.92	0.40	0.39	0.39			
7/7/2016	49	87.03	118.00	155.14	12.43	16.86	22.16	4183.34	4693.36	6903.05	0.33	0.22	0.09			
7/14/2016	56	58.84	56.49	61.25	8.41	8.07	8.75	4242.19	4749.84	6964.30	0.29	0.38	0.59			
7/21/2016	63	50.00	55.63	55.96	7.14	7.95	7.99	4292.19	4805.48	7020.26	0.49	0.49	0.51			
7/28/2016	70	49.98	49.99	49.99	7.14	7.14	7.14	4342.17	4855.46	7070.25	0.36	0.29	0.27			
8/4/2016	77	63.76	61.88	60.39	9.11	8.84	8.63	4405.93	4917.34	7130.64	0.46	0.34	0.34			
8/11/2016	84	49.99	49.99	49.99	7.14	7.14	7.14	4455.91	4967.32	7180.63	0.53	0.41	0.43			
8/18/2016	91	61.76	61.28	59.44	8.82	8.75	8.49	4517.67	5028.61	7240.07	0.40	0.56	0.63			

7.5A Mass Balance Calculations													
date	Time Days	Fe Reduced Weekly			Rate of Iron reduction			Total Iron reduced			Total Methane Produced		
		7.5 A-1 uM	7.5 A-2 uM	7.5 A-3 uM	7.5 A-1 uM/Day	7.5 A-2 uM/Day	7.5 A-3 uM/Day	7.5 A-1 uM/Day	7.5 A-2 uM/Day	7.5 A-3 uM/Day	7.5 A-1 μmoles	7.5 A-2 μmoles	7.5 A-3 μmoles
4/26/2016	7	16.9355	28.8727	107.219	2.41936	4.12467	15.317	16.9355	28.8727	107.219			
6/2/2016	14	30.6089	45.7567	10.5215	4.37269	6.53667	1.50307	47.5444	74.6294	117.74			
6/9/2016	21	25.8757	20.1129	53.5919	3.69653	2.87327	7.65599	73.42	94.7423	171.332	48.1391155	67.4927549	116.611802
6/16/2016	28	57.5196	8.90452	8.73987	8.21709	1.27207	1.24855	130.94	103.647	180.072			
6/23/2016	35	-15.213	21.0338	18.8933	-2.1733	3.00483	2.69905	115.727	124.681	198.966			
6/30/2016	42	22.1864	21.8022	21.8571	3.16948	3.1146	3.12244	137.913	146.483	220.823			
7/7/2016	49	18.6189	12.1427	5.11756	2.65985	1.73467	0.73108	156.532	158.625	225.94			
7/14/2016	56	15.9845	21.1984	33.1082	2.2835	3.02835	4.72974	172.517	179.824	259.048	114.581708	137.564049	188.468508
7/21/2016	63	27.6138	27.2571	28.3547	3.94483	3.89387	4.05068	200.13	207.081	287.403			
7/28/2016	70	20.0727	16.5052	14.9685	2.86752	2.35789	2.13836	220.203	223.586	302.372			
8/4/2016	77	25.8636	18.8933	19.2775	3.69479	2.69905	2.75393	246.067	242.48	321.649			
8/11/2016	84	29.7603	22.845	23.8329	4.25147	3.26357	3.40469	275.827	265.325	345.482			
8/18/2016	91	22.1864	31.5166	35.2486	3.16948	4.50237	5.03552	298.013	296.841	380.731	130.169214	150.223529	211.253199
								Acetate needed	4.06494	Acetate Needed	163.881981		

7.5C Mass Balance Calculations																
date	Time Days	Acetate Consumed Weekly			Rate of Acetate Oxidation			Total Acetate Consumed			Acetate consumption by Fe reducers					
		7.5C-1 uM	7.5C-2 uM	7.5C-3 uM	7.5C-1 uM/Day	7.5C-2 uM/Day	7.5C-3 uM/Day	7.5C-1 uM	7.5C-2 uM	7.5C-3 uM	7.5C-1 uM/Day	7.5C-2 uM/Day	7.5C-3 uM/Day			
4/26/2016	7	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A	N/A
6/2/2016	14	56.34	64.99	71.82	8.05	9.28	10.26	56.34	64.99	71.82	0.54	0.52	0.99			
6/9/2016	21	115.58	103.71	140.84	16.51	14.82	20.12	171.92	168.70	212.66	0.54	0.69	0.32			
6/16/2016	28	142.61	261.29	336.14	20.37	37.33	48.02	314.53	430.00	548.79	0.33	0.30	0.30			
6/23/2016	35	190.03	234.39	246.60	27.15	33.48	35.23	504.56	664.38	795.39	0.35	0.38	0.50			
6/30/2016	42	73.03	141.67	145.04	10.43	20.24	20.72	577.59	806.05	940.44	0.37	0.84	0.37			
7/7/2016	49	11.20	21.43	14.48	1.60	3.06	2.07	588.79	827.48	954.92	0.18	-0.08	0.17			
7/14/2016	56	5.01	0.00	8.91	0.72	0.00	1.27	593.80	827.48	963.83	0.30	0.29	0.38			
7/21/2016	63	0.00	0.00	0.00	0.00	0.00	0.00	593.80	827.48	963.83	0.59	0.69	0.61			
7/28/2016	70	-22.36	-26.37	-22.40	-3.19	-3.77	-3.20	571.44	801.11	941.43	0.09	0.19	0.20			
8/4/2016	77	17.89	21.09	17.92	2.56	3.01	2.56	589.33	822.21	959.35	0.35	0.34	0.32			
8/11/2016	84	-28.53	-37.06	-44.73	-4.08	-5.29	-6.39	560.80	785.15	914.62	-0.05	0.29	0.31			
8/18/2016	91	-15.38	-9.40	-4.28	-2.20	-1.34	-0.61	545.41	775.75	910.34	0.17	0.33	0.70			

