THE INFLUENCE OF OXYGEN DELIVERY AND OXYGEN UTILIZATION ON THE DETERMINANTS OF EXERCISE TOLERANCE

by

RYAN MICHAEL BROXTERMAN

B.A., Washburn University, 2009

M.S., Kansas State University, 2011

AN ABSTRACT OF A DISSERTATION

submitted in partial fulfillment of the requirements for the degree

DOCTOR OF PHILOSOPHY

Department of Anatomy and Physiology College of Veterinary Medicine

KANSAS STATE UNIVERSITY Manhattan, Kansas

2015

Abstract

The physiological mechanisms determining the tolerable duration of exercise dictate human physical accomplishments across all spectrums of life. Despite extensive study, these specific mechanisms, and their dependence on oxygen delivery and oxygen utilization, remain, a certain extent, undefined. The purpose of this dissertation was to test the overarching hypothesis that muscle contraction characteristics (i.e., intensity of contraction, muscle contractionrelaxation duty cycle, etc.) alter oxygen delivery and oxygen utilization, which directly influence the power-duration relationship and fatigue development, and therefore, exercise tolerance. To accomplish this, specific interventions of altered muscle contraction-relaxation duty cycle and blood flow occlusion were utilized. In the first investigation (Chapter 2), we utilized low and high muscle contraction-relaxation duty cycles to alter blood flow to the active skeletal muscle, demonstrating that critical power (CP) was reduced with the high muscle contraction-relaxation duty cycle due to a reduction in blood flow, while the curvature constant (W') was not altered. The second investigation (Chapter 3) utilized blood flow occlusion to show that CP was reduced and W' increased for blood flow occlusion exercise conditions compared to control blood flow exercise conditions. The final investigation (Chapter 4) utilized periods of blood flow occlusion during and post-exercise to reveal greater magnitudes of peripheral and central fatigue development during blood flow occlusion exercise compared to control blood flow exercise. Moreover, this investigation demonstrated that W' was significantly related to the magnitude of fatigue development. Collectively, alterations in oxygen delivery and oxygen utilization via muscle contraction characteristics and blood flow occlusion directly influence CP and the magnitude of fatigue development. However, W' does not appear to be influenced by manipulations in oxygen delivery and oxygen utilization, per se. Rather, W' may be determined by the magnitude of fatigue accrued during exercise, which is dependent upon oxygen delivery and oxygen utilization. The novel findings of the investigations presented in this dissertation highlight important physiological mechanisms that determine exercise tolerance and demonstrate the need for interventions that improve oxygen delivery and oxygen utilization in specific populations, such as those with chronic heart failure or chronic obstructive pulmonary disease, to improve exercise tolerance.

THE INFLUENCE OF OXYGEN DELIVERY AND OXYGEN UTILIZATION ON THE DETERMINANTS OF EXERCISE TOLERANCE

by

RYAN MICHAEL BROXTERMAN

B.A., Washburn University, 2009

M.S., Kansas State University, 2011

A DISSERTATION

submitted in partial fulfillment of the requirements for the degree

DOCTOR OF PHILOSOPHY

Department of Anatomy and Physiology College of Veterinary Medicine

KANSAS STATE UNIVERSITY Manhattan, Kansas

2015

Approved by:

Major Professor Dr. Thomas J. Barstow

Copyright

RYAN MICHAEL BROXTERMAN

2015

Abstract

The physiological mechanisms determining the tolerable duration of exercise dictate human physical accomplishments across all spectrums of life. Despite extensive study, these specific mechanisms, and their dependence on oxygen delivery and oxygen utilization, remain, a certain extent, undefined. The purpose of this dissertation was to test the overarching hypothesis that muscle contraction characteristics (i.e., intensity of contraction, muscle contractionrelaxation duty cycle, etc.) alter oxygen delivery and oxygen utilization, which directly influence the power-duration relationship and fatigue development, and therefore, exercise tolerance. To accomplish this, specific interventions of altered muscle contraction-relaxation duty cycle and blood flow occlusion were utilized. In the first investigation (Chapter 2), we utilized low and high muscle contraction-relaxation duty cycles to alter blood flow to the active skeletal muscle, demonstrating that critical power (CP) was reduced with the high muscle contraction-relaxation duty cycle due to a reduction in blood flow, while the curvature constant (W') was not altered. The second investigation (Chapter 3) utilized blood flow occlusion to show that CP was reduced and W' increased for blood flow occlusion exercise conditions compared to control blood flow exercise conditions. The final investigation (Chapter 4) utilized periods of blood flow occlusion during and post-exercise to reveal greater magnitudes of peripheral and central fatigue development during blood flow occlusion exercise compared to control blood flow exercise. Moreover, this investigation demonstrated that W' was significantly related to the magnitude of fatigue development. Collectively, alterations in oxygen delivery and oxygen utilization via muscle contraction characteristics and blood flow occlusion directly influence CP and the magnitude of fatigue development. However, W' does not appear to be influenced by manipulations in oxygen delivery and oxygen utilization, per se. Rather, W' may be determined by the magnitude of fatigue accrued during exercise, which is dependent upon oxygen delivery and oxygen utilization. The novel findings of the investigations presented in this dissertation highlight important physiological mechanisms that determine exercise tolerance and demonstrate the need for interventions that improve oxygen delivery and oxygen utilization in specific populations, such as those with chronic heart failure or chronic obstructive pulmonary disease, to improve exercise tolerance.

Table of Contents

List of Figures	V111
List of Tables	ix
Acknowledgements	X
Preface	xii
Chapter 1 - Introduction	1
References	5
Chapter 2 - Influence of duty cycle on the power-duration relationship for handgrip exercise	:
observations and potential mechanisms	10
Summary	11
Introduction	12
Methods	15
Results	23
Discussion	26
References	42
Chapter 3 - Influence of blood flow occlusion on muscle oxygenation characteristics and the	;
parameters of the power-duration relationship	49
Summary	50
Introduction	51
Methods	54
Results	59
Discussion	61
References	78
Chapter 4 - Influence of blood flow occlusion on the development of peripheral and central	
fatigue during small muscle mass handgrip exercise	83
Summary	84
Introduction	85
Methods	88
Results	94
Discussion	96

References	108
Chapter 5 - Conclusions	113
Appendix A - Curriculum Vitae	115

List of Figures

Figure 2.1 Displacement profiles for the 50% and 20% duty cycles.	35
Figure 2.2 A representative subject's power-duration relationship for the two duty cycles	36
Figure 2.3 Brachial artery blood flow responses for the two duty cycles at the same absolute	
power output.	37
Figure 2.4 Deoxygenated-[hemoglobin + myoglobin] response for the two duty cycles at the	;
same power output.	38
Figure 2.5 Mean electromyography response for the two duty cycles at the same power outp	ut.39
Figure 2.6 Estimated oxygen uptake response for the two duty cycles at the same power outp	out.
	40
Figure 2.7 Diagram of estimated oxygen uptake as a function of microvascular P_{O_2} for each	duty
cycle.	41
Figure 3.1 Individual subject constant-power and hyperbolic curve fit data	71
Figure 3.2 Mean CP and W' determined for control and occlusion handgrip exercise	72
Figure 3.3 Mean NIRS muscle oxygenation data for control exercise.	73
Figure 3.4 Mean NIRS muscle oxygenation data for occlusion exercise.	74
Figure 3.5 Mean NIRS muscle oxygenation data for control and occlusion at 110 %P _{peak}	75
Figure 3.6 Mean EMG data for control and occlusion.	76
Figure 3.7 Schematic diagram demonstrating the contribution of W' to mechanical- and non	
mechanical-work energy consumption processes.	77
Figure 4.1 Experimental design.	102
Figure 4.2 Mean NIRS muscle oxygenation data during each exercise trial.	103
Figure 4.3 Mean EMG data for each exercise trial.	104
Figure 4.4 Neuromuscular function for Control and Occlusion exercise trials.	105
Figure 4.5 Neuromuscular function for Control + Occlusion and Occlusion + Occlusion exer	rcise
trials	106
Figure 4.6 Relationship between W' and the change in neuromuscular function variables	107

List of Tables

Table 2.1 Brachial artery diameter and blood velocity data.	34
Table 3.1 End-exercise NIRS values for control and occlusion.	69
Table 3.2 Exercise onset deoxy-[Hb + Mb] kinetics parameters for control and occlusion	70

Acknowledgements

"If I have seen further it is by standing on the shoulders of giants." – Sir Isaac Newton

I am forever grateful to those whom I have interacted with throughout my career as a graduate student at Kansas State University, especially my mentors whose tutelage I will carry with me the rest of my life.

I would like to thank Dr. Thomas J. Barstow for his excellent mentorship and commitment to me as a graduate student. I will always try to emulate his deep passion for scientific research and academics in my own career. He is an outstanding person, whom I thank for all of the time and effort he so graciously shared with me

In addition to Dr. Barstow, I am grateful for the professional and scientific guidance from my committee members: Drs. David C. Poole, Craig A. Harms, Timothy I. Musch, Andrew M. Jones, and Brian Geisbrecht. I thank Drs. Musch and Poole for instilling in me a greater understanding of the philosophy of science and the importance of letting the scientific evidence speak for itself. I am grateful to Dr. Harms for continually highlighting the importance of answering meaningful questions. I am thankful to Dr. Jones for his willingness to serve on my committee and his high standard for scientific research. Additionally, I thank Dr. Brian Geisbrecht for serving as outside chair on my committee.

To the graduate students I have had the pleasure of interacting with, thank you. I want to specifically thank Samuel Wilcox, Jesse Craig, Josh Smith, Stephanie Kurti, Dr. Steven Copp, Dr. Daniel Hirai, Clark Holdsworth, and Scott Ferguson for their influence on my graduate student career. I would also like to extend a special thank you to Dr. Carl Ade for the many fun times we have had and his consistent motivation to make me a better person.

I am forever indebted to my family for their unending support throughout my life. You are truly special people that I am lucky to have had by my side every step of the way. My parents have inspired me to go after my dreams and to not let anyone prevent me from achieving them. I love you and thank you for all you have done for me. I owe a special thank you to my wife Carrie and my son Lane. They are the foundation of my life and the ones who have kept me focused through it all. I love you and I hope you know how much I appreciate you and your support throughout this journey. Ultimately, I would like to thank God for giving me these opportunities and wonderful people that have so deeply influenced my life.

Preface

Chapters 2-4 of this dissertation represent original research articles that have been published following or are currently in the peer-review process (citations may be found below). They are reproduced here with permission from the publishers.

Broxterman RM, Ade CJ, Wilcox SL, Schlup SJ, Craig JC, and Barstow TJ.

Influence of duty cycle on the power-duration relationship: observations and potential mechanism. *Respiratory Physiology and Neurobiology* 192: 102-111, 2014.

Broxterman RM, Ade CJ, Craig JC, Wilcox SL, Schlup SJ, and Barstow TJ.

Influence of blood flow occlusion on muscle oxygenation characteristics and the parameters of the power-duration relationship. *Journal of Applied Physiology* In press, doi:10.1152/japplphysiol.00875.2014.

Broxterman RM, Craig JC, Smith JR, Wilcox SL, Jia C, Warren S, and Barstow TJ. Influence of blood flow occlusion on the development of peripheral and central fatigue during small muscle mass handgrip exercise. *Journal of Physiology* In Revision.

Chapter 1 - Introduction

The physiological mechanisms determining the tolerable duration of activity have dictated human physical accomplishments throughout history. The notion of a relationship between the intensity of an exercise and the tolerable duration of that exercise dates back as early as the fourth century (59, 60). This relationship constrains human performance across all spectrums of life, from elite athletes to disease populations (such as those with chronic heart failure or chronic obstructive pulmonary disease) where activities of daily living and the quality of life are contingent upon the tolerable intensities of exercise.

The relationship between progressively increasing power outputs and decreasing exercise duration was first described by A.V. Hill in the early 1900s (25, 26) and formally characterized in 1965 by Monod and Scherrer (41). This robust power-duration relationship is now commonly characterized using a two-parameter hyperbolic mathematical model to obtain the asymptote (critical power (CP)) and the curvature constant (W') (27, 31, 58). CP represents the highest attainable steady-state for energy production without continually drawing upon W' (13, 15, 41, 42, 45, 55), while W' has been purported to be determined by intramuscular energy stores (38-40), the accumulation of fatigue inducing metabolites (12, 19, 21, 33), and/or the magnitude of the severe-intensity exercise domain (9, 55). Furthermore, CP is the highest intensity in which a physiological steady-state can be achieved for oxygen uptake (\dot{V}_{O_2}), blood flow, intramuscular concentrations of phosphocreatine ([PCr]), inorganic phosphate ([Pi]), and hydrogen ions ([H[†]]) (14, 33, 46). As such, CP demarcates the boundary between the heavy- and severe-intensity exercise domains, distinguishing sustainable and unsustainable intensities of exercise (45).

Elucidating the mechanisms of fatigue has been a primary focal point for understanding the physiological determinants of exercise tolerance. Fatigue is defined as a reversible decrease in the

ability to produce voluntary maximal force (2, 17, 24) and can be quantified as peripheral or central in origin. Peripheral fatigue occurs at or distal to the neuromuscular junction, while central fatigue occurs proximal to the neuromuscular junction (2, 24). Accumulating evidence suggests that exercise tolerance above CP is limited by the development of fatigue (3, 4, 10, 16, 23, 49, 50, 53). Amann et al. (7) merged the concepts of a "critical threshold" of muscle fatigue, a "sensory tolerance limit" of group III/IV muscle afferent feedback, and central motor drive into a paradigm describing the mechanisms determining exercise tolerance. This integrative paradigm highlights the importance of feedback from group III/IV muscle afferent fibers to the central nervous system regarding the physiological state of the working skeletal muscle (1, 34, 35) in determining the "critical threshold" of fatigue (6, 24) and the point where central motor drive becomes limited or limiting (54). This physiological paradigm is purported to limit the magnitude of fatigue developed during exercise as a component of homeostasis (5, 7, 50). Recently, Pethick et al. (44) demonstrated, that beyond a decrease in torque-generating capacity, fatigue also limits the ability of the neuromuscular system to adapt to external perturbation. Thus, the magnitude of fatigue developed during exercise dictates the exercise tolerance.

Oxygen delivery and oxygen utilization are important determinants of the power-duration relationship and the development of fatigue during exercise. CP has been demonstrated to be aerobic in nature and influenced by alterations in the fraction of inspired oxygen content (15, 42, 55). Although it has been demonstrated that blood flow can be impeded or occluded due to the increased intramuscular pressure accompanying muscle contraction (28, 36, 48, 52) and that rhythmic alterations in blood flow occur throughout the muscle contraction-relaxation cycle (8, 20, 48, 57), the influence of muscle contraction-relaxation alterations in blood flow on CP has not been investigated. Less is understood regarding the influence of oxygen delivery and oxygen utilization on W', but this parameter has traditionally been associated with anaerobic energy production due to early findings that interventions

affecting anaerobic energy production altered W' (18, 19, 30, 38, 39) and interventions affecting oxygen delivery and oxygen utilization did not alter W' (22, 29, 41, 42, 47). Moreover, these lines of thinking were greatly influenced by Monod and Scherrer (41) who in originally characterizing the power-duration relationship speculated on the influence of blood flow occlusion by stating, "Factor b [CP] is linked to circulatory conditions in the muscle. For when the dynamic work is performed under arterial cuff, the maximum work becomes constant whatever be the time after which exhaustion occurs. The maximum work is then equal to factor a [W']..." However, this speculation remains to be empirically tested and more recent evidence suggests that W' may be associated with the severe-intensity exercise domain (9, 55) and the \dot{V}_{O_2} slow component (19, 32, 43, 56). Moreover, W' has been shown to be decreased during hyperoxia compared to normoxia (55). The magnitude of peripheral fatigue developed during exercise has been demonstrated to be independent of alterations in oxygen delivery and oxygen utilization for large muscle mass activity (4, 49), but not for small muscle mass activity (11, 37, 51). Collectively, these findings suggest that oxygen delivery and oxygen utilization influence CP, while the influence on W' and the magnitude of fatigue development is equivocal.

The findings of Burnley et al. (10) that peripheral fatigue development occurs above CP, in combination with the fatigue paradigm of Amann et al. (7), suggest that CP may represent the exercise intensity above which exercise tolerance is limited by the attainment of the "sensory tolerance limit". Therefore, the mechanisms determining W' may be related to the magnitude of fatigue developed during severe-intensity exercise. In addition, a constant "sensory tolerance limit" for a given exercise condition would constrain the amount of work that could be performed above CP and the degree of intramuscular metabolic perturbation, which may explain the consistency in these variables associated with the complete utilization of W' (33, 41, 42, 46, 55).

There are currently no investigations that have altered the muscle contraction-relaxation duty cycle or utilized blood flow occlusion to examine the influence of oxygen delivery and oxygen utilization on the power-duration relationship and the magnitude of fatigue development. The purpose of this dissertation was to test the overarching hypothesis that muscle contraction characteristics (i.e., intensity of contraction, muscle contraction-relaxation duty cycle, etc.) alter oxygen delivery and oxygen utilization, which directly influence the power-duration relationship and fatigue development, and therefore, exercise tolerance.

References

- 1. **Adreani CM, Hill JM, and Kaufman MP**. Responses of group III and IV muscle afferents to dynamic exercise. *J Appl Physiol* 82: 1811-1817, 1997.
- 2. **Allen DG, Lamb GD, and Westerblad H**. Skeletal muscle fatigue: Cellular mechanisms. *Physiological Reviews* 88: 287-332, 2008.
- 3. **Amann M, and Dempsey JA**. Locomotor muscle fatigue modifies central motor drive in healthy humans and imposes a limitation to exercise performance. *J Physiol* 586: 161-173, 2008.
- 4. **Amann M, Eldridge MW, Lovering AT, Stickland MK, Pegelow DF, and Dempsey JA**. Arterial oxygenation influences central motor output and exercise performance via effects on peripheral locomotor muscle fatigue in humans. *J Physiol* 575: 937-952, 2006.
- 5. **Amann M, Proctor LT, Sebranek JJ, Eldridge MW, Pegelow DF, and Dempsey JA**. Somatosensory feedback from the limbs exerts inhibitory influence on central neural drive during whole body endurance exercise. *J Appl Physiol* 105: 1717-1724, 2008.
- 6. **Amann M, Proctor LT, Sebranek JJ, Pegelow DF, and Dempsey JA**. Opiod-mediated muscle afferents inhibit central motor drive and limit peripheral muscle fatigue development in humans. *J Physiol* 587: 271-283, 2009.
- 7. **Amann M, Venturelli M, Ives SJ, McDaniel J, Layec G, Rossman MJ, and Richardson RS**. Peripheral fatigue limits endurance exercise via a sensory feedback-mediated reduction in spinal motorneuronal output. *J Appl Physiol* 115: 355-364, 2013.
- 8. **Barcroft H, and Dornhorst AC**. The blood flow through the human calf during rhythmic exercise. *J Physiol* 109: 402-411, 1949.
- 9. **Burnley M, and Jones AM**. Oxygen uptake kinetics as a determinant of sports performance. *Eur J Sport Sci* 7: 63-79, 2007.
- Burnley M, Vanhatalo A, and Jones AM. Distinct profiles of neuromuscular fatigue during muscle contractions below and above the critical torque in humans. *J Appl Physiol* 113: 215-223, 2012.
- 11. **Christian RJ, Bishop DJ, Billaut F, and Girard O**. Peripheral fatigue is not critically regulated during maximal, intermittent, dynamic leg extensions. *J Appl Physiol* 117: 1063-1073, 2014.
- 12. Coats EM, Rossiter HB, Day JR, Miura A, Fukuba Y, and Whipp BJ. Intensity-dependent tolerance to exercise after attaining $\dot{V}O_{2max}$ in humans. *J Appl Physiol* 95: 483-490, 2003.

- 13. **Conley KE, Kemper WE, and Crowther GJ**. Limits to sustainable muscle performance: interaction between glycolysis and oxidative phosphorylation. *The Journal of Experimental Biology* 204: 3189-3194, 2001.
- 14. **Copp SW, Hirai DM, Musch TI, and Poole DC**. Critical speed in the rat: implications for hindlimb muscle blood flow distribution and fibre recruitment. *J Physiol* 588: 5077-5087, 2010.
- 15. **Dekerle J, Mucci P, and Carter H**. Influence of moderate hypoxia on tolerance to high-intensity exercise. *Eur J Appl Physiol* 112: 327-335, 2012.
- 16. **Duffield R, Green R, Castle P, and Maxwell N**. Precooling can prevent the reduction in self-paced exercise intensity in the heat. *Med Sci Sports Exerc* 42: 577-584, 2010.
- 17. **Enoka RM, and Duchateau J**. Muscle fatigue: what, why and how it influences muscle function. *J Physiol* 586.1: 11-23, 2008.
- 18. **Ferguson CS, Rossiter HB, Whipp BJ, Cathcart AJ, Murgatroyd SR, and Ward SA**. Effect of recovery duration from prior exhaustive exercise on the parameters of the power-duration relationship. *J Appl Physiol* 108: 866-874, 2010.
- 19. **Ferguson CS, Whipp BJ, Cathcart AJ, Rossiter HB, Turner AP, and Ward SA**. Effects of prior very-heavy intensity exercise on indices of aerobic function and high-intensity exercise tolerance. *J Appl Physiol* 103: 812-822, 2007.
- 20. **Folkow B, Gaskell P, and Waaler BA**. Blood flow through limb muscles during heavy rhythmic exercise. *Acto Physiologica* 80: 61-72, 1970.
- 21. **Fukuba Y, Miura A, Endo M, Kan A, Yanagawa K, and Whipp BJ**. The curvature constant parameter of the power-duration curve for varied-power exercise. *Med Sci Sports Exerc* 35: 1413-1418, 2003.
- 22. **Gaesser GA, and Wilson LA**. Effects of continuous and interval training on the parameters of the power-endurance time relationship for high-intensity exercise. *Int J Sports Med* 9: 417-421, 1988.
- 23. **Gagnon P, Saey D, Vivodtzev I, Laviolette L, Mainguy V, Milot J, Provencher S, and Maltais F.** Impact of preinduced quadriceps fatigue on exercise response in chronic obstructive pulmonary disease and healthy subjects. *J Appl Physiol* 107: 832-840, 2009.
- 24. **Gandevia SC**. Spinal and supraspinal factors in human muscle fatigue. *Physiol Rev* 81: 1725-1789, 2001.
- 25. **Hill AV**. The physiological basis of athletic records. *Nature* 116: 544-548, 1925.

- 26. **Hill AV**. Speed and energy requirement. In: *Muscular Movement in Man*. New York: McGraw-Hill, 1927, p. 41-44.
- 27. **Hill DW**. The critical power concept: A review. *Sports Med* 16: 237-254, 1993.
- 28. **Hoelting BD, Scheuermann BW, and Barstow TJ**. Effect of contraction frequency on leg blood flow during knee extension exercise in humans. *J Appl Physiol* 91: 671-679, 2001.
- 29. **Jenkins DG, and Quigley BM**. Endurance training enhances critical power. *Med Sci Sports Exerc* 24: 1283-1289, 1992.
- 30. **Jenkins DG, and Quigley BM**. The influence of high-intensity exercise training on the Wlim-Tlim relationship. *Med Sci Sports Exerc* 25: 275-282, 1993.
- Jones AM, Vanhatalo A, Burnley M, Morton RH, and Poole DC. Critical power: Implications for determination of VO_{2max} and exercise tolerance. *Med Sci Sports Exerc* 42: 1876-1890, 2010.
- 32. **Jones AM, Wilkerson DP, Burnley M, and Koppo K**. Prior heavy exercise enhances performance during subsequent perimaximal exercise. *Med Sci Sports Exerc* 35: 2085-2092, 2003.
- 33. **Jones AM, Wilkerson DP, DiMenna FJ, Fulford J, and Poole DC**. Muscle metabolic responses to exercise above and below the "critical power" assessed using ³¹P-MRS. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 294: 585-593, 2008.
- 34. **Kaufman MP, and Rybicki KJ**. Discharge properties of group III and IV muscle afferents: their responses to mechanical and metabolic stimuli. *Circulation Research* 61: I60-I65, 1987.
- 35. **Kennedy DS, Fitzpatrick SC, Gandevia SC, and Taylor JL**. Fatigue-related firing of muscle nociceptors reduces voluntary activation of ipsilateral but not contralateral lower limb muscles. *J Appl Physiol* 118: 408-418, 2015.
- 36. Lutjemeier BJ, Miura A, Scheuermann BW, Koga S, Townsend DK, and Barstow TJ. Muscle contraction-blood flow interactions during upright knee exercise in humans. *J Appl Physiol* 98: 1575-1583, 2005.
- 37. **Millet GY, Aubert D, Favier FB, Busso T, and Benoît H**. Effect of acute hypoxia on central fatigue during repeated isometric leg contractions. *Scand J Med Sci Sports* 19: 695-702, 2009.

- 38. **Miura A, Kino F, Kajitani S, Sato H, Sato H, and Fukuba Y**. The effect of oral creatine supplementation on the curvature constant parameter of the power-duration curve for cycle ergometry in humans. *Japanese Journal of Physiology* 49: 169-174, 1999.
- 39. **Miura A, Sato H, Sato H, Whipp BJ, and Fukuba Y**. The effect of glycogen depletion on the curvature constant parameter of the power-duration curve for cycle ergometry. *Ergonomics* 43: 133-141, 2000.
- 40. **Molé PA, Chung Y, Tran TK, Sailasuta N, Hurn R, and Jue T**. Myoglobin desaturation with exercise intensity in human gastronemius muscle. *Am J Physiol Regul Integr Comp Physiol* 277: R173 R180, 1999.
- 41. **Monod H, and Scherrer J**. The work capacity of a synergic muscular group. *Ergonomics* 8: 329-338, 1965.
- 42. **Moritani T, Nagata A, DeVries HA, and Muro M**. Critical power as a measure of physical work capacity and anaerobic threshold. *Ergonomics* 24: 339-350, 1981.
- 43. **Murgatroyd SR, Ferguson CS, Ward SA, Whipp BJ, and Rossiter HB**. Pulmonary O₂ uptake kinetics as a determinant of high-intensity exercise tolerance in humans. *J Appl Physiol* 110: 1598-1606, 2011.
- 44. **Pethick J, Winter SL, and Burnley M**. Fatigue reduces the complexity of knee extensors torque fluctuations during maximal and submaximal intermittent isometric contractions in man. *J Physiol* 2015.
- 45. **Poole DC**. Resolving the determinants of high-intensity exercise performance. *Exp Physiol* 94: 197-198, 2008.
- 46. **Poole DC, Ward SA, Gardner GW, and Whipp BJ**. Metabolic and respiratory profile of the upper limit for prolonged exercise in man. *Ergonomics* 31: 1265-1279, 1988.
- 47. **Poole DC, Ward SA, and Whipp BJ**. The effects of training on the metabolic and respiratory profile of high-intensity cycle ergometer exercise. *Eur J Appl Physiol* 59: 421-429, 1990.
- 48. **Robergs RA, Icenogle MV, Hudson TL, and Greene ER**. Temporal inhomogeneity in brachial artery blood flow during forearm exercise. *Med Sci Sports Exerc* 29: 1021-1027, 1997.
- 49. **Romer LM, Haverkamp HC, Amann M, Lovering AT, Pegelow DF, and Dempsey JA**. Effect of acute severe hypoxia on peripheral fatigue and endurance capacity in healthy humans. *Am J Physiol Regul Integr Comp Physiol* 292: 2007.

- 50. **Rossman MJ, Venturelli M, McDaniel J, Amann M, and Richardson RS**. Muscle mass and peripheral fatigue: a potential role for afferent feedback? *Acta Physiologica* 206: 242-250, 2012.
- 51. **Russ DW, and Kent-Braun JA**. Sex differences in human skeletal muscle fatigue are eliminated under ischemic conditions. *J Appl Physiol* 94: 2414-2422, 2003.
- 52. **Sadamoto T, Bonde-Petersen F, and Suzuki Y**. Skeletal muscle tension, flow, pressure, and EMG during sustained isometric contractions in humans. *European Journal of Applied Physiology and Occupational Physiology* 51: 395-408, 1983.
- 53. Saey D, Michaud A, Couillard A, Côté CH, Mador MJ, LeBlanc P, Jobin J, and Maltais F. Contractile fatigue, muscle morphometry, and blood lactate in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 171: 1109-1115, 2005.
- 54. **Taylor JL, Petersen N, Butler JE, and Gandevia SC**. Ischaemia after exercise does not reduce responses of human motoneurones to cortical or coriticospinal tract stimulation. *J Physiol* 525: 793-801, 2000.
- 55. **Vanhatalo A, Fulford J, DiMenna FJ, and Jones AM**. Influence of hyperoxia on muscle metabolic responses and the power-duration work relationship during severe-intensity exercise in humans: a ³¹P magnetic resonance spectroscopy study. *Exp Physiol* 95: 528-540, 2010.
- 56. **Vanhatalo A, Jones AM, and Burnley M**. Application of critical power in sport. *International Journal of Sports Physiology and Performance* 6: 128-136, 2011.
- 57. **Walloe L, and Wesche J**. Time course and magnitude of blood flow changes in the human quadriceps muscles during and following rhythmic exercise. *J Physiol* 405: 257-273, 1988.
- 58. **Whipp BJ, Huntsman DJ, Stoner N, Lamarra N, and Wasserman K**. A constant which determines the duration of tolerance to high intensity work. *Federation Proceedings* 41: 1591, 1982.
- 59. **Whipp BJ, Ward SA, and Hassall MWC**. Estimating the metabolic rate of marching Roman Legionaries. *J Physiol* 491P: 60, 1996.
- 60. **Whipp BJ, Ward SA, and Hassall MWC**. Paleo-bioenergetics: the metabolic rate of marching Roman legionaries. *British Journal of Sports Medicine* 32: 261-264, 1998.

Chapter 2 - Influence of duty cycle on the power-duration relationship for handgrip exercise: observations and potential mechanisms

Summary

The highest sustainable rate of aerobic metabolism [critical power (CP)] and the finite amount of work that can be performed above CP (W') were determined under two muscle contraction duty cycles. Eight men completed at least three constant-power handgrip tests to exhaustion to determine CP and W' for 50% and 20% duty cycles, while brachial artery blood flow (\dot{Q}_{BA}) and deoxygenated-[hemoglobin+myoglobin] (deoxy-[Hb+Mb]) were measured. CP was lower for the 50% duty cycle (3.9 \pm 0.9 W) than the 20% duty cycle (5.1 \pm 0.8 W; p < 0.001), while W' was not significantly different (50% duty cycle: 452 \pm 141 J vs. 20% duty cycle: 432 \pm 130 J; p > 0.05). At the same power output, \dot{Q}_{BA} and deoxy-[Hb+Mb] achieved higher end-exercise values for the 20% duty cycle (9.87 \pm 1.73 ml·s⁻¹; 51.7 \pm 4.7 μ M) than the 50% duty cycle (7.37 \pm 1.76 ml·s⁻¹, p < 0.001; 44.3 \pm 2.4 μ M, p < 0.03). These findings indicate that blood flow influences CP, but not W'.

Introduction

The notion of an increase in exercise duration with progressively decreasing power outputs dates back at least to the early 20th century (34, 35) and potentially as early as the 4th century (81, 82). This robust power-duration relationship is now commonly characterized using a hyperbolic mathematical model to obtain the asymptote (critical power, CP) and the curvature constant (W') (36, 43, 80). CP demarcates the boundary between the heavy- and severe-exercise intensity domains, as it is the highest intensity in which a physiological steady-state can be achieved [i.e., for oxygen uptake (\dot{V}_{O_2}) (60), blood flow (inferred from (14)), intramuscular concentrations of phosphocreatine [PCr], inorganic phosphate [Pi], and hydrogen ion [H⁺] (45)]. W' represents a finite work capacity that can be performed above CP and has traditionally been associated with 'anaerobic' metabolism (13, 21, 26, 50, 51, 53). This interpretation is supported by [Pi], [H⁺], and [PCr] consistently achieving 'critical levels' upon exhaustion (12, 45, 60, 75). Alternatively, W' may be determined by the magnitude of the severedomain (i.e., the range between CP and $\dot{V}_{\rm O_{2max}}$) (8, 75) and has been associated with the $\dot{V}_{\rm O_{2}}$ slow component (21, 44, 55, 76). This interpretation is supported by several studies that demonstrated a decrease in W' with interventions that increased CP (40, 74, 75). It has been speculated that these decreases in W' were a result of the interventions increasing CP disproportionately to $\dot{V}_{\rm O_{2max}}$ and therefore decreasing the magnitude of the severe-domain (8, 75). Although the mechanism(s) determining W' are not fully understood, it is clear that exercise tolerance for any activity performed at an intensity above CP is limited by the magnitude of W' with exhaustion ensuing upon complete utilization of W' if the power output is not lowered to an intensity equal to or below CP.

Monod and Scherrer (1965), in originally characterizing the power-duration relationship, suggested that CP is dependent upon the circulatory conditions in the muscle, while W' is determined by intramuscular 'anaerobic' (with the exception of O_2 stores) mechanisms. Subsequent experiments have

revealed that CP is dependent upon the rate of aerobic ATP production (i.e., O₂ delivery and O₂ utilization) (18, 36, 43, 54, 75), while W' (at least in part) is dependent upon 'anaerobic' ATP production (33, 41, 50, 51, 72). Thus, any intervention altering O₂ delivery (i.e., reduced blood flow) to the active skeletal muscle would be expected to alter CP, with presumably no (or little) affect on W'.

The increased intramuscular pressure accompanying muscle contraction can exhibit a profound influence on blood flow as a result of blood vessel compression, increased impedance to blood flow, and possible occlusion of blood flow (38, 47, 64, 68). The muscle contraction-relaxation cycle yields rhythmic alterations in intramuscular pressure, and therefore blood flow, with the majority of blood flow occurring during the relaxation period when intramuscular pressure is low (4, 25, 64, 79). Robergs et al. (1997) suggested that the relaxation period blood flow may be important in determining the attainment of a steady-state metabolic rate. The muscle contraction duty cycle (time under tension/total contraction time) directly impacts blood flow, such that with high duty cycles (high time under tension relative to total contraction time) blood flow to the active skeletal muscle becomes limited (6, 7), while blood flow is not compromised at low duty cycles (low time under tension relative to total contraction time) even with increased contraction frequencies (23, 56, 71). Collectively, these results demonstrate that the muscle contraction duty cycle directly influences blood flow to the active skeletal muscle.

We are aware of no study to date that has examined the influence of alterations in blood flow due to the muscle contraction-relaxation cycle on the parameters of the power-duration relationship.

Therefore, the aim of the current study was to manipulate blood flow using muscle contraction duty cycles in order to assess the dependence of CP and W' on blood flow. We hypothesized that 1) CP would be higher for the 20% duty cycle compared to the 50% duty cycle, while W' would remain unchanged. Further, when the same power output was repeated at both duty cycles, 2) blood flow would

be higher for the 20% duty cycle than the 50% duty cycle, but 3) deoxy-[Hb+Mb] and EMG measurements would achieve similar end-exercise values for both duty cycles.

Methods

Subjects

Eight healthy men (age: 24.8 ± 2.5 yrs, height: 173.7 ± 4.6 cm; weight: 77.1 ± 14.6 kg) volunteered to participate in this study. Subjects reported to the Human Exercise Physiology Laboratory with at least 24 h between testing sessions and having abstained from vigorous activity within that 24 h period. All experimental procedures in the present study were approved by the Institutional Review Board of Kansas State University and conformed to the standards set forth by the Declaration of Helsinki. Prior to testing, each subject was informed of the overall protocol along with the potential risks involved. Each subject then provided written informed consent and completed a health history evaluation.

Experimental Protocol

All testing was performed on a custom-built handgrip ergometer. The handrail of the ergometer was attached to a pneumatic cylinder by means of a cable-pulley system and provided a fixed linear displacement of 4 cm. Resistance was set by pressurizing the pneumatic cylinder and work was accomplished by compressing the air within the cylinder when the handrail was moved. Power output was calculated as P = Rdf·k⁻¹, where P is power in Watts (W), R is resistance in kg, d is displacement in meters, f is contraction frequency, and k is the constant 6.12 for the conversion of kg·m·min⁻¹ to W. When seated at the ergometer the subject grasped the handrail so that the forearms were at approximately heart level and the elbows were slightly bent. A contraction frequency of 20 contractions·min⁻¹ was utilized for both duty cycles so that each total contraction cycle duration was maintained at 3.0 s. Thus any set resistance would produce the same power output for both duty cycles. The 50% duty cycle consisted of a 1.5 s contraction period (in which the handrail was raised with concentric muscle contraction and lowered with eccentric muscle contraction) followed by a 1.5 s

relaxation period. The 20% duty cycle consisted of a 0.6 s contraction period (in which the handrail was raised with concentric muscle contraction and then immediately released) followed by a 2.4 s relaxation period (Figure 2.1). Both duty cycles had the same duration of concentric contraction and total contraction cycle, while the 20% duty cycle had no eccentric contraction period and therefore a longer duration of time without muscle tension. The eccentric contraction period duration was altered specifically to minimize any metabolic differences between duty cycles, while emphasizing blood flow differences (see Discussion). Audio recordings set with the specific timing for each duty cycle were used along with feedback provided by an investigator monitoring the tests to ensure correct timing. Subjects completed three familiarization trials per duty cycle prior to data collection to aid in correct, consistent production of the contraction-relaxation timing. All testing sessions were continued until exhaustion, determined as the inability to complete three consecutive contraction cycles.

A peak incremental test for each duty cycle was completed in a randomized order during the initial two testing sessions. These tests were initiated at 1.0 W and the power output was increased by $0.5~\mathrm{W\cdot min^{-1}}$ until exhaustion. The peak power (P_{peak}) was recorded as the highest power output for which at least 30 s of the stage was completed. The P_{peak} was utilized to determine the power outputs for the subsequent constant-power testing sessions that would elicit exhaustion between 2-15 min. Subjects completed a minimum of three randomly ordered constant-power tests per duty cycle in which the time-to-exhaustion (T_{lim}) was recorded. After the initial three constant-power tests, the data were fit with the two-parameter hyperbolic model and the goodness-of-fit was analyzed. If the goodness-of-fit data did not meet the a priori criteria (see Data Analysis) a fourth testing session was conducted in an attempt to lower the parameter standard error values. A power output that elicited exhaustion between 2-5 min for the 50% duty cycle was repeated for the 20% duty cycle, such that any differences in the physiologic

responses between duty cycles could be examined without the confounding influence of different power outputs (i.e., metabolic rates).

Measurements

Doppler ultrasound

The raw blood velocity profiles were measured using Doppler ultrasound (Vivid 3, GE Medical Systems, Milwaukee, WI, USA) operating in pulse wave mode at a Doppler frequency of 4.0 MHz with a phased linear array transducer probe operating at an imaging frequency of 6.7 MHz, and were stored for post-hoc analysis. For all testing sessions the Doppler gate was set to the full width of the brachial artery to ensure complete insonation and all Doppler velocity measurements were corrected for the angle of insonation, which was adjusted to be less than 60 degrees. Measurements were made 2-5 cm above the antecubital fossa to avoid the bifurcation of the brachial artery. A bifurcation was not seen in the two-dimensional image, suggesting that all Doppler measurements were made greater than 1 cm from the bifurcation, as previously utilized in our laboratory (2). Brachial artery diameters were measured in the transverse axis using two-dimensional sonography.

Near-infrared spectroscopy

The oxygenation characteristics of the flexor digitorum superficialis were determined using a frequency-domain multi-distance NIRS system (Oxiplex TS, ISS, Champaign, IL, USA). The principles and algorithms of the NIRS technology were reviewed by Gratton (29) and have previously been described by Ferreira et al. (23). Briefly, this device consists of eight light-emitting diodes (LED) operating at wavelengths of 690 and 830 nm (four LEDs per wavelength) with one detector fiber bundle and LED-detector separation distances of 2.0, 2.5, 3.0, and 3.5 cm. The NIRS data were collected at 50

Hz and stored for post-hoc analysis. After locating the flexor digitorum superficialis of the right arm using EMG and palpation, the NIRS probe was secured longitudinally along the belly of the muscle. The position of the probe was then marked with indelible ink for reproducible placement throughout the study. The NIRS probe was calibrated prior to each test according to the manufacturer's recommendations using a calibration block with known absorption and scattering coefficients. Calibration was confirmed on a separate block with different absorption and scattering coefficients.

Electromyography

Surface electromyography (EMG) measurements were obtained from the flexor digitorum superficialis in the left forearm. The single differential EMG electrode (Trigno EMG, Delsys Inc., Boston, MA, USA) consists of four silver contact bars (5 x 1 mm) arranged in a 2 x 2 orientation. The electrode was positioned over the belly of the muscle, as determined by palpation and strong electrical activity when the fingers were flexed, but not with ulnar or radial deviation. This site was then marked with indelible ink for reproducible placement of the electrode throughout the study. The EMG data were sampled at 1000 Hz and stored for post-hoc analysis.

Data Analysis

Determination of the power-duration relationship

The parameters of the power-duration relationship (CP and W') were determined with the two-parameter hyperbolic model t = W' / (P - CP), where t is time in s, W' is the finite work capacity in Joules (J), P is power in W, and CP is critical power in W. The data from the initial three constant-power tests were fit with the hyperbolic model and the goodness-of-fit was assessed. A fourth constant-power test was conducted if the parameter standard error (SE) was greater than 10% of the parameter

value for either CP or W'. When applied to whole-body exercise (i.e., cycling) this 'acceptable' margin of error allows for the anaerobic work capacity to be accurately estimated by W' (36). There has been no established margin of error for small muscle mass exercise (i.e., handgrip) and therefore we chose to use the 10% cutoff.

Doppler Ultrasound

Mean blood velocity (\dot{V}_{mean} ; cm·s⁻¹) was defined as the time-averaged mean velocity over each 3 s contraction cycle. Brachial artery blood flow (\dot{Q}_{BA}) was calculated using the product of \dot{V}_{mean} and vessel cross-sectional area (CSA). Brachial artery diameters were measured every minute throughout each test and were used to calculate vessel CSA in cm² (CSA = π r²). The \dot{Q}_{BA} data were analyzed using one contraction cycle (i.e., 3 s) at the time points 0 s, 46.5 s, 91.5 s, while three consecutive contraction cycles (i.e., 9 s) were utilized for the end of each subsequent minute, at the equivalent time within the 20% duty cycle to the time of end-exercise for the 50% duty cycle (matched-time), and at end-exercise. \dot{Q}_{BA} data were also measured 9 s post-exercise using a 3 s average.

NIRS

The NIRS data were processed using 3 s averages throughout each testing session. During the first minute of exercise and at 91.5 s the NIRS data were analyzed for each contraction cycle, while 3 consecutive contraction cycles were used for each subsequent minute, at 50% matched-time, and at end-exercise. At 9 s post-exercise the NIRS data were analyzed using a 3 s average. The deoxy-[hemoglobin+myoglobin] (deoxy-[Hb+Mb]) is relatively insensitive to changes in blood-volume (16, 22, 28) and has been used to reliably estimate the fractional oxygen extraction (17, 19, 22, 23, 28). The device used in the present study provides absolute concentrations (μM) for deoxy-[Hb+Mb] and

oxygenated-[hemoglobin+myoglobin] (oxy-[Hb+Mb]), which may be combined to provide total-[hemoglobin+myoglobin] (total-[Hb+Mb]). The dynamic reduced scattering coefficients were measured throughout the tests and were incorporated in all of the NIRS data calculations.

EMG

The raw EMG data were processed with a band-pass filter (30-300 Hz) and each electrical burst corresponding to a muscle contraction was detected using a custom-designed computer program. The EMG signal amplitude characteristics were analyzed via integrated EMG (iEMG), a measure of motor unit recruitment and motorneuron firing rate, which typically increases as the muscle fatigues (20, 27). The EMG signal frequency characteristics were analyzed via mean power frequency (MPF), a measure of the muscle action potential conduction velocity, which typically shifts to lower frequencies as the muscle fatigues (31). During the first minute of exercise and at 91.5 s the EMG data were analyzed for each contraction cycle, after which the end of each subsequent minute, 50% matched-time, and end-exercise were analyzed using three consecutive contraction cycles.

Estimation of oxygen consumption

To investigate the relationship between O_2 delivery and O_2 extraction across duty cycles we used the model put forth by Wagner and colleagues (65, 77, 78) which integrates perfusive O_2 delivery [Fick Principle, $\dot{V}_{O_2} = \dot{Q}$ (arterial-venous O_2 content difference), where \dot{Q} is blood flow] and diffusive O_2 capacity [Fick's Law of Diffusion, $\dot{V}_{O_2} = \dot{D}_{O_2}(P_{cap}O_2 - P_{mit}O_2)$, where \dot{D}_{O_2} is the oxygen diffusing capacity of the muscle, $P_{cap}O_2$ is the partial pressure of oxygen within the microcirculation, and $P_{mit}O_2$ is the partial pressure of oxygen within the mitochondria]. The intersection of these two relationships yields the $\dot{V}_{O_{2peak}}$ for those conditions. The mechanisms for the discrepancy in CP between duty cycles

in the current study can be further explored using this model under a few assumptions. The assumptions were held constant between duty cycles to reduce systematic error, so that any differences in the model would be attributable to differences in the deoxy-[Hb+Mb] and QBA values. It was assumed that the deoxy-[Hb+Mb] signal reflects only deoxy-[Hb] [n.b., we are aware that the signal contains deoxy-[Mb] as well (15)] and that the entire signal arises solely from the muscle (i.e., not from any intervening adipose or skin tissue). With these assumptions the deoxy-[Hb] may be converted into an estimated \dot{V}_{O_2} . The deoxy-[Hb] values are in units of µmole heme/l tissue, where the tissue is assumed to be muscle. These deoxy-[Hb] units can be converted into μmole heme/ℓ blood using the conversion 1.36% capillary blood volume/muscle volume [derived from 400 cap/mm², 28.3 µm² CSA, and a coefficient of 1.2 correcting for tortuosity and branching of the capillaries (63)]. These units can then be converted into mole O_2/ℓ blood assuming 1 mole O_2 /mole heme and further to ℓ O_2/ℓ blood using the conversion 22.4 ℓ O₂/mole O₂. \dot{V}_{O_2} values in ℓ O₂/min may then be obtained by multiplying this value by the measured \dot{Q}_{BA} values. The \dot{Q}_{BA} and deoxy-[Hb+Mb] responses for each duty cycle (Figures 3 and 4) were fit with exponential models which were then integrated to provide the \dot{V}_{O_2} response throughout each duty cycle.

Statistical analysis

 P_{peak} , CP, and W' were compared across duty cycles using paired t-tests. Main effects for \dot{Q}_{BA} , deoxy-[Hb+Mb], total-[Hb+Mb], \dot{V}_{O_2} , iEMG, and MPF were tested using two-way ANOVA with repeated measures (duty cycle x time) for the same power output constant-power tests for each duty cycle. When a significant main effect was detected, a Tukey's post-hoc analysis was conducted.

Differences were considered statistically significant when p < 0.05 and all data are presented as mean \pm SD unless otherwise noted.

Results

Power-duration relationship

As determined from the a priori goodness-of-fit criteria, the power-duration relationships were determined using four constant-power tests in all of the subjects for the 50% duty cycle and in four of the subjects for the 20% duty cycle. The resulting SE values as a percent of the parameter value were $4.6 \pm 6.1\%$ for CP and $12.7 \pm 8.7\%$ for W' with the 50% duty cycle and $1.6 \pm 1.4\%$ for CP and $10.8 \pm 11.7\%$ for W% with the 20% duty cycle. The coefficient of determination values for the 50% and 20% duty cycles were 0.98 ± 0.02 and 0.98 ± 0.02 , respectively. CP was significantly lower for the 50% duty cycle $(3.9 \pm 0.9 \text{ W})$ than the 20% duty cycle $(5.1 \pm 0.8 \text{ W}; p < 0.001)$, while W' was not significantly different (50% duty cycle: $452 \pm 141 \text{ J}$ and 20% duty cycle: $432 \pm 130 \text{ J}$; coefficient of variation = 13.9%) (Figure 2.2). There was a significant inverse correlation between the percent change in CP versus the percent change in W' between the 50% and 20% duty cycles (r = -0.83, p = 0.01), but not for the absolute changes in CP versus W' (r = -0.61, p = 0.11).

Equivalent power output tests

The P_{peak} from the incremental test was significantly lower for the 50% duty cycle $(5.7 \pm 0.7 \text{ W})$ compared to the 20% duty cycle $(6.7 \pm 0.8 \text{ W}; p < 0.001)$. The mean power output that was repeated for both duty cycles was $6.2 \pm 0.8 \text{ W}$, which equated to a significantly higher relative power output for the 50% duty cycle $(109 \pm 8.4 \text{ %P}_{peak})$ compared to the 20% duty cycle $(93.7 \pm 7.5 \text{ %P}_{peak}; p < 0.001)$. This power output was also significantly higher as a percentage of CP for the 50% duty cycle $(165 \pm 37.3 \text{ %})$ than the 20% duty cycle $(125 \pm 14.4\%; p = 0.003)$. The T_{lim} for the 50% duty cycle $(201 \pm 52.1 \text{ s})$ was significantly shorter than the 20% duty cycle $(501 \pm 314 \text{ s}; p = 0.017)$.

\dot{Q}_{BA}

The brachial artery diameter and \dot{V}_{mean} data are presented in Table 2.1. \dot{Q}_{BA} increased significantly between 91.5 s to end-exercise for the 20% duty cycle, while \dot{Q}_{BA} did not increase for the 50% duty cycle, such that at matched-time (50% duty cycle: $7.37 \pm 1.76 \, \text{ml} \cdot \text{s}^{-1}$; 20% duty cycle: $9.26 \pm 1.99 \, \text{ml} \cdot \text{s}^{-1}$; p = 0.001) and end-exercise (50% duty cycle: $7.37 \pm 1.76 \, \text{ml} \cdot \text{s}^{-1}$; 20% duty cycle: $9.87 \pm 1.73 \, \text{ml} \cdot \text{s}^{-1}$; p < 0.001) the 50% duty cycle \dot{Q}_{BA} was significantly lower than the 20% duty cycle (Figure 2.3A). At 9 s post-exercise \dot{Q}_{BA} was not significantly different from end-exercise within the 20% duty cycle, but had significantly increased above end-exercise within the 50% duty cycle (p = 0.008), such that \dot{Q}_{BA} was no longer significantly different between duty cycles (20% duty cycle: $11.3 \pm 2.8 \, \text{ml} \cdot \text{s}^{-1}$; 50% duty cycle: $10.6 \pm 3.4 \, \text{ml} \cdot \text{s}^{-1}$) (Figure 2.3B).

NIRS

The deoxy-[Hb+Mb] increased to a significantly higher value at end-exercise for the 20% duty cycle (51.7 \pm 4.7 μ M) compared to the 50% duty cycle (44.3 \pm 2.4 μ M; p = 0.03) (Figure 2.4A). At 9 s post-exercise there was no significant difference (albeit marginally) between duty cycles for the deoxy-[Hb+Mb] (20% duty cycle: 52.3. \pm 16.7 μ M; 50% duty cycle: 45.8 \pm 9.19 μ M; p = 0.054) (Figure 2.4B). Throughout exercise the total-[Hb+Mb] increased within each duty cycle and no significant difference was detected between duty cycles.

EMG

Within the 20% duty cycle, iEMG did not change significantly throughout the test (Figure 2.5A). During the 50% duty cycle, the iEMG progressively increased until end-exercise, resulting in significant differences between duty cycles at 120 s (p = 0.017), matched-time (p < 0.001), and end-exercise (p = 0.002). The MPF did not change significantly during the 20% duty cycle test, while it continually

decreased throughout the 50% duty cycle test (Figure 2.5B). This resulted in a significantly higher MPF for the 20% duty cycle at matched-time (p = 0.005) and end-exercise (p = 0.008).

 \dot{V}_{O_2}

The integration of the \dot{Q}_{BA} and deoxy-[Hb+Mb] values estimated \dot{V}_{O_2} data that qualitatively increased with similar time courses and amplitudes for both duty cycles until approximately 90 s (as seen for \dot{Q}_{BA} , Figure 2.3 and deoxy-[Hb+Mb], Figure 2.4), after which the 20% duty cycle \dot{V}_{O_2} increased beyond that of the 50% duty cycle (Figure 2.6). \dot{V}_{O_2} significantly increased between 91.5 s to end-exercise for the 20% duty cycle, while \dot{V}_{O_2} did not increase for the 50% duty cycle, such that at matched-time (50% duty cycle: $32 \pm 9 \text{ ml } O_2 \cdot \text{min}^{-1}$; 20% duty cycle: $43 \pm 11 \text{ ml } O_2 \cdot \text{min}^{-1}$; p = 0.003) and end-exercise (50% duty cycle: $32 \pm 9 \text{ ml } O_2 \cdot \text{min}^{-1}$; 20% duty cycle: $50 \pm 11 \text{ ml } O_2 \cdot \text{min}^{-1}$; p, 0.001) the 50% duty cycle \dot{V}_{O_2} was lower than the 20% duty cycle (Figure 2.7). The model analysis resulted in \dot{D}_{O_2} values of 1.02 ml⁻¹·min⁻¹·mmHg⁻¹ for the 50% duty cycle and 1.72 ml⁻¹·min⁻¹·mmHg⁻¹ for the 20% duty cycle. At 9 s post-exercise the \dot{V}_{O_2} values were not significantly different between duty cycles (50% duty cycle $50 \pm 13 \text{ ml } O_2 \cdot \text{min}^{-1}$ and 20% duty cycle $53 \pm 17 \text{ ml } O_2 \cdot \text{min}^{-1}$; p = 0.612).

Discussion

Consistent with our first hypothesis, CP was higher for the 20% duty cycle compared to the 50% duty cycle, while W' was not different between duty cycles. When the same power output was completed for both duty cycles, \dot{Q}_{BA} was higher for the 20% duty cycle compared to the 50% duty cycle, consistent with our second hypothesis. In contrast to our third hypothesis, however, deoxy-[Hb+Mb] achieved higher values for the 20% duty cycle, while the iEMG was lower and the MPF higher for the 20% duty cycle compared to the 50% duty cycle.

In characterizing the power-duration relationship, Monod and Scherrer (1965) considered CP to be dependent upon muscle blood flow (i.e., O_2 delivery). Since this seminal publication, CP has been demonstrated to be dependent upon O_2 delivery (blood flow x arterial O_2 content) by manipulating inspired O_2 concentrations to reveal that CP is lowered with hypoxia (18, 54) and elevated with hyperoxia (75). The current study has extended these findings by manipulating O_2 delivery via altered blood flow with the use of two different duty cycles for muscle contraction, demonstrating that CP is lower for the 50% duty cycle compared to the 20% duty cycle as a result of reduced blood flow. In addition, O_2 extraction was altered with duty cycle as the 50% duty cycle deoxy-[Hb+Mb] was lower compared to the 20% duty cycle. These differences in blood flow and deoxy-[Hb+Mb] measured between the duty cycles performed at the same power output, would be anticipated for the other exercise tests as well. Consistent with this, CP was lower for the 50% duty cycle compared to the 20% duty cycle. These findings support that CP reflects the highest rate of O_2 utilization which is matched by O_2 delivery, that the muscle contraction duty cycle directly influences CP.

The deterministic mechanisms of W' have traditionally been associated with intramuscular 'anaerobic' energy production [depletion of the intramuscular energy stores (45, 50, 51, 53) and/or metabolite accumulation (13, 21, 26, 45)]. In the current study, muscle contraction duty cycle-induced alterations in O_2 delivery (\dot{Q}_{BA}) and O_2 extraction (deoxy-[Hb+Mb]) did not alter W'. The findings of

the current study are consistent with hypoxia leading to a decrease in CP, while not altering W' (18, 54). In contrast, hyperoxia led to an increase in CP and, interestingly, a decrease in W' (75). A more recent definition of W' as a work capacity that is determined by the magnitude of the severe-intensity domain has emerged in the literature (8, 75). This definition postulates that the parameters determining the severe-intensity domain ($\dot{V}_{O_{2max}}$ and CP) dictate W'.

It is worth noting that this definition implies $\dot{V}_{\rm O_{2max}}$ is involved in determining W', despite the power-duration relationship being determined by (and assumed to hold true for) work rates in the extreme-intensity domain where exhaustion ensues (and theoretically W' is completely utilized) prior to the attainment of $\dot{V}_{\mathrm{O}_{2\mathrm{max}}}$. The magnitude of the severe-intensity domain was not determined in the current study, as the $\dot{V}_{\rm O_2}$ at CP was not measured. However, implications may be drawn from examining the power outputs associated with the boundaries of the severe-intensity domain. P_{peak} and CP were both approximately 1.0 W lower for the 50% duty cycle compared to the 20% duty cycle, which implies that the magnitude of the severe domain, and therefore W', was unaltered. This is consistent with hypoxia resulting in a concomitant 29 W reduction in the power output at $\dot{V}_{\rm O_{2peak}}$ with a 30 W reduction in CP and no change in W' (Dekerle et al., 2012). An unaltered magnitude of W' with a lower CP would necessitate a faster rate of W' utilization at the same power output and a decrease in time-to-exhaustion (as seen in the present study), as 'critical levels' (60) of the 'anaerobic' substances associated with fatigue would be achieved earlier in the exercise bout. Collectively, these findings support that the magnitude of W' is not dependent on O_2 delivery or O_2 extraction per se, rather these determine the rate at which W' is utilized, via alterations in $\dot{V}_{\rm O_{2max}}$ and CP.