7.5C Mass Balance Calculations													
date	Time Days	Fe Reduced Weekly			Rate of Iron reduction			Total Iron reduced			Total Methane Produced		
		7.5C-1 μM	7.5C-2 μM	7.5C-3 μM	7.5C-1 $\mu\text{M}/\text{Day}$	7.5C-2 $\mu\text{M}/\text{Day}$	7.5C-3 $\mu\text{M}/\text{Day}$	7.5C-1 $\mu\text{M}/\text{Day}$	7.5C-2 $\mu\text{M}/\text{Day}$	7.5C-3 $\mu\text{M}/\text{Day}$	7.5C-1 μmoles	7.5C-2 μmoles	7.5C-3 μmoles
4/26/2016	7	20.8108	52.9177	61.4247	2.97297	7.55967	8.77495	20.8108	52.9177	61.4247			
6/2/2016	14	30.2528	29.2649	55.3895	4.32183	4.1807	7.91278	51.0636	82.1826	116.814			
6/9/2016	21	30.2664	38.4989	18.192	4.32377	5.49985	2.59886	81.33	120.682	135.006	38.0539912	46.7917103	67.5335853
6/16/2016	28	18.6738	16.8626	16.9724	2.66769	2.40895	2.42463	100.004	137.544	151.979			
6/23/2016	35	19.3324	21.5277	27.7296	2.76177	3.07539	3.96137	119.336	159.072	179.708			
6/30/2016	42	20.7045	47.2133	20.7045	2.95779	6.74475	2.95779	140.041	206.285	200.413			
7/7/2016	49	9.94731	-4.2675	9.34359	1.42104	-0.6096	1.3348	149.988	202.018	209.756			
7/14/2016	56	16.9175	16.0394	21.0338	2.41679	2.29134	3.00483	166.906	218.057	230.79	65.4668869	100.150736	110.531115
7/21/2016	63	33.0473	38.6454	33.9803	4.72104	5.52077	4.85433	199.953	256.702	264.77			
7/28/2016	70	5.30898	10.9071	11.1815	0.75843	1.55816	1.59736	205.262	267.61	275.952			
8/4/2016	77	19.8264	19.0031	18.0152	2.83234	2.71473	2.5736	225.088	286.613	293.967			
8/11/2016	84	-2.7857	16.3138	17.631	-0.398	2.33054	2.51872	222.302	302.926	311.598			
8/18/2016	91	9.28871	18.4543	39.31	1.32696	2.63632	5.61572	231.591	321.381	350.908	63.0419443	95.0525556	115.396734
								Acetate needed	321.381	350.908	3.76617	Acetate Needed	91.1637447

Appendix D - Thermodynamic Calculations

				Activities of chemical species for thermo calculations										Reaction Quotient		Reaction Free Energy	
Date	ID	Temp °C	Temp K	pH	CH3COO- activity	HCO3- activity	H+ activity	Fe++ activity	CH4(aq) activity	Fe++ Q	methanog. Q	Fe++ ΔGr	methanog. ΔGr				
7/14/2016	6.0 A-1	20	293.15	6.32	6.66E-06	8.26E-03	4.79E-07	1.23E-03	5.74E-05	3.301E+72	0.0712456	-68.03199	-21.40026				
7/14/2016	6.0 A-2	20	293.15	6.31	6.59E-06	8.36E-03	4.90E-07	1.28E-03	5.93E-05	3.458E+72	0.0752772	-67.91922	-21.26611				
7/14/2016	6.0 A-3	20	293.15	6.33	6.77E-06	8.39E-03	4.68E-07	1.14E-03	5.13E-05	2.645E+72	0.0635209	-68.57182	-21.67997				
7/14/2016	6.0 C-1	20	293.15	6.16	1.22E-05	4.63E-03	6.92E-07	4.43E-05	5.80E-05	6.54E+57	0.0221011	-150.5456	-24.25308				
7/14/2016	6.0 C-2	20	293.15	6.28	1.29E-05	6.56E-03	5.25E-07	7.54E-04	5.60E-05	5.513E+69	0.0284938	-83.6183	-23.63387				
7/14/2016	6.0 C-3	20	293.15	6.24	1.20E-05	6.59E-03	5.75E-07	9.51E-04	5.94E-05	9.647E+69	0.0324798	-82.25458	-23.31476				
7/14/2016	7.5 A-1	20	293.15	6.34	6.84E-06	7.73E-03	4.57E-07	1.10E-03	2.75E-05	2.344E+72	0.0310944	-68.86714	-23.421				
7/14/2016	7.5 A-2	20	293.15	6.28	6.78E-06	7.79E-03	5.25E-07	1.13E-03	3.31E-05	3.722E+71	0.0380677	-73.35162	-22.92785				
7/14/2016	7.5 A-3	20	293.15	6.45	6.84E-06	7.63E-03	3.55E-07	1.12E-03	3.91E-05	1.194E+74	0.0436319	-59.28673	-22.59536				
7/14/2016	7.5 C-1	20	293.15	6.11	1.26E-05	5.80E-03	7.76E-07	6.47E-04	2.50E-05	3.645E+66	0.0114899	-101.4627	-25.84743				
7/14/2016	7.5 C-2	20	293.15	6.10	1.20E-05	6.05E-03	7.94E-07	7.72E-04	3.12E-05	1.218E+67	0.0156787	-98.52197	-25.08985				
7/14/2016	7.5 C-3	20	293.15	6.05	1.18E-05	0.005853	8.91E-07	6.53E-04	3.78E-05	5.397E+65	0.018693	-106.1179	-24.66127				

					Activities of chemical species for thermo calculations													
Date	ID	Temp °C	Temp K	pH	CH3COO-		HCO3-		H+		Fe++		CH4(aq)		Reaction Quotient		Reaction Free Energy	
					activity	activity	activity	activity	activity	activity	Q	Q	Fe++	methanog.	ΔGr	methanog.	ΔGr	
8/18/2016	6.0 A-1	20	293.15	7.53	8.74E-06	0.004703	2.95E-08	6.22E-05	6.46E-05	5.086E+79	0.0347693	-27.69505	-23.14874					
8/18/2016	6.0 A-2	20	293.15	7.55	8.75E-06	0.004535	2.82E-08	6.44E-05	7.76E-05	1.244E+80	0.040216	-25.51407	-22.79405					
8/18/2016	6.0 A-3	20	293.15	7.53	8.70E-06	0.004648	2.95E-08	7.82E-05	1.06E-04	3.104E+80	0.056778	-23.2865	-21.95348					
8/18/2016	6.0 C-1	20	293.15	6.85	1.03E-05	0.004466	1.41E-07	7.32E-05	7.34E-05	8.91E+69	0.0318876	-82.44801	-23.35961					
8/18/2016	6.0 C-2	20	293.15	7.12	8.68E-06	0.004443	7.59E-08	7.21E-05	8.47E-05	1.044E+74	0.0433501	-59.61444	-22.61115					
8/18/2016	6.0 C-3	20	293.15	7.11	9.33E-06	0.004666	7.76E-08	7.86E-05	1.19E-04	1.512E+74	0.0595436	-58.71199	-21.83756					
8/18/2016	7.5 A-1	20	293.15	7.56	8.75E-06	0.004499	2.75E-08	6.07E-05	3.69E-05	1.078E+80	0.0189773	-25.86383	-24.62448					
8/18/2016	7.5 A-2	20	293.15	7.54	8.74E-06	0.004508	2.88E-08	6.98E-05	5.65E-05	1.641E+80	0.029103	-24.83954	-23.58231					
8/18/2016	7.5 A-3	20	293.15	7.54	8.73E-06	0.004488	2.88E-08	7.21E-05	6.23E-05	2.111E+80	0.0320564	-24.22614	-23.34674					
8/18/2016	7.5 C-1	20	293.15	7.31	3.34E-05	0.004654	4.90E-08	4.17E-05	3.56E-05	2.651E+74	0.004961	-57.34306	-27.89437					
8/18/2016	7.5 C-2	20	293.15	7.23	3.39E-05	0.004578	5.89E-08	6.26E-05	5.36E-05	4.106E+74	0.0072426	-56.27672	-26.97217					
8/18/2016	7.5 C-3	20	293.15	7.10	3.47E-05	4.92E-03	7.94E-08	7.48E-05	6.51E-05	2.165E+73	0.0092363	-63.44886	-26.37956					

Equations		Log K (20°C)	ΔG° _r	ΔG° = -2.303RTlog(K)	ΔGr = ΔG° + RTln(Q)
CH3COO- + 15 H+ + 8 Goethite = 2 HCO3- + 12 H2O + 8 Fe++		84.6261	-475.005		
CH3COO- + H2O = HCO3- + CH4(aq)		2.6656	-14.96197		

Usable Energy					
Date	ID	Fe ⁺⁺	methanog.	Fe ⁺⁺	methanog.
		Δ GA	Δ GA	Δ GU	Δ GU
7/14/2016	6.0A-1 56	68.03	21.40	11.78	12.40
7/14/2016	6.0A-2 56	67.92	21.27	11.67	12.27
7/14/2016	6.0A-3 56	68.57	21.68	12.32	12.68
8/18/2016	6.0A-1 91	150.55	24.25	94.30	15.25
8/18/2016	6.0A-2 91	83.62	23.63	27.37	14.63
8/18/2016	6.0A-3 91	82.25	23.31	26.00	14.31
7/14/2016	6.0C-1 56	68.87	23.42	12.62	14.42
7/14/2016	6.0C-2 56	73.35	22.93	17.10	13.93
7/14/2016	6.0C-3 56	59.29	22.60	3.04	13.60
8/18/2016	6.0C-1 91	101.46	25.85	45.21	16.85
8/18/2016	6.0C-2 91	98.52	25.09	42.27	16.09
8/18/2016	6.0C-3 91	106.12	24.66	49.87	15.66
7/14/2016	7.5A-1 56	27.70	23.15	-28.55	14.15
7/14/2016	7.5A-2 56	25.51	22.79	-30.74	13.79
7/14/2016	7.5A-3 56	23.29	21.95	-32.96	12.95
8/18/2016	7.5A-1 91	82.45	23.36	26.20	14.36
8/18/2016	7.5A-2 91	59.61	22.61	3.36	13.61
8/18/2016	7.5A-3 91	58.71	21.84	2.46	12.84
7/14/2016	7.5C-1 56	25.86	24.62	-30.39	15.62
7/14/2016	7.5C-2 56	24.84	23.58	-31.41	14.58
7/14/2016	7.5C-3 56	24.23	23.35	-32.02	14.35
8/18/2016	7.5C-1 91	57.34	27.89	1.09	18.89
8/18/2016	7.5C-2 91	56.28	26.97	0.03	17.97
8/18/2016	7.5C-3 91	63.45	26.38	7.20	17.38

Appendix E - Microbial Data

Phylum Relative Abundance	
#OTU ID	Relative Abundance
k_Bacteria;p__Proteobacteria	55.3456%
k_Bacteria;p__Chloroflexi	10.1790%
k_Bacteria;p__Bacteroidetes	8.1015%
k_Bacteria;p__Acidobacteria	5.0703%
k_Bacteria;p__Verrucomicrobia	2.6123%
k_Bacteria;p__Firmicutes	2.3124%
Unassigned;Other	2.1901%
k_Bacteria;p__Nitrospirae	1.9410%
k_Bacteria;p__Chlorobi	1.9013%
k_Bacteria;p__Planctomycetes	1.6660%
k_Bacteria;p__Actinobacteria	0.8945%
k_Bacteria;p__Spirochaetes	0.8638%
k_Bacteria;p__WS3	0.7264%
k_Archaea;p__Euryarchaeota	0.6909%
k_Bacteria;p__Gemmatimonadetes	0.6678%
k_Archaea;p__Crenarchaeota	0.5884%
k_Bacteria;p__Elusimicrobia	0.5763%
k_Bacteria;p__GNO4	0.5164%
k_Bacteria;p__Armatimonadetes	0.2718%
k_Bacteria;p__OD1	0.2558%
k_Bacteria;p__TM6	0.2153%
k_Bacteria;p__Cyanobacteria	0.2053%
k_Bacteria;p__Lentisphaerae	0.2022%
k_Bacteria;p__Fibrobacteres	0.1784%
k_Bacteria;p__NC10	0.1652%
k_Bacteria;p__OP8	0.1576%
k_Bacteria;p__	0.1282%
k_Bacteria;p__Tenericutes	0.1279%
k_Bacteria;p__BRC1	0.1069%
k_Bacteria;p__GOUTA4	0.1001%
k_Bacteria;p__SR1	0.0943%
k_Bacteria;p__Caldithrix	0.0897%
k_Bacteria;p__OP11	0.0873%
k_Bacteria;p__OP3	0.0855%
k_Bacteria;p__Chlamydiae	0.0801%
k_Bacteria;p__MVS-104	0.0571%
k_Archaea;p__[Parvarchaeota]	0.0570%
k_Bacteria;p__SC4	0.0495%
k_Bacteria;p__WPS-2	0.0487%
k_Bacteria;p__WS5	0.0437%
k_Bacteria;p__NKB19	0.0427%
k_Bacteria;p__TPD-58	0.0424%
k_Bacteria;p__H-178	0.0387%
k_Bacteria;p__Caldiserica	0.0247%
k_Bacteria;p__WWE1	0.0229%