The $\dot{V}_{\rm O_{2peak}}$ values from the current study are similar to previously reported directly measured values for handgrip exercise of ~30-50 ml O_2 ·min⁻¹ (62, 73). From the model analysis, it was found that the change in $\dot{D}_{\rm O_2}$ (69 %) was double the change in $\dot{Q}_{\rm BA}$ (34 %) between duty cycles. The $\dot{D}_{\rm O_2}/\dot{Q}$ ratio is

indicative of O_2 extraction (59, 65), and therefore the increased D_{O_2}/\dot{Q} ratio for the 20% duty cycle in the current study would explain the increased deoxy-[Hb+Mb] compared to the 50% duty cycle. A possible mechanism for the increased diffusive O_2 capacity $[\dot{D}_{O_2} = A/T \times (solubility/\sqrt{molecular weight}), where <math>A$ is the surface area for diffusion and T is the thickness of the membrane across which diffusion occurs] for the 20% duty cycle may be due to enhanced longitudinal recruitment of capillary surface area (59). Increased red blood cell velocity and fractional O_2 extraction increase the length of a capillary involved in O_2 exchange ('longitudinal recruitment') (Poole et al. 2011), which may be a mechanism for increasing capillary surface area, and thus $\dot{D}_{\rm O_2}$, during exercise. In the current study, brachial artery blood velocity was significantly faster and brachial artery diameter was significantly larger for the 20% duty cycle compared to the 50% duty cycle (Table 2.1). Assuming that duty cycle did not alter the microvascular volume, the increased brachial artery blood flow (\dot{V}_{mean} x CSA) for the 20% duty cycle evinces an increased red blood cell velocity in the capillaries, leading to increased longitudinal recruitment. The increased O_2 extraction despite the increased blood flow for the 20% duty cycle suggests that red blood cell transit time was not limiting [consistent with Richardson et al. (1993b)], but rather served to augment O_2 extraction. The surface area for gas exchange in the capillary is also determined by capillary hematocrit, which is approximately 33% of systemic values at rest and increases to systemic values at maximal exercise (46). However, total-[Hb+Mb] was not significantly different between duty cycles, suggesting that microvascular hematocrit was similar between conditions. Thus, both the increased O_2 delivery and O_2 extraction for the 20% duty cycle likely contributed to the higher CP and estimated $\dot{V}_{\rm O_{2peak}}$ compared to the 50% duty cycle.

The underlying end-exercise metabolic state of the muscle was examined by analyzing the data immediately in recovery to remove the influence of the muscle contraction. The increase in \dot{Q}_{BA} with no physiological change in deoxy-[Hb+Mb] (despite slight statistical differences) during recovery for the

50% duty cycle above the end-exercise value suggests that $\dot{Q}_{\rm BA}$ was limited during exercise (possibly due to the decreased relaxation time), while the lack of increase during recovery for the 20% duty cycle suggests that this contraction style was not limiting. Importantly, both duty cycles were performed at the same power output and therefore similar metabolic rates would be expected. However, the $\dot{V}_{\rm O_2}$ model demonstrates that this was not the case, as the estimated $\dot{V}_{\rm O_2peak}$ was lower for the 50% duty cycle. The fact that the 50% duty cycle $\dot{V}_{\rm O_2}$ increased (as a result of the increase in $\dot{Q}_{\rm BA}$) in recovery to a value not different from the 20% duty cycle suggests that the same metabolic rate may have been 'wanted' by the muscles for both duty cycles, but the limitations imposed on O_2 delivery ($\dot{Q}_{\rm BA}$) and $\dot{Q}_{\rm C}$ extraction (deoxy-[Hb+Mb]) for the 50% duty cycle prevented this metabolic rate from being achieved. As a result, the aerobic energy contribution would be diminished for the 50% duty cycle (reflecting a decreased CP) while requiring a greater anaerobic energy contribution for any power output above CP. This would result in the faster utilization of W' and earlier occurrence of exhaustion, as seen in the present study.

Despite being performed at the same power output, the iEMG and MPF profiles differed between the 20% and 50% duty cycles. In contrast to the current study, Burnley et al. (9) reported that isometric knee-extension exercise at various intensities above CP yielded similar end-exercise EMG values. However, these findings may not directly relate to the current study due to differences in the mode of exercise (knee-extension *versus* handgrip), contraction style (isometric *versus* dynamic), and duty cycle (constant duty cycle *versus* altered duty cycle). The EMG profiles in the current study may have differed between duty cycles as a result of differences in the relative intensity and the duration of the tests, as these have been demonstrated to affect the EMG response (10, 57, 58). Amann (3) suggested that the accumulation of metabolites within the muscle may lead to increased firing rates of the group III/IV afferents, resulting in the inability to voluntarily produce the required force despite the muscle

being capable of generating it. In addition, exhaustion for supra-CP power outputs has been linked to the attainment of critical levels for intramuscular energy stores and metabolites ([PCr], $[H^+]$, and [Pi]) (75), which may directly impair force production (24). Therefore EMG differences between duty cycles in the current study may be a consequence of increased firing rates of the group III/IV afferents and/or the attainment of a critical level of intramuscular [PCr], $[H^+]$, and [Pi] at different levels of muscle activation. During handgrip exercise the *flexor digitorum superficialis* and *flexor digitorum profundus* are the primary muscles activated and used throughout the exercise test (52) and the fiber type composition of these muscles are ~50% Type I (39, 42). The EMG data from the current study suggest that the 50% duty cycle led to a greater recruitment of Type II fibers and/or induced more fatigue of these fibers than the 20% duty cycle. The limitation of O_2 delivery and O_2 extraction in the 50% duty cycle may have led to greater muscle fiber fatigue, thus requiring the recruitment of more muscle fibers to maintain the requisite power output. The current study demonstrated that at exhaustion the muscles were in different states of motor unit recruitment and action potential conduction velocities between the two duty cycles.

The findings of the present study have direct implications for past and future studies, as well as direct application for activities performed above CP. The different values for CP, \dot{Q}_{BA} , deoxy-[Hb+Mb], and EMG between duty cycles emphasize that comparisons among different protocols need to be made with regard to the specific contraction protocol so as to prevent misinterpretation. In application, altering the biomechanics of locomotion to provide a shorter duty cycle may lead to higher levels of sustainable performance. In running, decreasing the ground-contact time (i.e., duty cycle) may permit higher blood flow and O_2 extraction values, leading to improved performance. In a clinical setting, a shorter duty cycle in ambulation may allow higher intensities of 'exercise' to be maintained, such that activities of daily living become less fatiguing, increasing the patient's quality of life. For cycling, the

power output [but not the metabolic rate (5)] associated with CP has consistently been demonstrated to be lower with high pedal cadences (\geq 100 rpm) compared to low pedal cadences (\leq 60 rpm), while W' has not been affected when fitting the data with the hyperbolic model used in the current study (5, 11, 37, 49). The pedal frequencies used in these studies would vary the time under tension for the muscle, as the force generation by the muscle decreases with increasing pedal cadences (48, 70), and therefore would alter O_2 delivery by producing less blood flow impedance (30). The finding that W' was not altered by duty cycle in the current study is in line with the pedal cadence manipulation studies and cumulatively the data support that W' is independent of O_2 delivery. Importantly, the subjects of the studies examining the effect of pedal cadence on the power-duration relationship were non-cyclists. As trained cyclists tend to select higher pedal cadences ((30, 61, 69) than non-cyclists (30), it is not known if experienced cyclists would demonstrate similar decreases in CP with high pedal cadences.

Several experimental limitations are pertinent when interpreting the findings from the current study. In order to vary the duty cycle while maintaining the 3 s contraction cycle duration, the eccentric contraction component was omitted for the 20% duty cycle. The additional eccentric contraction of the 50% duty cycle may have contributed to metabolic differences between duty cycles. However, this does not seem likely as the estimated $\dot{V}_{\rm O_{2peak}}$ (along with peak $\dot{Q}_{\rm BA}$ and deoxy-[Hb+Mb]) was higher for the 20% duty cycle. In addition, eccentric contraction O_2 consumption is approximately 20% that of concentric contraction (1) and concentric contraction efficiency (measured as ATP/contraction) is ~15%, while eccentric efficiency is ~35% (67). Furthermore, the maintenance of tension by the muscle requires less energy than the development of tension (32, 66). Therefore, the major energy requiring component (concentric contraction) was the same between duty cycles, while the less energy demanding component (eccentric contraction) differed. Another limitation of the current study was that $\dot{V}_{\rm O_2}$ was not directly measured. Rather it was estimated using the deoxy-[Hb+Mb] and $\dot{Q}_{\rm BA}$ values. We recognize

that these assumptions may have limited the accuracy of the absolute values. Nevertheless, any contribution of these assumptions to the detected differences was minimized by holding the assumptions constant between duty cycles. The integration of the deoxy-[Hb+Mb] and \dot{Q}_{BA} responses allowed for the estimation of time course changes in \dot{V}_{O_2} for both duty cycles. However, due to the limited number of deoxy-[Hb+Mb] and \dot{Q}_{BA} data points for the exponential fits, no statistical kinetic analyses were conducted to prevent over interpretation of the data.

In conclusion, the current study reveals that a relatively long muscle contraction duty cycle imposes limitations on blood flow and O_2 extraction that ultimately leads to a decrease in exercise tolerance. CP was lower for the 50% duty cycle compared to the 20% duty cycle, while W' was not different. The 20% duty cycle resulted in elevated \dot{Q}_{BA} and deoxy-[Hb+Mb] compared to the 50% duty cycle and upon removal of the contraction impedance \dot{Q}_{BA} and deoxy-[Hb+Mb] increased for the 50% duty cycle to values not different from those of the 20% duty cycle. These findings suggest that O_2 delivery and O_2 extraction were limited for the 50% duty cycle, possibly due to a decreased relaxation time and less longitudinal recruitment of the capillary surface area, respectively. The EMG values also differed between duty cycles, such that at exhaustion the muscles were in different states of motor unit recruitment and action potential conduction velocities. The 50% duty cycle appears to have limited the physiological determinants of CP, resulting in a greater utilization of W' per contraction and less W' restoration between contractions at the same power output. This resulted in a decreased exercise tolerance compared to the 20% duty cycle. The findings of the current study support the notion that CP is determined by the highest sustainable rate of aerobic ATP production, which in turn is influenced by O_2 delivery and O_2 extraction. In contrast, the magnitude of W' appears to be determined (at least in part) by mechanisms that are independent of O_2 delivery and O_2 extraction, while the rate of W utilization is affected by O_2 delivery and O_2 extraction via alterations in CP.

Table 2.1 Brachial artery diameter and blood velocity data.

	50% duty cycle		20% duty cycle	
	Diameter (cm)	$\dot{V}_{mean}(cm \cdot s^{-1})$	Diameter (cm)	$\dot{V}_{mean}(cm \cdot s^{-1})$
Baseline	0.45 ± 0.04	11.4 ± 4.56	0.46 ± 0.03	12.1 ± 6.85
46.5 s	0.46 ± 0.04	35.6 ± 10.6^{b2}	0.46 ± 0.03	36.6 ± 6.56^{b2}
91.5 s	0.46 ± 0.03	41.6 ± 9.62^{b2}	0.47 ± 0.03	41.3 ± 7.62^{b2}
Matched-time	_	_	0.48 ± 0.03	$50.1 \pm 11.7^{a1,b2,c2,d1}$
End-exercise	$0.48 \pm 0.03^{b1,c1}$	43.0 ± 6.88^{b2}	$0.51 \pm 0.04^{a2,b2,c2,d2}$	$49.1 \pm 8.18^{a1,b2,c2,d1}$

 \dot{V}_{mean} , mean blood velocity; Matched-time, equivalent time-point within the 20% duty cycle test to that of end-exercise for the 50% duty cycle. ^a significantly different from 50% duty cycle. ^b significantly different from baseline within duty cycle. ^c significantly different from 46.5 s within duty cycle. ^d significantly different from 91.5 s within duty cycle. Level of significance: $^1p < 0.05$ and $^2p < 0.001$.

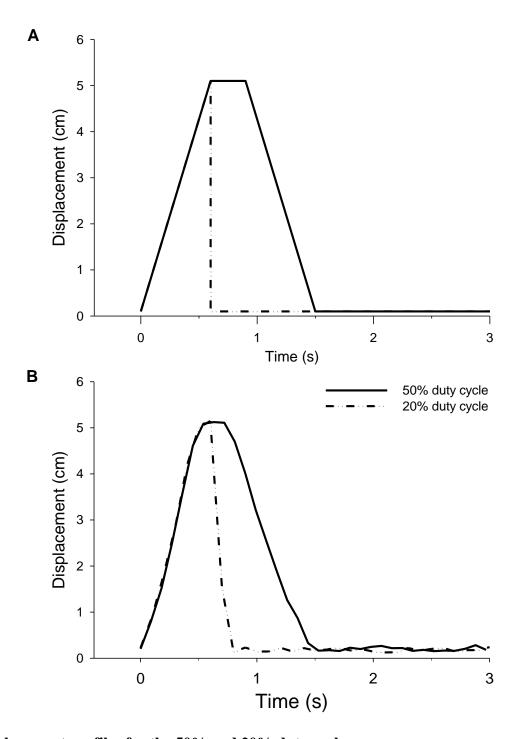


Figure 2.1 Displacement profiles for the 50% and 20% duty cycles.

Schematic representation of the specific contraction components for each duty cycle (Panel A). The 50% duty cycle consisted of a 0.6 s concentric contraction period, a 0.3 s isometric transition period, a 0.6 s eccentric contraction period, and a 1.5 s relaxation period. The 20% duty cycle consisted of a 0.6 s concentric contraction period and a 2.4 s relaxation period. A displacement profile for a representative subject throughout a contraction cycle for each duty cycle is shown in Panel B.

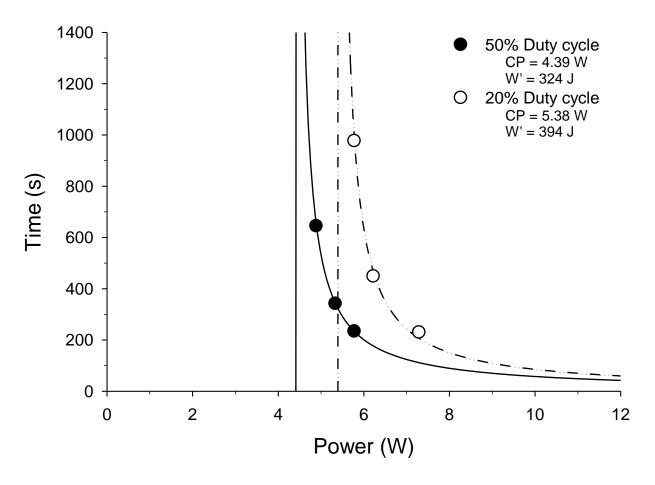


Figure 2.2 A representative subject's power-duration relationship for the two duty cycles.

Two-parameter hyperbolic fits to the 50% duty cycle (solid line) and the 20% duty cycle (dashed line) data are shown. The asymptote of each model represents critical power (CP) and the curvature constant represents W'.

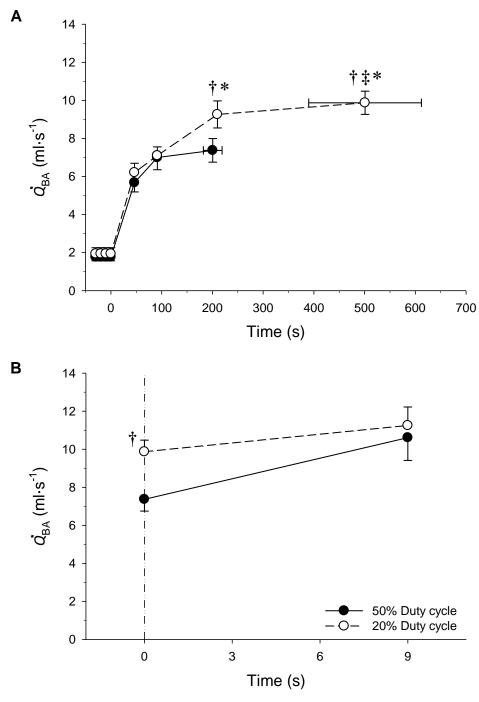


Figure 2.3 Brachial artery blood flow responses for the two duty cycles at the same absolute power output.

Mean and standard error exercise brachial artery blood flow (\dot{Q}_{BA}) data (Panel A), and end-exercise and 9 s post-exercise (Panel B) for the 50% and 20% duty cycles. † significantly different from the 50% duty cycle end-exercise, p < 0.001. ‡ significantly different from the 50% end-exercise time, p < 0.001. * significantly different from 20% duty cycle at 91.5 s, p < 0.001.

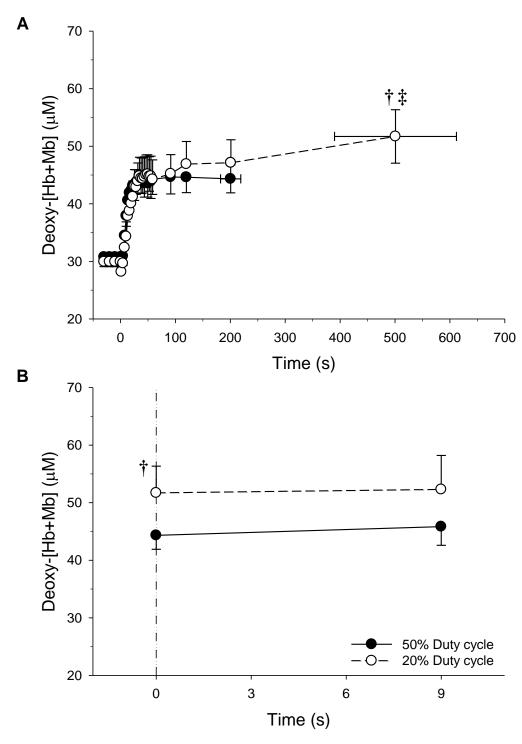
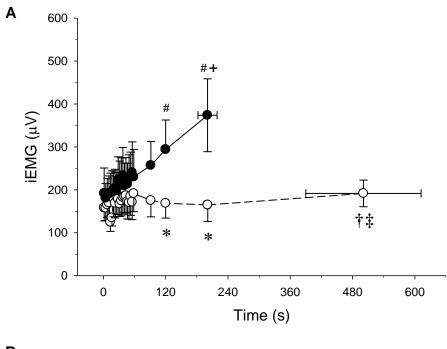


Figure 2.4 Deoxygenated-[hemoglobin + myoglobin] response for the two duty cycles at the same power output.

Mean and standard error deoxygenated-[hemoglobin+myoglobin] (deoxy-[Hb+Mb]) data (Panel A), and end-exercise and 9 s post-exercise (Panel B) for the 50% and 20% duty cycles. \dagger significantly different from 50% end-exercise, p < 0.05. \ddagger significantly different from the 50% duty cycle end-exercise time, p < 0.001.



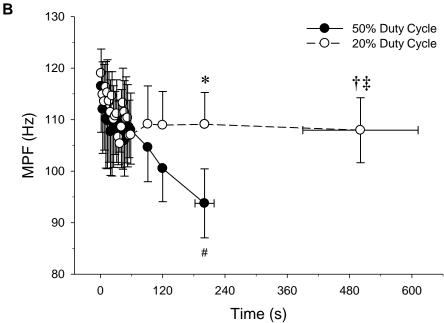


Figure 2.5 Mean electromyography response for the two duty cycles at the same power output.

Group mean and standard error integrated electromyography (iEMG) (Panel A) and mean power frequency (MPF) (Panel B). † significantly different from the 50% duty cycle end-exercise, p < 0.01. ‡ significantly different from 50% end-exercise time, p < 0.001. * significantly different from the 50% duty cycle at the same time points, p < 0.05. # significantly different from the 50% duty cycle at 60s, p < 0.05. + significantly different from the 50% duty cycle at 90s and 120 s, p < 0.05.

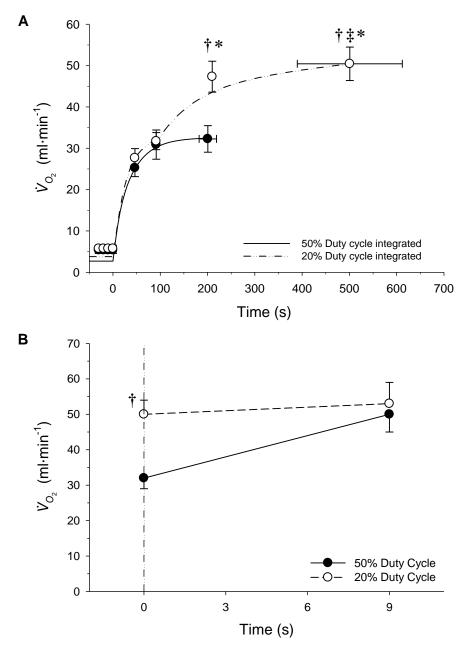


Figure 2.6 Estimated oxygen uptake response for the two duty cycles at the same power output.

Mean and standard error exercise estimated oxygen uptake (\dot{V}_{O_2} ; determined from brachial artery blood flow and deoxygenated-[hemoglobin+myoglobin]; see Methods for assumptions) (Panel A), and end-exercise and 9 s post-exercise (Panel B) for the 50% and 20% duty cycles. The integrated \dot{V}_{O_2} response was determined from the integration of exponential model fits to the brachial artery blood flow and deoxygenated-[hemoglobin/myoglobin] data. † significantly different from the 50% duty cycle end-exercise, p < 0.001. ‡ significantly different from the 50% end-exercise time, p < 0.001. * significantly different from 20% duty cycle at 91.5 s, p < 0.001.

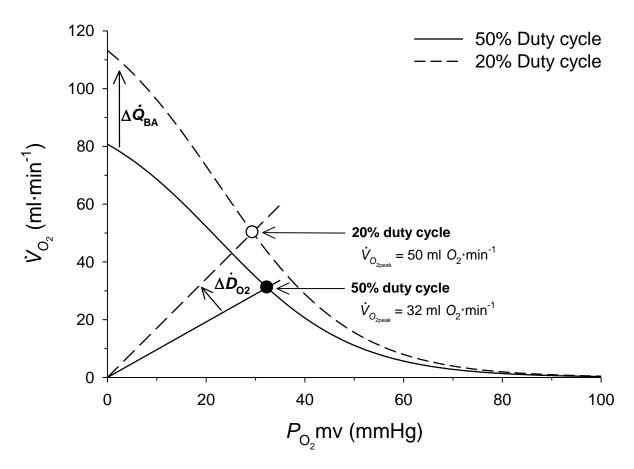


Figure 2.7 Diagram of estimated oxygen uptake as a function of microvascular $P_{\rm O_2}$ for each duty cycle.

The model integrates perfusive oxygen delivery (Fick Principle, curved lines) and diffusive oxygen delivery (Fick's Law, straight lines from origin) to yield $\dot{V}_{O_{2peak}}$ values for the specific conditions of each duty cycle. Exercise was carried out to the limit of tolerance (T_{lim}) at the same absolute power output (6.5 ± 0.9 W) for each duty cycle. \dot{V}_{O_2} was estimated using brachial artery blood flow (\dot{Q}_{BA}) and the deoxygenated-[hemoglobin+myoglobin] (deoxy-[Hb+Mb]) (See Discussion for details). The percent change in \dot{Q}_{BA} and the diffusive oxygen capacity (\dot{D}_{O_2}) from the 50% duty cycle to the 20% duty cycle was estimated to be 69 % and 34 %, respectively, which together predicted a 56 % higher $\dot{V}_{O_{2peak}}$ for the 20% duty cycle. The 20% duty cycle allowed for a higher \dot{Q}_{BA} (possibly due to the longer relaxation time) and a higher \dot{D}_{O_2} (possibly due to increased longitudinal capillary recruitment) compared to the 50% duty cycle.

References

- 1. **Abbott BC, Bigland B, and Ritchie JM**. The physiological cost of negative work. *J Physiol* 117: 380-390, 1952.
- 2. **Ade CJ, Broxterman RM, Wong BJ, and Barstow TJ**. Anterograde and retrograde blood velocity profiles in the intact human cardiovascular system. *Exp Physiol* 97: 849-860, 2012.
- 3. **Amann M**. Central and Peripheral Fatigue: Interaction during Cycling Exercise in Humans. *Med Sci Sports Exerc* 43: 2039-2045, 2011.
- 4. **Barcroft H, and Dornhorst AC**. The blood flow through the human calf during rhythmic exercise. *J Physiol* 109: 402-411, 1949.
- 5. **Barker T, Poole DC, Noble LM, and Barstow TJ**. Human critical power-oxygen uptake relationship at different pedalling frequencies. *Exp Physiol* 91: 621-632, 2006.
- 6. **Bellemare F, Wight D, Lavigne CM, and Grassino A**. Effect of tension and timing of contraction on the blood flow of the diaphragm. *J Appl Physiol* 54: 1597-1606, 1983.
- 7. **Buchler B, Magder S, and Roussos C**. Effects of contraction frequency and duty cycle on diaphragmatic blood flow. *J Appl Physiol* 58: 265-273, 1985.
- 8. **Burnley M, and Jones AM**. Oxygen uptake kinetics as a determinant of sports performance. *Eur J Sport Sci* 7: 63-79, 2007.
- 9. **Burnley M, Vanhatalo A, and Jones AM**. Distinct profiles of neuromuscular fatigue during muscle contractions below and above the critical torque in humans. *J Appl Physiol* 113: 215-223, 2012.
- 10. Camic CL, Housh TJ, Johnson GO, Hendrix CR, Zuniga JM, Mielke M, and Schmidt RJ. An EMG frequency-based test for estimating the neuromuscular fatigue threshold during cycle ergometry. *Eur J Appl Physiol* 108: 337-345, 2010.
- 11. **Carnevale TJ, and Gaesser GA**. Effects of pedaling speed on the power-duration relationship for high-intensity exercise. *Med Sci Sports Exerc* 23: 242-246, 1991.
- 12. **Chidnok W, Fulford J, Bailey SJ, DiMenna FJ, Skiba PF, Vanhatalo A, and Jones AM**. Muscle metabolic determinants of exercise tolerance following exhaustion: relationship to the "critical power". *J Appl Physiol* 115: 243-250, 2013.
- 13. Coats EM, Rossiter HB, Day JR, Miura A, Fukuba Y, and Whipp BJ. Intensity-dependent tolerance to exercise after attaining $\dot{V}O_{2max}$ in humans. *J Appl Physiol* 95: 483-490, 2003.

- 14. **Copp SW, Hirai DM, Musch TI, and Poole DC**. Critical speed in the rat: implications for hindlimb muscle blood flow distribution and fibre recruitment. *J Physiol* 588: 5077-5087, 2010.
- 15. **Davis ML, and Barstow TJ**. Estimated contribution of hemoglobin and myoglobin to near infrared spectroscopy. *Respiratory Physiology & Neurobiology* 186: 180-187, 2013.
- 16. **De Blasi RA, Cope M, Elwell C, Safoue F, and Ferrari M**. Noninvasive measurement of human forearm oxygen consumption by near infrared spectroscopy. *European Journal of Applied Physiology and Occupational Physiology* 67: 20-25, 1993.
- 17. **De Blasi RA, Ferrari M, Natali A, Conti G, Mega A, and Gasparetto A**. Noninvasive measurement of of forearm blood flow and oxygen consumption by near-infrared spectroscopy. *J Appl Physiol* 76: 1388-1393, 1994.
- 18. **Dekerle J, Mucci P, and Carter H**. Influence of moderate hypoxia on tolerance to high-intensity exercise. *Eur J Appl Physiol* 112: 327-335, 2012.
- 19. **DeLorey DS, Kowalchuk JM, and Paterson DH**. Relationship between pulmonary O₂ uptake kinetics and muscle deoxygenation during moderate intensity exercise. *J Appl Physiol* 95: 113-120, 2003.
- 20. **Enoka RM, and Stuart DG**. Neurobiology of muscle fatigue. *J Appl Physiol* 72: 1631-1648, 1992.
- 21. **Ferguson CS, Whipp BJ, Cathcart AJ, Rossiter HB, Turner AP, and Ward SA**. Effects of prior very-heavy intensity exercise on indices of aerobic function and high-intensity exercise tolerance. *J Appl Physiol* 103: 812-822, 2007.
- 22. **Ferrari M, Binzoni T, and Quaresima V**. Oxidative metabolism in muscle. *Philosophical Transactions of the Royal Society Biological Sciences* 352: 677-683, 1997.
- 23. **Ferreira LF, Lutjemeier BJ, Townsend DK, and Barstow TJ**. Effects of pedal frequency on estimated muscle microvascular O₂ extraction. *J Appl Physiol* 96: 558-563, 2006.
- 24. **Fitts RH**. The cross-bridge cycle and skeletal muscle fatigue. *J Appl Physiol* 104: 551-558, 2008.
- 25. **Folkow B, Gaskell P, and Waaler BA**. Blood flow through limb muscles during heavy rhythmic exercise. *Acto Physiologica* 80: 61-72, 1970.
- 26. **Fukuba Y, Miura A, Endo M, Kan A, Yanagawa K, and Whipp BJ**. The curvature constant parameter of the power-duration curve for varied-power exercise. *Med Sci Sports Exerc* 35: 1413-1418, 2003.