Genus Relative Abundance									
#OTU ID	pH.6.C.1	pH.6.C.2	pH.6.C.3	pH.7.5.A.1	pH.7.5.A.2	pH.7.5.A.3	pH.7.5.C.1	pH.7.5.C.2	pH.7.5.C.3
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfuromonadales;f__Geobacteraceae;g__Geobacter	0.2248	0.2571	0.2165	0.1161	0.1283	0.1107	0.1345	0.1719	0.1457
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Rhodocyclales;f__Rhodocyclaceae;g__Dechloromonas	0.0349	0.0341	0.0266	0.1255	0.0665	0.0539	0.0743	0.0629	0.0669
k__Bacteria;p__Bacteroidetes;c__Bacteroidia;o__Bacteroidales;f__g__	0.0671	0.0735	0.0781	0.0411	0.0394	0.0388	0.0431	0.0616	0.0620
k__Bacteria;p__Chloroflexi;c__Anaerolineae;o__GCA004.f__g__	0.0331	0.0379	0.0268	0.0338	0.0336	0.0353	0.0377	0.0465	0.0400
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Burkholderiales;f__Comamonadaceae;g__	0.0199	0.0201	0.0343	0.0724	0.1182	0.1156	0.0077	0.0049	0.0042
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Gallionellales;f__Gallionellaceae;g__Gallionella	0.0395	0.0024	0.0783	0.0010	0.0014	0.0017	0.0473	0.0035	0.0021
Unassigned\Other\Other\Other	0.0218	0.0224	0.0234	0.0343	0.0254	0.0237	0.0258	0.0234	0.0230
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__Pseudomonadales;f__Moraxellaceae;g__Acinetobacter	0.0000	0.0001	0.0000	0.0883	0.0985	0.1128	0.0002	0.0000	0.0000
k__Bacteria;p__Chloroflexi;c__Anaerolineae;o__Anaerolineales;f__Anaerolinaceae;g__Anaerolinea	0.0160	0.0203	0.0090	0.0102	0.0125	0.0129	0.0126	0.0164	0.0165
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfuromonadales;f__Pelobacteraceae;g__	0.0308	0.0288	0.0211	0.0193	0.0169	0.0157	0.0130	0.0194	0.0170
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacteriales;f__Syntrophobacteraceae;g__	0.0117	0.0111	0.0107	0.0111	0.0112	0.0109	0.0134	0.0118	0.0148
k__Bacteria;p__Chloroflexi;c__Anaerolineae;o__envOP512.f__g__	0.0096	0.0106	0.0083	0.0093	0.0112	0.0101	0.0103	0.0134	0.0108
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfuromonadales;f__Geobacteraceae;g__	0.0091	0.0088	0.0110	0.0188	0.0222	0.0204	0.0256	0.0313	0.0289
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__SBla14.f__g__	0.0028	0.0005	0.0016	0.0010	0.0008	0.0010	0.0334	0.0016	0.0026
k__Bacteria;p__Verrucomicrobia;c__[Pedosphaerae]o__[Pedosphaerales]f__Ellin515.g__	0.0137	0.0103	0.0093	0.0080	0.0080	0.0089	0.0104	0.0116	0.0131
k__Bacteria;p__Chloroflexi;c__Anaerolineae;o__Anaerolineales;f__Anaerolinaceae;g__	0.0075	0.0116	0.0082	0.0098	0.0087	0.0087	0.0073	0.0091	0.0076
k__Bacteria;p__Acidobacteria;c__Acidobacteria-6;o__Jii1-15.f__g__	0.0102	0.0121	0.0079	0.0075	0.0064	0.0060	0.0087	0.0084	0.0101
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacteriales;f__Syntrophaceae;g__	0.0079	0.0097	0.0080	0.0171	0.0104	0.0097	0.0104	0.0119	0.0112
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Myxococcales;f__g__	0.0067	0.0091	0.0057	0.0077	0.0068	0.0089	0.0079	0.0088	0.0087
k__Bacteria;p__Nitrospirae;c__Nitrospirae;o__Nitrospirales;f__Nitrospiraceae;g__Nitrospira	0.0058	0.0055	0.0059	0.0047	0.0060	0.0057	0.0063	0.0071	0.0071
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__f__g__	0.0055	0.0074	0.0069	0.0063	0.0080	0.0072	0.0102	0.0083	0.0102
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Myxococcales;f__Myxococaceae;g__Anaeromyxobacter	0.0106	0.0177	0.0048	0.0053	0.0060	0.0064	0.0046	0.0073	0.0078
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfobacteriales;f__Desulfobulbaceae;g__	0.0086	0.0023	0.0180	0.0035	0.0054	0.0058	0.0277	0.0074	0.0054
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfarculales;f__Desulfarculaceae;g__	0.0044	0.0048	0.0061	0.0070	0.0085	0.0062	0.0098	0.0104	0.0087
k__Bacteria;p__Acidobacteria;c__BPC102;o__B110.f__g__	0.0071	0.0049	0.0041	0.0060	0.0053	0.0048	0.0058	0.0069	0.0074
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__f__g__	0.0066	0.0068	0.0039	0.0047	0.0058	0.0042	0.0056	0.0067	0.0073
k__Bacteria;p__Verrucomicrobia;c__[Pedosphaerae]o__[Pedosphaerales]f__g__	0.0063	0.0066	0.0043	0.0041	0.0060	0.0044	0.0051	0.0073	0.0086
k__Bacteria;p__Chlorobi;c__Ignavibacteria;o__Ignavibacteriales;f__Ignavibacteriaceae;g__	0.0077	0.0064	0.0067	0.0040	0.0046	0.0043	0.0064	0.0058	0.0059
k__Bacteria;p__Chloroflexi;c__Anaerolineae;o__SIA-15.f__g__	0.0066	0.0055	0.0043	0.0068	0.0061	0.0057	0.0070	0.0066	0.0063
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacteriales;f__Syntrophobacteraceae;g__Syntrophobacter	0.0055	0.0043	0.0052	0.0064	0.0086	0.0082	0.0072	0.0094	0.0081
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Burkholderiales;f__Comamonadaceae;g__Hydrogenophaga	0.0048	0.0047	0.0097	0.0006	0.0003	0.0007	0.0202	0.0202	0.0178
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__BPC076.f__g__	0.0034	0.0028	0.0032	0.0034	0.0035	0.0039	0.0046	0.0056	0.0057
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfuromonadales;f__Desulfuromonadaceae;g__	0.0042	0.0040	0.0047	0.0061	0.0093	0.0064	0.0093	0.0109	0.0100

#OTU ID	Genus Relative Abundance	Relative Abundance	Marsh_sed B	Mud_2	pH_6.A.1	pH_6.A.2	pH_6.A.3
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__MBNT15;f__g__		0.52%	0.0106	0.0109	0.0016	0.0041	0.0070
k__Bacteria;p__Acidobacteria;c__BPC102;o__MVS-40;f__g__		0.50%	0.0104	0.0120	0.0023	0.0029	0.0031
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Hydrogenophilales;f__Hydrogenophilaceae;g__Thiobacillus		0.48%	0.0128	0.0140	0.0015	0.0019	0.0030
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__f__g__		0.44%	0.0105	0.0099	0.0015	0.0038	0.0031
k__Bacteria;p__Nitrospirae;c__Nitrospira;o__Nitrospirales;f__Thermodesulfovibrionaceae;g__GOUTA19		0.43%	0.0055	0.0060	0.0036	0.0020	0.0020
k__Bacteria;p__Acidobacteria;c__Solibacteres;o__Solibacterales;f__g__		0.43%	0.0081	0.0089	0.0015	0.0030	0.0031
k__Bacteria;p__Bacteroidetes;c__[Saprospirae];o__[Saprospirales];f__Chitinophagaceae;g__		0.43%	0.0137	0.0100	0.0017	0.0015	0.0023
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__MND1;f__g__		0.39%	0.0156	0.0055	0.0009	0.0020	0.0025
k__Bacteria;p__Planctomycetes;c__Planctomycetia;o__Pirellulales;f__Pirellulaceae;g__		0.38%	0.0040	0.0066	0.0017	0.0019	0.0031
k__Bacteria;p__Chlorobi;c__BSV26;o__PK329;f__g__		0.38%	0.0054	0.0051	0.0019	0.0027	0.0034
k__Bacteria;p__Actinobacteria;c__OPB41;o__f__g__		0.37%	0.0008	0.0012	0.0029	0.0042	0.0064
k__Bacteria;p__Firmicutes;c__Bacilli;o__Bacillales;f__Bacillaceae;g__Bacillus		0.37%	0.0042	0.0110	0.0023	0.0033	0.0024
k__Archaea;p__Crenarchaeota;c__Thaumarchaeota;o__Nitrososphaerales;f__Nitrososphaeraceae;g__Candidatus Nitrososphaera		0.37%	0.0054	0.0050	0.0015	0.0035	0.0041
k__Bacteria;p__Bacteroidetes;c__Bacteroidia;o__Bacteroidales;f__Rikenellaceae;g__Bvii28		0.36%	0.0000	0.0000	0.0026	0.0084	0.0093
k__Bacteria;p__Chlorobi;c__SJA-28;o__f__g__		0.36%	0.0120	0.0077	0.0030	0.0020	0.0020
k__Bacteria;p__Proteobacteria;c__Alphaproteobacteria;o__Rhizobiales;f__Hyphomicrobiaceae;g__Rhodoplanes		0.35%	0.0067	0.0075	0.0019	0.0020	0.0031
k__Bacteria;p__Gemmatimonadetes;c__Gemm-1;o__f__g__		0.34%	0.0107	0.0080	0.0007	0.0022	0.0024
k__Bacteria;p__Proteobacteria;c__Alphaproteobacteria;o__Sphingomonadales;f__Sphingomonadaceae;g__Kaistobacter		0.33%	0.0078	0.0070	0.0007	0.0021	0.0020
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacteriales;f__Syntrophorhabdaceae;g__		0.32%	0.0035	0.0043	0.0024	0.0027	0.0023
k__Bacteria;p__Eusimicrobia;c__Eusimicrobia;o__Eusimicrobiales;f__g__		0.31%	0.0005	0.0000	0.0017	0.0034	0.0074
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;Other;Other;Other		0.31%	0.0018	0.0013	0.0031	0.0015	0.0016
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Myxococcales;f__Haliangiaceae;g__		0.31%	0.0093	0.0063	0.0006	0.0027	0.0022
k__Bacteria;p__GND4;c__GN15;o__f__g__		0.31%	0.0045	0.0062	0.0018	0.0029	0.0019
k__Bacteria;p__WS3;c__PRR-12;o__Sediment-1;f__g__		0.30%	0.0091	0.0099	0.0015	0.0013	0.0011
k__Bacteria;p__Nitrospirae;c__Nitrospira;o__Nitrospirales;f__FW;g__4-29		0.30%	0.0070	0.0082	0.0020	0.0019	0.0017
k__Bacteria;p__Proteobacteria;c__Epsilonproteobacteria;o__Campylobacteriales;f__Campylobacteraceae;g__Sulfurospirillum		0.30%	0.0000	0.0000	0.0093	0.0029	0.0028
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacteriales;f__Syntrophaceae;g__Syntrophus		0.29%	0.0000	0.0002	0.0046	0.0046	0.0038
k__Bacteria;p__Acidobacteria;c__Holophagae;o__Holophagales;f__Holophagaceae;g__		0.29%	0.0002	0.0000	0.0101	0.0096	0.0027
k__Bacteria;p__Firmicutes;c__Clostridia;o__Clostridiales;f__Ruminococcaceae;g__		0.29%	0.0004	0.0009	0.0029	0.0024	0.0030
k__Bacteria;p__Proteobacteria;c__Alphaproteobacteria;o__Rhodospirillales;f__Rhodospirillaceae;g__		0.28%	0.0070	0.0042	0.0012	0.0019	0.0020
k__Bacteria;p__Planctomycetes;c__Phycisphaerae;o__MSBL9;f__g__		0.28%	0.0073	0.0071	0.0018	0.0010	0.0021
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__Legionellales;f__Coxiellaceae;g__		0.27%	0.0016	0.0009	0.0011	0.0016	0.0010
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Burkholderiales;f__Comamonadaceae;Other		0.27%	0.0010	0.0010	0.0019	0.0010	0.0018

Genus Relative Abundance									
#OTU ID	pH.6.C.1	pH.6.C.2	pH.6.C.3	pH.7.5.A.1	pH.7.5.A.2	pH.7.5.A.3	pH.7.5.C.1	pH.7.5.C.2	pH.7.5.C.3
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__MBNT15;f__g__	0.0051	0.0069	0.0051	0.0028	0.0039	0.0041	0.0030	0.0028	0.0048
k__Bacteria;p__Acidobacteria;c__BPC102;o__MVS-40;f__g__	0.0037	0.0044	0.0039	0.0050	0.0047	0.0046	0.0042	0.0042	0.0052