- 27. **Fulco CS, Lewis SF, Frykman PN, Boushel R, Smith S, Harman EA, Cymerman A, and Pandolf KB**. Muscle fatigue and exhaustion during dynamic leg exercise in normoxia and hypobaric hypoxia. *J Appl Physiol* 81: 1891-1900, 1996.
- 28. **Grassi B, Pogliaghi S, Rampichini S, Quaresima V, Ferrari M, Marconi C, and Cerretelli P.** Muscle oxygenation and pulmonary gas exchange kinetics during cycling exercise ontransitions in humans. *J Appl Physiol* 95: 149-158, 2003.
- 29. **Gratton E, Fantini S, Franceschini MA, Gratton G, and Fabiani M**. Measurements of scattering and absorption changes in muscle and brain. *Philosophical Transactions of the Royal Society Biological Sciences* 352: 727-735, 1997.
- 30. **Hagberg JM, Mullin JP, Giese MD, and Spitznagel E**. Effect of pedaling rate on submaximal exercise responses of competitive cyclists. *J Appl Physiol* 51: 447-451, 1981.
- 31. **Hagg GM**. Interpretation of EMG spectral alterations and alteration indexes at sustained contraction. *J Appl Physiol* 73: 1211-1217, 1992.
- 32. **Hamann JJ, Kluess HA, Buckwalter JB, and Clifford PS**. Blood flow response to muscle contractions is more closely related to metabolic rate than contractile work. *J Appl Physiol* 98: 2096-2100, 2005.
- 33. Heubert RAP, Billat VL, Chassaing P, Bocquet V, Morton RH, Koralsztein J-P, and di Prampero PE. Effect of a previous sprint on the parameters of the wok-time to exhaustion relationship in high intensity cycling. *Int J Sports Med* 26: 583-592, 2005.
- 34. **Hill AV**. The physiological basis of athletic records. *Nature* 116: 544-548, 1925.
- 35. **Hill AV**. Speed and energy requirement. In: *Muscular Movement in Man*. New York: McGraw-Hill, 1927, p. 41-44.
- 36. **Hill DW**. The critical power concept: A review. *Sports Med* 16: 237-254, 1993.
- 37. **Hill DW, Smith JC, Leuschel JL, Chasteen SD, and Miller SA**. Effect of pedal cadence on parameters of the power-time relationship. *Int J Sports Med* 16: 82-87, 1995.
- 38. **Hoelting BD, Scheuermann BW, and Barstow TJ**. Effect of contraction frequency on leg blood flow during knee extension exercise in humans. *J Appl Physiol* 91: 671-679, 2001.
- 39. **Hwang K, Huan F, and Kin DJ**. Muscle fibre types of the lumbrical, interossei, flexor, and extensor muscles moving the index finger. *Journal of Plastic Surgery and Hand Surgery* 47: 268-272, 2013.

- 40. **Jenkins DG, and Quigley BM**. Endurance training enhances critical power. *Med Sci Sports Exerc* 24: 1283-1289, 1992.
- 41. **Jenkins DG, and Quigley BM**. The influence of high-intensity exercise training on the Wlim-Tlim relationship. *Med Sci Sports Exerc* 25: 275-282, 1993.
- 42. **Johnson MA, Polgar J, Weightman D, and Appleton D**. Data on the distribution of fibre types in thirty-six human muscels: An autopsy study. *Journal of the Neurological Sciences* 18: 111-129, 1973.
- 43. **Jones AM, Vanhatalo A, Burnley M, Morton RH, and Poole DC**. Critical power: Implications for determination of $\dot{V}O_{2max}$ and exercise tolerance. *Med Sci Sports Exerc* 42: 1876-1890, 2010.
- 44. **Jones AM, Wilkerson DP, Burnley M, and Koppo K**. Prior heavy exercise enhances performance during subsequent perimaximal exercise. *Med Sci Sports Exerc* 35: 2085-2092, 2003.
- 45. **Jones AM, Wilkerson DP, DiMenna FJ, Fulford J, and Poole DC**. Muscle metabolic responses to exercise above and below the "critical power" assessed using ³¹P-MRS. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 294: 585-593, 2008.
- 46. **Kindig CA, Richardson TE, and Poole DC**. Skeletal muscle capillary hemodynamics from rest to contractions: implications for oxygen transfer. *J Appl Physiol* 92: 2513-2520, 2002.
- 47. **Lutjemeier BJ, Miura A, Scheuermann BW, Koga S, Townsend DK, and Barstow TJ**. Muscle contraction-blood flow interactions during upright knee exercise in humans. *J Appl Physiol* 98: 1575-1583, 2005.
- 48. **McCartney N, Heigenhauser GJF, and Jones NL**. Power output and fatigue of human muscle in maximal cycling exercise. *Journal of Applied Physiology: respiratory, environmental, and exercise physiology* 55: 218-224, 1983.
- 49. **McNaughton L, and Thomas D**. Effects of differing pedalling speeds on the power-duration relationship of high intensity cycle ergometry. *Int J Sports Med* 17: 287-292, 1996.
- 50. **Miura A, Kino F, Kajitani S, Sato H, Sato H, and Fukuba Y**. The effect of oral creatine supplementation on the curvature constant parameter of the power-duration curve for cycle ergometry in humans. *Japanese Journal of Physiology* 49: 169-174, 1999.

- 51. **Miura A, Sato H, Sato H, Whipp BJ, and Fukuba Y**. The effect of glycogen depletion on the curvature constant parameter of the power-duration curve for cycle ergometry. *Ergonomics* 43: 133-141, 2000.
- 52. **Mizuno M, Secher NH, and Quistorff B**. ³¹P-NMR spectroscopy, rsEMG, and histochemical fiber types of human wrist flexor muscles. *J Appl Physiol* 76: 531-538, 1994.
- 53. **Monod H, and Scherrer J**. The work capacity of a synergic muscular group. *Ergonomics* 8: 329-338, 1965.
- 54. **Moritani T, Nagata A, DeVries HA, and Muro M**. Critical power as a measure of physical work capacity and anaerobic threshold. *Ergonomics* 24: 339-350, 1981.
- 55. **Murgatroyd SR, Ferguson CS, Ward SA, Whipp BJ, and Rossiter HB**. Pulmonary O₂ uptake kinetics as a determinant of high-intensity exercise tolerance in humans. *J Appl Physiol* 110: 1598-1606, 2011.
- 56. **Osada T, and Radegran G**. Femoral artery inflow in relation to external and total work rate at different knee extensor contraction rates. *J Appl Physiol* 92: 1325-1330, 2002.
- 57. **Perry SR, Housh TJ, Weir JP, Johnson GO, Bull AJ, and Ebersole KT**. Mean power frequency and amplitude of the mechanomyographic and electromyographic signals during incremental cycle ergometry. *Journal of Electromyography and Kinesiology* 11: 299-305, 2001.
- 58. **Petrofsky JS**. Frequency and Amplitude Analysis of the EMG During Exercise on the Bicycle ergometer. *Eur J Appl Physiol* 41: 1-15, 1979.
- 59. **Poole DC, Copp SW, Hirai DM, and Musch TI**. Dynamics of muscle microcirculatory and blood-myocyte O₂ flux during contractions. *Acta Physiologica* 202: 293-310, 2011.
- 60. **Poole DC, Ward SA, Gardner GW, and Whipp BJ**. Metabolic and respiratory profile of the upper limit for prolonged exercise in man. *Ergonomics* 31: 1265-1279, 1988.
- 61. **Pugh LGCE**. The relation of oxygen intake and speed in competition cycling and comparative observations on the bicycle ergometer. *J Physiol* 241: 795-808, 1974.
- 62. **Richards JC, Crecelius AR, Kirby BS, Larson DG, and Dinenno FA**. Muscle contraction duration and fibre recruitment influence blood flow and oxygen consumption independent of contractile work during steady-state exercise in humans. *Exp Physiol* 97: 750-761, 2012.
- 63. **Richardson RS, Poole DC, Knight DR, Wagner PD, Hogan MC, and Mathieu-Costello O**. Red blood cell transit time in man: Theoretical effects of capillary density. In: *Oxygen Transport to Tissue XVI*. New York: Plenum Press, 1993, p. 521-532.

- 64. **Robergs RA, Icenogle MV, Hudson TL, and Greene ER**. Temporal inhomogeneity in brachial artery blood flow during forearm exercise. *Med Sci Sports Exerc* 29: 1021-1027, 1997.
- 65. Roca J, Agusti AGN, Alonso A, Poole DC, Viegas C, Barbera JA, Rodriguez-Roisin R, Ferrer A, and Wagner PD. Effects of training on muscle O₂ transport at $\dot{V}O_{2max}$. *J Appl Physiol* 73: 1067-1076, 1992.
- 66. **Russ DW, Elliott MA, Vandenborne K, Walter GA, and Binder-Macleod SA**. Metabolic costs of isometric force generation and maintenance of human skeletal muscle. *Am J Physiol Endrocrinol Metab* 282: E448-E457, 2002.
- 67. **Ryschon TW, Fowler MD, Wysong RE, Anthony AR, and Balaban RS**. Efficiency of human skeletal muscle in vivo: comparison of isometric, concentric, and eccentric muscle action. *J Appl Physiol* 83: 867-874, 1997.
- 68. **Sadamoto T, Bonde-Petersen F, and Suzuki Y**. Skeletal muscle tension, flow, pressure, and EMG during sustained isometric contractions in humans. *European Journal of Applied Physiology and Occupational Physiology* 51: 395-408, 1983.
- 69. **Sargeant AJ**. Human power output and muscle fatigue. *Int J Sports Med* 15: 116, 1994.
- 70. **Sargeant AJ, Hoinville E, and Young A**. Maximum leg force and power output during short-term dynamic exercise. *Journal of Applied Physiology: respiratory, environmental, and exercise physiology* 51: 1175-1182, 1981.
- 71. **Sjogaard G, Hanses EA, and Osada T**. Blood flow and oxygen uptake increase with total power during five different knee-extension contraction rates. *J Appl Physiol* 93: 1676-1684, 2002.
- 72. **Smith JC, Stephens DP, Hall EL, Jackson AW, and Earnest CP**. Effect of oral creatine ingestion on parameters of the work rate-time relationship and time to exhaustion in high-intensity cycling. *Eur J Appl Physiol* 77: 360-365, 1998.
- 73. Van Beekvelt MCP, Shoemaker JK, Tschakovsky ME, Hopman MTE, and Hughson RL. Blood flow and muscle oxygen uptake at the onset and end of moderate and heavy dynamic forearm exercise. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 280: R1741-R1747, 2001.
- 74. **Vanhatalo A, Doust JH, and Burnley M**. A 3-min all-out cycling test is sensitive to a change in critical power. *Med Sci Sports Exerc* 40: 1693-1699, 2008.

- 75. **Vanhatalo A, Fulford J, DiMenna FJ, and Jones AM**. Influence of hyperoxia on muscle metabolic responses and the power-duration work relationship during severe-intensity exercise in humans: a ³¹P magnetic resonance spectroscopy study. *Exp Physiol* 95: 528-540, 2010.
- 76. Vanhatalo A, Poole DC, DiMenna FJ, Bailey SJ, and Jones AM. Muscle fiber recruitment and the slow component of O₂ uptake: constant work rate vs. all-out sprint exercise. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 300: R700-R707, 2011.
- 77. **Wagner PD**. Determinants of maximal oxygen transport and utilization. *Annual Review of Physiology* 58: 21-50, 1996.
- 78. **Wagner PD**. Modeling O₂ transport as an integrated system limiting $\dot{V}O_{2MAX}$. *Computer Methods and Programs in Biomedicine* 101: 109-114, 2011.
- 79. **Walloe L, and Wesche J**. Time course and magnitude of blood flow changes in the human quadriceps muscles during and following rhythmic exercise. *J Physiol* 405: 257-273, 1988.
- 80. **Whipp BJ, Huntsman DJ, Stoner N, Lamarra N, and Wasserman K**. A constant which determines the duration of tolerance to high intensity work. *Federation Proceedings* 41: 1591, 1982.
- 81. **Whipp BJ, Ward SA, and Hassall MWC**. Estimating the metabolic rate of marching Roman Legionaries. *J Physiol* 491P: 60, 1996.
- 82. **Whipp BJ, Ward SA, and Hassall MWC**. Paleo-bioenergetics: the metabolic rate of marching Roman legionaries. *British Journal of Sports Medicine* 32: 261-264, 1998.

Chapter 3 - Influence of blood flow occlusion on muscle oxygenation characteristics and the parameters of the power-duration relationship

Summary

It was previously postulated that blood flow occlusion during exercise would reduce critical power (CP) to 0 Watts (W), while not altering the curvature constant (W'). We empirically assessed the influence of blood flow occlusion on CP, W', and muscle oxygenation characteristics. Ten healthy men (age: 24.8 ± 2.6 yrs; height: 180 ± 5 cm; weight: 84.6 ± 10.1 kg) completed four constant-power handgrip exercise tests during both control blood flow (control) and blood flow occlusion (occlusion) for the determination of the power-duration relationship. Occlusion CP (-0.7 \pm 0.4 W) was significantly (p < 0.001) lower than control CP (4.1 \pm 0.7 W) and significantly (p < 0.001) lower than 0 W. Occlusion W' (808 \pm 155 J) was significantly (p < 0.001) different from control W' (558 \pm 129 J) and all ten subjects demonstrated an increased occlusion W' with a mean increase of ~49 %. The current findings support the aerobic nature of CP. The findings also demonstrate that the amount of work that can be performed above CP is constant for a given condition, but can vary across conditions. Moreover, this amount of work that can be performed above CP does not appear to be the determinant of W', but rather a consequence of the depletion of intramuscular energy stores and/or the accumulation of fatigue inducing metabolites which limit exercise tolerance and determine W'.

Introduction

The robust nature of the power-duration relationship (and its equivalents for other modes of exercise) has been well established (26, 27, 52). Nevertheless, the precise physiological mechanisms of the curvature constant (W'), and to a lesser degree critical power (CP), have remained elusive. The growing body of evidence supports that CP represents the highest attainable steady-state for aerobic energy production without continually drawing on W' and, as such, demarcates the boundary between the heavy- and severe-intensity exercise domains (4, 12, 38, 39, 42, 51). It is also evident that W' is a constant term that determines T_{lim} for severe-intensity exercise (21, 55). Intramuscular energy stores (35, 36, 38), the accumulation of fatigue inducing metabolites (7, 18, 21, 28), and/or the magnitude of the severe-intensity domain (5, 51) have all been postulated to determine W'. Building evidence supports that complete utilization of W' is associated with consistent muscle [PCr], [Pi], and [H⁺] perturbations, which may limit the amount of work performed above CP (28, 43, 51). Additionally, the rate of W' utilization (but not the magnitude of W') is influenced by manipulations in O₂ delivery and O₂ extraction via alterations in muscle contraction duty cycle (4). While much has been revealed regarding the mechanisms of CP and W', it is not clear how blood flow occlusion influences each parameter.

In originally characterizing the power-duration relationship, Monod and Scherrer (38) speculated on the influence of blood flow occlusion by stating, "Factor b [CP] is linked to circulatory conditions in the muscle. For when the dynamic work is performed under arterial cuff, the maximum work becomes constant whatever be the time after which exhaustion occurs. The maximum work is then equal to factor a [W']..." Implicit in this statement, is that CP must be 0 Watts (W) with blood flow occlusion for the maximum work performed to be equal to W' and W' to remain constant. This suggests that CP is independent of anaerobic energy production and that the O₂ trapped in the arm with occlusion (i.e., bound to myoglobin and within the vasculature) is not sufficient to measurably contribute to CP.

Furthermore, this statement implies that W' is not determined by aerobic energy production nor the clearance of H⁺ from the muscle vascular space with perfusion (at least beyond the conditions of the occluded limb). A constant W' also implies that intracellular pathways for energy production are unaltered, if indeed similar intramuscular metabolic perturbations do occur for the complete utilization of W' across interventions. Thus, blood flow occlusion exercise is a unique model to empirically test the statement of Monod and Scherrer (38), while providing further insight into the mechanisms determining the power-duration relationship.

Exercise across work rates within the severe-intensity domain (42) is characterized by the concomitant progressive increases in oxygen uptake (\dot{V}_{O_2}) and blood flow, the depletion of intramuscular phosphocreatine ([PCr]), and the accumulation of inorganic phosphate ([Pi]) and hydrogen ions ($[H^+]$), all of which achieve similar values at the limit of exercise tolerance (T_{lim}) (3, 9, 28, 32, 38, 43, 49, 51). Moreover, Vanhatalo et al. (51) demonstrated that [PCr], [Pi], and $[H^+]$ at T_{lim} during knee-extension exercise are independent of augmented O₂ delivery via inspired hyperoxic gas. Consistent with these muscle physiological responses being independent of O₂ delivery, similar skeletal muscle electromyography characteristics (EMG) are expressed between normoxic and hypoxic exercise (15, 41). In contrast, fractional O₂ extraction (indicated by deoxygenated-[hemoglobin + myoglobin] (deoxy-[Hb + Mb]) (10, 14, 19, 20, 22)) at T_{lim} appears to be scaled to O₂ delivery, such that it is greater with hypoxia (41) and less with hyperoxia (51) than normoxia, respectively. However, it remains to be determined if similar EMG characteristics and fractional O2 extractions are attained at Tlim for severeintensity domain power outputs within a given O₂ delivery. Thus, it remains to be determined if reductions in O₂ delivery via blood flow occlusion (rather than inspired hypoxic gas) result in similar EMG characteristics and attainment of fractional O₂ extraction.

Therefore, the aim of the current study was to assess the influence of blood flow occlusion on the parameters of the power-duration relationship (CP and W'), EMG characteristics, and muscle oxygenation characteristics. According to the prediction of Monod and Scherrer (38) our primary hypotheses were that with blood flow occlusion, 1) CP would not be significantly different from 0 W and 2) W' would not be significantly different from control. Second, we hypothesized that within each condition 3) deoxy-[Hb + Mb] and 4) EMG characteristics would not be significantly different at $T_{\rm lim}$.

Methods

Experimental Procedures

All experimental procedures in the present study were approved by the Institutional Review Board at Kansas State University and conformed to the standards set forth by the *Declaration of Helsinki*. Prior to providing written informed consent and completion of a health history questionnaire, subjects were informed of the protocol and potential risks of participation. Testing sessions were separated by at least 24 h and subjects were instructed to abstain from vigorous activity during the 24 h prior to testing, in addition to abstaining from caffeine and alcohol consumption during the 2 and 12 h prior to testing, respectively.

A previously described custom-built two-handed handgrip ergometer (4) was utilized for all testing sessions. The ergometer was attached to a pneumatic cylinder by means of a cable-pulley system, which provided a fixed linear displacement of 4 cm per handgrip contraction. Resistance was controlled via pressurization of the pneumatic cylinder and work was accomplished by compressing the air within the pneumatic cylinder. Power output was calculated as $P = Rdf \cdot k^{-1}$, where P is power in W, R is resistance in kg, d is displacement in meters (m), f is contraction frequency, and k is the constant 6.12 for the conversion of kg·m·min⁻¹ to W. Alterations in power output were accomplished via alterations in resistance, as d and f were held constant. The ergometer was calibrated prior to the study. Subjects were seated in front of the ergometer and grasped the handrail such that both forearms were approximately at heart level. Exercise was performed using a 50% contraction duty cycle (1.5 s contraction: 1.5 s relaxation) at a rate of 20 contractions·min⁻¹. An audio recording with the specific timing was used in conjunction with feedback provided by an investigator to ensure correct timing. Prior to data collection, subjects completed three testing sessions for familiarization with the contraction protocol. All testing sessions were continued until T_{lim}, determined as the inability to successfully complete three consecutive contraction cycles.

An incremental power output test was first performed for the determination of peak power (P_{peak}). This protocol was initiated at 1.0 W and the power output was increased by 0.5 W·min⁻¹ until T_{lim}. The greatest power output for which at least half of the stage was completed was used as P_{peak}. Each subject also completed four constant-power tests for both the control brachial artery blood flow (control) and occluded brachial artery blood flow (occlusion) conditions in a randomized order. The P_{peak} was utilized to determine power outputs that would elicit T_{lim} within ~1 – 15 min (43). The control constant-power tests were performed at 80, 90, 110, and 130 % P_{peak} and the occlusion constant-power tests were performed at ~17 (1 W for all subjects), 35, 80, and 110 %P_{peak}. Pilot work in our laboratory demonstrated these intensities elicited similar T_{lim} values for occlusion constant-power tests as those recommended for control constant-power tests. The 80 and 110 %P_{peak} were conducted in both conditions to examine the influence of blood flow, independent of differences in power output. Brachial artery blood flow was occluded with a vascular cuff positioned around the brachial region of each arm, which was rapidly inflated (< 0.3 s) to suprasystolic pressures (≥ 275 mmHg) at the onset of exercise and remained inflated until T_{lim} (E20 Rapid Cuff Inflator, Hokanson, Bellevue, WA, USA). Blood flow occlusion was verified by the absence of a radial pulse and the cuff pressures were continuously monitored to ensure ≥ 275 mmHg.

Measurements and data analysis

The power-duration relationship for both control and occlusion was determined by fitting the power output and T_{lim} data from the constant-power tests with the two-parameter hyperbolic model:

$$t = W' / (P - CP)$$

where t is T_{lim} in s, W' is the finite work capacity in Joules (J), P is the power output in W, and CP is critical power in W.

A frequency-domain multi-distance near-infrared spectroscopy (NIRS) system was used to measure the oxygenation characteristics of the flexor digitorum superficialis of the right forearm during each testing session (Oxiplex TS, ISS, Champaign, IL, USA). Detailed descriptions of the principles and algorithms of the NIRS technology have previously been described (20, 23). Briefly, this device consists of one detector fiber bundle and eight light-emitting diodes (LED) operating at wavelengths of 690 and 830 nm (four LEDs per wavelength). The LED-detector fiber bundle separation distances are 2.0, 2.5, 3.0, and 3.5 cm. This NIRS device measures and incorporates the dynamic reduced scattering coefficients to provide absolute concentrations (µM) for deoxy-[Hb + Mb], oxygenated-[Hb + Mb] (oxy-[Hb+Mb]), total-[Hb + Mb], and %Saturation-[Hb + Mb] (%Sat-[Hb + Mb]). The deoxy-[Hb + Mb] is relatively insensitive to changes in blood-volume (10, 19, 22) and has been used to reliably estimate the fractional oxygen extraction (10, 13, 14, 19, 20, 22, 34). The NIRS probe was calibrated prior to each test according to the manufacturer's recommendations. The belly of the flexor digitorum superficialis of the right arm was identified using palpation and EMG. The NIRS probe was secured along the belly of the flexor digitorum superficialis and was wrapped with an elastic bandage to prevent movement of the probe. The position of the NIRS probe was marked with indelible ink for reproducible placement throughout the study. The NIRS data were collected at 50 Hz and stored for post-hoc analysis. The NIRS data were analyzed using 1 s time-binned mean values and 9 s time-binned mean values at T_{lim}. A kinetics analysis was conducted for the deoxy-[Hb + Mb] data over the initial 60 s of exercise using a mono-exponential model:

$$\text{deoxy-[Hb + Mb]}(t) = \text{deoxy-[Hb + Mb]}(b) + A(1 - e^{-(t - \text{TD})/\tau})$$

where deoxy-[Hb + Mb](t) is the deoxy-[Hb + Mb] at any point in time, deoxy-[Hb + Mb](b) is the baseline deoxy-[Hb + Mb] before the onset of exercise, A is the amplitude of the deoxy-[Hb + Mb]

response, TD is the time delay before the start of the increase in deoxy-[Hb + Mb], and τ is the time constant of the increase in deoxy-[Hb + Mb].

Surface EMG (Trigno EMG, Delsys Inc., Boston, MA, USA) measurements were obtained from the flexor digitorum superficialis of the left forearm during each testing session. The belly of the left flexor digitorum superficialis was identified by palpation and strong electrical activity when the fingers were flexed, but not with ulnar or radial deviation. The position of the EMG sensor was marked with indelible ink for reproducible placement throughout the study. The EMG data were collected at 1000 Hz and stored for post-hoc analysis. The raw EMG data were processed with a band-pass filter (0.05 – 400 Hz) and each electrical burst corresponding to a muscle contraction was detected using a custom-designed computer program. For each muscle contraction, the signal amplitude characteristics were analyzed via root mean square (RMS) to provide an index of muscle activation and motorneuron firing rate. RMS values were normalized to the first minute value of each test. The frequency characteristics were analyzed via median power frequency (MedPF) to provide an index of the muscle action potential conduction velocity.

Data analysis

All curve fitting procedures and statistical analyses were performed using a commercially available software package (SigmaPlot and SigmaStat, Systat Software Inc., Point Richmond, CA, USA). Student's paired t-tests were used to compare the CP and W' parameters measured for control and occlusion. Occlusion CP was compared to 0 W using a one-sample t-test. One-way ANOVAs with repeated measures were used to compare NIRS or EMG measurements within each condition. Two-way ANOVAs with repeated measures (condition x intensity) were used to compare both the NIRS and EMG measurements for the tests performed at 80 and 110 %P_{peak}. Tukey's post-hoc analyses were conducted

when significant main effects were detected and effect sizes were calculated for both CP and W'. Differences were considered statistically significant when p < 0.05. 95% confidence intervals (CI) around CP and W' were determined for each subject in both conditions. All data are presented as mean \pm SD unless otherwise noted.

Results

Ten healthy men (age: 24.8 ± 2.6 yrs; height: 180 ± 5 cm; weight: 84.6 ± 10.1 kg) completed the study. The P_{peak} from the incremental ramp test was 6.1 ± 0.9 W. The durations of the constant-power tests were: $80 \% P_{peak}$: 759 ± 243 ; $90 \% P_{peak}$: 471 ± 156 ; $110 \% P_{peak}$: 211 ± 45 ; $130 \% P_{peak}$: 122 ± 32 s for control and $17 \% P_{peak}$: 494 ± 118 ; $35 \% P_{peak}$: 301 ± 49 ; $80 \% P_{peak}$: 143 ± 24 ; $110 \% P_{peak}$: 102 ± 10 s for occlusion. The time to achieve T_{lim} was significantly shorter for occlusion compared to control at both $80 \% P_{peak}$ (143 ± 24 s vs. 759 ± 243 s, p < 0.002) and $110 \% P_{peak}$ (102 ± 10 s vs. 211 ± 45 s, p < 0.001).

Individual subject hyperbolic model fits are presented in Figure 1. The determination of the control power-duration relationship in one subject was found to be greatly weighted towards a specific constant-power test, as previously described (4, 12, 38, 39, 42, 51). Therefore, this subject's control power-duration relationship was determined using three constant-power tests (4, 12, 38, 39, 42, 51). The mean hyperbolic model fits (r^2) were 0.98 ± 0.02 for control and 0.99 ± 0.01 for occlusion. Occlusion CP (-0.7 ± 0.4 W; mean 95% CI = -1.3 to 0.0 W) was significantly (p < 0.001, ES = 6.0) lower than control CP (4.1 ± 0.7 W; mean 95% CI = 3.4 to 4.7 W) and significantly (p < 0.001, ES = 1.75) lower than 0 W (Figure 2). Occlusion W' (808 ± 155 J; mean 95% CI = 533 to 1082 J) was significantly (p < 0.001, ES = 1.4) different from control W' (558 ± 129 J; mean 95% CI = 158 to 1582 J) and all 10 subjects demonstrated an increased occlusion W' with a mean increase of -49% (Figure 2).

The NIRS measurements were not significantly different at T_{lim} within each blood flow condition across the different intensities, except for %Sat-[Hb + Mb] between 35 and 110% within occlusion (Figures 3 and 4; Table 1). Total-[Hb + Mb] increased significantly above baseline for all control exercise tests and the values at T_{lim} were not significantly different among these tests. In contrast, total-[Hb + Mb] did not significantly increase above baseline during the occlusion tests (Figure 4), demonstrating complete blood flow occlusion of the limb. For the 80 and 110 %P_{peak} tests, deoxy-[Hb +

Mb] at T_{lim} was significantly greater for occlusion than control and Oxy-[Hb + Mb], Total-[Hb + Mb0, and %Sat-[Hb + Mb] were significantly lower for occlusion than control (Figure 5; Table 1). The deoxy-[Hb + Mb] kinetics analyses results are presented in Table 2. There were no significant differences in the TD or τ between control and occlusion, while the A was significantly greater for occlusion than control at both 80 and 110 % P_{peak} .