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Hydrogenophilales;f__Hydrogenophilaceae;g__Thiobacillus	0.0034	0.0041	0.0025	0.0030	0.0026	0.0028	0.0049	0.0050	0.0058
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__f__g__	0.0032	0.0029	0.0033	0.0029	0.0037	0.0015	0.0040	0.0061	0.0049
k__Bacteria;p__Nitrospirae;c__Nitrospirae;o__Nitrospirales;f__Thermodesulfobivibrionaceae;g__GOUTA19	0.0074	0.0028	0.0076	0.0028	0.0042	0.0031	0.0049	0.0045	0.0045
k__Bacteria;p__Acidobacteria;c__Solibacteres;o__Solibacterales;f__g__	0.0049	0.0038	0.0020	0.0030	0.0034	0.0037	0.0043	0.0056	0.0047
k__Bacteria;p__Bacteroidetes;c__[Saprosiriae];o__[Saprosiriales];f__Chitinophagaceae;g__	0.0024	0.0026	0.0037	0.0030	0.0038	0.0031	0.0050	0.0037	0.0034
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__MND1;f__g__	0.0035	0.0033	0.0018	0.0026	0.0022	0.0028	0.0036	0.0041	0.0037
k__Bacteria;p__Planctomycetes;c__Planctomycetia;o__Pirellulales;f__Pirellulaceae;g__	0.0045	0.0047	0.0028	0.0030	0.0042	0.0030	0.0048	0.0045	0.0049
k__Bacteria;p__Chlorobi;c__BSV26;o__PK329;f__g__	0.0051	0.0043	0.0041	0.0045	0.0028	0.0027	0.0029	0.0042	0.0038
k__Bacteria;p__Actinobacteria;c__OP841;o__f__g__	0.0040	0.0058	0.0042	0.0039	0.0053	0.0032	0.0027	0.0042	0.0036
k__Bacteria;p__Firmicutes;c__Bacilli;o__Bacillales;f__Bacillaceae;g__Bacillus	0.0029	0.0027	0.0035	0.0030	0.0024	0.0020	0.0040	0.0035	0.0051
k__Archaea;p__Crenarchaeota;c__Thaumarchaeota;o__Nitrososphaerales;f__Nitrososphaeraeae;g__Candidatus Nitrososphaera	0.0057	0.0041	0.0059	0.0023	0.0030	0.0021	0.0024	0.0036	0.0035
k__Bacteria;p__Bacteroidetes;c__Bacteroidia;o__Bacteroidales;f__Rikenellaceae;g__Bvii28	0.0082	0.0092	0.0020	0.0011	0.0013	0.0030	0.0009	0.0031	0.0018
k__Bacteria;p__Chlorobi;c__SJA-28;o__f__g__	0.0033	0.0026	0.0028	0.0035	0.0020	0.0029	0.0017	0.0024	0.0027
k__Bacteria;p__Proteobacteria;c__Alphaproteobacteria;o__Rhizobiales;f__Hyphomicrobiaceae;g__Rhodoplanes	0.0033	0.0033	0.0028	0.0024	0.0018	0.0032	0.0036	0.0032	0.0036
k__Bacteria;p__Gemmatimonadetes;c__Gemm-1;o__f__g__	0.0028	0.0033	0.0015	0.0021	0.0024	0.0027	0.0032	0.0032	0.0030
k__Bacteria;p__Proteobacteria;c__Alphaproteobacteria;o__Sphingomonadales;f__Sphingomonadaceae;g__Kaistobacter	0.0032	0.0031	0.0026	0.0023	0.0021	0.0028	0.0027	0.0042	0.0030
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacteriales;f__Syntrophorhabdaceae;g__	0.0021	0.0032	0.0025	0.0028	0.0030	0.0044	0.0039	0.0040	0.0045
k__Bacteria;p__Elusimicrobia;c__Elusimicrobia;o__Elusimicrobiales;f__g__	0.0033	0.0037	0.0044	0.0030	0.0022	0.0017	0.0043	0.0033	0.0049
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;Other;Other;Other	0.0146	0.0045	0.0106	0.0004	0.0002	0.0006	0.0012	0.0008	0.0014
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Myxococcales;f__Hallangiaceae;g__	0.0029	0.0033	0.0015	0.0017	0.0020	0.0023	0.0021	0.0031	0.0036
k__Bacteria;p__GN04;c__GN15;o__f__g__	0.0024	0.0030	0.0019	0.0027	0.0031	0.0023	0.0035	0.0038	0.0031
k__Bacteria;p__WS3;c__PRR-12;o__Sediment-1;f__g__	0.0023	0.0024	0.0026	0.0022	0.0010	0.0027	0.0021	0.0021	0.0022
k__Bacteria;p__Nitrospirae;c__Nitrospirae;o__Nitrospirales;f__FW;g__4-29	0.0023	0.0016	0.0035	0.0021	0.0015	0.0015	0.0023	0.0031	0.0033
k__Bacteria;p__Proteobacteria;c__Epsilonproteobacteria;o__Campylobacteriales;f__Campylobacteraceae;g__Sulfurospirillum	0.0027	0.0024	0.0050	0.0020	0.0039	0.0027	0.0014	0.0028	0.0036
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacteriales;f__Syntrophaceae;g__Syntrophus	0.0027	0.0057	0.0047	0.0022	0.0012	0.0019	0.0030	0.0039	0.0027
k__Bacteria;p__Acidobacteria;c__Holophagae;o__Holophagales;f__Holophagaceae;g__	0.0043	0.0022	0.0023	0.0018	0.0014	0.0022	0.0007	0.0012	0.0021
k__Bacteria;p__Firmicutes;c__Clostridia;o__Clostridiales;f__Ruminococcaceae;g__	0.0040	0.0048	0.0057	0.0021	0.0017	0.0023	0.0033	0.0035	0.0036
k__Bacteria;p__Proteobacteria;c__Alphaproteobacteria;o__Rhodospirillales;f__Rhodospirillaceae;g__	0.0029	0.0024	0.0029	0.0023	0.0020	0.0017	0.0026	0.0022	0.0032
k__Bacteria;p__Planctomycetes;c__Phycisphaerae;o__MSBL9;f__g__	0.0016	0.0025	0.0019	0.0015	0.0022	0.0015	0.0028	0.0030	0.0024
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__Legionellales;f__Coxiellaceae;g__	0.0010	0.0017	0.0023	0.0015	0.0041	0.0087	0.0033	0.0043	0.0054
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Burkholderiales;f__Comamonadaceae;Other	0.0048	0.0061	0.0101	0.0019	0.0018	0.0017	0.0014	0.0014	0.0024

Genus Relative Abundance

#OTU ID	Relative Abundance	Marsh_sed_B	Mud_2	pH_6.A.1	pH_6.A.2	pH_6.A.3
k_Bacteria;p_Firmicutes;c_Clostridia;o_Clostridiales;f_[Acidaminobacteraceae];g_Fusibacter	0.27%	0.0002	0.0001	0.0002	0.0001	0.0001
k_Bacteria;p_Proteobacteria;c_Gammaproteobacteria;o_Xanthomonadales;f_Sinobacteraceae;g__	0.27%	0.0097455	0.005134	0.0010456	0.0011358	0.0014762
k_Bacteria;p_Verrucomicrobia;c_[Pedosphaerae];o_[Pedosphaerales];f_auto67_4W;g__	0.26%	0.0014438	0.007224	0.0010001	0.001467	0.0015254
k_Bacteria;p_Proteobacteria;c_Alphaproteobacteria;o_Rhizobiales;f__g__	0.26%	0.0069783	0.0080872	0.0012729	0.0015144	0.0013778
k_Bacteria;p_Spirochaetes;c_Spirochaetes;o_Spirochaetales;f_Spirochaetaceae;g_Treponema	0.26%	0	0	0.0036823	0.0049217	0.004035
k_Bacteria;p_Spirochaetes;c_MVP-15;o_PL-11810;f__g__	0.25%	6.016E-05	0	0.0005001	9.465E-05	0.0004921
k_Bacteria;p_Proteobacteria;c_Deltaproteobacteria;o_Syntrophobacteriales;f_Syntrophaceae;g_Desulfobacca	0.25%	0.0029477	0.0039527	0.0017275	0.0012304	0.0019191
k_Archaea;p_Euryarchaeota;c_Methanomicrobia;o_Methanosarcinales;f_Methanosetaceae;g_Methanoseta	0.25%	0	4.543E-05	0.0010001	0.0014197	0.0015254
k_Bacteria;p_Acidobacteria;c_Acidobacteria-6;o_CCU21;f__g__	0.24%	0.0073994	0.005134	0.0007274	0.0010884	0.0017715
k_Bacteria;p_Acidobacteria;c_o_f__g__	0.24%	0.0099862	0.0039527	0.0006819	0.0013251	0.0011318
k_Bacteria;p_Bacteroidetes;c_Sphingobacteriia;o_Sphingobacteriales;f__g__	0.23%	0.0019852	0.0015902	0.0013184	0.0014197	0.002362
k_Bacteria;p_Proteobacteria;c_Deltaproteobacteria;o_MIZ46;f__g__	0.23%	0.0003008	9.087E-05	0.0015457	0.0015617	0.0015746
k_Bacteria;p_Chloroflexi;c_Anaerolineae;o_SBR1031;f_SHA-31;g__	0.22%	0.0060158	0.0068151	0.0007274	0.0015144	0.0009349
k_Bacteria;p_Bacteroidetes;c_Cytophagia;o_Cytophagales;f_Cytophagaceae;g__	0.22%	0.0026469	0.0035438	0.0015457	0.0015617	0.0013286
k_Bacteria;p_Spirochaetes;c_Spirochaetes;o_Spirochaetales;f_Spirochaetaceae;g__	0.22%	0.0047525	0.0022263	0.0006819	0.0012304	0.0020667
k_Bacteria;p_Bacteroidetes;c_Sphingobacteriia;o_Sphingobacteriales;f_Sphingobacteriaceae;g__	0.22%	0.0067377	0.0050432	0.0006819	0.0005679	0.0006397
k_Bacteria;p_WS3;c_PRR-12;o_Sediment-1;f_PRR-10;g__	0.21%	0.0058353	0.0046343	0.0004091	0.0008518	0.0009842
k_Bacteria;p_Firmicutes;c_Clostridia;o_Clostridiales;f_Veillonellaceae;g__	0.21%	0	0	0.0036823	0.003218	0.0032969
k_Bacteria;p_GN04;c_o_f__g__	0.20%	0.0055345	0.0063153	0.0014093	0.0008992	0.0008857
k_Bacteria;p_Proteobacteria;c_Deltaproteobacteria;o_Desulfobacteriales;f_Desulfobulbaceae;g_Desulfobulus	0.20%	0.0003609	4.543E-05	0.0009547	0.0010411	0.001181
k_Bacteria;p_Proteobacteria;c_Betaproteobacteria;o_Rhodocyclales;f_Rhodocyclaceae;g_Other	0.19%	0.0004211	0.000318	0.0015457	0.0010411	0.0018659
k_Bacteria;p_Bacteroidetes;c_Cytophagia;o_Cytophagales;f_Flammeovirgaceae;g__	0.19%	0.0033688	0.0004089	0.000591	0.0013724	0.0013778
k_Bacteria;p_Lentisphaerae;c_Lentisphaerae;o_Victivallales;f_Victivallaceae;g__	0.19%	0	0	0.0035914	0.0043538	0.0036906
k_Bacteria;p_OD1;c_ZB2;o_f__g__	0.18%	0	4.543E-05	0.0027731	0.0035966	0.002362
k_Bacteria;p_Proteobacteria;c_Deltaproteobacteria;o_NB1;f__g__	0.17%	0.0059556	0.0043617	0.0003182	0.0012777	0.0011318
k_Bacteria;p_Acidobacteria;c_S035;o_f__g__	0.17%	0.0058954	0.0053612	0.0007274	0.0005679	0.0003937
k_Bacteria;p_Proteobacteria;c_Betaproteobacteria;o_Ellin6067;f__g__	0.17%	0.0055345	0.004498	0.0003182	0.0009938	0.0007873
k_Bacteria;p_Proteobacteria;c_Betaproteobacteria;o_Thiobacteriales;f__g__	0.17%	0.0001805	0.0001817	4.546E-05	0.0003313	9.842E-05
k_Bacteria;p_TM6;c_SBRH58;o_f__g__	0.17%	0	0	0.0009092	0.0065307	0.0027556
k_Bacteria;p_Proteobacteria;c_Gammaproteobacteria;o_Methylcoccales;f_Methylcocccaceae;g_Methylomonas	0.16%	0.0003609	0.0001817	0.0100014	0.0003313	0.000246
k_Bacteria;p_Proteobacteria;c_Deltaproteobacteria;o_Desulfuromonadales;f_Pelobacteraceae;g_Other	0.16%	0.0001805	4.543E-05	0.0009092	0.001467	0.0009349
k_Bacteria;p_Proteobacteria;c_Gammaproteobacteria;o_Methylcoccales;f_Methylcocccaceae;g_Other	0.16%	0.0020454	0.0039073	0.0008183	0.0008992	0.0009349
k_Bacteria;p_NC10;c_12-24;o_JH-WHS47;f__g__	0.16%	0.0035493	0.002408	0.0006819	0.0010884	0.0015746
k_Bacteria;p_Chloroflexi;c_Anaerolineae;o_CFB-26;f__g__	0.15%	0.0052939	0.0034075	0.0003637	0.0009465	0.0005413