No statistically significant differences for control RMS at T_{lim} were detected, while the MedPF at T_{lim} was significantly less for 110 % P_{peak} compared to both 80 % P_{peak} (p < 0.001) and 90 % P_{peak} (p < 0.005). Occlusion RMS at T_{lim} was significantly greater for 110 % P_{peak} compared to 17 % P_{peak} (p < 0.001). There were no significant differences in occlusion MedPF at T_{lim} (Figure 6). RMS at T_{lim} was significantly lower for occlusion compared to control at 110 % P_{peak} (p = 0.012), but there was no significant difference at 80% P_{peak} between occlusion and control (p = 0.106). Occlusion MedPF at T_{lim} was significantly lower than control at 80% P_{peak} (p < 0.001) and there was no significant difference at $110\%P_{peak}$ between occlusion and control (p = 0.43).

Discussion

This study examined the influence of blood flow occlusion on the parameters of the power-duration relationship during handgrip exercise. Handgrip exercise during blood flow occlusion was well-described by the two-parameter hyperbolic model. In contrast to the primary hypotheses, occlusion CP was lower than 0 W and occlusion W' was greater than control W'. These results support the aerobic nature of CP, as the reduction in O_2 delivery with occlusion reduced CP. These results also support that W' is relatively constant within a given O_2 delivery condition, but can vary across O_2 delivery conditions. Additionally, this study presents novel findings regarding muscle oxygenation characteristics during control and occluded severe-intensity handgrip exercise. In agreement with our third hypothesis, deoxy-[Hb + Mb] at T_{lim} was similar within each condition. In contrast to our fourth hypothesis, differences in EMG characteristics at T_{lim} were detected within each condition.

The power-duration relationship

A two-parameter hyperbolic model may be used to empirically describe the decrease in T_{lim} with increasing power outputs to yield the parameters CP and W' (38, 43, 55). The asymptote in this model represents CP, which distinguishes an exercise intensity above which a physiological steady-state is not attained and exercise T_{lim} is limited by W'. As such, CP represents the highest sustainable rate of aerobic ATP production for which W' will not be continuously utilized (4, 12, 38, 39, 42, 51). The curvature constant in this model represents W', modeled as a finite amount of work that can be performed above CP that when fully expended results in T_{lim} (21, 38, 43, 55). However, mounting evidence suggests that W' is not solely a finite amount of work, per se. Rather it appears that similar "limiting" muscle metabolic perturbations in [PCr], [Pi], and [H⁺] are attained, that in turn constrain the maximal amount of work that can be performed above CP (6, 28, 43, 51). Despite the precise mechanisms of CP and W' not being fully understood, it is clear that exercise intensities above CP are

predictably limited by W' and that the limit of exercise tolerance occurs at similar physiological states for these intensities.

Deterministic mechanisms of CP

To date, evidence suggests that CP represents the highest attainable steady-state for aerobic energy production without continuously drawing upon W' (4, 12, 38, 39, 42, 51). It was with this interpretation of CP in mind that Monod and Scherrer (38) postulated that the maximum amount of work performed under blood flow occlusion would be equal to W', which would necessitate a CP equal to 0 W. In contrast to this prediction, the current study demonstrated that the calculated occlusion CP was actually reduced to a power output slightly, but significantly less than 0 W. While a negative CP is only theoretical and not physiologically attainable, nonetheless this estimate provides mechanistic insight into CP. An occlusion CP equal to 0 W (i.e., rest) would indicate that indefinite resting occlusion would not hinder the ability of the skeletal muscle to perform contractions. Previous studies have demonstrated hemoglobin and myoglobin deoxygenation and the subsequent depletion of [PCr] during resting occlusion (25, 31). Thus, after sufficient depletion of aerobic energy sources, resting metabolism is sustained by anaerobic energy production and the depletion of [PCr] and accumulation of [Pi] suggests that W' is utilized during resting occlusion (i.e., 0 W). As CP is the highest sustainable rate of aerobic ATP production without drawing continuously upon W', it would not be expected that occlusion CP be equal 0 W, as without blood flow there is no sustainable rate of aerobic ATP production. Rather, CP would be expected to be a theoretical value of negative power for which the magnitude is proportional to the resting metabolic rate.

Deterministic mechanisms of W'

W' was significantly greater with blood flow occlusion in all ten subjects (mean increase ~49%). A greater W' with occlusion may be due to differences in metabolic economy between occlusion and control exercise (30, 31) altering the degree of metabolic perturbation for a given amount of mechanical work. Moreover, as T_{lim} during occlusion appears to be the result of the accumulation of inhibitory byproducts of glycolytic ATP turnover (31), it may be that a greater degree of accumulation is tolerated during occlusion exercise and thus, more work above CP may be performed. Furthermore, if the 250 J increase in W' for occlusion arose solely from oxidative energy production, it would require a greater O₂ extraction of 12 ml (assuming $0.000239 \text{ kcal} \cdot \text{J}^{-1}$ and $5 \text{ kcal} \cdot \text{l}^{-1} \cdot \text{O}_2$). The greater deoxy-[Hb + Mb] for occlusion than for control suggests that a small portion of the increase in W' may be due to an increased O₂ extraction. However, the greater deoxy-[Hb + Mb] for occlusion (~64 μM) than for control (~50 μM) would amount only to 0.05 ml O₂ (assuming 150 g flexor digitorum superficialis and 1060 g·l⁻¹ muscle density), which represents 0.4 % of the 12 ml O₂ needed for the increase in W' to be the result of O₂ extraction. Moreover, no relationship between the increase in deoxy-[Hb + Mb] and W' was detected. Thus, it does not appear that the increased W' during occlusion is the result of the greater fractional O₂ extraction."

The results of the current study lend credence to W' not being determined by a finite amount of work per se, but rather by other mechanisms such as the rate of attainment of "limiting" muscle metabolic perturbations (7, 21, 28, 43, 51), the magnitude of the severe domain (5, 51), and/or the \dot{V}_{O_2} slow component (40). Modeling the current data reveals that while W' is utilized in its entirety for exercise intensities above CP, the proportion of W' that contributes directly to external work is not constant across this range of power outputs (Figure 7). For example, at rest (i.e., 0 W) under occlusion, W' will be used in its entirety to support factors distinct from external work (i.e., resting cellular processes, ion handling, etc.). This is demonstrated by resting blood flow occlusion leading to a

desaturation in myoglobin (46), increased adenosine diphosphate ([ADP]), and a decreased [PCr] (2, 25). With increasing power outputs, a greater proportion of W' would be utilized for external work, as the proportion of energy turnover via external work will increase (Figure 7). Thus, at sufficiently high power outputs (i.e., right side of Figure 7 bottom) the majority of W' is associated with external work. However, it appears that some of the energy derived from the utilization of W' still contributes to the factors that are distinct from external work, including the internal work of handgrip contraction. This model may explain why W' (determined as the amount of external work performed above CP) is constant when determined under normal blood flow conditions, as power outputs utilized to determine the normal power-duration relationship are on the upper-right portion of the curve in Figure 7 bottom. This is consistent with previous suggestions that W' is determined by an integration of multiple mechanisms, rather than a single mechanism alone (7, 17).

Muscle oxygenation characteristics

In the current study, fractional O_2 extraction was significantly greater for occlusion than for control. This is consistent with previous reports of alterations in the fractional O_2 extraction with varying inspired O_2 concentrations (41, 46, 51). These alterations in fractional O_2 extraction may be the consequence of the Bohr Effect. In the current study, an elevated $[H^+]$ with occlusion (31) may have facilitated oxyhemoglobin dissociation, resulting in the greater fractional O_2 extraction (50).

Fick's law of diffusion states that the flux of O_2 (\dot{V}_{O_2}) is dependent on the diffusivity of oxygen (\dot{D}_{O_2}) and the P_{O_2} gradient between the microvasculature and the mitochondria [$\dot{V}_{O_2} = \dot{D}_{O_2} x$ ($P_{O_2mv} - P_{O_2mit}$)]. Exercise hyperemia increases microvascular hematocrit, and therefore enhances \dot{D}_{O_2} (16, 24, 29, 33). The similar peak plateaus in microvascular hematocrit (i.e., total-[Hb + Mb]) in the current study for control exercise suggest that the peak microvascular hematocrit, and therefore \dot{D}_{O_2} , is

constrained for a given condition. The peak O_2 diffusion gradient $(P_{O_2mv}-P_{O_2mit})$ during severe exercise would also be constrained, as intramuscular P_{O_2} (P_{O_2 mit) achieves similar low values during exercise (37, 44-46) (assuming similar P_{O_2mv} at T_{lim} for a given condition). These constraints directly impact aerobic energy production, thus dictating $\dot{V}_{O_{2max}}$ (44, 53, 54), and presumably CP (4, 12, 39, 51). CP in the current study was greatly attenuated during occlusion, due presumably, in part, to the prevented increase in microvascular hematocrit (i.e., no increase total-[Hb + Mb] above baseline) and the expected exacerbated fall in P_{O2mv}. As such, W' would be utilized earlier and at a faster rate in the occlusion exercise bouts compared to control, thus resulting in the earlier attainment of Tlim. Moreover, the similar nadirs above zero in oxy-[Hb + Mb] at T_{lim} for occlusion exercise suggest that complete deoxygenation of Hb and/or Mb did not occur, consistent with previous findings that Hb and Mb do not fully desaturate during blood flow occlusion exercise (11, 31, 47, 48). These similar nadir values of deoxygenation may result from the continual fall in P_{O_2mv} (decrease in oxy-[Hb + Mb]) as oxygen is utilized and not replenished during occlusion exercise, until the capillary-mitochondria P_{O2} gradient achieves equilibrium with the inherent resistance to O_2 diffusion $(1/D_{O_2})$. At this point, further O_2 flux into the mitochondria would be prevented (i.e., oxy-[Hb + Mb] nadir). Additionally, the accumulation of metabolic byproducts (i.e., $[H^+]$) may inhibit oxidative phosphorylation (8) despite the presence of oxygen remaining in the microcirculation (i.e., oxy-[Hb + Mb] above 0 µM). Therefore severe-intensity exercise tolerance appears to be dependent upon perfusive and diffusive O₂ supply dictating CP, along with the magnitude of W' and its rate of utilization (i.e., the power output).

During occlusion exercise, the Fick principle ($\dot{V}_{O_2} = \dot{Q} \ x \ (a-v)O_2 diff$) dictates that \dot{V}_{O_2} is proportional to O_2 extraction. Interestingly, the time constant (τ) for the change in the deoxy-[Hb + Mb] was not significantly different between conditions at the same power output, while the amplitude was

greater for occlusion. Consistent with this, Vanhatalo et al. (51) demonstrated an unchanged τ for deoxy-[Hb + Mb] between hyperoxic and normoxic knee-extension exercise. These results suggest that O_2 delivery, per se, does not determine the τ for deoxy-[Hb + Mb]. Rather, it appears that at the onset of exercise sufficient levels of O_2 are present within the muscle and surrounding tissue so as to not limit O_2 flux despite O_2 delivery interventions. It is not until later ($\sim >30$ s) into the exercise bout that the O_2 delivery interventions begin to exert influence on fractional O_2 extraction.

Motor unit recruitment and firing characteristics

In the current study, RMS and MedPF at T_{lim} were not always similar within each condition. Moreover, significant differences were detected between control and occlusion RMS at 80 %P_{peak} and MedPF at 110 %P_{peak}. These findings differ from those of prior studies demonstrating similar EMG characteristics between normoxia and hypoxia (41) (15). The current results may differ from previous findings due to greater magnitude of O_2 delivery reduction with occlusion compared to hypoxia. Together these results suggest that EMG characteristics appear not to be influenced by reductions in O_2 delivery up to a certain point, but further reductions in O_2 delivery past this point result in EMG characteristics differences.

Implications of current findings

The findings of the current study further demonstrate the integration of perfusive and diffusive O_2 delivery in determining aerobic energy production, and therefore CP. It is this established CP that dictates the intensities at which W' will be continually utilized until fatigue ensues. Specifically, the current findings indicate that reductions in blood flow (i.e., O_2 delivery) lower CP which results in the utilization of W' and fatigue at lower intensities. Cumulatively, current evidence

suggests that alteration in O_2 delivery via changes in blood flow or inspired O_2 concentrations directly affect CP and W' utilization (4, 12, 39, 51).

Limitations

Several experimental limitations must be considered when interpreting the current data. It is not known from the current data if similar metabolic perturbations are occurring within the muscle during occlusion and control. Therefore, it remains to be determined if the same limiting muscle metabolic perturbations are being achieved at T_{lim} during blood flow occlusion exercise as during control. With this, the current study and many previous studies have utilized small muscle mass exercise (e.g., handgrip and knee-extension). It remains to be determined if large muscle mass exercise (i.e., wholebody exercise such as cycling and running) elicits similar intramuscular metabolic responses for power outputs within the severe-intensity domain. Furthermore, the current study is not able to determine which other non-mechanical-work energy consuming processes may be utilizing W'. Previous publications have demonstrated that a substantial amount of energy is utilized for ion pumping during muscular contraction (1). However, it is not known if these previous findings are applicable in the extreme case of blood flow occlusion exercise. As such, it currently is not known how the relationship between W' and the severe-intensity domain is altered with blood flow occlusion. As [PCr], [Pi], and [H⁺] were not measured in the current study, it cannot be certain that the consistent muscle oxygenation coincided with consistent concentration of these intramuscular metabolites. Prior evidence suggests that consistent values for these metabolites are to be expected despite alterations in O₂ delivery (28, 49, 51). However, it remains to be determined if these values are altered with blood flow occlusion. The noise inherent in EMG measurements limits the confidence in conclusively interpreting the data. As a result, the EMG data are not presented to precisely state the EMG characteristics of the muscle, but rather to

suggest that the muscle may not be achieving similar EMG characteristics at T_{lim} within and between O_2 delivery conditions. Future studies are needed to more completely evaluate the EMG responses for each condition.

Conclusions

The current study demonstrated a greater fractional O₂ extraction during occlusion exercise compared to control. Moreover, muscle oxygenation characteristics attained similar values within a given O₂ delivery condition. Additionally, EMG characteristics appear to be independent of O₂ delivery, but rather to be determined by the performed power output. Cumulatively, current evidence suggests that intramuscular [PCr], [Pi], [H⁺] at T_{lim} are similar and independent of alterations in O₂ delivery, but that fractional O₂ extraction is directly influenced by alterations in O₂ delivery. Moreover, the reduction of O₂ delivery with blood flow occlusion exercise decreased the estimated CP below 0 W and led to an increased W' compared to control. It appears that the resting metabolic rate and/or the internal work of handgrip exercise may result in the estimated apparent CP being below 0 W during occlusion. The findings of the current study support the aerobic nature of CP. Additionally, the findings demonstrate that the amount of work that can be performed above CP is constant for a given condition, but can vary across conditions. Moreover, this amount of work that can be performed above CP does not appear to be the determining mechanism of W', but rather a consequence of the depletion of intramuscular energy stores and/or the accumulation of fatigue inducing metabolites which limit exercise tolerance and determine W'.

Table 3.1 End-exercise NIRS values for control and occlusion.

	Deoxy-[Hb + Mb]	Oxy-[Hb + Mb]	Total-[Hb + Mb]	Sat-[Hb + Mb]	
	(μΜ)	(µM)	(μM)	(%)	
	Control				
$80 \% P_{\text{peak}}$	47.0 ± 14.7	64.7 ± 20.2	111.70 ± 13.2	57.4 ± 14.5	
$90 \% P_{\text{peak}}$	50.0 ± 12.1	57.2 ± 9.5	107.2 ± 12.2	53.7 ± 8.6	
$110 \% P_{\text{peak}}$	50.6 ± 15.7	59.5 ± 10.7	110.0 ± 16.1	54.7 ± 10.5	
$130\%P_{\mathrm{peak}}$	51.3 ± 15.1	31.2 ± 18.4	112.4 ± 19.5	54.5 ± 11.7	
	Occlusion				
$17 \% P_{\text{peak}}$	63.9 ± 14.6	21.0 ± 17.4	84.2 ± 13.7	24.0 ± 18.2	
35 % <i>P</i> _{peak}	70.0 ± 21.9	16.8 ± 14.0	86.8 ± 11.9	$20.5 \pm 17.7^{\rm C}$	
$80 \% P_{\text{peak}}$	$62.8 \pm 15.5^{\dagger}$	$26.7 \pm 11.8^{\dagger}$	$89.5 \pm 12.6^{\dagger}$	$30.1\pm13.3^{\dagger}$	
$110\%P_{\text{peak}}$	$59.2 \pm 17.7^{\dagger}$	$28.4 \pm 13.2^{\dagger}$	$87.6 \pm 15.2^{\dagger}$	$33.0 \pm 16.2^{\dagger}$	

Deoxy-[Hb + Mb], deoxygenated hemoglobin + myoglobin; Oxy-[Hb + Mb], oxygenated hemoglobin + myoglobin; Total-[Hb + Mb], total hemoglobin + myoglobin; Sat-[Hb + Mb], percent saturation of hemoglobin + myoglobin; P_{peak} , peak power from the incremental maximal exercise test. † significantly (p < 0.05) different from control at the same % P_{peak} . Significantly (p < 0.05) different from 110 % P_{peak} within condition.

Table 3.2 Exercise onset deoxy-[Hb + Mb] kinetics parameters for control and occlusion

	y0	A	τ	TD		
_	(μM)	(μM)	(s)	(s)		
_	Control					
$80 \% P_{\text{peak}}$	26.7 ± 4.8	17.8 ± 8.4	10.2 ± 3.4	8.8 ± 3.0^{A}		
$90 \% P_{\text{peak}}$	26.7 ± 5.3	21.8 ± 11.5	11.6 ± 12.2	6.9 ± 1.5		
$110 \% P_{\text{peak}}$	28.1 ± 6.7	22.1 ± 11.2	7.8 ± 2.6	6.7 ± 1.8		
$130 \% P_{\text{peak}}$	27.0 ± 5.7	24.0 ± 11.6	12.0 ± 11.6	4.5 ± 2.4		
_						
_	Occlusion					
$17 \% P_{\text{peak}}$	26.1 ± 5.4	38.1 ± 15.7	32.2 ± 14.3	11.2 ± 3.1		
$35 \% P_{\text{peak}}$	$28.7 \pm 7.0^{\rm C}$	38.0 ± 17.4	17.4 ± 7.1^{B}	$10.4 \pm 2.7^{\rm C}$		
$80 \% P_{\text{peak}}$	27.6 ± 5.5	$33.4 \pm 10.1^{\dagger}$	8.7 ± 2.6^{B}	8.4 ± 2.0^{BC}		
$110 \% P_{\text{peak}}$	23.0 ± 4.6	$34.0 \pm 16.0^{\dagger}$	12.1 ± 9.9^{B}	4.9 ± 3.6^{B}		

Data were analyzed over the first 60 s of exercise with a mono-exponential model. Deoxy-[Hb + Mb], deoxygenated hemoglobin + myoglobin; y0, baseline concentration; A, amplitude of response; τ , time constant of response; TD, time delay from exercise onset to the increase in response. † significantly (p < 0.05) different from control. A significantly (p < 0.05) different from 130 % P_{peak} within condition. Significantly (p < 0.05) different from 110 % P_{peak} within condition.

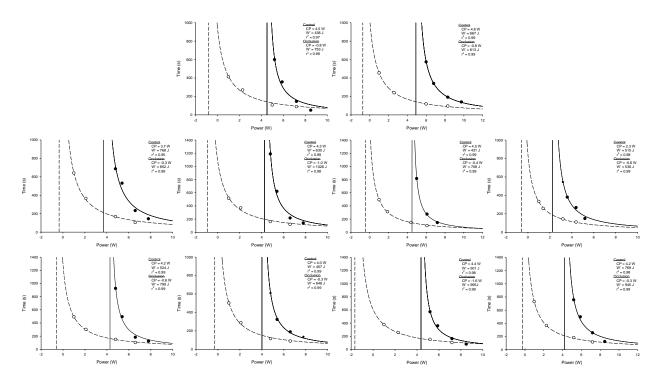
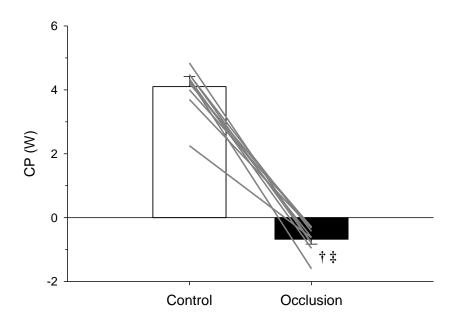


Figure 3.1 Individual subject constant-power and hyperbolic curve fit data.

Subjects completed four constant-power tests for both control and occlusion at power outputs selected to elicit exhaustion in $\sim 1-15$ min. Each subject's constant-power data were fit with a two-parameter hyperbolic model to provide parameter estimated for critical power (CP) and the curvature constant (W').



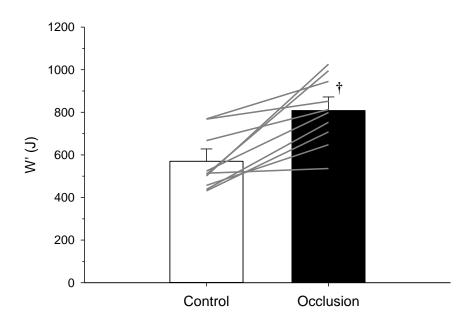


Figure 3.2 Mean CP and W' determined for control and occlusion handgrip exercise.

Critical power (CP) and the curvature constant (W') were determined for both control blood flow (control) and brachial artery blood flow occlusion (occlusion). The grey lines indicate individual subject responses. \dagger significantly different from control (p < 0.001). \ddagger significantly different from 0 Watts (p < 0.0001).

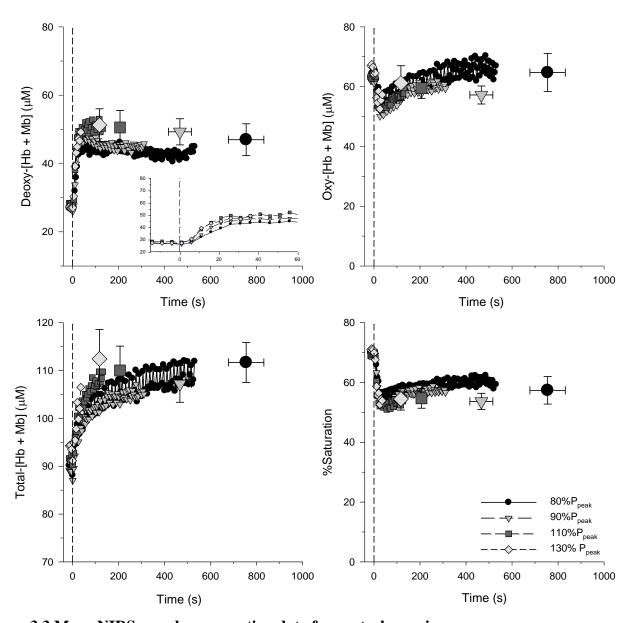


Figure 3.3 Mean NIRS muscle oxygenation data for control exercise.

Deoxygenated-[hemoglobin + myoglobin] (deoxy-[Hb + Mb]), oxygenated (oxy-[Hb + Mb]), total-[Hb + Mb], and percent saturation (%Sat-[Hb + Mb]) data during 80, 90, 110, and 130 %peak power (%P_{peak}). No significant differences were detected between intensities at the limit of exercise tolerance (T_{lim}). Graph insert: deoxy-[Hb + Mb] during the initial 60 s of exercise onset.

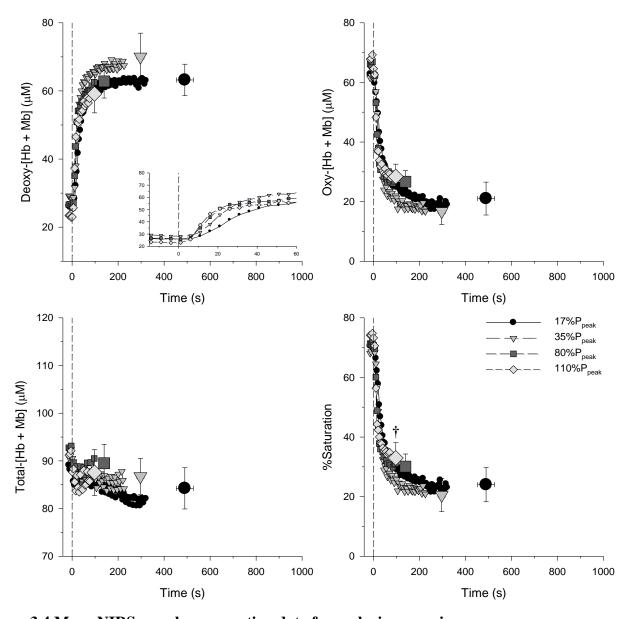


Figure 3.4 Mean NIRS muscle oxygenation data for occlusion exercise.

Deoxygenated-[hemoglobin + myoglobin] (deoxy-[Hb + Mb]), oxygenated (oxy-[Hb + Mb]), total-[Hb + Mb], and percent saturation (%Sat-[Hb + Mb]) data during 17, 35, 80, and 110 %peak power (%P_{peak}). \dagger significantly (p < 0.05) different from 110% P_{peak} at the limit of exercise tolerance (T_{lim}). Total-[Hb + Mb] did not significantly increase above baseline for occlusion. Graph insert: deoxy-[Hb + Mb] during the initial 60 s of exercise onset.

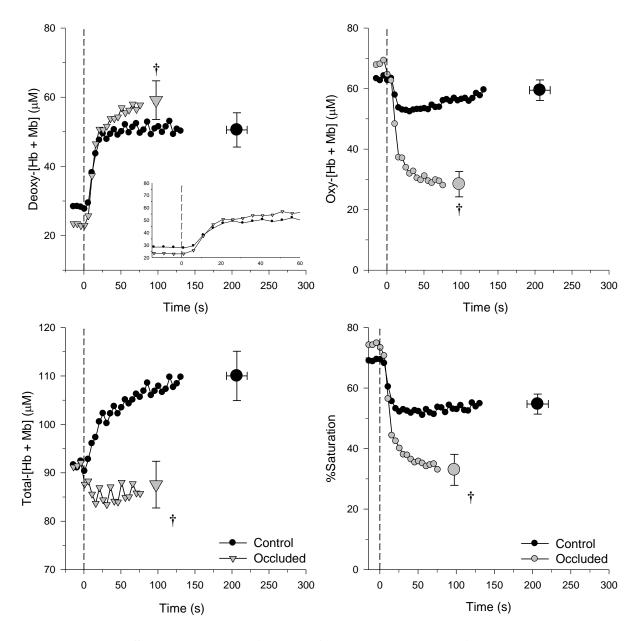


Figure 3.5 Mean NIRS muscle oxygenation data for control and occlusion at 110 % P_{peak} . Deoxygenated-[hemoglobin + myoglobin] (deoxy-[Hb + Mb]), oxygenated (oxy-[Hb + Mb]), total-[Hb + Mb], and percent saturation (%Sat-[Hb + Mb]) at 110 %peak power (% P_{peak}). † significantly (p < 0.05) different from control. Total-[Hb + Mb] did not significantly increase above baseline for occlusion. Graph insert: deoxy-[Hb + Mb] during the initial 60 s of exercise onset.

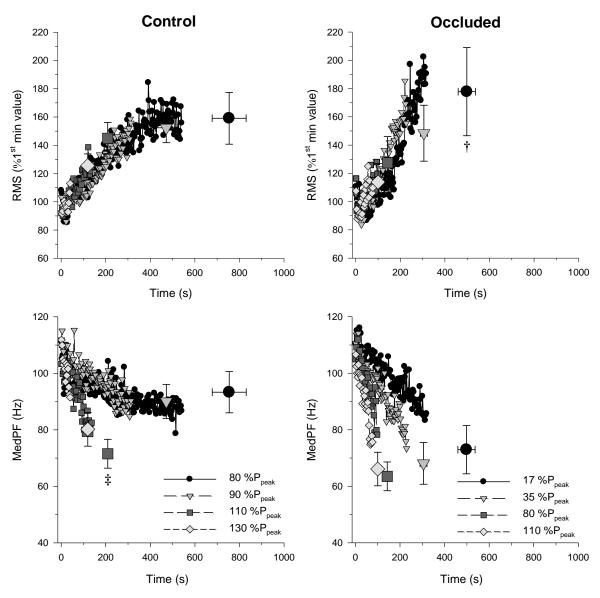
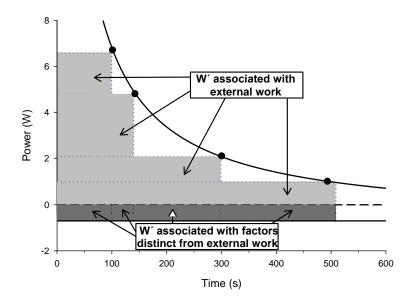


Figure 3.6 Mean EMG data for control and occlusion.

Root mean square (RMS) and median power frequency (MedPF) data at all intensities for control and occlusion. † significantly different from 17 % peak power (P_{peak}). ‡ significantly different from 80 and 90 % P_{peak} within control.



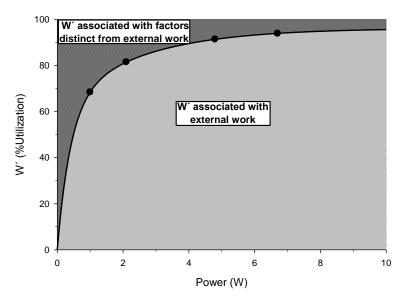


Figure 3.7 Schematic diagram demonstrating the contribution of W' to mechanical- and non-mechanical-work energy consumption processes.

Top: Critical power (CP; the horizontal asymptote) and curvature constant (W') for blood flow occlusion handgrip exercise. CP was significantly below 0 Watts (W). The area defined by the power output and the duration of exercise between CP and 0 W represents the amount of W' associated with factors distinct from external work (i.e., ion handling, resting metabolic rate, internal work, etc.) and the area between 0 W and power output is assumed to represent the amount of W' associated with external work. Below: The mean amount of W' associated with external work and factors distinct from external work (i.e., ion handling, resting metabolic rate, internal work, etc.) at the four occlusion exercise intensities which accounted for the amount of W' positioned between CP and 0 W. This demonstrates the nature of W' and that varying proportions of W' are utilized for different processes depending on the intensity of the exercise.

References

- 1. **Bergstrom M, and Hultman E**. Energy cost and fatigue during intermittent electrical stimulation of human skeletal muscle. *J Appl Physiol* 65: 1500-1505, 1988.
- 2. **Boushel R, Pott F, Madsen P, Radegran G, Nowak M, Quistorff B, and Secher N**. Muscle metabolism from near infrared spectroscopy during rhythmic handgrip in humans. *Eur J Appl Physiol* 79: 41-48, 1998.
- 3. **Broxterman RM, Ade CJ, Poole DC, Harms CA, and Barstow TJ**. A single test for the determination of the parameters of the speed-time relationship for running. *Respiratory Physiology & Neurobiology* 185: 380-385, 2013.
- 4. **Broxterman RM, Ade CJ, Wilcox SL, Schlup SJ, Craig JC, and Barstow TJ**. Influence of duty cycle on the power-duration relationship: Observations and potential mechanisms. *Respiratory Physiology & Neurobiology* 192: 102-111, 2014.
- 5. **Burnley M, and Jones AM**. Oxygen uptake kinetics as a determinant of sports performance. *Eur J Sport Sci* 7: 63-79, 2007.
- 6. **Chidnok W, Fulford J, Bailey SJ, DiMenna FJ, Skiba PF, Vanhatalo A, and Jones AM**. Muscle metabolic determinants of exercise tolerance following exhaustion: relationship to the "critical power". *J Appl Physiol* 115: 243-250, 2013.
- 7. Coats EM, Rossiter HB, Day JR, Miura A, Fukuba Y, and Whipp BJ. Intensity-dependent tolerance to exercise after attaining $\dot{V}O_{2max}$ in humans. *J Appl Physiol* 95: 483-490, 2003.
- 8. **Conley KE, Kemper WE, and Crowther GJ**. Limits to sustainable muscle performance: interaction between glycolysis and oxidative phosphorylation. *The Journal of Experimental Biology* 204: 3189-3194, 2001.
- 9. **Copp SW, Hirai DM, Musch TI, and Poole DC**. Critical speed in the rat: implications for hindlimb muscle blood flow distribution and fibre recruitment. *J Physiol* 588: 5077, 2010.
- 10. De Blasi RA, Cope M, Elwell C, Safoue F, and Ferrari M. Noninvasive measurement of human forearm oxygen consumption by near infrared spectroscopy. *European Journal of Applied Physiology and Occupational Physiology* 67: 20-25, 1993.
- 11. **De Blasi RA, Cope M, and Ferrari M**. Oxygen consumption of human skeletal muscle by near infrared spectroscopy during tourniquet-induced ischemia in maximal voluntary contraction. *Adv Exp Med Biol* 317: 771-777, 1992.

- 12. **Dekerle J, Mucci P, and Carter H**. Influence of moderate hypoxia on tolerance to high-intensity exercise. *Eur J Appl Physiol* 112: 327-335, 2012.
- 13. **DeLorey DS, Kowalchuk JM, and Paterson DH**. Effects of prior heavy-intensity exercise on pulmonary O₂ uptake and muscle deoxygenation kinetics in young and older adult humans. *J Appl Physiol* 97: 998-1005, 2004.
- 14. **DeLorey DS, Kowalchuk JM, and Paterson DH**. Relationship between pulmonary O₂ uptake kinetics and muscle deoxygenation during moderate intensity exercise. *J Appl Physiol* 95: 113-120, 2003.
- 15. **Donnelly J, and Green S**. Effect of hypoxia on the dynamic response of hyperaemia in the contracting human calf muscle. *Exp Physiol* 98: 81-93, 2013.
- 16. **Federspiel WJ, and Popel AS**. A theoretical analysis of the effect of the particulate nature of blood and oxygen release in capillaries. *Microvasc Res* 32: 164-189, 1986.
- 17. **Ferguson CS, Rossiter HB, Whipp BJ, Cathcart AJ, Murgatroyd SR, and Ward SA**. Effect of recovery duration from prior exhaustive exercise on the parameters of the power-duration relationship. *J Appl Physiol* 108: 866-874, 2010.
- 18. **Ferguson CS, Whipp BJ, Cathcart AJ, Rossiter HB, Turner AP, and Ward SA**. Effects of prior very-heavy intensity exercise on indices of aerobic function and high-intensity exercise tolerance. *J Appl Physiol* 103: 812-822, 2007.
- 19. **Ferrari M, Binzoni T, and Quaresima V**. Oxidative metabolism in muscle. *Philosophical Transactions of the Royal Society Biological Sciences* 352: 677-683, 1997.
- 20. **Ferreira LF, Lutjemeier BJ, Townsend DK, and Barstow TJ**. Effects of pedal frequency on estimated muscle microvascular O₂ extraction. *J Appl Physiol* 96: 558-563, 2006.
- 21. **Fukuba Y, Miura A, Endo M, Kan A, Yanagawa K, and Whipp BJ**. The curvature constant parameter of the power-duration curve for varied-power exercise. *Med Sci Sports Exerc* 35: 1413-1418, 2003.
- 22. **Grassi B, Pogliaghi S, Rampichini S, Quaresima V, Ferrari M, Marconi C, and Cerretelli P.** Muscle oxygenation and pulmonary gas exchange kinetics during cycling exercise ontransitions in humans. *J Appl Physiol* 95: 149-158, 2003.
- 23. **Gratton E, Fantini S, Franceschini MA, Gratton G, and Fabiani M**. Measurements of scattering and absorption changes in muscle and brain. *Philosophical Transactions of the Royal Society Biological Sciences* 352: 727-735, 1997.

- 24. **Groebe K, and Thews G**. Calculated intra- and extracellular PO₂ gradients in heavily working red muscle. *Am J Physiol Heart Circ Physiol* 28: 1990.
- 25. Hamaoka T, Iwane H, Shimomitsu T, Katsumura T, Murase N, Nishio S, Osada T, Kurosawa Y, and Chance B. Noninvasive measures of oxidative metabolism on working human muscles by near-infrared spectroscopy. *J Appl Physiol* 81: 1410-1417, 1996.
- 26. **Hill DW**. The critical power concept: A review. *Sports Med* 16: 237-254, 1993.
- 27. Jones AM, Vanhatalo A, Burnley M, Morton RH, and Poole DC. Critical power: Implications for determination of VO_{2max} and exercise tolerance. *Med Sci Sports Exerc* 42: 1876-1890, 2010.
- 28. Jones AM, Wilkerson DP, DiMenna FJ, Fulford J, and Poole DC. Muscle metabolic responses to exercise above and below the "critical power" assessed using ³¹P-MRS. American Journal of Physiology Regulatory, Integrative and Comparative Physiology 294: 585-593, 2008.
- 29. **Kindig CA, Richardson TE, and Poole DC**. Skeletal muscle capillary hemodynamics from rest to contractions: implications for oxygen transfer. *J Appl Physiol* 92: 2513-2520, 2002.
- 30. **Krustrup P, Ferguson RC, Kjaer M, and Bangsbo J**. ATP and heat production in human skeletal muscle during dynamic exercise: higher efficiency of anaerobic than aerobic ATP resynthesis. *J Physiol* 549: 255-269, 2003.
- 31. **Lanza IR, Wigmore DM, Befroy DE, and Kent-Braun JA**. In vivo ATP production during free-flow and ischaemic muscle contractions in humans. *J Physiol* 577: 353-367, 2006.
- 32. **Linnarsson D, Karlsson J, Fagraeus L, and Saltin B**. Muscle metabolites and oxygen deficit with exercise in hypoxia and hyperoxia. *J Appl Physiol* 36: 399-402, 1974.
- 33. **Malvin GM, and Wood SC**. Effects of capillary red cell density on gas conductance of frog skin. *J Appl Physiol* 73: 224-233, 1992.
- 34. **Mancini DM, Bolinger L, Li H, Kendrick K, Chance B, and Wilson JR**. Validation of near-infrared spectroscopy in humans. *J Appl Physiol* 77: 2740-2747, 1994.
- 35. **Miura A, Kino F, Kajitani S, Sato H, Sato H, and Fukuba Y**. The effect of oral creatine supplementation on the curvature constant parameter of the power-duration curve for cycle ergometry in humans. *Japanese Journal of Physiology* 49: 169-174, 1999.

- 36. **Miura A, Sato H, Sato H, Whipp BJ, and Fukuba Y**. The effect of glycogen depletion on the curvature constant parameter of the power-duration curve for cycle ergometry. *Ergonomics* 43: 133-141, 2000.
- 37. **Molé PA, Chung Y, Tran TK, Sailasuta N, Hurn R, and Jue T**. Myoglobin desaturation with exercise intensity in human gastronemius muscle. *Am J Physiol Regul Integr Comp Physiol* 277: R173 R180, 1999.
- 38. **Monod H, and Scherrer J**. The work capacity of a synergic muscular group. *Ergonomics* 8: 329-338, 1965.
- 39. **Moritani T, Nagata A, DeVries HA, and Muro M**. Critical power as a measure of physical work capacity and anaerobic threshold. *Ergonomics* 24: 339-350, 1981.
- 40. **Murgatroyd SR, Ferguson CS, Ward SA, Whipp BJ, and Rossiter HB**. Pulmonary O₂ uptake kinetics as a determinant of high-intensity exercise tolerance in humans. *J Appl Physiol* 110: 1598-1606, 2011.
- 41. **Osawa T, Kime R, Hamaoka T, Katsumura T, and Yamamoto M**. Attenuation of muscle deoxygenation precedes EMG threshold in normoxia and hypoxia. *Med Sci Sports Exerc* 43: 1406-1413, 2011.
- 42. **Poole DC**. Resolving the determinants of high-intensity exercise performance. *Exp Physiol* 94: 197-198, 2008.
- 43. **Poole DC, Ward SA, Gardner GW, and Whipp BJ**. Metabolic and respiratory profile of the upper limit for prolonged exercise in man. *Ergonomics* 31: 1265-1279, 1988.
- 44. **Richardson RS**. What governs skeletal muscle $\dot{V}O_{2max}$? New evidence. *Med Sci Sports Exerc* 32: 100-107, 2000.
- 45. **Richardson RS, Newcomer SC, and Noyszewski EA**. Skeletal muscle intracellular PO2 assessed by myoglobin desaturation: response to graded exercise. *J Appl Physiol* 91: 2679-2685, 2001.
- 46. **Richardson RS, Noyszewski EA, Kendrick KF, Leigh JS, and Wagner PD**. Myoglobin O₂ desaturation during exercise. *J Clin Invest* 96: 1916-1926, 1995.
- 47. Roca J, Agusti AGN, Alonso A, Poole DC, Viegas C, Barbera JA, Rodriguez-Roisin R, Ferrer A, and Wagner PD. Effects of training on muscle O₂ transport at VO_{2max}. *J Appl Physiol* 73: 1067-1076, 1992.

- 48. **Roca J, Hogan MC, Story D, Bebout DE, Haab P, Gonzalez R, Ueno O, and Wagner PD**. Evidence for tissue diffusion limitation of $\dot{V}O_{2max}$ in normal humans. *J Appl Physiol* 67: 291-299, 1989.
- 49. **Skiba PF, Chidnok W, Vanhatalo A, and Jones AM**. Modeling the expenditure and reconstitution of work capacity above critical power. *Medicine and science in sports* 44: 1526-1532, 2012.
- 50. **Stringer W, Wasserman K, Casaburi R, Porszasz J, Maehara K, and French W**. Lactic acidosis as a facilitator of oxyhemoglobin dissociation during exercise. *J Appl Physiol* 76: 1462-1467, 1994.
- 51. **Vanhatalo A, Fulford J, DiMenna FJ, and Jones AM**. Influence of hyperoxia on muscle metabolic responses and the power-duration work relationship during severe-intensity exercise in humans: a ³¹P magnetic resonance spectroscopy study. *Exp Physiol* 95: 528-540, 2010.
- 52. **Vanhatalo A, Jones AM, and Burnley M**. Application of critical power in sport. *International Journal of Sports Physiology and Performance* 6: 128-136, 2011.
- 53. **Wagner PD**. Central and peripheral aspects of oxygen transport and adaptations with exercise. *Sports Med* 11: 133-142, 1991.
- 54. **Wagner PD**. Determinants of maximal oxygen transport and utilization. *Annual Review of Physiology* 58: 21-50, 1996.
- 55. **Whipp BJ, Huntsman DJ, Stoner N, Lamarra N, and Wasserman K**. A constant which determines the duration of tolerance to high intensity work. *Federation Proceedings* 41: 1591, 1982.

Chapter 4 - Influence of blood flow occlusion on the development of peripheral and central fatigue during small muscle mass handgrip exercise

Summary

The influence of the muscle metabolic milieu on peripheral and central fatigue is currently unclear. Moreover, the relationship between peripheral and central fatigue and the curvature constant (W') have not been investigated. Six men (age: 25 ± 4 years, body mass: 82 ± 10 kg, height: 179 ± 4 cm) completed four constant power handgrip tests to exhaustion under conditions of control exercise (Con), blood flow occlusion exercise (Occ), Con with 5 min post-exercise blood flow occlusion (Con + Occ), and Occ with 5 min post-exercise blood flow occlusion (Occ + Occ). Neuromuscular fatigue measurements and W' were obtained for each subject. Each trial resulted in significant peripheral and central fatigue. Significantly greater peripheral (-79.7 \pm 5.1 % vs. -22.7 \pm 6.0 %) and central (-42.6 \pm 3.9 % vs. -4.9 ± 2.0 %) fatigue occurred for Occ than for Con. In addition, significantly greater peripheral (-83.0 \pm 4.2 % vs. -69.0 \pm 6.2 %) and central (-65.5 \pm 14.6 % vs. -18.6 \pm 4.1 %) fatigue occurred for Occ + Occ than for Con + Occ. W' was significantly related to the magnitude of peripheral (r = 0.85) and central (r = 0.60) fatigue. The current findings demonstrate that blood flow occlusion exacerbated the development of both peripheral and central fatigue and that post-exercise blood flow occlusion prevented the recovery of both peripheral and central fatigue. Moreover, the current findings suggest that W' may be determined by the magnitude of fatigue permitted to develop during exercise.

Introduction

The tolerance of exercise within the severe-intensity domain is well described as the hyperbolic power-duration relationship. The asymptote of this relationship is critical power (CP) and represents the highest attainable steady-state for aerobic energy production without continually drawing upon the second parameter of this relationship, the curvature constant (W') (14, 21, 44, 45, 47, 58). The precise deterministic mechanisms of W' have remained elusive, yet it appears that W' represents a finite anaerobic capacity that when completely utilized, results in similar amounts of work performed above CP and similar end-exercise intramuscular perturbations (i.e., phosphocreatine [PCr], inorganic phosphate [Pi], and hydrogen ion [H⁺]) (35, 44, 45, 48, 58). The consistency of these variables suggests that the mechanisms determining W' must be constant within a given exercise condition. Moreover, the hyperbolic nature of the power-duration relationship implies that exercise tolerance for any power output within the severe-intensity domain is determined by the same mechanisms. Furthermore, the robust hyperbolic nature of the power-duration relationship across exercise modalities (13-15, 17, 33, 35, 45, 48, 58) and species (19, 40), where the determinants of exercise tolerance likely differ (i.e., central cardiovascular limitations, convective O₂ transport limitations, diffusive O₂ transport limitations, etc.), suggests a mechanism of exercise tolerance regulation that is common to severe-intensity exercise. Importantly, Burnley et al. (16) demonstrated that knee-extension critical torque (the isometric exercise equivalent to CP) represents a "critical threshold" for neuromuscular fatigue development, suggesting that exercise tolerance within the severe-intensity domain may be determined by the magnitude of fatigue development or degree of system impairment tolerated.

Accumulating evidence suggests that "high-intensity" (more precisely, severe-intensity (16)) exercise tolerance is limited by the attainment of a specific level of peripheral muscle fatigue (5, 6, 24, 27, 49, 51, 53). Amann et al. (10) elegantly merged the concepts of a "critical threshold" of muscle fatigue, a "sensory tolerance limit" of group III/IV muscle afferent feedback, and central motor drive

into a paradigm describing the mechanism determining exercise tolerance. This paradigm highlights the important function of the feedback from group III/IV muscle afferent fibers to the central nervous system regarding the physiological state of the working skeletal muscles (1, 36, 37) in determining the "critical threshold" of fatigue (8, 28) and the point where central motor drive becomes limited or limiting (57). Recently, Pethick et al. (46) demonstrated that beyond a decrease in torque-generating capacity, fatigue also limits the ability of the neuromuscular system to adapt to external perturbation. It has been postulated that these mechanisms serve to limit the magnitude of fatigue developed during exercise, presumably as a component of homeostasis (7, 10, 51).

The findings of Burnley et al. (16) in combination with the fatigue paradigm of Amann et al. (10), suggest that CP may represent the exercise intensity above which exercise tolerance is limited by the attainment of the "sensory tolerance limit". Therefore, the mechanisms determining W' may be related to the magnitude of fatigue developed during severe-intensity exercise. A constant "sensory tolerance limit" would constrain the amount of work that could be performed and the degree of intramuscular metabolic perturbation prior to fatigue within the severe-intensity domain. This constraint may explain the consistency in the amount of work performed and the intramuscular metabolic perturbations associated with the complete utilization of W' (35, 44, 45, 48, 58). Previous research suggests that peripheral fatigue (and therefore the "sensory tolerance limit") is constant for whole-body exercise (i.e., cycling) across normoxic, hypoxic, and hyperoxic conditions (6, 49). However, the results are equivocal for smaller muscle mass exercise, as the sensory tolerance limit has been demonstrated to be similar (42) and different (18, 52) with varying O_2 delivery conditions. Thus, the "sensory tolerance" limit" may be regulated differently between large and small muscle mass exercise and the magnitude of fatigue permitted to develop by magnitude of the "sensory tolerance limit" during severe-intensity exercise may be a determining mechanism of W'.

To date, we are unaware of a study that has assessed the magnitude of fatigue development and the "sensory tolerance limit" using small muscle mass (handgrip) exercise with reductions in O₂ delivery (via blood flow occlusion) during and post-exercise in order to determine the influence of the muscle metabolic milieu on peripheral and central fatigue. Moreover, we are aware of no study that has examined the relationship between the magnitude of fatigue developed during exercise and W'.

Therefore, the current study utilized handgrip exercise with periods of blood flow occlusion during and post-exercise to determine the influence of O₂ delivery on the development of peripheral and central fatigue. Furthermore, the current study assessed the relationship between the magnitude of fatigue development and the magnitude of W'. We tested the hypotheses that 1) peripheral and central fatigue development would be significantly exacerbated during exercise with blood flow occlusion compared to control exercise, 2) there would be no significant recovery of peripheral and central fatigue during post-exercise blood flow occlusion, and 3) a greater magnitude of peripheral fatigue developed during exercise would be associated with a greater magnitude of W'.

Methods

Ethical approval

All experimental procedures were approved by the Institutional Review Board of Kansas State

University and conformed to the standards set by the *Declaration of Helsinki*. Written informed consent
was attained after subjects were informed of the overall protocol and the potential risks of participation.

Subjects were free of overt cardiovascular or metabolic disease, determined via medical health history
evaluation.

Experimental design

After thorough familiarization with the handgrip contraction and fatigue assessment protocol, subjects completed a total of five testing sessions. Testing sessions were separated by at least 24 h and the subjects were instructed to abstain from vigorous activity during the 24 h prior to testing. Additionally, subjects were instructed to abstain from caffeine and alcohol consumption during the 2 and 12 h, respectively, prior to testing. All testing was conducted using a custom-built two-handed handgrip ergometer (14), which was calibrated prior to the study. The ergometer was attached to a pneumatic cylinder by means of a cable-pulley system, which provided a fixed linear displacement of 4 cm per handgrip contraction. Resistance was controlled via pressurization of the pneumatic cylinder and work was accomplished by compressing the air within the pneumatic cylinder. Power output was calculated as $P = Rdf \cdot k^{-1}$, where P is power in W, R is resistance in kg, d is displacement in meters (m), f is contraction frequency, and k is the constant 6.12 for the conversion of kg·m·min⁻¹ to W. Alterations in power output were accomplished via alterations in resistance (air pressure), as d and f were held constant. Subjects were seated in front of the ergometer and grasped the handrail such that both forearms were approximately at heart level. Exercise was performed using a 50% contraction duty cycle (1.5 s contraction: 1.5 s relaxation) at a rate of 20 contractions min⁻¹. An audio recording with the

specific timing was used in conjunction with feedback provided by an investigator to ensure correct timing. All testing sessions were continued until the limit of tolerance (T_{lim}), determined as the inability to successfully complete three consecutive contraction cycles.

Subjects completed an incremental power output test $(1.0 \text{ W} + 0.5 \text{ W} \cdot \text{min}^{-1})$ to determine peak power (Ppeak) during the first testing session. Ppeak was determined as the greatest power output for which at least half of the stage was completed. Subjects subsequently completed four constant-power testing sessions at 85 %P_{peak}. The protocols were randomly ordered conditions of control exercise (Con), blood flow occlusion exercise (Occ), control exercise with 5 minutes post-exercise blood flow occlusion (Con + Occ), and blood flow occlusion exercise with 5 minutes post-exercise blood flow occlusion (Occ + Occ). Brachial artery blood flow was occluded with a vascular cuff positioned around the brachial region of each arm, which was rapidly inflated (< 0.3 s) to suprasystolic pressures (≥ 275 mmHg) at the onset of exercise and remained inflated until the appropriate time within the specific protocol (E20 Rapid Cuff Inflator, Hokanson, Bellevue, WA, USA). Blood flow occlusion was verified by the absence of a radial pulse and the cuff pressures were continuously monitored to ensure ≥ 275 mmHg. Neuromuscular function was assessed prior to and following each protocol (Figure 4.1). In a recent study (13), the parameters of the power-duration relationship were determined for each of the current subjects for control and blood flow occlusion conditions (Con CP, Con W', Occ CP, Occ W'), affording the opportunity to examine the relationships between the parameters of neuromuscular fatigue and the power-duration relationship. Importantly, P_{peak} (5.8 \pm 0.9 W vs. 6.1 \pm 1.1 W, p = 0.1) and T_{lim} at similar power outputs (5.2 \pm 0.9 W vs. 5.3 \pm 0.9 W, p = 0.2) were not statistically different (459 \pm 154 s vs. 470 ± 140 s, p = 0.8), suggesting that the physiological determinants of exercise tolerance had not changed for the subjects. In addition, all subjects reported no changes in whole-body and handgrip muscle training status.

Near-infrared spectroscopy

Oxygenation characteristics were measured during the pre-exercise and exercise portions of each protocol using a frequency-domain multi-distance NIRS system (Oxiplex TS, ISS, Champaign, IL, USA). Detailed descriptions of the principles and algorithms of the NIRS technology have previously been described (26, 31). Briefly, this NIRS device consists of one detector fiber bundle and eight lightemitting diodes (LED) operating as wavelengths of 690 and 830 nm (four LEDs per wavelength). The LED-detector fiber bundle separation distances are 2.0, 2.5, 3.0, and 3.5 cm. This NIRS device measures and incorporates the dynamic reduced scattering coefficients to provide absolute concentrations (µM) for deoxygenated-[hemoglobin + myoglobin] (deoxy-[Hb + Mb]), oxygenated-[Hb + Mb] (oxy-[Hb + Mb]), total-[Hb + Mb], and %Saturation-[Hb + Mb] (%Sat-[Hb + Mb]). The deoxy-[Hb + Mb] is relatively insensitive to changes in blood-volume (20, 25, 30) and has been used to reliably estimate the fractional oxygen extraction (20, 22, 23, 25, 26, 30, 41). The NIRS device was calibrated prior to each test according to the manufacturer's recommendations. The flexor digitorum superficialis of the left arm was identified using palpation and EMG. The NIRS probe was secured along the belly of the muscle with a Velcro strap and an elastic bandage. The position of the probe was marked with indelible ink to assess movement of the probe during the testing session and for reproducible placement of the probe throughout the study. NIRS data were collected at 50 Hz and analyzed using 9 s timebinned mean values.

Electromyography

Surface EMG measurements were obtained during the pre-exercise and exercise portions of each protocol using a commercially available system (Trigno EMG, Delsys Inc., Boston, MA, USA). The

EMG sensor consists of four silver electrodes (5 x 1mm) arranged in a 2 x 2 orientation used to make single differential EMG measurements. The flexor digitorum superficialis of the right arm was identified by palpation and strong EMG activity when the fingers were flexed, but not with ulnar or radial deviation. The sensor was secured along the belly of the muscle using adhesive surgical tape and the position marked with indelible ink. The EMG data were collected at a sampling rate of 1000 Hz and band-pass filtered (0.05 – 400 Hz) using fifth-order Butterworth filter. The EMG signal corresponding to each muscle contraction was detected using code 'developed in house' (MATLAB R2011a, The Mathworks, Natick, MA, USA). The amplitude characteristics were analyzed via integrated electromyography (iEMG) to provide an index of muscle activation and motorneuron firing rate. The frequency characteristics were analyzed via median power frequency (MedPF) to provide an index of the muscle action potential conduction velocity. The EMG data were analyzed using 9 s time-binned mean values.

Neuromuscular function

Neuromuscular function testing was conducted with the subjects standing at the dynamometer, such that the shoulders were in-line with the dynamometer and the right arm was resting on a platform at shoulder level with the elbow fully extended. The handgrip dynamometer was attached to a calibrated force transducer (LBG1, BLH Electronics, Waltham, MA, USA) that was fixed to the platform to prevent movement. Force was sampled at 1000 Hz and displayed on a computer screen (LabVIEW, National Instruments, Austin, TX, USA). Adhesive stimulation electrodes (4 x 6 cm) were attached to the antebrachial region of the right arm for electrical stimulation of the flexor digitorum superficialis. The anode was positioned proximal to the olecranon process on the posterior brachial region of the arm and the cathode was positioned over the median nerve on the anterior antebrachial region of the arm.

During the familiarization session, the cathode location that provided the greatest force development with electrical stimulation was determined. The positions of the electrodes were marked with indelible ink for reproducible placement throughout the study. The flexor digitorum superficialis was stimulated using a high-voltage constant-current electrical stimulator (DS7AH, Digitimer, Welwyn Garden City, UK). Paired stimuli (doublets) were delivered at 400 V with 100 µs square-wave pulse durations and a 10 ms pulse interval. Stimulation intensity was initiated at 50 mA and was increased in 10 mA increments until the measured force and compound muscle action potential (M-wave) no longer increased. The stimulator current was then increased by a further 19 ± 4 % to ensure the stimuli were supramaximal (range 140 – 230 mA). Subjects subsequently performed a series of six, 3 s maximal voluntary contractions (MVCs), separated by 30 s (~2.75 min total duration). Doublet muscle stimulations were delivered 5 s prior to each MVC, 1.5 s into the MVC, and 5 s after each MVC to obtain measurements of unpotentiated, superimposed, and potentiated twitch forces, respectively. Neuromuscular assessment was completed prior to exercise and following the end of the protocol for the testing session (Figure 1). In all cases, neuromuscular assessment was initiated < 45 s after the cessation of the protocol. MVC was measured as the greatest force attained prior to the superimposed muscle doublet stimulation. Superimposed twitch force was measured as the increment in force following the delivery of doublet stimulation during the MVC. Voluntary activation (VA) was calculated using twitch interpolation (11, 12, 56) corrected for when the superimposed doublet stimulation did occur at MVC:

$$\% VA = \left[1 - \left(\frac{\text{force prior to superimposed twitch}}{\text{MVC}}\right) \cdot \left(\frac{\text{superimposed twitch force}}{\text{potentiated twitch force}}\right)\right] \cdot 100.$$

Potentiated twitch force (Q_{tw}) was measured as the greatest force produced with double stimulation 5 s after the MVC. The last four MVCs of each six MVC series were utilized for data analysis, as the degree of potentiation was lessened after the first two MVCs (3, 50).