Genus Relative Abundance

#OTU ID	pH.6.C.1	pH.6.C.2	pH.6.C.3	pH.7.5.A.1	pH.7.5.A.2	pH.7.5.A.3	pH.7.5.C.1	pH.7.5.C.2	pH.7.5.C.3
k__Bacteria;p__Firmicutes;c__Clostridia;o__Clostridiales;f__[Acidaminobacteraceae];g__Fusibacter	0.0008	0.0011	0.0012	0.0038	0.0053	0.0078	0.0042	0.0059	0.0075
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__Xanthomonadales;f__Sinobacteraceae;g__	0.0023	0.0024	0.0019	0.0011	0.0023	0.0015	0.0018	0.0027	0.0028
k__Bacteria;p__Verrucomicrobia;c__[Pedosphaerae];o__[Pedosphaerales];f__auto57_4W;g__	0.0033	0.0027	0.0036	0.0017	0.0023	0.0022	0.0025	0.0024	0.0036
k__Bacteria;p__Proteobacteria;c__Alphaproteobacteria;o__Rhizobiales;f__g__	0.0020	0.0017	0.0015	0.0022	0.0019	0.0013	0.0021	0.0021	0.0019
k__Bacteria;p__Spirochaetes;c__Spirochaetes;o__Spirochaetales;f__Spirochaetaceae;g__Treponema	0.0024	0.0019	0.0044	0.0025	0.0015	0.0031	0.0018	0.0029	0.0028
k__Bacteria;p__Spirochaetes;c__MWP-15;o__PL-11810;f__g__	0.0004	0.0002	0.0006	0.0092	0.0055	0.0125	0.0037	0.0007	0.0008
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Syntrophobacterales;f__Syntrophaceae;g__Desulfobacca	0.0020	0.0021	0.0042	0.0011	0.0020	0.0018	0.0033	0.0033	0.0031
k__Archaea;p__Euryarchaeota;c__Methanomicrobia;o__Methanosarcinales;f__Methanosarcetaceae;g__Methanosarata	0.0010	0.0002	0.0006	0.0050	0.0047	0.0094	0.0031	0.0037	0.0028
k__Bacteria;p__Acidobacteria;c__Acidobacteria-6;o__CCU21;f__g__	0.0020	0.0032	0.0010	0.0014	0.0010	0.0018	0.0017	0.0029	0.0020
k__Bacteria;p__Acidobacteria;c__o__f__g__	0.0021	0.0018	0.0020	0.0015	0.0014	0.0019	0.0015	0.0012	0.0027
k__Bacteria;p__Bacteroidetes;c__Sphingobacterii;o__Sphingobacteriales;f__g__	0.0014	0.0022	0.0015	0.0018	0.0027	0.0025	0.0034	0.0048	0.0035
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__MIZ46;f__g__	0.0040	0.0033	0.0040	0.0027	0.0025	0.0035	0.0028	0.0028	0.0013
k__Bacteria;p__Chloroflexi;c__Anaerolineae;o__SBR1031;f__SHA-31;g__	0.0022	0.0024	0.0015	0.0018	0.0015	0.0011	0.0015	0.0012	0.0018
k__Bacteria;p__Bacteroidetes;c__Cytophagia;o__Cytophagales;f__Cytophagaceae;g__	0.0020	0.0009	0.0010	0.0027	0.0021	0.0023	0.0027	0.0036	0.0030
k__Bacteria;p__Spirochaetes;c__Spirochaetes;o__Spirochaetales;f__Spirochaetaceae;g__	0.0014	0.0019	0.0004	0.0026	0.0025	0.0025	0.0019	0.0035	0.0032
k__Bacteria;p__Bacteroidetes;c__Sphingobacterii;o__Sphingobacteriales;f__Sphingobacteriaceae;g__	0.0019	0.0017	0.0017	0.0016	0.0018	0.0026	0.0014	0.0021	0.0017
k__Bacteria;p__Bacteroidetes;c__PRR-12;o__Sediment-1;f__PRR-10;g__	0.0011	0.0020	0.0012	0.0016	0.0016	0.0023	0.0016	0.0027	0.0029
k__Bacteria;p__Firmicutes;c__Clostridia;o__Clostridiales;f__Veillonellaceae;g__	0.0023	0.0031	0.0041	0.0033	0.0012	0.0016	0.0006	0.0016	0.0011
k__Bacteria;p__GN04;c__o__f__g__	0.0020	0.0018	0.0014	0.0012	0.0012	0.0011	0.0009	0.0017	0.0019
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfobacteriales;f__Desulfobulbaceae;g__Desulfobulbus	0.0011	0.0015	0.0010	0.0041	0.0027	0.0037	0.0042	0.0034	0.0028
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Rhodocyclales;f__Rhodocyclaceae;Other	0.0009	0.0011	0.0005	0.0018	0.0014	0.0019	0.0047	0.0032	0.0063
k__Bacteria;p__Bacteroidetes;c__Cytophagia;o__Cytophagales;f__Flammeovirgaceae;g__	0.0021	0.0016	0.0010	0.0021	0.0024	0.0019	0.0025	0.0033	0.0028
k__Bacteria;p__Lentisphaerae;c__[Lentisphaeria];o__Victivallales;f__Victivallaceae;g__	0.0022	0.0026	0.0047	0.0012	0.0009	0.0014	0.0005	0.0007	0.0006
k__Bacteria;p__OD1;c__ZB2;o__f__g__	0.0031	0.0025	0.0041	0.0008	0.0001	0.0009	0.0031	0.0006	0.0008
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__NB1-1;f__g__	0.0013	0.0013	0.0013	0.0015	0.0006	0.0017	0.0013	0.0005	0.0016
k__Bacteria;p__Acidobacteria;c__S035;o__f__g__	0.0014	0.0010	0.0017	0.0012	0.0009	0.0009	0.0009	0.0013	0.0018
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Ellin6067;f__g__	0.0018	0.0026	0.0006	0.0008	0.0015	0.0009	0.0007	0.0014	0.0013
k__Bacteria;p__Proteobacteria;c__Betaproteobacteria;o__Thiobacteriales;f__g__	0.0011	0.0000	0.0002	0.0001	0.0007	0.0009	0.0055	0.0014	0.0127
k__Bacteria;p__TM6;c__SBRH58;o__f__g__	0.0005	0.0008	0.0003	0.0004	0.0030	0.0011	0.0014	0.0015	0.0039
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__Methylococcales;f__Methylococcaceae;g__Methylomonas	0.0007	0.0002	0.0004	0.0003	0.0002	0.0002	0.0093	0.0002	0.0004
k__Bacteria;p__Proteobacteria;c__Deltaproteobacteria;o__Desulfuromonadales;f__Pelobacteraceae;Other	0.0023	0.0023	0.0030	0.0016	0.0015	0.0015	0.0020	0.0029	0.0023
k__Bacteria;p__Proteobacteria;c__Gammaproteobacteria;o__Methylococcales;f__Methylococcaceae;Other	0.0010	0.0007	0.0011	0.0015	0.0015	0.0016	0.0015	0.0016	0.0027
k__Bacteria;p__NC10;c__12-24;o__JH-WHS47;f__g__	0.0008	0.0014	0.0009	0.0015	0.0012	0.0019	0.0014	0.0012	0.0020

Appendix F - Sorption Calculations

Reactor	Days	pH	Goethite moles	>(s)FeOH moles	>(w)FeOH moles	tot Fe++ moles	Fe++ sorbed fraction	sorbed Fe++ mmoles	sorbed Fe++/goeth mmol/mol
6A 1	56	6.32	0.004206	3.79E-06	1.51E-04	0.0022	0.02849	0.062678	14.9020447
6A 1	91	6.16	0.004704	4.23E-06	1.69E-04	6.92E-05	0.1295	0.0089614	1.905059524
6A 2	56	6.31	0.003605	3.24E-06	1.30E-04	0.00231	0.02322	0.0536382	14.87883495
6A 2	91	6.28	0.002803	2.52E-06	1.01E-04	0.00129	0.0275	0.035475	12.65608277
6A 3	56	6.33	0.004006	3.61E-06	1.44E-04	0.00205	0.02899	0.0594295	14.83512232
6A 3	91	6.24	0.002603	2.34E-06	9.37E-05	0.00164	0.02015	0.033046	12.69535152
6C 1	56	6.34	0.004406	3.97E-06	1.59E-04	0.00194	0.03433	0.0666002	15.11579664
6C 1	91	6.11	0.003805	3.42E-06	1.37E-04	0.00108	0.02847	0.0307476	8.080840999
6C 2	56	6.28	0.004406	3.97E-06	1.59E-04	0.002	0.03026	0.06052	13.7358148
6C 2	91	6.1	0.003505	3.15E-06	1.26E-04	0.0013	0.02269	0.029497	8.415691869
6C 3	56	6.45	0.004405	3.96E-06	1.59E-04	0.00197	0.03951	0.0778347	17.66962543
6C 3	91	6.05	0.003905	3.51E-06	1.41E-04	0.00109	0.02421	0.0263889	6.757720871
7.5A 1	56	7.53	0.009806	8.83E-06	0.000353	0.0001	0.701	0.0701	7.148684479
7.5A 1	91	6.85	0.009706	8.74E-06	0.000349	0.000116	0.6122	0.0710152	7.316628889
7.5A 2	56	7.55	0.009806	8.83E-06	0.000353	0.000103	0.6955	0.0716365	7.305374261
7.5A 2	91	7.12	0.009706	8.74E-06	0.000349	1.14E-04	0.6547	0.0746358	7.689655883
7.5A 3	56	7.53	0.009706	8.74E-06	0.000349	1.26E-04	0.6498	0.0818748	8.435483206
7.5A 3	91	7.11	0.009606	8.65E-06	0.000346	1.26E-04	0.6299	0.0793674	8.262273579
7.5C 1	56	7.56	0.009806	8.83E-06	0.000353	9.73E-05	0.7078	0.06886894	7.023142974
7.5C 1	91	7.31	0.009806	8.83E-06	0.000353	6.73E-05	0.7688	0.05174024	5.276385886
7.5C 2	56	7.54	0.009806	8.83E-06	0.000353	1.11E-04	0.6797	0.0754467	7.693932286
7.5C 2	91	7.23	0.009706	8.74E-06	0.000349	1.01E-04	0.6881	0.0694981	7.160323511
7.5C 3	56	7.54	0.009806	8.83E-06	0.000353	1.15E-04	0.672	0.07728	7.880889251
7.5C 3	91	7.1	0.009606	8.65E-06	0.000346	1.21E-04	0.6367	0.0770407	8.020060379