Statistical analysis

All statistical analyses were performed using a commercially available software package (SigmaStat, Systat Software Inc., Point Richmond, CA, USA). Two-way ANOVAs with repeated measures (trial x time) were used to compare main effects for all of the NIRS variables at baseline and end-exercise. One-way ANOVAs with a repeated measure were used to compare main effects for T_{lim} and then EMG variables at end-exercise. Tukey's post-hoc analyses were conducted when main effects were detected. Student's paired t-tests were used to compare pre- and post-exercise Q_{tw} , MVC, %VA within each exercise test and the %change in Q_{tw} , MVC, %VA for Con vs. Occ and Con + Occ vs. Occ + Occ. Linear regression analyses were used to describe the relationship between W' and MVC, Q_{tw} , and %VA. The α -level was set at 0.05 a priori. All data are presented as mean \pm SD, unless otherwise noted.

Results

Six recreationally active men (age: 25 ± 4 years, body mass: 82 ± 10 kg, height: 179 ± 4 cm) volunteered to participate in the study. The P_{peak} from the incremental power test was 6.1 ± 1.1 W and $85 \% P_{peak}$ was 5.2 ± 0.9 W. The T_{lim} for the trials were: Con: 472 ± 150 s, Con + Occ: 446 ± 165 s, Occ: 131 ± 12 s, Occ + Occ: 134 ± 25 s. T_{lim} was significantly (p < 0.001) shorter for exercise during blood flow occlusion than for exercise during control blood flow. Occ CP (-0.7 \pm 0.5 W) was significantly lower than Con CP (3.9 ± 0.8 W), while Occ W' (810 ± 205 J) was significantly greater than Con W' (550 ± 127 J).

NIRS

For all muscle oxygenation measurements at end-exercise, both Occ and Occ + Occ values were significantly different from both Con and Con + Occ, while there were no significant differences within each exercise condition (Figure 4.2). Baseline muscle oxygenation values were not significantly different between trials. Deoxy-[Hb + Mb] at end-exercise was significantly greater than baseline for all trials. End-exercise total-[Hb + Mb] was significantly greater than baseline for Con and Con + Occ, while there was no significant difference between end-exercise and baseline for Occ and Occ + Occ. Oxy-[Hb + Mb] was significantly lower at end-exercise compared to baseline for Occ and Occ + Occ, but no significance differences were detected for Con or Con + Occ. %Sat-[Hb + Mb] was significantly lower at end-exercise compared to baseline for all trials (Figure 4.2).

EMG

EMG measurements were not significantly different between trials for the first 9 s of exercise.

There were no significant differences detected for EMG measurements at end-exercise within each

exercise condition (control or occlusion). MedPF was significantly lower at end-exercise for occlusion exercise than control exercise, while no significant difference was detected for iEMG (Figure 4.3).

Neuromuscular function and W'

For all exercise trials, post-exercise neuromuscular fatigue measurements were significantly lower than pre-exercise values. The reductions in Q_{tw} , MVC, and %VA were significantly greater for Occ than for Con (Figure 4.4) and for Occ + Occ than for Con + Occ (Figure 4.5). W' was significantly related to the pre- to post-exercise reduction in MVC (r = 0.87, p < 0.001), Q_{tw} (r = 0.85, p < 0.001), and %VA (r = 0.60, p = 0.04) (Figure 4.6).

Discussion

The purpose of the current study was to determine the influence of reductions in O_2 delivery (via blood flow occlusion) on the development of peripheral and central fatigue during handgrip exercise. It was demonstrated that blood flow occlusion during exercise exacerbated the development of both peripheral and central fatigue. Moreover, continued blood flow occlusion after the cessation of exercise prevented the recovery of (or worsened the magnitude of) peripheral and central fatigue. These results suggest that the "sensory tolerance limit" for handgrip exercise is not constant, as different magnitudes of fatigue were measured across O_2 delivery conditions. The current study is the first to identify a significant relationship between the magnitude of fatigue developed during exercise and the magnitude of W'. This relationship suggests that the magnitude of fatigue permitted to develop during severe-intensity exercise may be a determining mechanism of W'.

Influence of O_2 delivery on fatigue

It has been postulated that the voluntary termination of severe-intensity exercise is the result of attaining a "sensory tolerance limit" (28). Accumulating evidence suggests that the ensemble group III/IV afferent input from the active locomotor muscles plays a vital role in determining this "sensory tolerance limit" (2, 4, 6, 8, 9, 37, 51) and the subsequent reduction in central motor drive (57). This reduction in central motor drive is hypothesized to be a protective mechanism that constrains the magnitude of fatigue development within the muscle by limiting intramuscular perturbations (6). The consistency of peripheral fatigue development (and therefore the "sensory tolerance limit") during exercise is not without some degree of ambiguity. It appears that the "sensory tolerance limit" for large muscle mass activity (e.g., cycling) is constant and does not vary with alterations in O_2 delivery (6, 49). In contrast, a constant "sensory tolerance limit" is not consistently found for small muscle mass activity (e.g., knee-extension and handgrip exercise). Christian et al. (18) demonstrated a greater magnitude of

peripheral fatigue development during knee-extension exercise in hypoxia than normoxia. Russ and Kent-Braun (52) demonstrated that ischemic handgrip exercise resulted in greater peripheral fatigue development than control handgrip exercise. However, Millet et al. (42) demonstrated similar levels of peripheral fatigue with knee-extension exercise during normoxic and hypoxic exercise with and without ischemia. The current study demonstrated that reductions in O₂ delivery (vial blood flow occlusion) exacerbated the magnitude of fatigue development through both peripheral and central origins for small muscle mass handgrip exercise. Moreover, the current study demonstrated a greater magnitude of peripheral fatigue was incurred during occlusion exercise, despite lower iEMG values. The iEMG signal has previously been used as a surrogate measure of central motor drive (10). In the paradigm of Amann et al. (10) the "sensory tolerance limit" is attained via increases in afferent feedback from the active muscle and central motor drive. This suggests that occlusion exercise augmented the influence of afferent feedback relative to central motor drive, such that the greater "sensory tolerance limit" for occlusion exercise was attained despite central motor drive being relatively low. Thus, for "small" muscle mass exercise it appears be that the "sensory tolerance limit" may be sensitive to alterations in O₂ delivery or that the magnitude of fatigue permitted to develop during exercise is regulated differently between large and small muscle mass exercise.

It was demonstrated in the current study that recovery of peripheral and central fatigue is prevented (and may be worsened) if blood flow occlusion is maintained post-exercise. It is well documented that the accumulation of intramuscular metabolites during exercise increases the firing frequency of group III/IV afferents (1, 36), which, in turn, have been implicated as determinants of the magnitude of fatigue developed during exercise (2, 37). Recently, Kennedy et al. (37) demonstrated that an ischemic period after a fatiguing knee-extension protocol decreased VA, presumably due to activity of group III/IV muscle afferents. Consistent with this, it was demonstrated in the current study that

post-exercise blood flow occlusion resulted in the persistence (or further development) of not only central fatigue, but also peripheral fatigue.

Moreover, blood flow occluded exercise resulted in significantly greater levels of peripheral and central fatigue. Interestingly, peripheral fatigue appears to be more sensitive to the reduction in O₂ delivery than central fatigue. Central fatigue has been demonstrated to be influenced by cerebral O₂ delivery, as it was demonstrated that central fatigue is exacerbated during exercise with blood flow occlusion when cerebral O₂ delivery is reduced via hypoxia (42, 43). Thus, central fatigue may have been less sensitive to the effects of occlusion exercise, as cerebral oxygenation was likely not challenged during the current study. Cumulatively, these findings support the notion that the concentration of intramuscular metabolites contributes to the magnitude of fatigue developed within the muscle, by affecting the firing frequency of group III/IV muscle afferents.

Despite an apparent difference in fatigue regulation between large and small muscle mass exercise, cycling, knee-extension, and handgrip exercise have all been demonstrated to hold true to the hyperbolic power-duration relationship, even with alterations in O₂ delivery (13-15, 45, 48, 58). This implies that the determinants of exercise tolerance are regulated within each muscle mass and O₂ delivery condition, and that the mechanisms of regulation may vary across muscle mass and O₂ delivery conditions. However, some commonality in exercise tolerance regulation must nonetheless exist, as there appears to be no deviation from the hyperbolic power-duration relationship.

Relationship between fatigue and W'

The findings of the current study demonstrate that the magnitude of fatigue accrued during handgrip exercise is significantly related to the magnitude of W'. W' has repeatedly been associated with a finite amount of work that can be performed above CP for a given exercise condition (44, 45, 48).

The relationship between W' and fatigue suggests that the mechanisms determining when fatigue occurs (via a reduction in central motor drive) constrain the amount of work that can be performed above CP. Thus, the greater amount of fatigue that is allowed to develop, the greater the amount work that can be performed. For example, Broxterman et al. (13) demonstrated a significantly greater W' for exercise with blood flow occlusion than for control exercise. The findings of the current study suggest that the greater W' with blood flow occlusion exercise may likely be due to the greater magnitude of fatigue permitted to develop. It has also been demonstrated that W' is associated with the attainment of consistent intramuscular metabolite concentrations at end-exercise (48, 58). The magnitude of fatigue permitted to develop during exercise would constrain the amount of intramuscular metabolic perturbation. This may explain the consistent levels measured within given exercise conditions. However, it does not appear that intramuscular metabolic perturbations at end-exercise vary with O₂ delivery conditions (32, 58). Thus, it appears that different magnitudes of fatigue and amounts of work can be performed for given intramuscular metabolic perturbations. This may arise from differences in efficiency and energy yield as a result of O₂ delivery to the muscle (38, 39, 55, 58). Consistent with this, W' (determined as the amount of work performed above CP) was decreased in hyperoxia, while no change in the end-exercise intramuscular metabolic perturbations was measured (58). Importantly, the attainment of these consistent end-exercise intramuscular metabolite concentrations may not be a direct determining mechanism of W', as these concentrations may be attained and maintained for several minutes before the limit of exercise tolerance (see Figure 2 in ref. (58)). Moreover, it appears that specific exercise training protocols alter peripheral and central fatigue characteristics (59), which may potentially explain alterations in W' with exercise training (34, 54). However, as CP and W' were not measured in the study by Zgahal et al. (59) it cannot be known if the reported changes in peripheral and central fatigue were related to alterations in the magnitude of W'. The findings of the current study

suggest that the amount of work available or the degree of intramuscular perturbation may not mechanistically determine W'. Rather, the amount of fatigue permitted to develop during exercise may determine W' and therefore exercise tolerance above CP.

Limitations

It has previously been demonstrated that compression block influences afferent activity (29). Thus, the vascular cuffing used in the current study potentially could have altered afferent feedback due to nerve compression. However, compression block is typically performed by occluding blood flow for ~20 min. In the current study, blood flow occlusion was not initiated until the onset of exercise and the total occlusion duration never exceeded 20 min. There were no measurements of intramuscular metabolite concentrations or afferent firing in the current study. Therefore, inferences were made from previous studies demonstrating no effect of inspired O₂ concentration on end-exercise intramuscular metabolite concentrations (32, 58), and that group III/IV afferent firing increases with metabolite accumulation (1, 36). Lastly, no comparisons were made between fatigue measurements from the post-exercise blood flow occlusion data and fatigue measurements obtained immediately post-exercise. This was purposeful in order to prevent attributing the difference in fatigue between these conditions to occlusion, as it is not known what the fatigue measurements would be 5 minutes after the cessation of exercise.

Conclusion

This study provides further evidence that the magnitude of fatigue development is not constant for small muscle mass exercise. Moreover, the current study demonstrated that post-exercise blood flow occlusion prevented the recovery of both peripheral and central fatigue, presumably due to the persisting

(or worsening) intramuscular metabolic milieu. In combination, it appears that fatigue is regulated differently between large and small muscle mass exercise and that the stimulation of group III/IV muscle afferents via intramuscular metabolites contributes to the development of fatigue. The current study is the first to provide evidence of a relationship between the magnitude of fatigue development during exercise and the magnitude of W'. This evidence suggests that W' may be determined by the magnitude of fatigue accrued during exercise, which may constrain the amount of work that can be performed and the intramuscular metabolic perturbations for severe-intensity exercise.

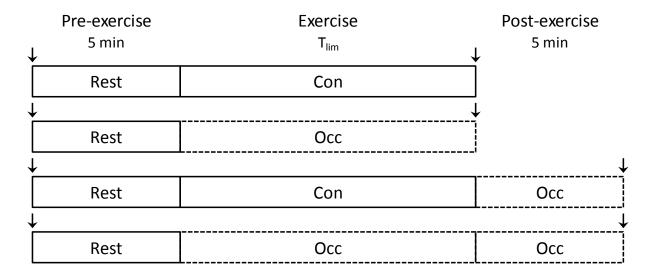


Figure 4.1 Experimental design.

Control (Con) and occluded (Occ) brachial artery blood flow. The arrows signify when neuromuscular function testing was conducted.

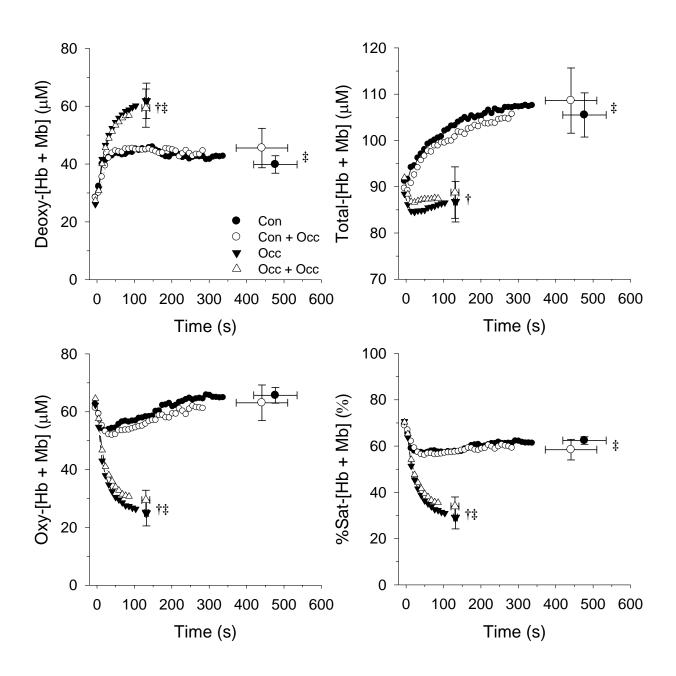


Figure 4.2 Mean NIRS muscle oxygenation data during each exercise trial.

Deoxygenated-[hemoglobin + myoglobin] (deoxy-[Hb + Mb]), total-[Hb + Mb], oxygenated (oxy-[Hb + Mb]), and percent saturation (%Sat-[Hb + Mb]) data during each exercise trial. No significant differences were detected within control or occlusion exercise data. † Occ and Occ + Occ end-exercise data significantly different from Con and Con + Occ at end-exercise. ‡ end-exercise significantly different from baseline within exercise condition.

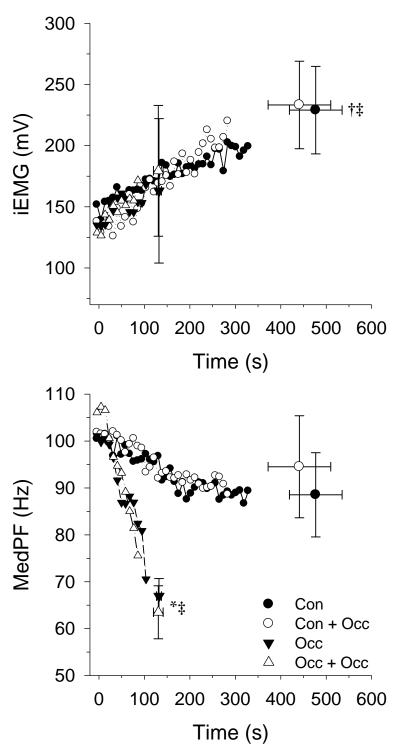


Figure 4.3 Mean EMG data for each exercise trial.

Integrated EMG (iEMG) and median power frequency (MedPF) data for each exercise trial. † Con and Con + Occ significantly different from Occ at end-exercise. ‡ end-exercise significantly different from initial 9 s value within exercise condition. * Occ and Occ + Occ significantly different from Con and Con + Occ at end-exercise.

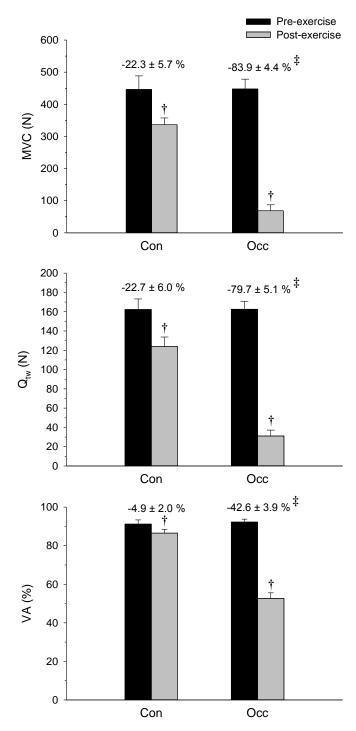


Figure 4.4 Neuromuscular function for Control and Occlusion exercise trials.

Maximal voluntary contraction (MVC), potentiated twitch force (Q_{tw}) , and voluntary activation (%VA) determined pre- and post-exercise for control (Con) and occlusion (Occ) blood flow conditions. The percent change from pre- to post-exercise is presented above the respective exercise trial bar graph. † significantly different from pre-exercise. ‡ significantly different from Con percent change.

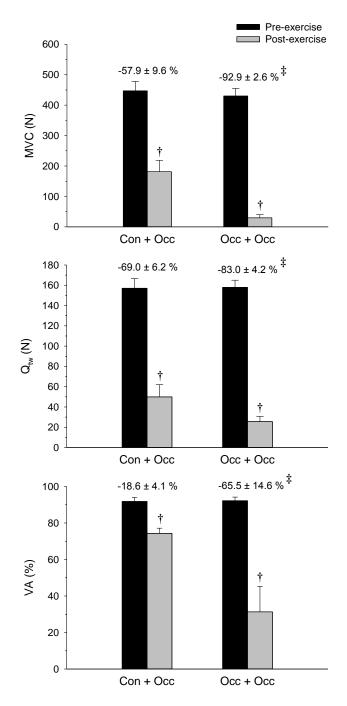


Figure 4.5 Neuromuscular function for Control + Occlusion and Occlusion + Occlusion exercise trials.

Maximal voluntary contraction (MVC), potentiated twitch force (Q_{tw}) , and voluntary activation (%VA) determined pre- and post-exercise during control blood flow exercise with post-exercise blood flow occlusion (Con + Occ) and blood flow occlusion exercise with post-exercise blood flow occlusion (Occ + Occ). The percent change from pre- to post-exercise is presented above the respective exercise trial bar graph. \dagger significantly different from pre-exercise. \ddagger significantly different from Con + Occ percent change.

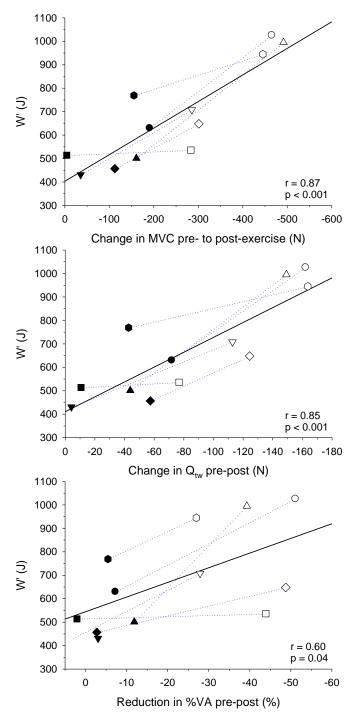


Figure 4.6 Relationship between W' and the change in neuromuscular function variables.

Maximal voluntary contraction (MVC), potentiated twitch force (Q_{tw}), and voluntary activation (%VA) determined from neuromuscular function testing. The solid symbols represent Con data and the open symbols represent Occ data, while distinct symbol shapes represent individual subjects.

References

- 1. **Adreani CM, Hill JM, and Kaufman MP**. Responses of group III and IV muscle afferents to dynamic exercise. *J Appl Physiol* 82: 1811-1817, 1997.
- 2. **Amann M**. Central and Peripheral Fatigue: Interaction during Cycling Exercise in Humans. *Med Sci Sports Exerc* 43: 2039-2045, 2011.
- 3. **Amann M, Blain G, Proctor LT, Sebranek JJ, Pegelow DF, and Dempsey JA**. Implications of group III and IV muscle afferents for high-intensity endurance exercise performance in humans. *J Physiol* 589: 5299-5309, 2011.
- 4. **Amann M, and Calbet JA**. Convective oxygen transport and fatigue. *J Appl Physiol* 104: 861-870, 2008.
- 5. **Amann M, and Dempsey JA**. Locomotor muscle fatigue modifies central motor drive in healthy humans and imposes a limitation to exercise performance. *J Physiol* 586: 161-173, 2008.
- 6. **Amann M, Eldridge MW, Lovering AT, Stickland MK, Pegelow DF, and Dempsey JA**. Arterial oxygenation influences central motor output and exercise performance via effects on peripheral locomotor muscle fatigue in humans. *J Physiol* 575: 937-952, 2006.
- 7. **Amann M, Proctor LT, Sebranek JJ, Eldridge MW, Pegelow DF, and Dempsey JA**. Somatosensory feedback from the limbs exerts inhibitory influence on central neural drive during whole body endurance exercise. *J Appl Physiol* 105: 1717-1724, 2008.
- 8. **Amann M, Proctor LT, Sebranek JJ, Pegelow DF, and Dempsey JA**. Opiod-mediated muscle afferents inhibit central motor drive and limit peripheral muscle fatigue development in humans. *J Physiol* 587: 271-283, 2009.
- 9. **Amann M, Romer LM, Pegelow DF, Jacques AJ, Hess CJ, and Dempsey JA**. Effects of arterial oxygen content on peripheral locomotor muscle fatigue. *J Appl Physiol* 101: 119-127, 2006.
- 10. **Amann M, Venturelli M, Ives SJ, McDaniel J, Layec G, Rossman MJ, and Richardson RS**. Peripheral fatigue limits endurance exercise via a sensory feedback-mediated reduction in spinal motorneuronal output. *J Appl Physiol* 115: 355-364, 2013.
- 11. **Behm DG, St-Pierre DMM, and Perez D**. Muscle inactivation: assessment of interpolated twitch technique. *J Appl Physiol* 81: 2267-2273, 1996.
- 12. **Belanger AY, and McComas AJ**. Extent of motor unit activation during effort. *J Appl Physiol* 51: 1131-1135, 1981.

- 13. **Broxterman RM, Ade CJ, Craig JC, WIlcox SL, Schlup SJ, and Barstow TJ**. Influence of blood flow occlusion on muscle deoxygenation characteristics and the parameters of the power-duration relationship. *J Appl Physiol* 2015.
- 14. **Broxterman RM, Ade CJ, Wilcox SL, Schlup SJ, Craig JC, and Barstow TJ**. Influence of duty cycle on the power-duration relationship: Observations and potential mechanisms. *Respiratory Physiology & Neurobiology* 192: 102-111, 2014.
- 15. **Burnley M**. Estimation of critical torque using intermittent isometric maximal voluntary contractions of the quadriceps in humans. *J Appl Physiol* 106: 975-983, 2009.
- 16. Burnley M, Vanhatalo A, and Jones AM. Distinct profiles of neuromuscular fatigue during muscle contractions below and above the critical torque in humans. *J Appl Physiol* 113: 215-223, 2012.
- 17. **Cheng C-F, Yang Y-S, Lin H-M, Lee C-L, and Wang C-Y**. Determination of critical power in trained rowers using a three-minute all-out rowing test. *Eur J Appl Physiol* 112: 1251-1260, 2012.
- 18. **Christian RJ, Bishop DJ, Billaut F, and Girard O**. Peripheral fatigue is not critically regulated during maximal, intermittent, dynamic leg extensions. *J Appl Physiol* 117: 1063-1073, 2014.
- 19. **Copp SW, Hirai DM, Musch TI, and Poole DC**. Critical speed in the rat: implications for hindlimb muscle blood flow distribution and fibre recruitment. *J Physiol* 588: 5077-5087, 2010.
- 20. **De Blasi RA, Cope M, Elwell C, Safoue F, and Ferrari M**. Noninvasive measurement of human forearm oxygen consumption by near infrared spectroscopy. *European Journal of Applied Physiology and Occupational Physiology* 67: 20-25, 1993.
- 21. **Dekerle J, Mucci P, and Carter H**. Influence of moderate hypoxia on tolerance to high-intensity exercise. *Eur J Appl Physiol* 112: 327-335, 2012.
- 22. **DeLorey DS, Kowalchuk JM, and Paterson DH**. Effects of prior heavy-intensity exercise on pulmonary O₂ uptake and muscle deoxygenation kinetics in young and older adult humans. *J Appl Physiol* 97: 998-1005, 2004.
- 23. **DeLorey DS, Kowalchuk JM, and Paterson DH**. Relationship between pulmonary O₂ uptake kinetics and muscle deoxygenation during moderate intensity exercise. *J Appl Physiol* 95: 113-120, 2003.
- 24. **Duffield R, Green R, Castle P, and Maxwell N**. Precooling can prevent the reduction in self-paced exercise intensity in the heat. *Med Sci Sports Exerc* 42: 577-584, 2010.

- 25. **Ferrari M, Binzoni T, and Quaresima V**. Oxidative metabolism in muscle. *Philosophical Transactions of the Royal Society Biological Sciences* 352: 677-683, 1997.
- 26. **Ferreira LF, Lutjemeier BJ, Townsend DK, and Barstow TJ**. Effects of pedal frequency on estimated muscle microvascular O₂ extraction. *J Appl Physiol* 96: 558-563, 2006.
- 27. **Gagnon P, Saey D, Vivodtzev I, Laviolette L, Mainguy V, Milot J, Provencher S, and Maltais F.** Impact of preinduced quadriceps fatigue on exercise response in chronic obstructive pulmonary disease and healthy subjects. *J Appl Physiol* 107: 832-840, 2009.
- 28. **Gandevia SC**. Spinal and supraspinal factors in human muscle fatigue. *Physiol Rev* 81: 1725-1789, 2001.
- 29. **Garland SJ**. Role of small diameter afferents in reflex inhibition during human muscle fatigue. *J Physiol* 435: 547-558, 1991.
- 30. Grassi B, Pogliaghi S, Rampichini S, Quaresima V, Ferrari M, Marconi C, and Cerretelli P. Muscle oxygenation and pulmonary gas exchange kinetics during cycling exercise ontransitions in humans. *J Appl Physiol* 95: 149-158, 2003.
- 31. **Gratton E, Fantini S, Franceschini MA, Gratton G, and Fabiani M**. Measurements of scattering and absorption changes in muscle and brain. *Philosophical Transactions of the Royal Society Biological Sciences* 352: 727-735, 1997.
- 32. **Hogan MC, Richardson RS, and Haseler LJ**. Human muscle performance and PCr hydrolysis with varied inspired oxygen fractions: a ³¹P-MRS study. *J Appl Physiol* 86: 1367-1373, 1999.
- 33. **Hughson RL, Orok CJ, and Staudt LE**. A high velocity treadmill running test to assess endurance running potential. *Int J Sports Med* 5: 23-25, 1984.
- 34. **Jenkins DG, and Quigley BM**. The y-intercept of the critical power function as a measure of anaerobic work capacity. *Ergonomics* 34: 13-22, 1991.
- 35. **Jones AM, Wilkerson DP, DiMenna FJ, Fulford J, and Poole DC**. Muscle metabolic responses to exercise above and below the "critical power" assessed using ³¹P-MRS. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 294: 585-593, 2008.
- 36. **Kaufman MP, and Rybicki KJ**. Discharge properties of group III and IV muscle afferents: their responses to mechanical and metabolic stimuli. *Circulation Research* 61: I60-I65, 1987.

- 37. **Kennedy DS, Fitzpatrick SC, Gandevia SC, and Taylor JL**. Fatigue-related firing of muscle nociceptors reduces voluntary activation of ipsilateral but not contralateral lower limb muscles. *J Appl Physiol* 118: 408-418, 2015.
- 38. **Krustrup P, Ferguson RC, Kjaer M, and Bangsbo J**. ATP and heat production in human skeletal muscle during dynamic exercise: higher efficiency of anaerobic than aerobic ATP resynthesis. *J Physiol* 549: 255-269, 2003.
- 39. **Lanza IR, Wigmore DM, Befroy DE, and Kent-Braun JA**. In vivo ATP production during free-flow and ischaemic muscle contractions in humans. *J Physiol* 577: 353-367, 2006.
- 40. **Lauderdal MA, and Hinchcliff KW**. Hyperbolic relationship time-to-fatigue and workload. *Equine Veterinary Journal Supplements* 30: 586-590, 1999.
- 41. **Mancini DM, Bolinger L, Li H, Kendrick K, Chance B, and Wilson JR**. Validation of near-infrared spectroscopy in humans. *J Appl Physiol* 77: 2740-2747, 1994.
- 42. **Millet GY, Aubert D, Favier FB, Busso T, and Benoît H**. Effect of acute hypoxia on central fatigue during repeated isometric leg contractions. *Scand J Med Sci Sports* 19: 695-702, 2009.
- 43. **Millet GY, Mauthalib M, Jubeau M, Laursen PB, and Nosaka K**. Severe hypoxia affects exercise performance independently of afferent feedback and peripheral fatigue. *J Appl Physiol* 112: 1335-1344, 2012.
- 44. **Monod H, and Scherrer J**. The work capacity of a synergic muscular group. *Ergonomics* 8: 329-338, 1965.
- 45. **Moritani T, Nagata A, DeVries HA, and Muro M**. Critical power as a measure of physical work capacity and anaerobic threshold. *Ergonomics* 24: 339-350, 1981.
- 46. **Pethick J, Winter SL, and Burnley M**. Fatigue reduces the complexity of knee extensors torque fluctuations during maximal and submaximal intermittent isometric contractions in man. *J Physiol* 2015.
- 47. **Poole DC**. Resolving the determinants of high-intensity exercise performance. *Exp Physiol* 94: 197-198, 2008.
- 48. **Poole DC, Ward SA, Gardner GW, and Whipp BJ**. Metabolic and respiratory profile of the upper limit for prolonged exercise in man. *Ergonomics* 31: 1265-1279, 1988.
- 49. **Romer LM, Haverkamp HC, Amann M, Lovering AT, Pegelow DF, and Dempsey JA**. Effect of acute severe hypoxia on peripheral fatigue and endurance capacity in healthy humans. *Am J Physiol Regul Integr Comp Physiol* 292: 2007.

- 50. **Romer LM, Lovering AT, Haverkamp HC, Pegelow DF, and Dempsey JA**. Effect of inspiratory muscle work on peripheral fatigue of locomotor muscles in healthy humans. *J Physiol* 571: 425-439, 2006.
- 51. **Rossman MJ, Venturelli M, McDaniel J, Amann M, and Richardson RS**. Muscle mass and peripheral fatigue: a potential role for afferent feedback? *Acta Physiologica* 206: 242-250, 2012.
- 52. **Russ DW, and Kent-Braun JA**. Sex differences in human skeletal muscle fatigue are eliminated under ischemic conditions. *J Appl Physiol* 94: 2414-2422, 2003.
- 53. **Saey D, Michaud A, Couillard A, Côté CH, Mador MJ, LeBlanc P, Jobin J, and Maltais F**. Contractile fatigue, muscle morphometry, and blood lactate in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 171: 1109-1115, 2005.
- 54. **Sawyer BJ, Stokes DG, Womack CJ, Morton RH, Weltman A, and Gaesser GA**. Strength training increases endurance time to exhaustion during high-intensity exercise despite no change in critical power. *J Strength Cond Res* 28: 601-609, 2014.
- 55. **Stellingweff T, LeBlanc PJ, Hollidge MG, Heigenhauser GJF, and Spriet LL**. Hyperoxia decreases muscle glycogenolysis, lactate production, and lactate efflux during steady-state exercise. *Am J Physiol Endrocrinol Metab* 290: E1180-E1190, 2006.
- 56. **Strojnik V, and Komi PV**. Neuromuscular fatigue after maximal stretch-shortening cycle exercise. *J Appl Physiol* 84: 344-350, 1998.
- 57. **Taylor JL, Petersen N, Butler JE, and Gandevia SC**. Ischaemia after exercise does not reduce responses of human motoneurones to cortical or coriticospinal tract stimulation. *J Physiol* 525: 793-801, 2000.
- 58. **Vanhatalo A, Fulford J, DiMenna FJ, and Jones AM**. Influence of hyperoxia on muscle metabolic responses and the power-duration work relationship during severe-intensity exercise in humans: a ³¹P magnetic resonance spectroscopy study. *Exp Physiol* 95: 528-540, 2010.
- 59. Zghal F, Cottin F, Kenoun I, Rebaï H, Moalla W, Dogui M, Tabka Z, and Martin V.
 Improved tolerance of peripheral fatigue by the central nervous system after endurance training.
 Eur J Appl Physiol 2015.

Chapter 5 - Conclusions

The purpose of this dissertation was to test the overarching hypothesis that muscle contraction characteristics (i.e., intensity of contraction, muscle contraction-relaxation duty cycle, etc.) alter oxygen delivery and oxygen utilization, which directly influence the power-duration relationship and fatigue development, and therefore, exercise tolerance. In support of this hypothesis, alterations in oxygen delivery and oxygen utilization via muscle contraction characteristics and blood flow occlusion directly influenced CP. These results further support that CP is aerobic in nature. W' was not affected with alterations in muscle contraction-relaxation duty cycle, but was increased during blood flow occlusion exercise compared to control blood flow exercise. The magnitude of fatigue development was also greater during blood flow occlusion exercise compared to control blood flow exercise.

Importantly, a significant relationship was observed between the magnitude of fatigue developed and the magnitude of W'. These results suggest that W' is not influenced directly by manipulations in oxygen delivery and oxygen utilization, per se. Rather, manipulations in oxygen delivery and oxygen utilization influence the magnitude of fatigue accrued during exercise, which dictates the magnitude of W' measured as the amount of work performed above CP, the amount of energy store depletion, or the degree of intramuscular metabolic perturbation. Therefore, CP represents the threshold above which exercise tolerance becomes predictably limited. This exercise tolerance appears to be determined by the magnitude of fatigue permitted to develop, via the attainment of the "sensory tolerance limit" and the consequential limiting of central motor drive. The findings presented highlight important physiological mechanisms that determine exercise tolerance and demonstrate that characteristics of muscle contraction affect exercise tolerance by influencing oxygen delivery and oxygen utilization. Furthermore, these findings demonstrate that enhancing oxygen delivery and oxygen utilization should be a primary focus

of interventions designed to improve exercise tolerance, particularly for improving activity tolerance in diseased patients, such as chronic heart failure.

Appendix A - Curriculum Vitae

CURRICULUM VITAE Ryan Michael Broxterman, M.S.

Date of Birth: September 8, 1986

Place of Birth: Topeka, KS, USA

Citizenship: USA

Married; 1 son

Address: Department of Kinesiology

1A Natatorium

Kansas State University Manhattan, KS 66506-0302

Phone: 785-221-1165 (Cell)

Email: rbrox@ksu.edu

Education:

2011 M.S. in Kinesiology, Kansas State University, Manhattan, KS, 2011

Thesis: "A Single Test for the Determination of the Velocity: Time-to-

Exhaustion Relationship"

Advisor: Thomas J. Barstow, Ph.D.

Committee: David C. Poole, Ph.D.; Craig A. Harms, Ph.D.

2009 B.A. (Summa cum laude) in Physical Education emphasis Exercise

Physiology, Washburn University, Topeka, KS

Academic Appointments:

2013 – Present Adjunct Instructor, Department of Kinesiology, Kansas State University,

Manhattan, KS

2011 – Present Graduate Research Assistant, Department of Kinesiology, Kansas State

University, Manhattan, KS

2012 – 2013 Adjunct Instructor, Department of Kinesiology, Washburn University,

Topeka, KS

2010 – 2011	Adjunct Instructor, Department of Biological Science, Manhattan Area Technical College, Manhattan, KS
2010 - 2011	Instructor of Multicultural Academic Program Success (MAPS), Department of Kinesiology, Kansas State University, Manhattan, KS
2009 – 2011	Graduate Teaching Assistant, Department of Kinesiology, Kansas State University, Manhattan, KS

Teaching Experience:

_	
2015 – Present	Kansas State University, Department of Kinesiology, KIN 609 "Environmental Physiology." Adjunct Instructor.
2013 – Present	Kansas State University, Department of Kinesiology, KIN 815 "Research Methods." Adjunct Instructor.
2013 – Present	Kansas State University, Department of Kinesiology, KIN 603 "Advanced Cardiovascular Physiology." Adjunct Instructor.
2012 – 2013	Washburn University, KS, Department of Kinesiology, KN 320 "Motor Learning." Adjunct Instructor.
2011 – 2012	Kansas State University, KS, Department of Kinesiology, "Biomechanics." Developed laboratory manual.
2011 – 2012	Kansas State University, KS, Department of Kinesiology, "Biobehavioral Bases of Physical Activity." Online course development.
2010 – 2012	Kansas State University, KS, Department of Kinesiology. "Exercise Physiology." Laboratory.
2009 – 2012	Kansas State University, KS, Department of Kinesiology. "Biomechanics." Laboratory.
2009 – 2012	Kansas State University, KS, Department of Kinesiology. "Adaptive Physical Activity." Activity Class.
2010 – 2011	Manhattan Area Technical College, KS, Department of Biological Science. "Anatomy and Physiology." Lecture and Laboratory.
2009 – 2011	Kansas State University, KS, Department of Kinesiology. "Biobehavioral Bases of Physical Activity." Laboratory.

2009 – 2010 Kansas State University, KS, Department of Kinesiology. "Public Health." Laboratory.

Invited Lecturer:

- 1. "Anatomy and Biomechanics of the Shoulder Girdle." Department of Kinesiology Biomechanics Lecture, Kansas State University, 2011.
- 2. "Anatomy and Biomechanics of the Hip and Knee Joints." Department of Kinesiology Biomechanics Lecture, Kansas State University, 2011.
- 3. "Health Benefits of Exercise and Fitness." Department of Kinesiology Biobehavioral Bases of Physical Activity Lecture, Kansas State University, 2012.
- 4. "Exercise and the Environment." Department of Kinesiology, Biobehavioral Bases of Physical Activity Lecture, Kansas State University, 2012.

Honors and Awards:

2015	GRA of the Year, Chapter of Golden Key International Honour Society, Kansas State University
2015	Graduate Award for Outstanding Academics, Alumni Association, Kansas State University
2015	Doctoral Scholar Award, American Kinesiology Association
2014	Finalist, GRA of the Year, Chapter of Golden Key International Honour Society, Kansas State University
2014	Distinguished Doctoral Student, Department of Kinesiology, \$300, Kansas State University
2014	College of Veterinary Medicine Dr. Albert L. Burroughs Memorial Award, \$1,000, Kansas State University
2014	Graduate Student Travel Award, \$500, Kansas State University
2013	College of Veterinary Medicine Frank Blecha Award, \$800, Kansas State University
2013	Graduate Student Travel Award, \$450, Kansas State University

2012	Outstanding Graduate Student, Department of Kinesiology, Kansas State University
2012	College of Veterinary Medicine Frank Blecha Award, \$1000, Kansas State University
2012	Graduate Student Travel Award, \$75, Kansas State University
2011	Graduate Student Travel Award, \$75, Kansas State University
2010	Graduate Student Travel Award, \$75, Kansas State University
2009	B.A. Major of the Year, Department of Health, Physical Education, & Exercise Science, \$1000, Washburn University
2009	Transformational Experience Travel Award, \$1000, Washburn University
2008	Helen Hocker Scholarship for outstanding Physical Education student, \$1000, Washburn University

Laboratory Experience:

2009 – Present Human Exercise Physiology Laboratory. Lab Directors: Thomas J. Barstow, Ph.D. & Craig A. Harms, Ph.D.

Research Grant Awards, Awarded:

2014 - 2015 American College of Sports Medicine. "Physiological responses during simulated partial-gravity ambulation." Total costs: \$5,000.

Research Grant Awards, Submitted:

American College of Sports Medicine. "Impact of muscle blood flow manipulations on neuromuscular fatigue" Total costs: \$5,000.

Professional Memberships:

2009 – present	American College of Sports Medicine
2010 – present	American Physiological Society
2013 – present	American College of Sports Medicine Central States

Journals Reviewed: Journal of Applied Physiology; European Journal of Applied

Physiology; Medicine and Science in Sports and Exercise; European Journal of Sports Science; Online Journal of Sports Medicine; American Journal of Physiology – Regulatory, Integrative, and Comparative

Physiology;

Committees (Departmental): Department Head Search Committee, Dr. Craig A. Harms, 2013.

Invited Presentations:

1. "Determination of appropriate physiological measurements for determining EVA taskfailure." Department of Animal Sciences and Industry, Kansas State University, 2014.

Scientific Meeting Presentations:

- 1. <u>Broxterman, R.M.</u>, C.J. Ade, G.L. Gadbury, D. Schinstock, S. Warren, and T.J. Barstow. Gender differences in laboratory assessment and simulated EVA performance. NASA HRP, Galveston, TX, 2013.
- 2. <u>Broxterman, R.M.</u>, C.J. Ade, S.L. Wilcox, J.C. Craig, and T.J. Barstow. Determination of appropriate physiological measurements for predicting EVA task-failure. NASA HRP, Galveston, TX, 2014.

Publications:

Research Papers, Peer Reviewed

- 1. Ade, C.J., <u>R.M. Broxterman</u>, B.J. Wong, and T.J. Barstow. Antegrade and retrograde blood velocity profiles in the intact cardiovascular system. *Experimental Physiology* 97.7: 849-860, 2012.
- 2. <u>Broxterman, R.M.</u>, C.J. Ade, C.A. Harms, D.C. Poole, and T.J. Barstow. A single test for the determination of the speed: time-to-exhaustion relationship. *Respiratory Physiology and Neurobiology* 185: 380-385, 2012.
- 3. Gude, D., R.M. Broxterman, C.J. Ade, T.J. Barstow, T. Nelson, W. Song, and S. Warren. Automated hand forearm ergometer data collection system. International Conference of the IEEE Engineering in Medicine and Biology Society, San Diego, CA, 2012.

- 4. Song, W., C.J. Ade, <u>R.M. Broxterman</u>, T.J. Barstow, T. Nelson, and S. Warren. Activity recognition in planetary navigation field tests using classification algorithms applied to accelerometer data. International Conference of the IEEE Engineering in Medicine and Biology Society, San Diego, CA, 2012.
- 5. Ade, C.J., <u>R.M. Broxterman</u>, and T.J. Barstow. Effects of body posture and exercise training on cardiorespiratory responses to exercise. *Respiratory Physiology and Neurobiology* 188: 39-48, 2013.
- 6. <u>Broxterman, R.M.</u>, C.J. Ade, S.L. Wilcox, S.J. Schlup, J.C. Craig, and T.J. Barstow. Influence of duty cycle on the power-duration relationship for handgrip exercise: Observations and potential mechanisms. *Respiratory Physiology and Neurobiology* 192: 102-111, 2014.
- 7. Day, D., X. Dong, W. Kuhn, D. Gruenbacher, B. Natarajan, T. Sobering, M. Taj-Eldin, S. Warren, T.J. Barstow, <u>R.M., Broxterman</u>, and A. Stonstreet. Biomedical sensing and wireless technologies for long duration EVAs and precursor scout missions. International Conference of the IEEE Engineering in Medicine and Biology Society, Big Sky, MT, 2014.
- 8. Kuehl, P., C. Jia, D. Gude, <u>R.M. Broxterman</u>, T.J. Barstow, and S. Warren. Real-time processing of electromyograms in an anutomated hand-forearm ergometer data collection and analysis system. International Conference of the IEEE Engineering in Medicine and Biology Society, Chicago, IL, 5759-5759, 2014.
- 9. Smith, J.R., C.J. Ade, <u>R.M. Broxterman</u>, B.C. Skutnik, T.J. Barstow, B.J. Wong, and C.A. Harms. Influence of respiratory muscle fatigue on brachial artery blood flow during cycling exercise. *European Journal of Applied Physiology*. 114(8): 1767-1777, 2014.
- 10. Ade, C.J., <u>R.M. Broxterman</u>, J.C. Craig, S.J. Schlup, S.L. Wilcox, and T.J. Barstow. Relationship between simulated extravehicular activity tasks and measurements of physical performance. *Respiratory Physiology and Neurobiology*. 203: 19-27, 2014.
- 11. <u>Broxterman, R.M.</u>, C.J. Ade, J.C. Craig, S.L. Wilcox, S.J. Schlup, and T.J. Barstow. The relationship between critical speed and the respiratory compensation point: coincidence or equivalence. *European Journal of Sport Science*. DOI: 10.1080/17461391.2014.966764, 2014.
- 12. Ade, C.J., R.M. Broxterman, and T.J. Barstow. $\dot{V}_{O_{2max}}$ and microgravity exposure: convective versus diffusive O_2 transport. *Medicine and Science in Sports and Exercise*. DOI: 10.1249/MSS.0000000000000557, 2014.
- 13. <u>Broxterman, R.M.</u>, C.J. Ade, T. Barker, and T.J. Barstow. Influence of pedal cadence on the respiratory compensation point and its relation to critical power. *Respiratory Physiology and Neurobiology*. DOI: 10.1016/j.resp.2014.12.008, 2014.

- 14. <u>Broxterman, R.M.</u>, C.J. Ade, J.C. Craig, S.L. Wilcox, S.J. Schlup, and T.J. Barstow. Influence of blood flow occlusion on muscle oxygenation characteristics and the parameters of the power-duration relationship. *Journal of Applied Physiology*. DOI: 10.1152/japplphysiol.00875.2014.
- 15. Ade, C.J., <u>R.M. Broxterman</u>, J.C. Craig, S.J. Schlup, S.L. Wilcox, and T.J. Barstow. Upper body aerobic exercise as a possible predictor of lower body performance. *Aerospace Medicine and Human Performance*. Accepted.
- 16. Ade, C.J., <u>R.M. Broxterman</u>, J.C. Craig, S.J. Schlup, S.L. Wilcox, S. Warren, P. Kuhl, D. Gude, C. Jia, and T.J. Barstow. Predicting performance on simulated Lunar and Martian based exploration tasks: A feasibility study. *PLOSONE*. In review.
- 17. <u>Broxterman, R.M.</u>, S.L. Wilcox, J.C. Craig, C. Jia, S. Warren, and T.J. Barstow. Influence of blood flow occlusion on the development of peripheral and central fatigue during small muscle mass handgrip exercise. J Physiol. In revision.

Published Letters

- 1. Ade, C.J., <u>R.M. Broxterman</u>, and T.J. Barstow. Critical velocity and maximal lactate steady state: better determinants of 2-hour marathon. *Journal of Applied Physiology* 110(1): 287-288, 2011.
- 2. Craig, J.C., <u>R.M. Broxterman</u>, and T.J. Barstow. Considerations for identifying the boundaries of sustainable performance. *Medicine and Science in Sports and Exercise*. Accepted.

Abstracts

- 1. <u>Broxterman, R.M.</u>, P.G. Wagner, and P.A. Bender. Comparison of RPE to blood lactate levels in cyclists based on mileage per year. American College of Sports Medicine, Seattle, WA, 2009.
- Broxterman, R.M., T. Barker, and T.J. Barstow. Respiratory compensation point oxygen uptake relationship at different pedaling frequencies. American College of Sports Medicine, Baltimore, MD, 2010.
- 3. <u>Broxterman, R.M.</u>, C.J. Ade, and T.J. Barstow. A single test for the determination of critical velocity. American College of Sports Medicine, Denver, CO, 2011.

- 4. Bopp, C.M., B.J. Wong, C.J. Ade, <u>R.M. Broxterman</u>, S.L. Wilcox, and T.J. Barstow. Ibuprofen alters hyperemic responses within skeletal muscle, but not cutaneous, microvasculature during post-occlusive reactive hyperemia. American College of Sports Medicine, Denver, CO, 2011.
- 5. Ade, C.J, <u>R.M. Broxterman</u>, B.J. Wong, T.J. Barstow. Brachial and femoral artery blood velocity profiles are quasi-parabolic during physiologic stress. Experimental Biology, Washington D.C., 2011.
- 6. Ade, C.J., <u>R.M. Broxterman</u>, S. Warren, R.D. Taylor, T.J. Barstow. Development of standardized exercise tests for predicting planetary task performance. The International Academy of Astronautics Humans in Space Symposium, Houston, TX, 2011.
- 7. Ade, C.J., R.M. Broxterman, G. L. Gadbury, D. Schinstock, S. Warren, and T.J. Barstow. Standardized exercise test to evaluate planetary mission readiness. NASA Human Research Program Workshop, Houston, TX, 2012.
- 8. <u>Broxterman, R.M.</u>, C.J. Ade, G.L. Gadbury, D. Schinstock, S. Warren, and T.J. Barstow. 10-km Walkback Performance Predicted From Standardized Exercise Tests. NASA Human Research Program Workshop, Houston, TX, 2012.
- 9. Ade, C.J., R.M. Broxterman, G. L. Gadbury, D. Schinstock, S. Warren, and T.J. Barstow. Physiological responses during simulated planetary field test. American College of Sports Medicine, San Francisco, CA, 2012.
- 10. <u>Broxterman, R.M.</u>, C.J. Ade, G.L. Gadbury, D. Schinstock, S. Warren, and T.J. Barstow. Predictors of 10 km performance. American College of Sports Medicine, San Francisco, CA, 2012.
- 11. Ade, C.J., <u>R.M. Broxterman</u>, S.J. Schlup, S.L. Wilcox, and T.J. Barstow. Influence of duty cycle on muscle deoxy-[Hb+Mb] during ramp hand grip exercise. American Physiological Society Intersociety Meeting: Integrative Biology of Exercise, Westminster, CO, 2012.
- 12. <u>Broxterman, R.M.</u>, C.J. Ade, S.L. Wilcox, S.J. Schulp, and T.J. Barstow. Influence of altered duty cycle on critical power during hand grip exercise. American Physiological Society Intersociety Meeting: Integrative Biology of Exercise, Westminster, CO, 2012.
- 13. <u>Broxterman, R.M.</u>, C.J. Ade, S.L. Wilcox, S.J. Schlup, and T.J. Barstow. Gender differences in laboratory assessment and simulated EVA performance. NASA Human Research Program Workshop, Galveston, TX, 2013.
- 14. Song, W., C.J. Ade, <u>R.M. Broxterman</u>, T. Nelson, D. Gude, T.J. Barstow, and S. Warren. Classification algorithms applied to accelerometer data as a means to identify subject activities related to planetary navigation tasks. NASA Human Research Program Workshop, Galveston, TX, 2013.

- 15. Gude, D., <u>R.M. Broxterman</u>, C.J. Ade, T.J. Barstow, T. Nelson, W. Song, and S. Warren. Automated hand-forearm ergometer data collection system. NASA Human Research Program Workshop, Galveston, TX, 2013.
- 16. Ade, C.J., <u>R.M. Broxterman</u>, S.L. Wilcox, and T.J. Barstow. Determinants of maximal O₂ consumption: modeling the effects of 365 days aboard ISS. NASA Human Research Program Workshop, Galveston, TX, 2013.
- 17. <u>Broxterman, R.M.</u>, C.J. Ade, S.L. Wilcox, S.J. Schlup, and T.J. Barstow. Influence of oxygen delivery on the parameters of the power-duration relationship. American College of Sports Medicine, Indianapolis, IN, 2013.
- 18. Schlup, S.J., C.J. Ade, <u>R.M. Broxterman</u>, S.L. Wilcox, J.C. Craig, and T.J. Barstow. Kinetics of leg and capillary blood flow response to knee extension exercise. American College of Sports Medicine, Indianapolis, IN, 2013.
- 19. Wilcox, S.L., <u>R.M. Broxterman</u>, C.J. Ade, S.J. Schlup, J.C. Craig, Y. Mendoza, L. Chavez, and T.J. Barstow. The relationship between physiologic parameters in upper versus lower body exercise. American College of Sports Medicine, Indianapolis, IN, 2013.
- 20. Craig, J.C., C.J. Ade, <u>R.M. Broxterman</u>, S.L. Wilcox, S.J. Schlup, and T.J. Barstow. The relationship between critical speed and the respiratory compensation point. American College of Sports Medicine, Indianapolis, IN, 2013.
- 21. Ade, C.J., R.M. Broxterman, S.J. Schlup, S.L. Wilcox, J.C. Craig, J. Bernard, and T.J. Barstow. Effects of retrograde shear on the kinetics of adjustment of blood flow and vascular conductance to hand grip exercise. American College of Sports Medicine, Indianapolis, IN, 2013.
- 22. Smith J.R., C.J. Ade, <u>R.M. Broxterman</u>, B.C. Skutnik, and C.A. Harms. The influence of respiratory muscle fatigue on inactive arm blood flow during cycling exercise. American College of Sports Medicine, Indianapolis, IN, 2013.
- 23. <u>Broxterman, R.M.</u>, C.J. Ade, J.C. Craig, S.L. Wilcox, and T.J. Barstow. Muscle oxygenation characteristics within the contraction-relaxation cycle for handgrip exercise. American College of Sports Medicine, Orlando, FL, 2014.
- 24. Wilcox, S.L., <u>R.M. Broxterman</u>, and T.J. Barstow. Predicting "near linear" \dot{V}_{O_2} responses via integration with variable parameters. American College of Sports Medicine, Orlando, FL, 2014.
- 25. Craig, J.C., <u>R.M. Broxterman</u>, and T.J. Barstow. Influence of adipose tissue thickness (ATT) on NIRS-derived total [Hb+Mb] at four sites. American College of Sports Medicine, Orlando, FL, 2014.

- 26. <u>Broxterman, R.M.</u>, C.J. Ade, S.L. Wilcox, J.C. Craig, and T.J. Barstow. Lunar and Mars simulated extravehicular activity (EVA) evoked physiological responses. Experimental Biology, San Diego, CA, 2014.
- 27. Kuehl, P., C. Jia, D. Gude, <u>R.M. Broxterman</u>, T.J. Barstow, and S. Warren. Real-time processing of electromyograms in an automated hand-forearm ergometer data collection and analysis system. NASA Human Research Program Workshop, Galveston, TX, 2014.
- 28. Jia, C. P. Kuehl, D. Gude, <u>R.M. Broxterman</u>, T.J. Barstow, and S. Warren. Improved algorithms for EMG burst identification and processing. NASA Human Research Program Workshop, Galveston, TX, 2014.
- 29. <u>Broxterman, R.M.</u>, C.J. Ade, S.L. Wilcox, J.C. Craig, and T.J. Barstow. Determination of appropriate physiological measurements for predicting EVA task-failure. NASA Human Research Program Workshop, Galveston, TX, 2014.
- 30. Craig, J.C., <u>R.M. Broxterman</u>, and T.J. Barstow. Effect of beetroot juice supplementation on conduit artery and microvascular hemodynamics during small muscle mass handgrip exercise. Experimental Biology, Boston, MA, 2015.
- 31. Craig, J.C., <u>R.M. Broxterman</u>, and T.J. Barstow. Beetroot supplementation and small muscle mass handgrip exercise: Effect on central and peripheral fatigue. American College of Sports Medicine, San Diego, CA, 2015.
- 32. Smith, J.R., <u>R.M Broxterman</u>, C.J. Ade, T.J. Barstow, and C.A. Harms. The effect of Nacetylcysteine on peripheral hemodynamics and fatigue during exercise. Experimental Biology, Boston, MA, 2015.
- 33. <u>Broxterman, R.M.</u>, S.L. Wilcox, J.C., Craig, C. Jia, S. Warren, and T.J. Barstow. Influence of ischemia on peripheral and central fatigue during handgrip exercise. Experimental Biology, Boston, MA, 2015.
- 34. <u>Broxterman, R.M.</u>, C.J. Ade, J.C. Craig, S.L. Wilcox, P.F. Skiba, and T.J. Barstow. Modeling the utilization and reconstitution of W' within the contraction-relaxation cycle for handgrip exercise. American College of Sports Medicine, San Diego, CA, 2015.
- 35. Ade, C.J., <u>R.M. Broxterman</u>, and T.J. Barstow. Standardized "pre-flight" exercise tests to predict performance during extravehicular activities in a lunar environment. NASA Human Research Program Workshop, Galveston, TX, 2015.
- 36. <u>Broxterman, R.M.</u>, C.J. Ade, W.J. Wagner, S.L. Wilcox, J.C. Craig, S. Warren, D. Schinstock, and T.J. Barstow. Development of an offload hoist system for the simulation of microgravity during activity. NASA Human Research Program Workshop, Galveston, TX, 2015.

37. Noel, J.A., G.D. McCoy, K.J. Phelps, <u>R.M. Broxterman</u>, T.J. Barstow, and J.M. Gonzalez. Effect of ractopamine-HCl on muscle fiber types and finishing barrow fatigue. American Meat Science Association Reciprocal Meat Conference, Lincoln, Nebraska, 2015.